

# Supporting information

## Anti-EGF receptor aptamer-guided co-delivery of anti-cancer siRNAs and quantum dots for theranosis of triple negative breast cancer

Min Woo Kim<sup>1,2</sup>, Hwa Yeon Jeong<sup>1</sup>, Seong Jae Kang<sup>1</sup>, In Ho Jeong<sup>1</sup>, Moon Jung Choi<sup>1</sup>, Young Myoung You<sup>1</sup>, Chan Su Im<sup>1</sup>, In Ho Song<sup>4</sup>, Tae Sup Lee<sup>4</sup>, Jin Suk Lee<sup>5</sup>, Aeju Lee<sup>2,3</sup> and Yong Serk Park<sup>1\*</sup>

<sup>1</sup>Department of Biomedical Laboratory Science, Yonsei University, Wonju, Republic of Korea;

<sup>2</sup>International Research Organization for Advance Science and Technology (IROAST), Kumamoto University, Kumamoto, Japan;

<sup>3</sup>Magnesium Research Center, Kumamoto University, Kumamoto, Japan

<sup>4</sup>Division of RI-Convergence Research, Korea Institute of Radiological and Medical Science, Seoul, Republic of Korea;

<sup>5</sup>Department of Anatomy, Yonsei University Wonju Collage of Medicine, Wonju, Republic of Korea;

\*Corresponding Author: Professor YS Park, Department of Biomedical Laboratory Science, Yonsei University, Wonju, Gangwon 220-710, Republic of Korea

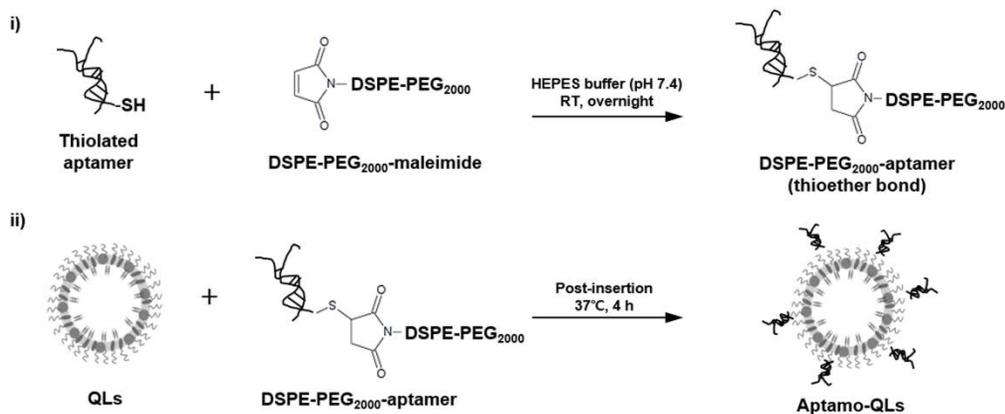
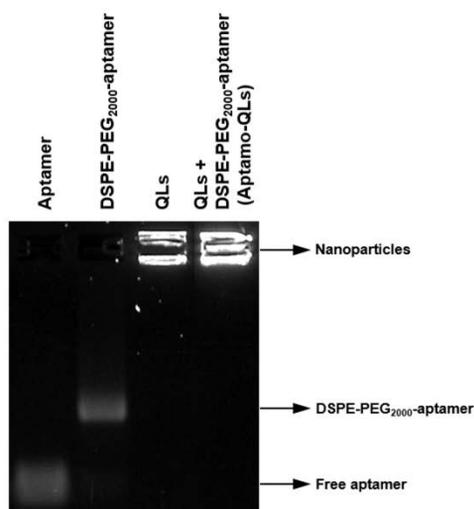
Telephone & Fax: 82-33-760-2448 / 82-33-760-2561

