

Supporting information

Anti-EGF receptor aptamer-guided co-delivery of anti-cancer siRNAs and quantum dots for theranosis of triple negative breast cancer

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Table S1. The vesicular size, ζ -potential, and Q-dot incorporation rate of lipoplexes prepared at varied mole% of DSPE-mPEG₂₀₀₀

DSPE-mPEG₂₀₀₀/Lipoplexes (mole%)	Vesicle size (nm)	Polydispersity Index (PDI)	ζ-potential (mV)	Q-dot incorporation rate (%)	Q-dot loading capacity (%)
0/100	385 ± 79	0.41 ± 0.36	51.8 ± 1.3	67.9 ± 1.2	11.3 ± 0.2
1/99	339 ± 74	0.32 ± 0.23	22.9 ± 3.1	75.0 ± 6.3	12.5 ± 1.1
4/96	175 ± 5	0.25 ± 0.04	8.0 ± 3.4	94.9 ± 5.8	15.8 ± 0.9
8/92	185 ± 5	0.22 ± 0.03	0.2 ± 0.5	93.7 ± 4.0	15.5 ± 0.7

The values were measured using a Zetasizer Nano-ZS90. Each value indicates the mean ± standard deviation (S.D.) of five measurements.

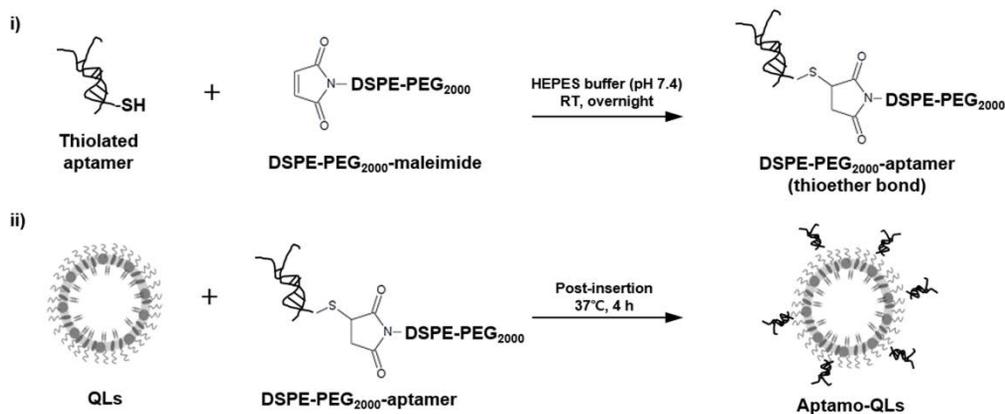
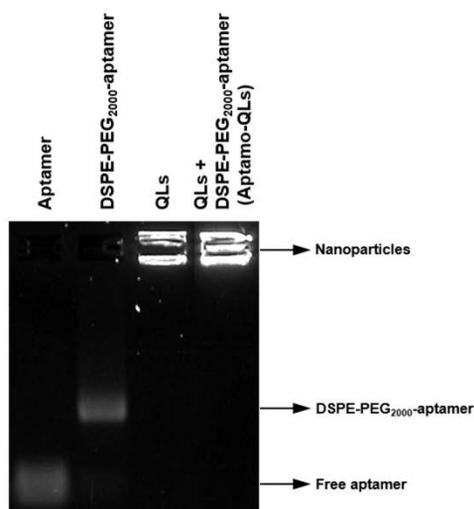
A**Reaction Scheme****B**

Figure S1. Anti-EGFR aptamer conjugation to DSPE-PEG₂₀₀₀-maleimide. (A) Thiolated aptamers were conjugated to DSPE-PEG₂₀₀₀-maleimide and post-inserted into QLs. (B) The post-insertion efficiency was measured by electrophoresis using a 1.5% agarose gel. Lane 1: free aptamer, lane 2: DSPE-PEG₂₀₀₀-aptamer, lane 3: QLs, and lane 4: QLs inserted with DSPE-PEG₂₀₀₀-aptamer.

