A Wash-Free Homogeneous Colorimetric Immunoassay Method

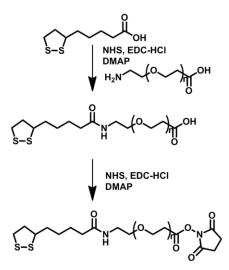
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Scheme S1. Synthesis of the PEG linker whose one side is thioctic acid and the other side is terminated in *N*-hydroxysucinimide (NHS)-activated carboxylic acid. (DMAP is the catalyst)

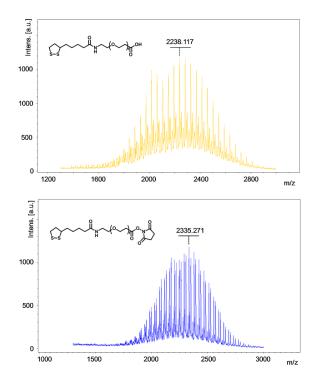


Figure S1. MALDI-TOF-MS data for the intermediate product and the final product indicated in Scheme S1.

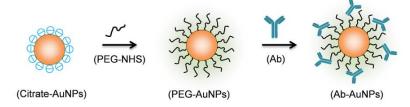


Figure S2. The preparation protocol of antibody (Ab)-functionalized AuNPs starting from citrate-stabilized AuNPs.

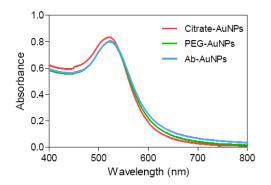


Figure S3. UV-vis absorption spectra of citrate-AuNPs, PEG-AuNPs, and Ab-AuNPs. The graduate red-shift of the absorption bands indicates the success of the step-by-step functionalization shown in Figure S2.

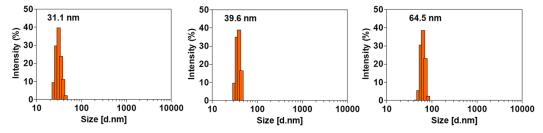


Figure S4. DLS measurements for citrate-AuNPs, PEG-AuNPs, and Ab-AuNPs.

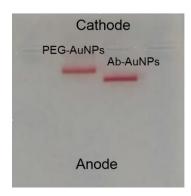


Figure S5. Gel electrophoresis measurement for PEG-AuNPs and Ab-AuNPs.

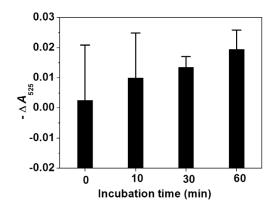


Figure S6. Histogram of $-\Delta A_{525}$ versus different incubation times of Ab-AuNPs and 100 ng/ml of IgG. $-\Delta A_{525} = A - A_0$, where *A* is the absorbance of Ab-AuNP incubate with IgG for different time at 525 nm after adding Au growth solutions, and A_0 is that adding Au growth solution to Ab-AuNP and IgG mixture before incubation. The error bars represent the standard deviations of three independent measurements.

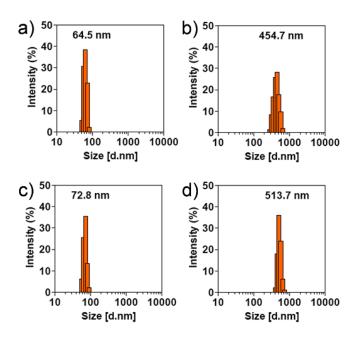


Figure S7. DLS measurements for the AuNPs samples. a) Ab-AuNPs; b) Ab-AuNPs incubated with target protein (human IgG, 100 ng/mL); c) Ab-AuNPs added with Au growth solutions (NH₂OH (20 mM) and HAuCl₄ (120 μ M)); d) The target protein-treated Ab-AuNPs added with the Au growth solutions. The concentration of Ab-AuNPs is 0.5 nM.

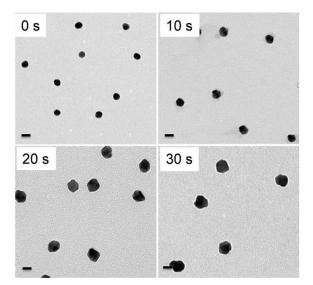


Figure S8. TEM images of the Ab-AuNP intermediates prepared by stopping the nanoparticle growth with mercaptopropionic acid (2 mM) after 0, 10, 20, and 30 s of the reaction.