E-mail: parkys@yonsei.ac.kr

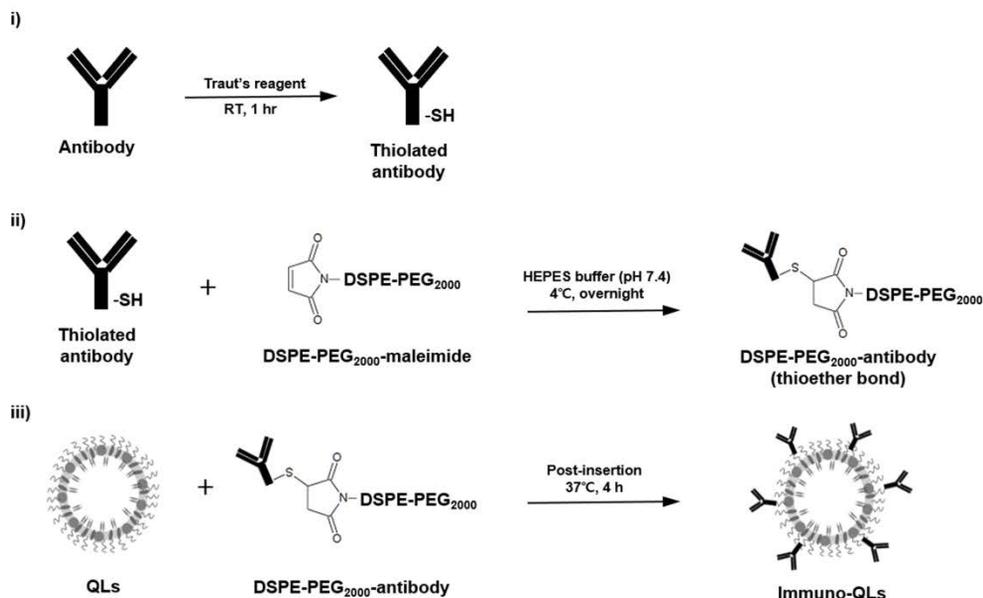
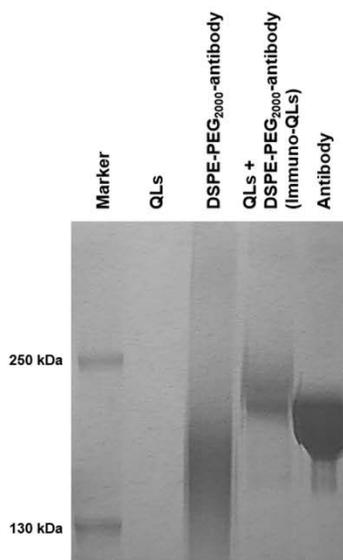
**Table S1. The vesicular size,  $\zeta$ -potential, and Q-dot incorporation rate of lipoplexes prepared at varied mole% of DSPE-mPEG<sub>2000</sub>**

<b>DSPE-mPEG<sub>2000</sub>/Lipoplexes (mole%)</b>	<b>Vesicle size (nm)</b>	<b>Polydispersity Index (PDI)</b>	<b><math>\zeta</math>-potential (mV)</b>	<b>Q-dot incorporation rate (%)</b>	<b>Q-dot loading capacity (%)</b>
0/100	385 ± 79	0.41 ± 0.36	51.8 ± 1.3	67.9 ± 1.2	11.3 ± 0.2
1/99	339 ± 74	0.32 ± 0.23	22.9 ± 3.1	75.0 ± 6.3	12.5 ± 1.1
4/96	175 ± 5	0.25 ± 0.04	8.0 ± 3.4	94.9 ± 5.8	15.8 ± 0.9
8/92	185 ± 5	0.22 ± 0.03	0.2 ± 0.5	93.7 ± 4.0	15.5 ± 0.7

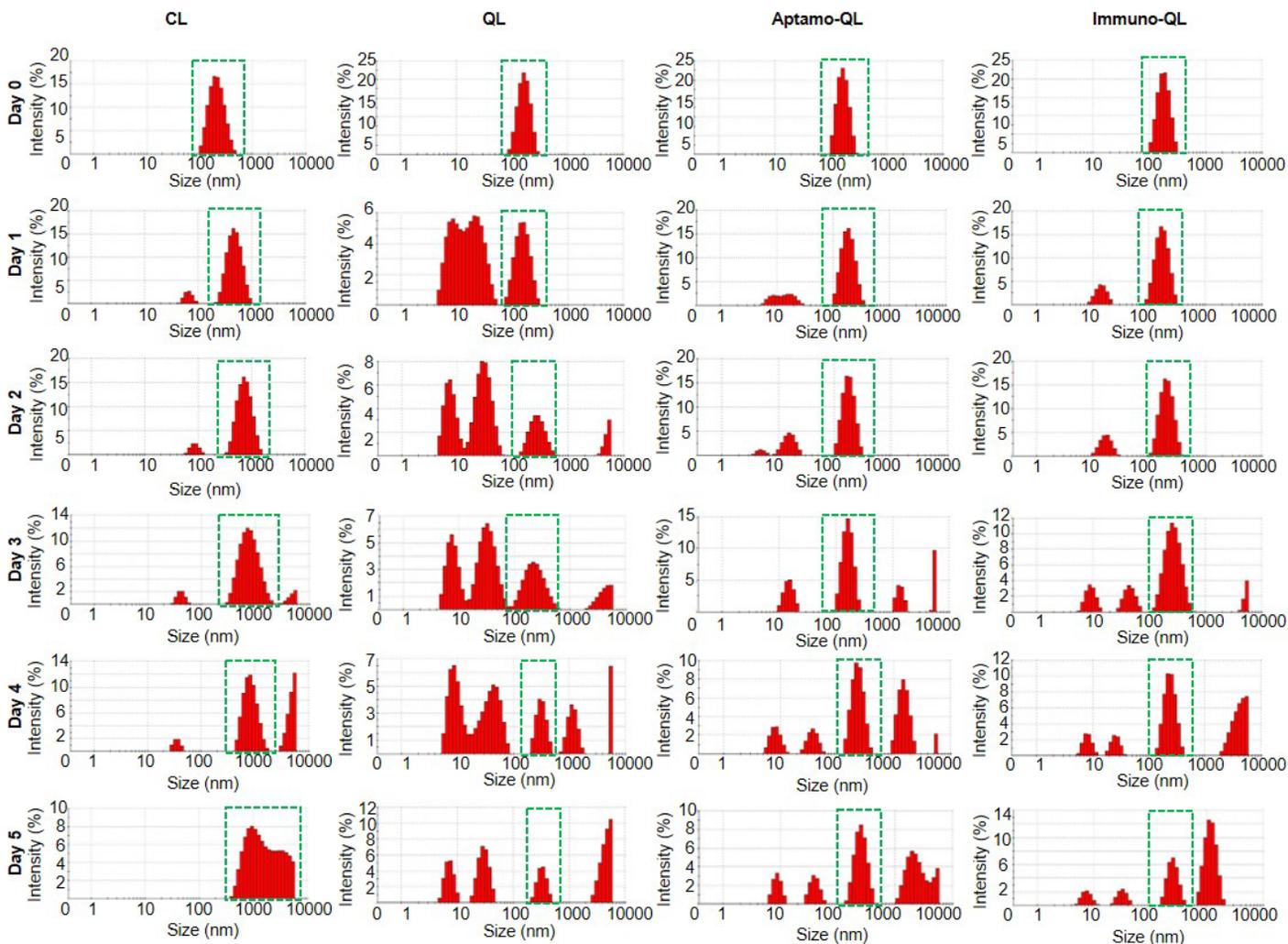
The values were measured using a Zetasizer Nano-ZS90. Each value indicates the mean ± standard deviation (S.D.) of five measurements.