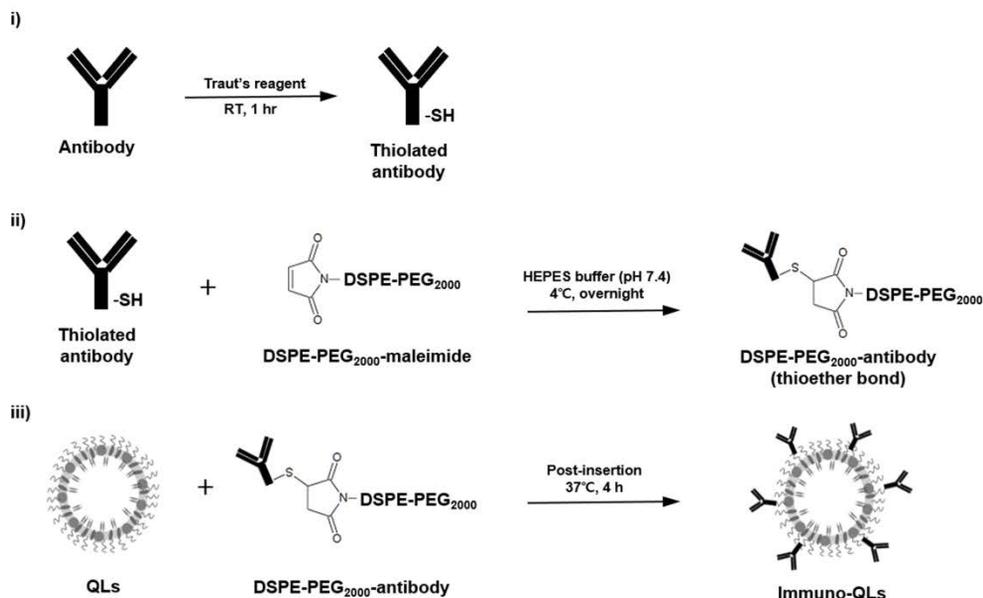
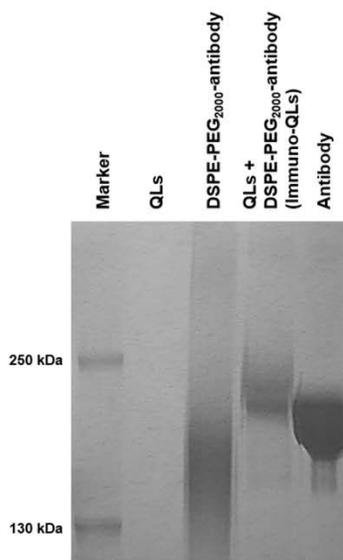
A**Reaction Scheme****B**

Figure S2. Anti-EGFR antibody conjugation to DSPE-PEG₂₀₀₀-maleimide. (A) A procedure of antibody thiolation, antibody conjugation to DSPE-PEG₂₀₀₀-maleimide, and insertion of antibody-lipid conjugate into QLs is illustrated. (B) Antibody conjugation to DSPE-PEG₂₀₀₀-maleimide and insertion of the conjugate into QLs were verified by 6% SDS-PAGE. Lane 1: marker, lane 2: QLs, lane 3: DSPE-PEG₂₀₀₀-antibody, lane 4: DSPE-PEG₂₀₀₀-antibody-inserted QLs, and lane 5: free antibody.

