

Figure S1. CAFs are associated with ferroptosis resistance in PDAC. PDAC cells were cultured with CAF-CM or PCS-CM, and after (A-B) Erastin, RSL3, or IKE treatment, CCK-8 was used to measure cell viability. (C-D) Colony formation by PANC-1 and MIAPaCa-2 cells was evaluated by clonal formation assay after treatment with vehicle, RSL3, or IKE. (E-F) Cell death ratios in PANC-1 and MIAPaCa-2 cells were evaluated by cell death assays after treatment with vehicle, RSL3, or IKE. * P < 0.05, ** P < 0.01, *** P < 0.001.



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9 Figure S2. Exocrine cysteine was essential for CAF-induced ferroptosis resistance in PDAC. (A) Cell viability was measured after treatment with CAF-CM, three repeated freeze-10 11 thaw cycles of CAF-CM, PSC-CM, or three repeated freeze-thaw cycles of PSC-CM using 12 Erastin. (B) Cell viability after culture of PANC-1 cells with cysteine, angelic acid, creatine, or 13 glutamine, measured by CCK-8 assays. The MIAPaCa-2 cell line was cultured with Ctrl or 14 cysteine and treated with Erastin, (C) clonal formation assay to assess cell colony formation 15 capacity, and (D-E) Cell death assay to evaluate the cell death ratio of PANC-1 and MIAPaCa-16 2 cells. (F) Representative IHC staining of Ki67 in mouse xenograft tumors. Scale bar = 100microns. * P < 0.05, ** P < 0.01, *** P < 0.001. 17







Figure S4. The knockout of ASCT2 significantly impairs the uptake of cysteine by tumors. (A) TCGA database mining of EAAT3, ASCT1, ASCT2 expression in tumor and paracancerous tissues. (B) Expression of SLC7A11, EAAT3, ASCT1 and ASCT2 knockdown efficiency of the siRNAs in cells shown by western blotting. (C) Cell viability in PANC-1 and MIAPaCa-2 cells was measured by CCK-8 assays with siSLC7A11, siEAAT3, siASCT1 or siASCT2. (D) Cysteine levels in PANC-1 and MIAPaCa-2 cells with siSLC7A11, siEAAT3, siASCT1 or siASCT2. * P < 0.05, *** P < 0.001.



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38 Figure S5. Pancreatic cancer cells require exogenous cysteine-dependent GSH synthesis

to avert ferroptosis. (A-B) Micrographs of cell death after drug treatment. (C) Flow cytometry
and CCK-8 assays were used to measure cell death and cell viability, respectively. (D) ROS

41 levels, assessed by flow cytometry. (E) MDA contents. (F) GSH levels. T, tumor tissue; N,

42 paracancerous tissue. * P < 0.05, ** P < 0.01, *** P < 0.001.



45 Figure S6. GCLC inhibitor (BSO) decreased intracellular GSH level. (A) GSH levels. (B)

- 46 Cell death measured by flow cytometry with BSO treatment. (C) colony formation assays with
- 47 BSO treatment. *** P < 0.001.



50 Figure S7. CAFs release cysteine inducing cisplatin resistance in PDAC. (A) Cell viability 51 measured by CCK-8 assays. (B) GSH levels. (C) Cell death measured by flow cytometry. (D) 52 Proliferation measured by EdU assays. (E) Expression of γ H2AX in cells shown by western 53 blotting. (F) ICP-MS measurement of intracellular cisplatin content. (G) Statistics on the ratio 54 of cisplatin content to tumor size in tumor tissue. ** P < 0.01, *** P < 0.001_o





57 Figure S8. Stromal transsulfuration pathway is regulated by TGF- β /SMAD3/ATF4 58 signaling. (A) Expression of CBS and CTH in cells shown by RT-PCR. (B) Expression of CBS 59 and CTH in cells shown by western blotting. (C) Expression of α -SMA shown by RT-PCR and 60 immunofluorescence. (D) Expression of CBS in cells after PDAC-CM treatment shown by RT-61 PCR. ** P < 0.01, *** P < 0.001.







66 ATF4 in cells shown by western blotting and RT-PCR. *** P < 0.001.



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Figure S10. TGF-β/SMAD3/ATF4 signaling pathway promotes ferroptosis and cisplatin
resistance in PDAC. (A) Cysteine secretion measured after inhibition of the TGF β/SMAD3/ATF4 signaling pathway. (B) Cell viability after treatment with different drugs,
measured by CCK-8 assays. (C-D) Colony formation by cells after treatment with Erastin or

- 72 Cisplatin. ** P < 0.01, *** P < 0.001.
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Table S1 Primer sequences.

Name	Forward primer (5' to 3')	Reverse primer (5' to 3')
CBS	ACAGAGCTCACACTTCA	GGTCTGGAGACCGCTGTAAG
	GGC	
CTH	GCATGGGGGGTACTTGACA	TGCACTTTGACTGAGCTCCC
	CA	
SLC7A11	AAAGCCTGTTGTGTCCAC	AGAAAATCTGGATCCGGGCG
	CA	
α-SMA	AGCGTGGCTATTCCTCCG	GCAGTGGCCATCTCATTTTCA
	TT	
ATF4	TCGTCCTGGTGGGATCTA	TCTGGCATGGTTTCCAGGTC
	GG	
β-actin	AGCGAGCATCCCCCAAA	GGGCACGAAGGCTCATCATT
	GTT	