**A****Reaction Scheme****B**

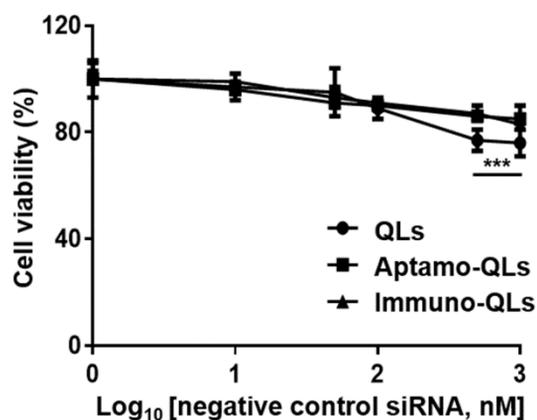
**Figure S1. Anti-EGFR aptamer conjugation to DSPE-PEG<sub>2000</sub>-maleimide.** (A) Thiolated aptamers were conjugated to DSPE-PEG<sub>2000</sub>-maleimide and post-inserted into QLs. (B) The post-insertion efficiency was measured by electrophoresis using a 1.5% agarose gel. Lane 1: free aptamer, lane 2: DSPE-PEG<sub>2000</sub>-aptamer, lane 3: QLs, and lane 4: QLs inserted with DSPE-PEG<sub>2000</sub>-aptamer.

**A****Reaction Scheme****B**

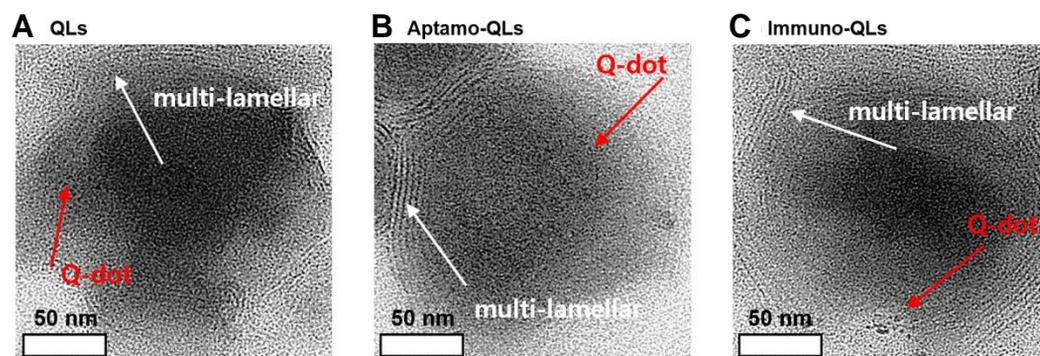
**Figure S2. Anti-EGFR antibody conjugation to DSPE-PEG<sub>2000</sub>-maleimide.** (A) A procedure of antibody thiolation, antibody conjugation to DSPE-PEG<sub>2000</sub>-maleimide, and insertion of antibody-lipid conjugate into QLs is illustrated. (B) Antibody conjugation to DSPE-PEG<sub>2000</sub>-maleimide and insertion of the conjugate into QLs were verified by 6% SDS-PAGE. Lane 1: marker, lane 2: QLs, lane 3: DSPE-PEG<sub>2000</sub>-antibody, lane 4: DSPE-PEG<sub>2000</sub>-antibody-inserted QLs, and lane 5: free antibody.