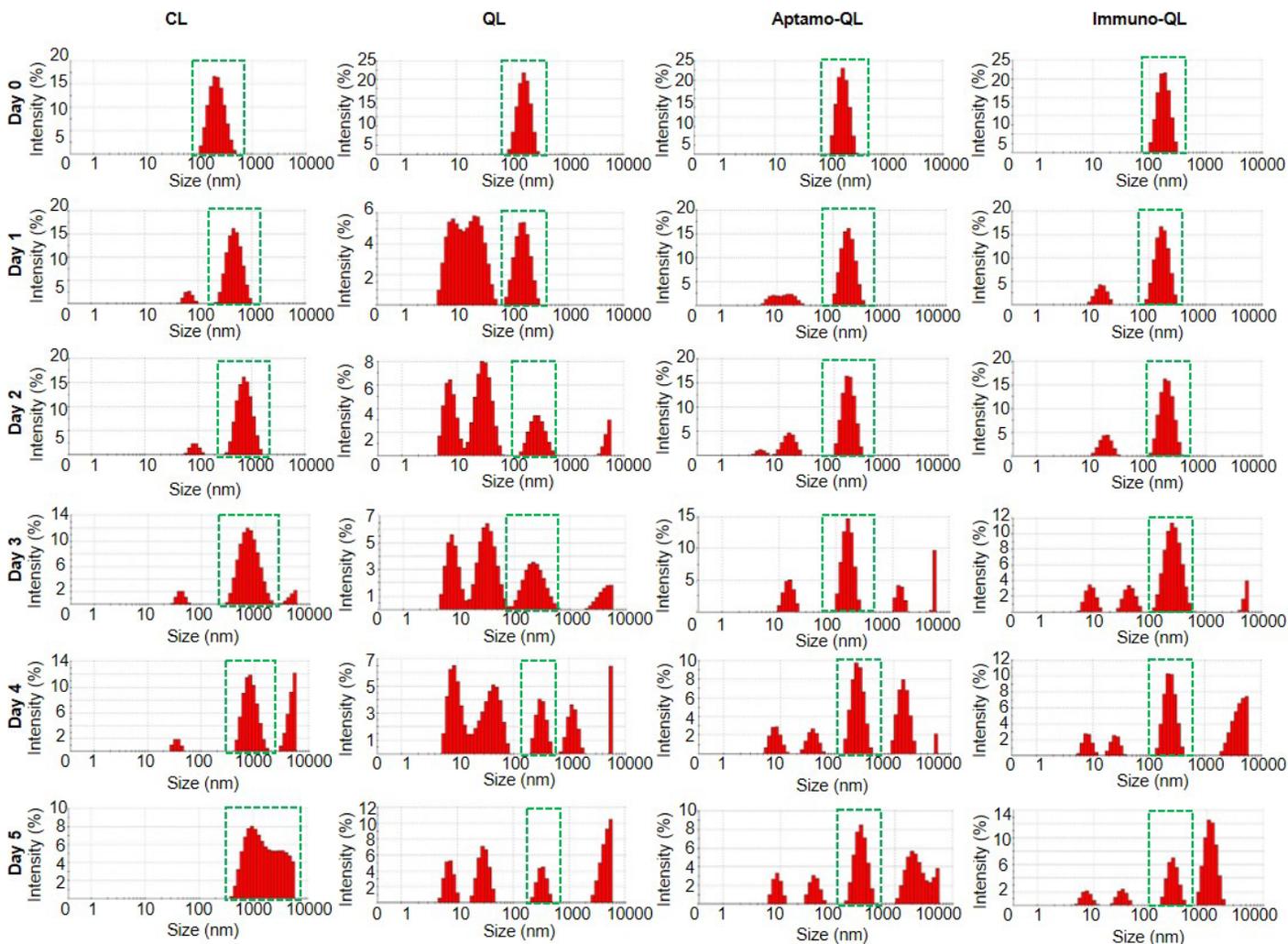


Figure S3. Size changes of aptamo-QLs and immuno-QLs in FBS. The various QL formulations were incubated in the presence of 50% FBS at 37°C and their changes in size were examined with a particle analyzer.

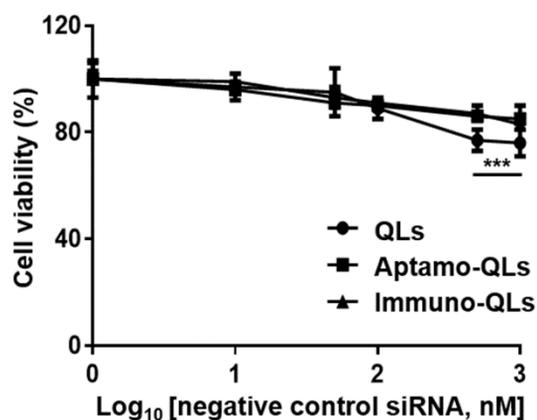


Figure S4. *In vitro* cytotoxicity of aptamo-QLs and immuno-QLs. MDA-MB-231 cells were treated with varied concentrations of prepared negative control siRNA lipoplexes and further cultured for 48 h. Cell viability was measured by the CCK-8 assay. Each error bar represents the mean \pm S.D. of five independent experiments. *** $p < 0.001$ vs. untreated control.

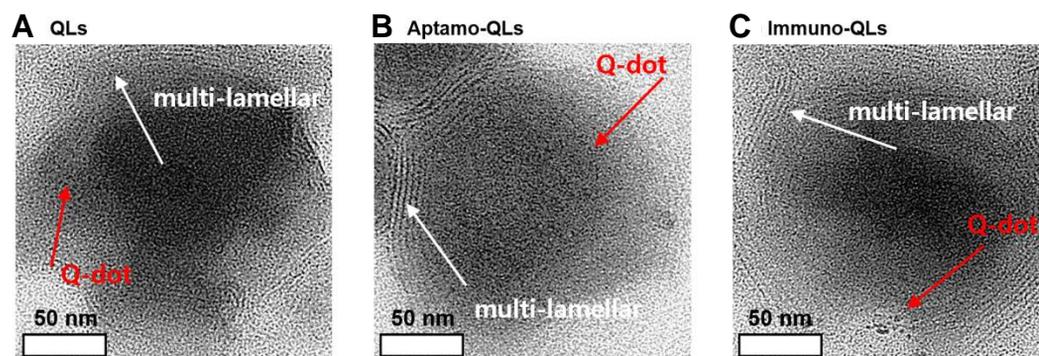


Figure S5. TEM images of aptamo-QLs and immuno-QLs. Representative transmission electron microscopy (TEM) images of QLs (A), aptamo-QLs (B), and immuno-QLs (C) are shown. All lipoplexes showed multi-lamellar structures (white arrow) and Q-dots incorporated in the lipoplex layers (red arrow). Bar = 50 nm.