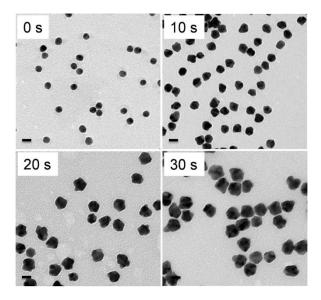


Figure S9. TEM images of the Ab-AuNP intermediates in the presence of detection target and the AuNP solutions were prepared by stopping the nanoparticle growth with mercaptopropionic acid (2 mM) after 0, 10, 20, and 30 s of the reaction.

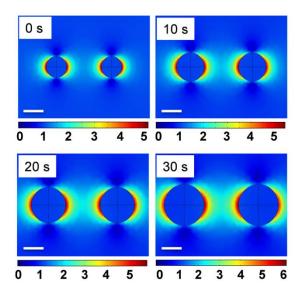


Figure S10. Near-field distributions of the normalized electric field $(|\mathbf{E}|/|\mathbf{E}_{inc}|)$ of AuNPs corresponding to various radii of the AuNPs after 0, 10, 20, and 30 s in Figure S6. The excitation wavelength of the laser is 633 nm. The scale bars are 15 nm.

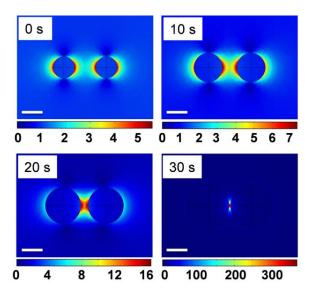


Figure S11. Near-field distributions of the normalized electric field $(|\mathbf{E}|/|\mathbf{E}_{inc}|)$ of AuNPs corresponding to the radii of the AuNPs after 0, 10, 20, and 30 s in Figure S7. The excitation wavelength of the laser is 633 nm. Stronger field enhancement is achieved since the width of the gap becomes smaller. The scale bars are 15 nm.

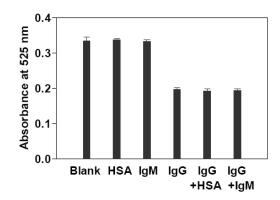


Figure S12. Specificity of the AuNP-based colorimetric immunoassay for IgG. The concentrations of each protein are prepared to be 100 ng/mL. The error bars represent the standard deviations of three independent measurements.

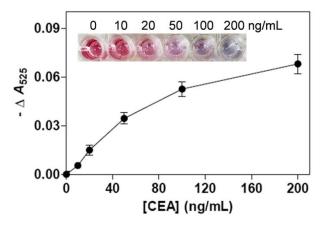


Figure S13. Plots of $-\Delta A_{525}$ versus various concentrations of cancer biomarker CEA as well as corresponding color change in the AuNPs samples. Error bars show the standard deviations of three independent measurements.

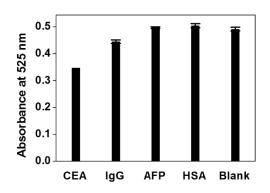


Figure S14. Specificity of the AuNP-based colorimetric immunoassay for CEA. The concentrations of each protein are prepared to be 100 ng/mL. The error bars represent the standard deviations of three independent measurements.

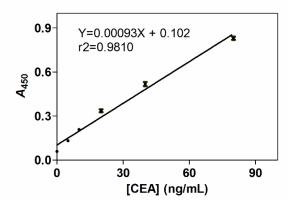


Figure S15. HRP-based ELISA for the detection of various concentrations of CEA spiked in fetal bovine sera. The OD values at 450 nm were recorded by a conventional microplate reader. Error bars show the standard deviations of three independent measurements.

 Table S1. Comparison of normalized electric-field magnitude and intensity for different sized mono-dispersed AuNPs.

Particle size	$\max \mathbf{E} / \mathbf{E}_{inc} $	$\max \mathbf{E} ^2 / \mathbf{E}_{inc} ^2$
15 nm	5.11	26.11
20 nm	5.4	29.16
25 nm	5.76	33.18
29 nm	6.15	37.82

Table S2. Comparison of normalized electric-field magnitude and intensity in the nano-gap for AuNPs oligomers composed of different sized AuNPs.

