Development and evaluation of a CEACAM6-targeting theranostic nanomedicine for photoacoustic-based diagnosis and chemotherapy of metastatic cancer

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Supplementary Figure S1

(A) Size of tHSA-NPs measured by Dynamic Light Scattering (DLS): Thiolated HSA(tHSA)-NPs (1:25) without DOX: 184.84 ± 2.42 nm, -32.80 ± 4.48 mV.

(B) Encapsulation efficiency corresponding to thiolation ratio (2-IT mole ratio per albumin molecule): 12.5, 61.08 ± 1.07; 25.0, 19.55 ± 0.11; 37.5, 16.91 ± 0.53.

(A) Size of tHSA-NPs measured by Dynamic Light Scattering (DLS); (B) Encapsulation efficiency corresponding to thiolation ratio (2-IT mole ratio per albumin molecule)
Immunohistochemistry images of normal and A549 AR-metastasized lung tissues. (A) Expressions of CD31 (neovascularization factor) and (B) CEACAM6 were higher in metastasized lung compared to normal lung.
Supplementary Figure S3

*In vivo* photoacoustic image of control (no injection) group using a Vevo LAZR Imaging System
(A) Cellular targeting of CEACAM6-targeting tHSA-NPs (C6-tHSA-NPs) to MCF-7 adherent and MCF-7 AR cells under the shaking media condition for 30 min. Cell nucleus, F-actin and nanoparticles were shown as blue, green, and red, respectively. The rightmost column represents merged images of all. (B) Flow cytometry data for quantification of cellular targeting in the same condition.
Cell viability assay with parental MDA-MB-231 adherent and MDA-MB-231 AR cells. Cells were treated at a concentration corresponding to 100 nM DOX for 6 h and further cultured up to 72 h in fresh medium. Bar with dense line and sparse line represent MDA-MB-231 adherent and MDA-MB-231 AR cells, respectively.
DOX uptake images to A549 AR cell nucleus under the shaking condition mimicking *in vivo* blood flow. Diffusion of free DOX was more efficient than DOX delivered by D-tHSA-NPs in this condition. Cells were treated at a concentration corresponding to 10 μM DOX for 1 h.
Serum stability and degradation by glutathione of tHSA-NPs. 50 (vol.%) FBS and glutathione was treated with concentration of 1.5 mg/ml to tHSA-NPs for 1 hour. Gel retardation assay was performed with SDS-PAGE and protein was stained with coomassie blue.
Supplementary Figure S8

(A) Photoacoustic contrast ability and (B) its quantification graph of the ICG-containing nanoparticles (I@NPs and I+D@NPs) dispersed in PBS and 50% FBS solution. The result shows that there are no significant changes of PA contrast ability after 24 hours in serum condition. (C) ICG mass in nanoparticles (I@NPs/PBS, 37 °C) over time.

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(A-B) *In vivo* tumor growth inhibition of D@C6-NPs and other groups. Three days after the intravenous injection of A549 AR cells, the first therapy was conducted. After 3 days, the second therapy was performed. Metastatic nodules in BALB/c nude mouse lungs were photographed and the number of metastatic nodules was quantified. D@C6-NPs group displayed statistical significance (ANOVA, *p*<0.01).
(A-B) Stability of photoacoustic contrast ability of I@NPs for multiple laser irradiation. The result shows that there is no reduction in the photoacoustic signal even after multiple laser exposures.
Supplementary Figure S11

(A) Ex vivo PA images of both I@NPs and I@C6-NPs from normal and metastasized lungs and its quantification data. L7_4 transducer was used to obtain ex vivo photoacoustic images. Lungs were extracted 24 h after the injection of both I@NPs and I@C6-NPs.