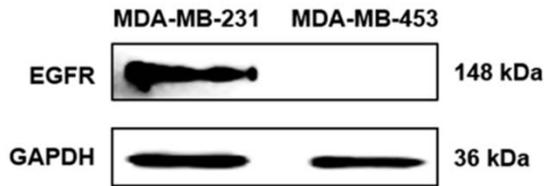


Figure S6. EGFR protein level in MDA-MB-231 and MDA-MB-453 cell lines. Expression of the EGF receptor in MDA-MB-231 and MDA-MB-453 cells was verified by western blotting.

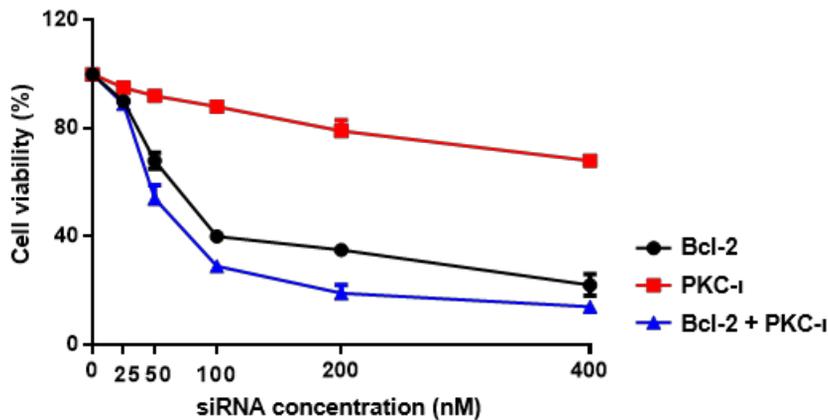


Figure S7. *In vitro* cytotoxicity of Bcl-2 and PKC- ι siRNAs in MDA-MB-231 cells. MDA-MB-231 cells were incubated with QLs encapsulating varied concentrations of Bcl-2 siRNA, PKC- ι siRNA, or Bcl-2/PKC- ι siRNA (1:1 molar ratio) for 48 h. Viability of the transfected cells was measured by the CCK-8 assay.