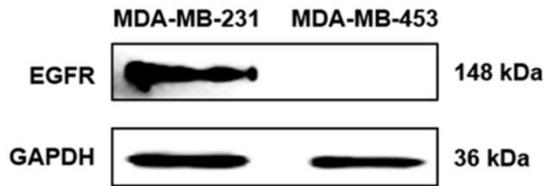
**Figure S3. Size changes of aptamo-QLs and immuno-QLs in FBS.** The various QL formulations were incubated in the presence of 50% FBS at 37°C and their changes in size were examined with a particle analyzer.



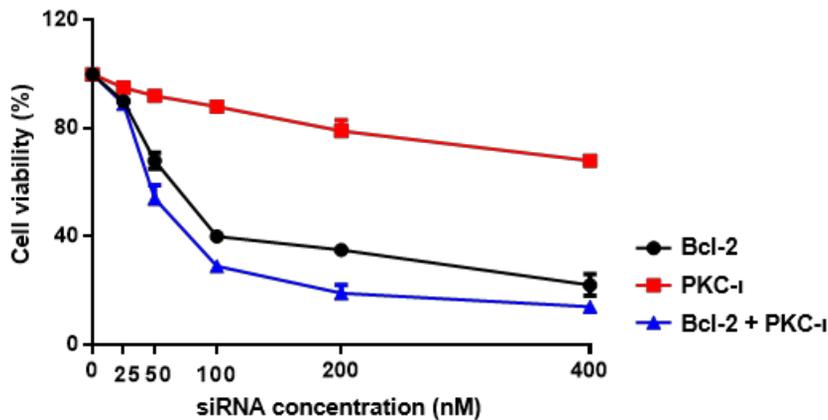
**Figure S4. *In vitro* cytotoxicity of aptamo-QLs and immuno-QLs.** MDA-MB-231 cells were treated with varied concentrations of prepared negative control siRNA lipoplexes and further cultured for 48 h. Cell viability was measured by the CCK-8 assay. Each error bar represents the mean  $\pm$  S.D. of five independent experiments. \*\*\* $p < 0.001$  vs. untreated control.



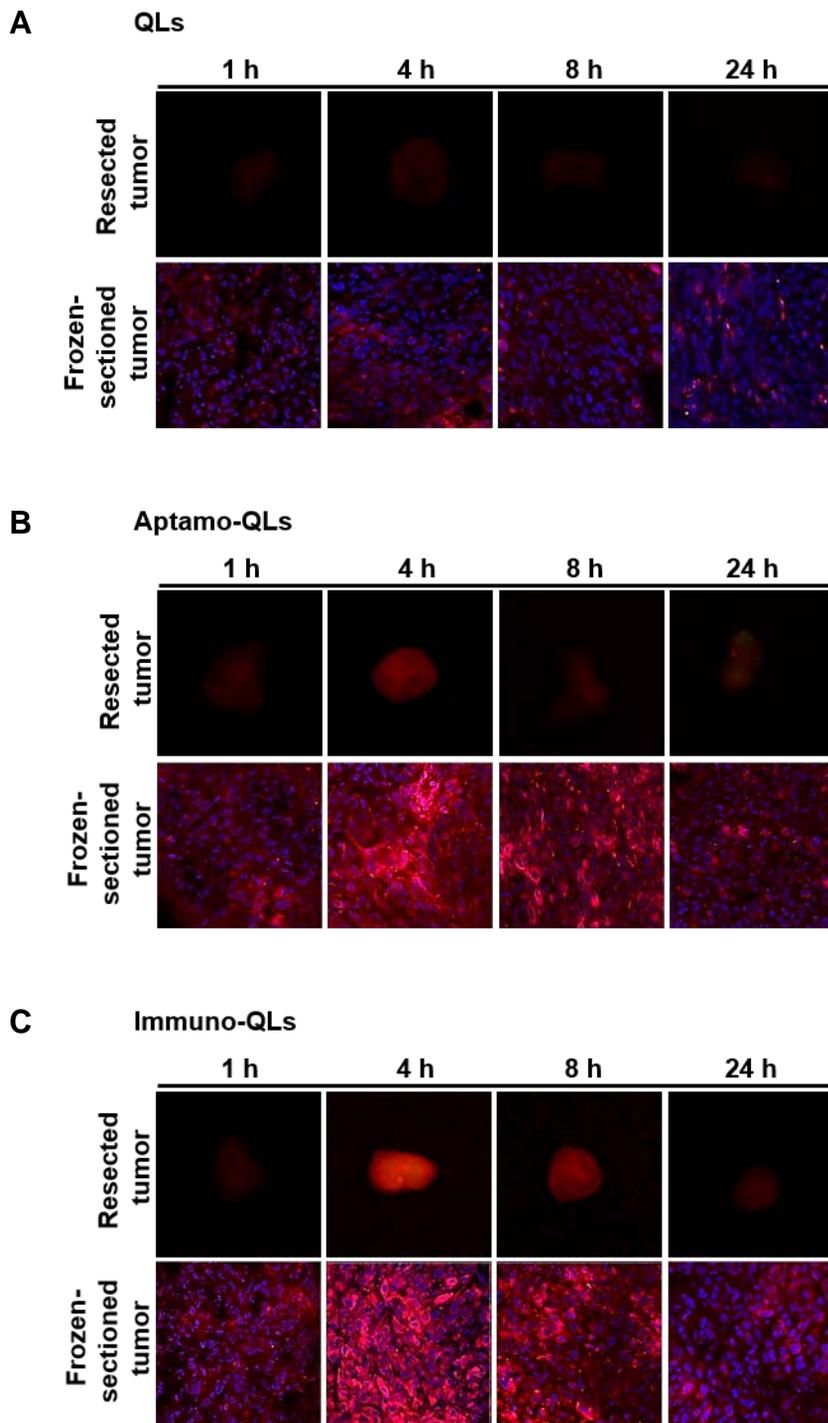
**Figure S5. TEM images of aptamo-QLs and immuno-QLs.** Representative transmission electron microscopy (TEM) images of QLs (A), aptamo-QLs (B), and immuno-QLs (C) are shown. All lipoplexes showed multi-lamellar structures (white arrow) and Q-dots incorporated in the lipoplex layers (red arrow). Bar = 50 nm.