Particle size	$\max \mathbf{E} / \mathbf{E}_{inc} $	$\max \mathbf{E} ^2 / \mathbf{E}_{inc} ^2$
15 nm	5.55	30.80
20 nm	7.34	53.88
25 nm	16.3	265.69
29 nm	336	1.13×10^{5}

target	nanomaterial	linear range	detection limit	real sample	ref
RIgG	AuNRs	0-20 ng/ml	1.5 ng/ml	N/A	S1
IgG	Au-patterning nanostructures	N/A	66.7 pM	N/A	S2
	nanostructures				
CEA	AuNPs	0-120 ng/ml	3 ng/ml	serum	S3
IgG	Au	0.1-400 ng/ml	0.04 ng/mL	serum	S4
	Nanoclusters				
PSA	AuNPs	1.11×10^{3} -	111aM	N/A	S5
		$1.11 \times 10^{7} \text{ aM}$			
IgG	Au nanoplates	N/A	10 pg/mL	N/A	S 6
CRP	AuNPs	N/A	50 ng/mL	serum	S 7
PSA and p24	AuNPs	N/A	10 ⁻¹⁸ g/mL	serum	11
AFP	AuNPs	0.1-5.0 ng/mL	0.01 ng/mL	serum	S8
AFP	gold-coated	N/A	0.01 ng/mL	N/A	S9
	magnetic				
	nanoparticle				
P53	AuNPs	N/A	5 pg/mL	N/A	S10
IgG	AuNPs	0 -100 ng/mL	3.74 ng/mL	N/A	This study
CEA	AuNPs	0 -100 ng/mL	5.66 ng/mL	serum	This study

Table S3. Comparison of AuNP-based immunoassays for various detection targets.

Table S4. The $-\Delta A_{525}$ values using the proposed AuNP-based colorimetric immunoassay for the clinical samples collected from 6 healthy individuals and 18 patients suffering from cancer.

Healthy		Cancer					
Patient ID	$-\Delta A_{525}$	Patient ID	$-\Delta A_{530}$	Patient ID	$-\Delta A_{525}$	Patient ID	$-\Delta A_{530}$
401543	0.002 ± 0.001	401546	0.033 ± 0.0045	378416	0.018± 0.005	379668	0.023 ± 0.002
400578	0.001 ± 0.003	401542	0.026 ± 0.0048	396435	0.015 ± 0.002	391794	0.022 ± 0.0055
401588	0.001 ± 0.006	402971	0.051 ± 0.006	390360	0.002 ± 0.0008	398770	0.028 ± 0.004
341426	-0.008 ± 0.002	254757	0.014 ± 0.0047	401020	-0.02 ± 0.0001	401090	0.011 ± 0.001
401596	-0.03 ± 0.002	389433	0.029 ± 0.0055	386163	-0.015 ± 0.00045	303556	0.029 ± 0.002
401155	0.003 ± 0.0008	210843	0.050 ± 0.002	401259	0.004 ± 0.0003	399281	0.030 ± 0.002

Sample No.	Patient ID	Age	Sex
A1	401543	45	Male
A2	400578	60	Male
A3	401588	50	Male
A4	341426	80	Male
A5	401596	66	Male
A6	401155	23	Female

Table S5. Clinical information of the 6 healthy individuals.

Table S6. Clinical information of the 18 patients suffering from various types of cancer and their diagnostic results by the clinical standard strategies and conventional ELISA.

Sample No.	Patient ID	Age	Sex	Diagnosis	CEA level (ng/mL)
B1	401546	75	Male	Rectal Carcinoma	55.2
B2	401542	59	Male	Lung Carcinoma	48.5
B3	402971	61	Male	Osteocarcinoma	86.8
B4	254757	48	Female	Breast Carcinoma	6.8
B5	389433	46	Male	Colon Carcinoma	35.2
B6	210843	51	Female	Colon Carcinoma	92.5
C1	378416	56	Female	Breast Carcinoma	15.3
C2	396435	56	Female	Ovarian Carcinoma	25.0
C3	390360	65	Female	Breast Carcinoma	Undetectable
C4	401020	58	Male	Lung Carcinoma	Undetectable
C5	386163	57	Male	Pancreatic Head Carcinoma	Undetectable
C6	401259	69	Male	Hepatocirrhosis	Undetectable
D1	379668	61	Male	Colon Carcinoma	28.3
D2	391794	68	Male	Colon Carcinoma	34.5
D3	398770	78	Male	Lung Carcinoma	45.0
D4	401090	50	Male	Anal Carcinoma	8.4
D5	303556	66	Male	Colon Carcinoma	35.8
D6	399281	48	Female	Hysterocarcinoma	32.9

Reference

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