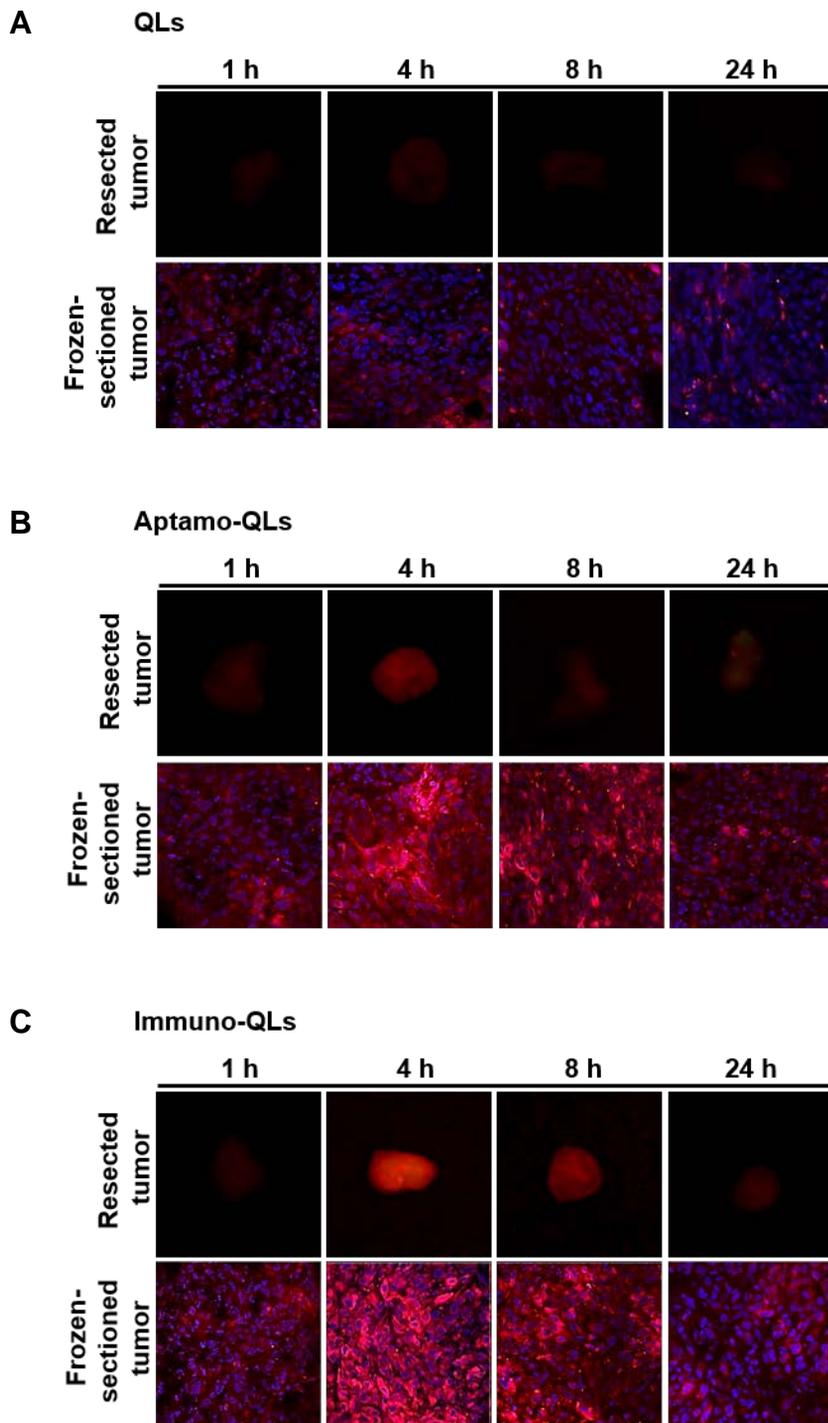


Figure S8. Fluorescence imaging of resected tumors and their frozen-sectioned tissues in mice treated with various QL formulations. After fluorescence imaging, mice intravenously treated with QLs (A), aptamo-QLs (B), and immuno-QLs (C) (0.2 mg lipid/mouse, n = 3) were sacrificed. The fluorescence intensity of resected tumors was observed using the Maestro 2 *in vivo* imaging system, and the frozen-sectioned tumor tissue areas were observed using a confocal microscope at 1, 4, 8, and 24 h time points ($\times 100$).