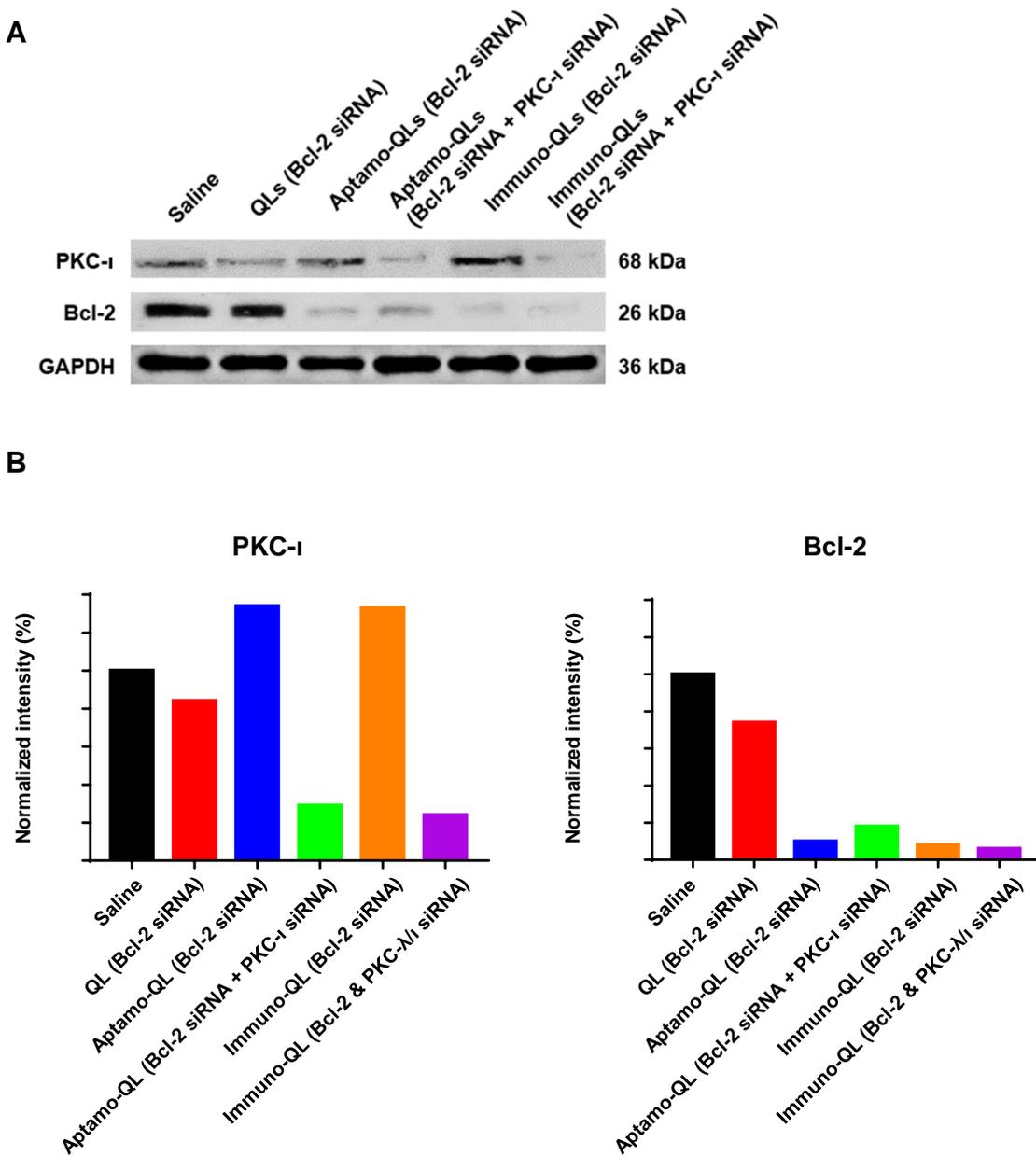
**Figure S6.** EGFR protein level in MDA-MB-231 and MDA-MB-453 cell lines. Expression of the EGF receptor in MDA-MB-231 and MDA-MB-453 cells was verified by western blotting.