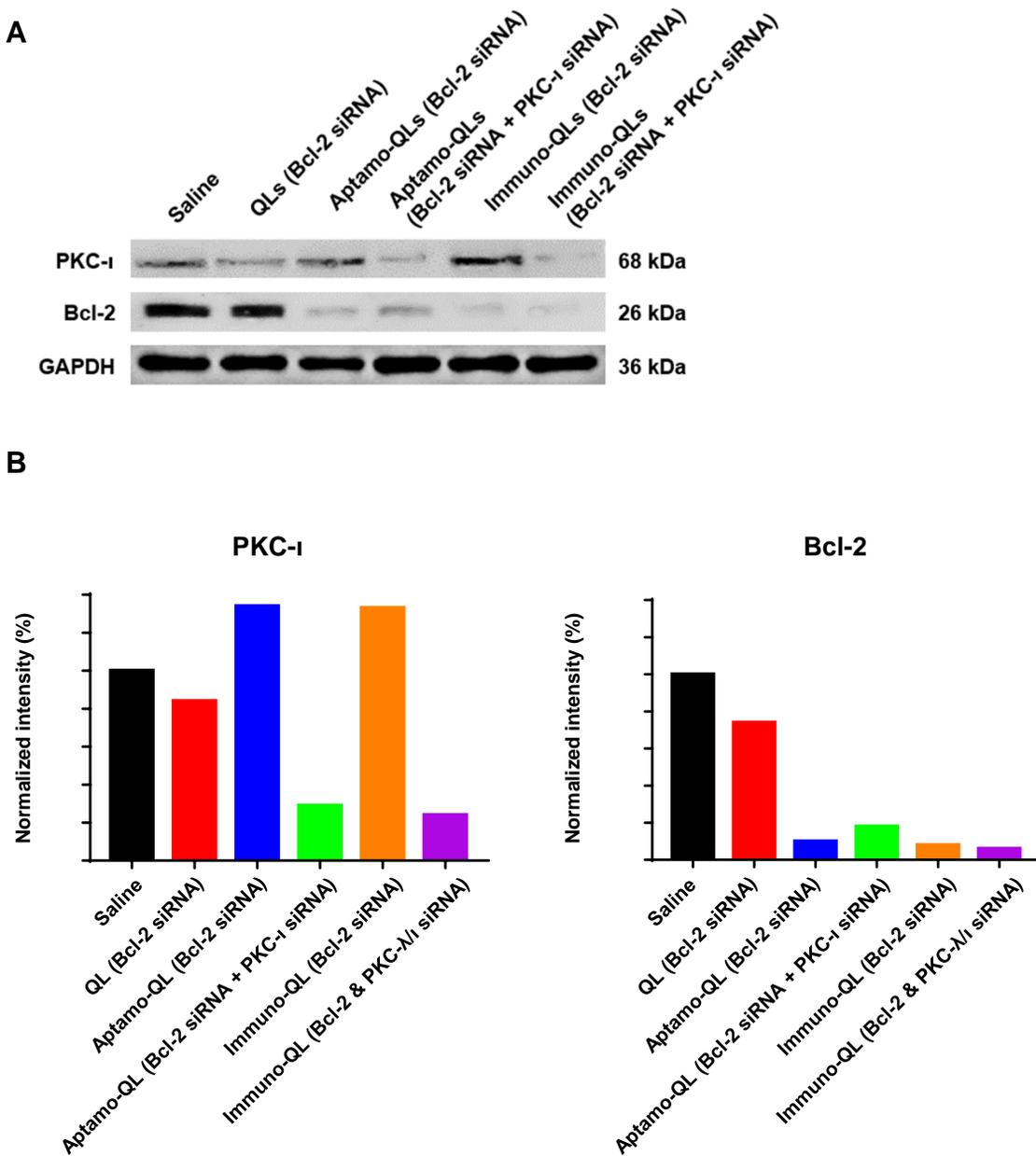


Figure S9. Reduction of Bcl-2 and PKC- ι expression by *in vivo* siRNA transfection.

(A) The tumor-carrying mice were intravenously administered with various QLs encapsulating Bcl-2 and/or PKC- ι siRNAs (10 mg/kg) three times at three-day intervals. At the 10th day post-injection, the mouse tumors were excised, and Bcl-2 and PKC- ι protein expression in the tumors were compared by western blotting analysis (n = 2). (B) The relative band intensities of PKC- ι and Bcl-2 were normalized with GAPDH expression.

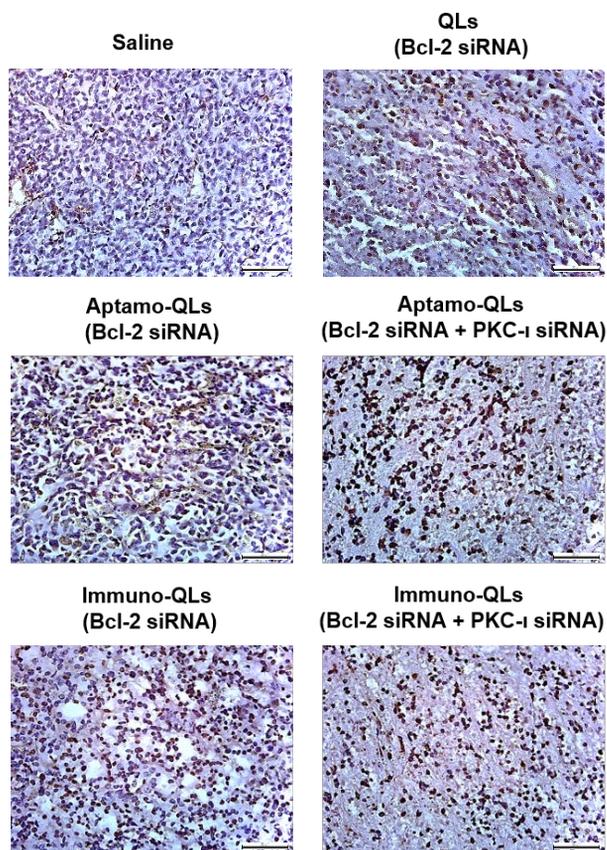


Figure S10. TUNEL staining of tumor sections from mice treated with various QL formulations. TUNEL-positive nuclei in representative tumor sections are shown in brown (n=6). Magnification; $\times 400$. The scale bar is 50 μm long.

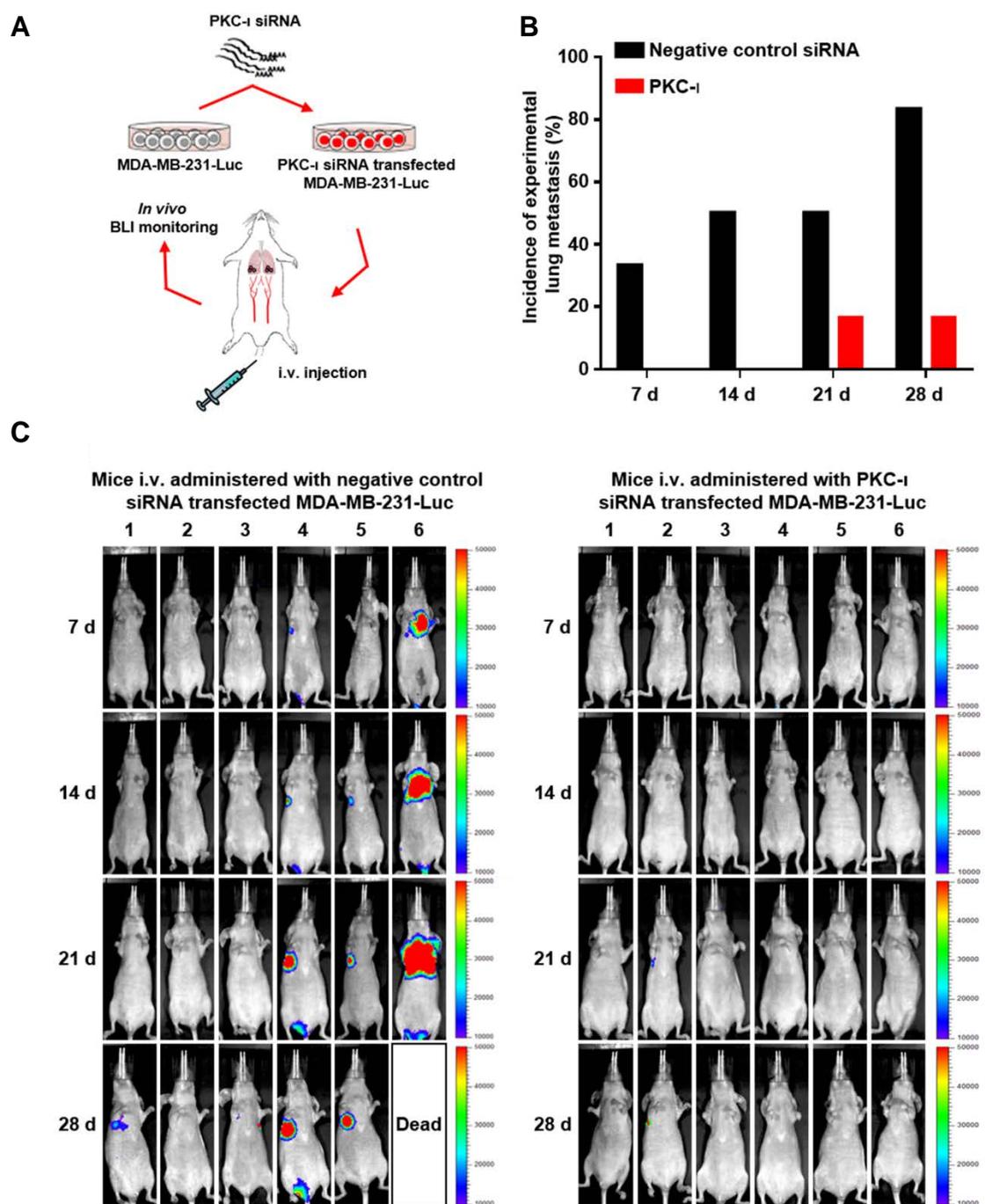


Figure S11. Reduction of pulmonary metastasis by PKC- ι transfection. (A) Schematic of metastatic lung colonization assay after intravenous administration of MDA-MB-231-Luc (2×10^6 cells in $100 \mu\text{L}$ PBS) with or without PKC- ι transfection. (B) The incidences of experimental lung metastasis were compared in the two groups. (C) The mice intravenously administered with PKC- ι siRNA-transfected MDA-MB-231 cells stably expressing luciferase were monitored by IVIS for 4 weeks using a luciferase assay ($n = 6$).