**Figure S7.** *In vitro* cytotoxicity of Bcl-2 and PKC- $\iota$  siRNAs in MDA-MB-231 cells. MDA-MB-231 cells were incubated with QLs encapsulating varied concentrations of Bcl-2 siRNA, PKC- $\iota$  siRNA, or Bcl-2/PKC- $\iota$  siRNA (1:1 molar ratio) for 48 h. Viability of the transfected cells was measured by the CCK-8 assay.

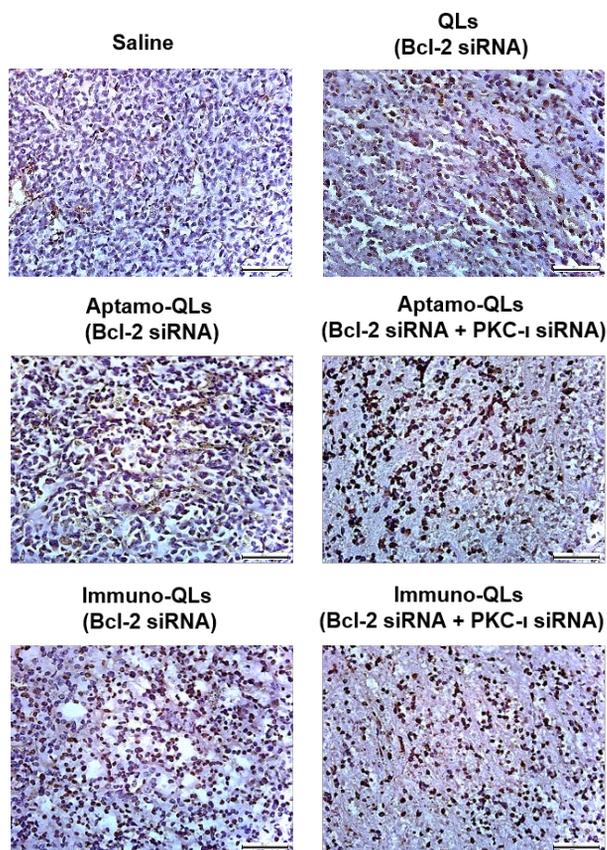


**Figure S8. Fluorescence imaging of resected tumors and their frozen-sectioned tissues in mice treated with various QL formulations.** After fluorescence imaging, mice intravenously treated with QLs (A), aptamo-QLs (B), and immuno-QLs (C) (0.2 mg lipid/mouse, n = 3) were sacrificed. The fluorescence intensity of resected tumors was observed using the Maestro 2 *in vivo* imaging system, and the frozen-sectioned tumor tissue areas were observed using a confocal microscope at 1, 4, 8, and 24 h time points ( $\times 100$ ).