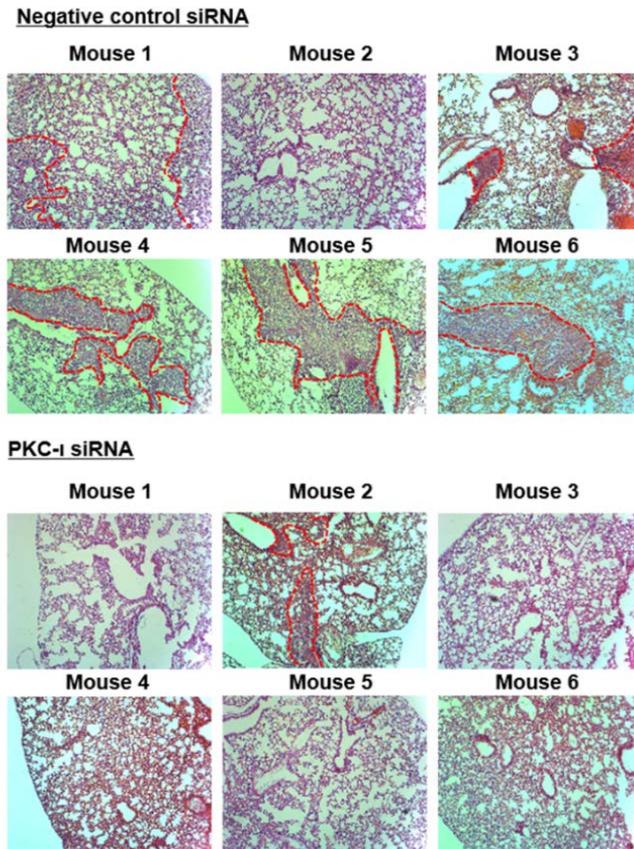


Figure S12. Histological sections of lung tissues from mice intravenously administered with PKC- ι siRNA-transfected MDA-MB-231-Luc. After 4 BLI monitoring by IVIS for 4 weeks using a luciferase assay (n = 6), representative images of lung tissue sections of control (upper) and PKC- ι siRNA-transfected (lower) groups were taken after H&E staining. The tissues in red dotted lines are pulmonary metastasized tumors ($\times 200$).

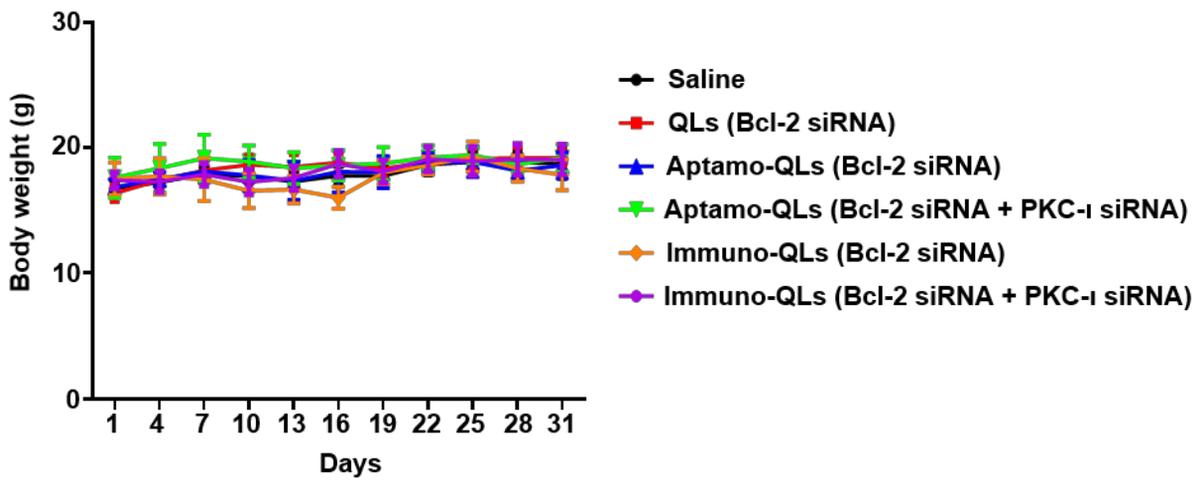


Figure S13. Body weight changes in mice treated with aptamo-QLs and immuno-QLs.

Mouse body weights were measured after intravenous administration of the various QLs encapsulating siRNAs (10 mg/kg) three times at three-day intervals (n = 5).

Table S2. Components of lipoplexes

	Components							
	Lipoplex : Q-dots (w/w ratio)	Lipoplex : siRNA (N/P ratio)	DMKE (mole%)	Cholesterol (mole%)	DSPE-mPEG2000 (mole%)	Post-insertion		
						DSPE- mPEG2000 (mole%)	DSPE-PEG2000- Apt (mole%)	DSPE- PEG2000-Ab (mole%)
CLs	-	4:1	50	50	-	-	-	-
QLs	5:1	4:1	48	48	4	-	-	-
Aptamo-QLs	5:1	4:1	46	46	4	3.8	0.2	-
Immuno-QLs	5:1	4:1	46	46	4	3.8	-	0.2