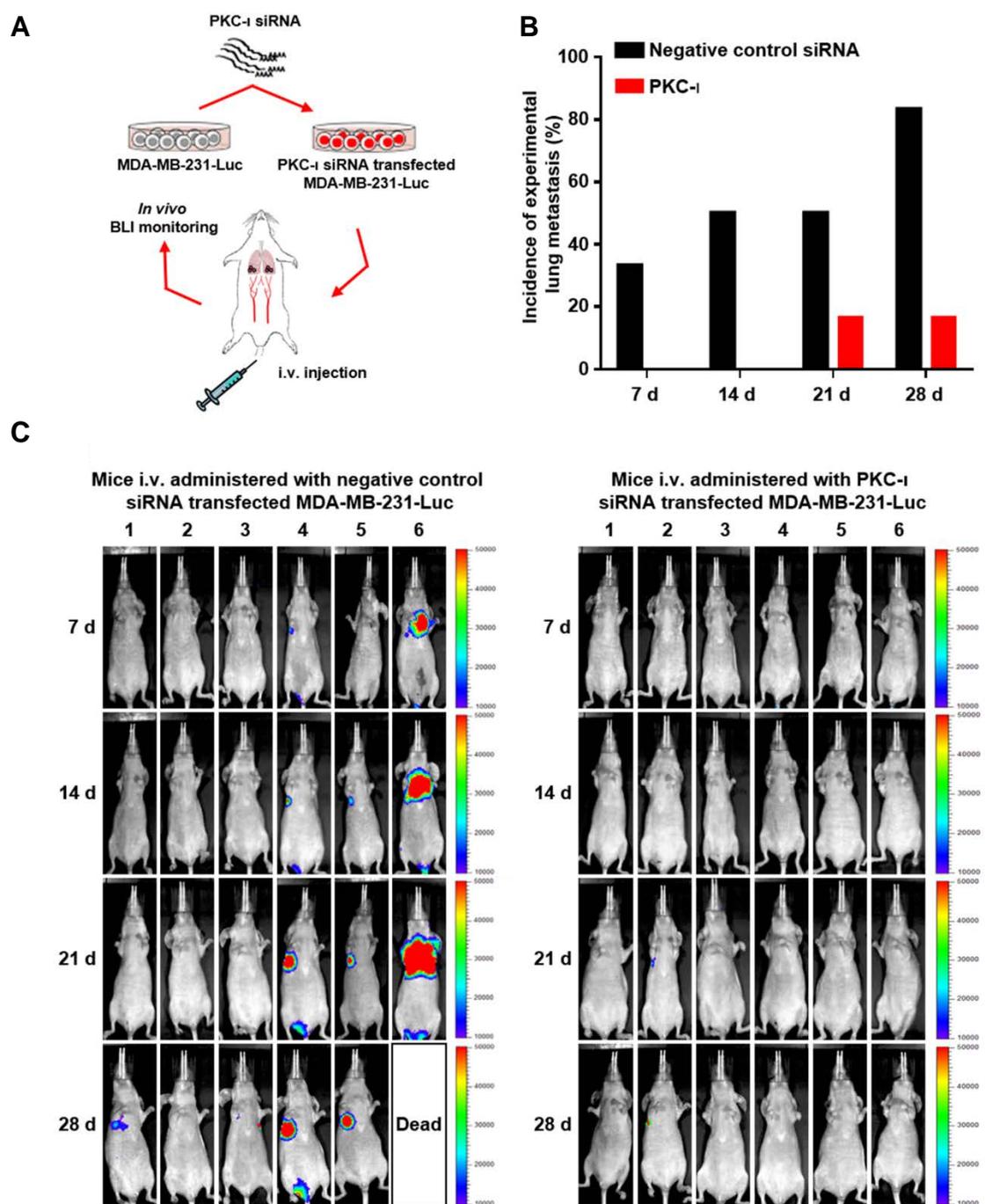


**Figure S9. Reduction of Bcl-2 and PKC- $\iota$  expression by *in vivo* siRNA transfection.**

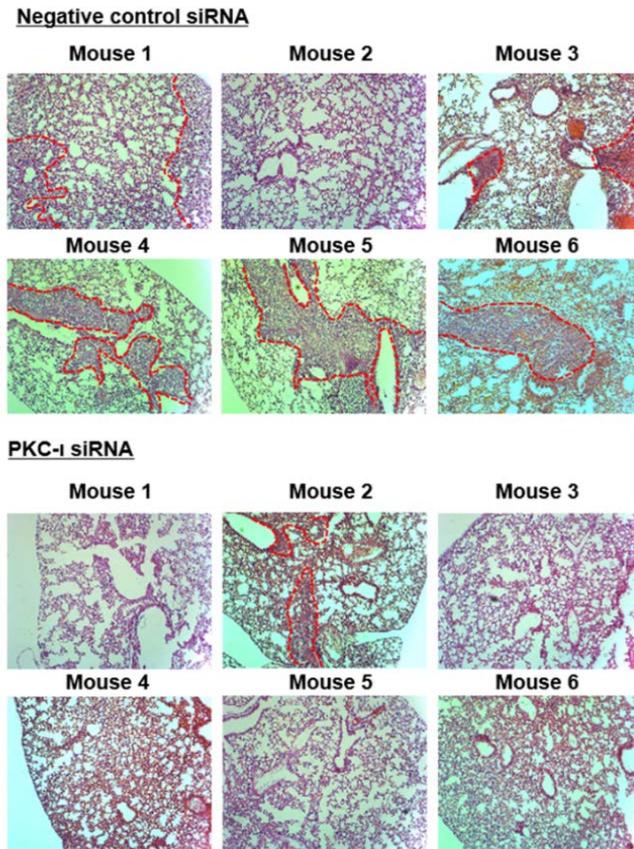
(A) The tumor-carrying mice were intravenously administered with various QLs encapsulating Bcl-2 and/or PKC- $\iota$  siRNAs (10 mg/kg) three times at three-day intervals. At the 10<sup>th</sup> day post-injection, the mouse tumors were excised, and Bcl-2 and PKC- $\iota$  protein expression in the tumors were compared by western blotting analysis (n = 2). (B) The relative band intensities of PKC- $\iota$  and Bcl-2 were normalized with GAPDH expression.



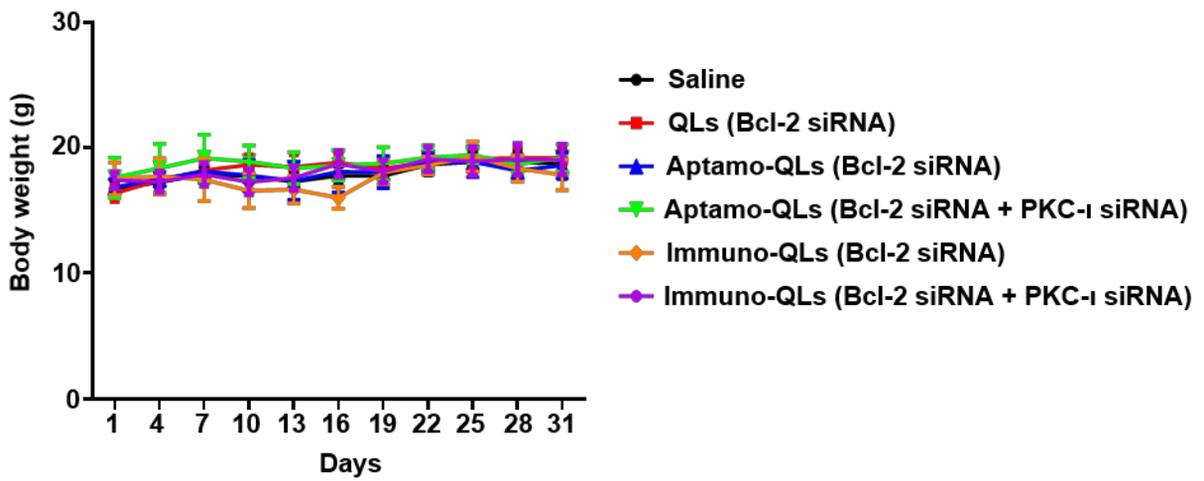
**Figure S10. TUNEL staining of tumor sections from mice treated with various QL formulations.** TUNEL-positive nuclei in representative tumor sections are shown in brown (n=6). Magnification;  $\times 400$ . The scale bar is 50  $\mu\text{m}$  long.



**Figure S11. Reduction of pulmonary metastasis by PKC- $\iota$  transfection.** (A) Schematic of metastatic lung colonization assay after intravenous administration of MDA-MB-231-Luc ( $2 \times 10^6$  cells in  $100 \mu\text{L}$  PBS) with or without PKC- $\iota$  transfection. (B) The incidences of experimental lung metastasis were compared in the two groups. (C) The mice intravenously administered with PKC- $\iota$  siRNA-transfected MDA-MB-231 cells stably expressing luciferase were monitored by IVIS for 4 weeks using a luciferase assay ( $n = 6$ ).



**Figure S12. Histological sections of lung tissues from mice intravenously administered with PKC- $\iota$  siRNA-transfected MDA-MB-231-Luc.** After 4 BLI monitoring by IVIS for 4 weeks using a luciferase assay (n = 6), representative images of lung tissue sections of control (upper) and PKC- $\iota$  siRNA-transfected (lower) groups were taken after H&E staining. The tissues in red dotted lines are pulmonary metastasized tumors ( $\times 200$ ).



**Figure S13. Body weight changes in mice treated with aptamo-QLs and immuno-QLs.**

Mouse body weights were measured after intravenous administration of the various QLs encapsulating siRNAs (10 mg/kg) three times at three-day intervals (n = 5).

**Table S2. Components of lipoplexes**

	Components							
	Lipoplex : Q-dots (w/w ratio)	Lipoplex : siRNA (N/P ratio)	DMKE (mole%)	Cholesterol (mole%)	DSPE-mPEG2000 (mole%)	Post-insertion		
						DSPE- mPEG2000 (mole%)	DSPE-PEG2000- Apt (mole%)	DSPE- PEG2000-Ab (mole%)
CLs	-	4:1	50	50	-	-	-	-
QLs	5:1	4:1	48	48	4	-	-	-
Aptamo-QLs	5:1	4:1	46	46	4	3.8	0.2	-
Immuno-QLs	5:1	4:1	46	46	4	3.8	-	0.2