

Supplementary Material

Supplemental Figures

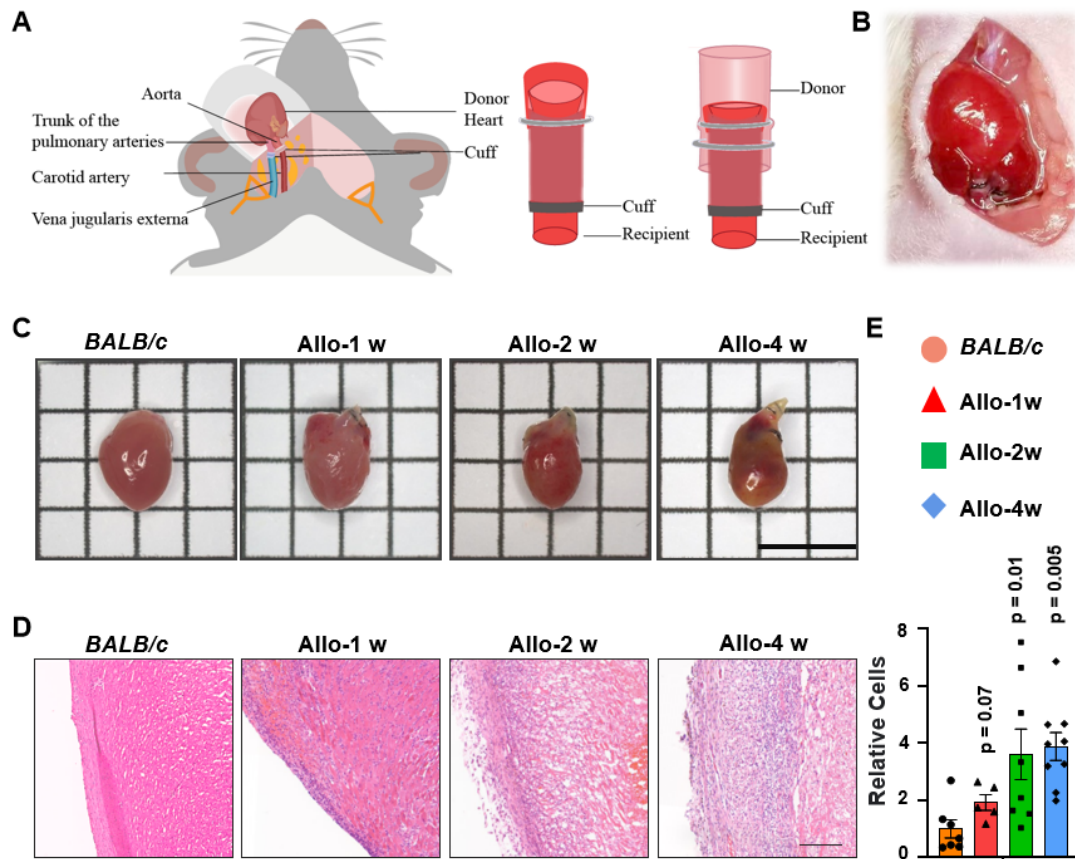


Figure S1. **A**, Sketch of the mouse heterotopic cervical heart transplantation. **B**, Construction of the mouse heterotopic cervical heart transplantation model. **C**, Cardiac allografts at indicated time points. **D**, HE staining of cardiac allograft slices at indicated time points. **E**, Quantification of relative cell number, cell number of BALB/c group as the criterion (1.0); Number of mice in each group: n = 7(control), n = 5 (Allo-1 w), n = 8 (Allo-2 w), n = 9 (Allo-4 w), one-way ANOVA test. Scale bars, 1 mm (C), and 200 μ m (D).

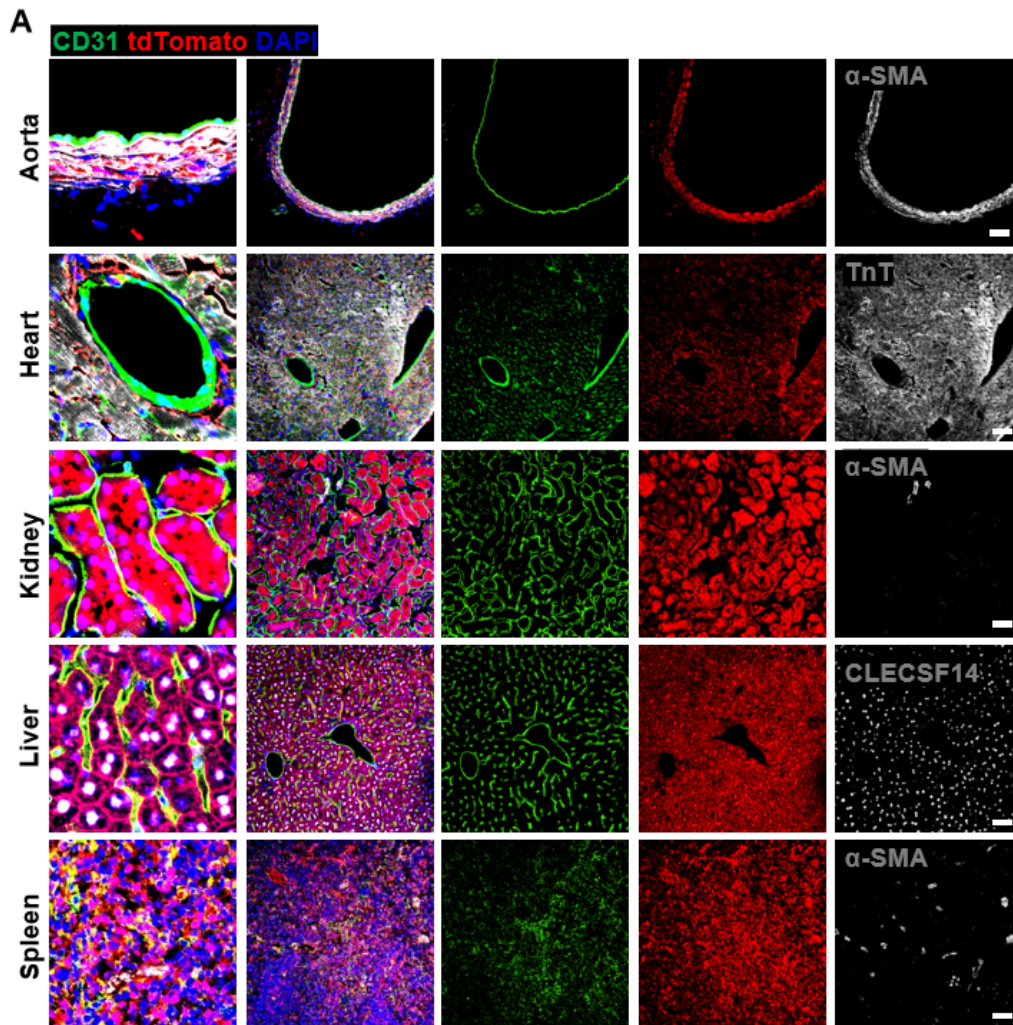


Figure S2. A, Immunofluorescence staining to verify the efficiency of tdTomato-labeled CAG⁺ cells in Cag-Cre;R26-tdTomato mice. Scale bar: 100 μ m.

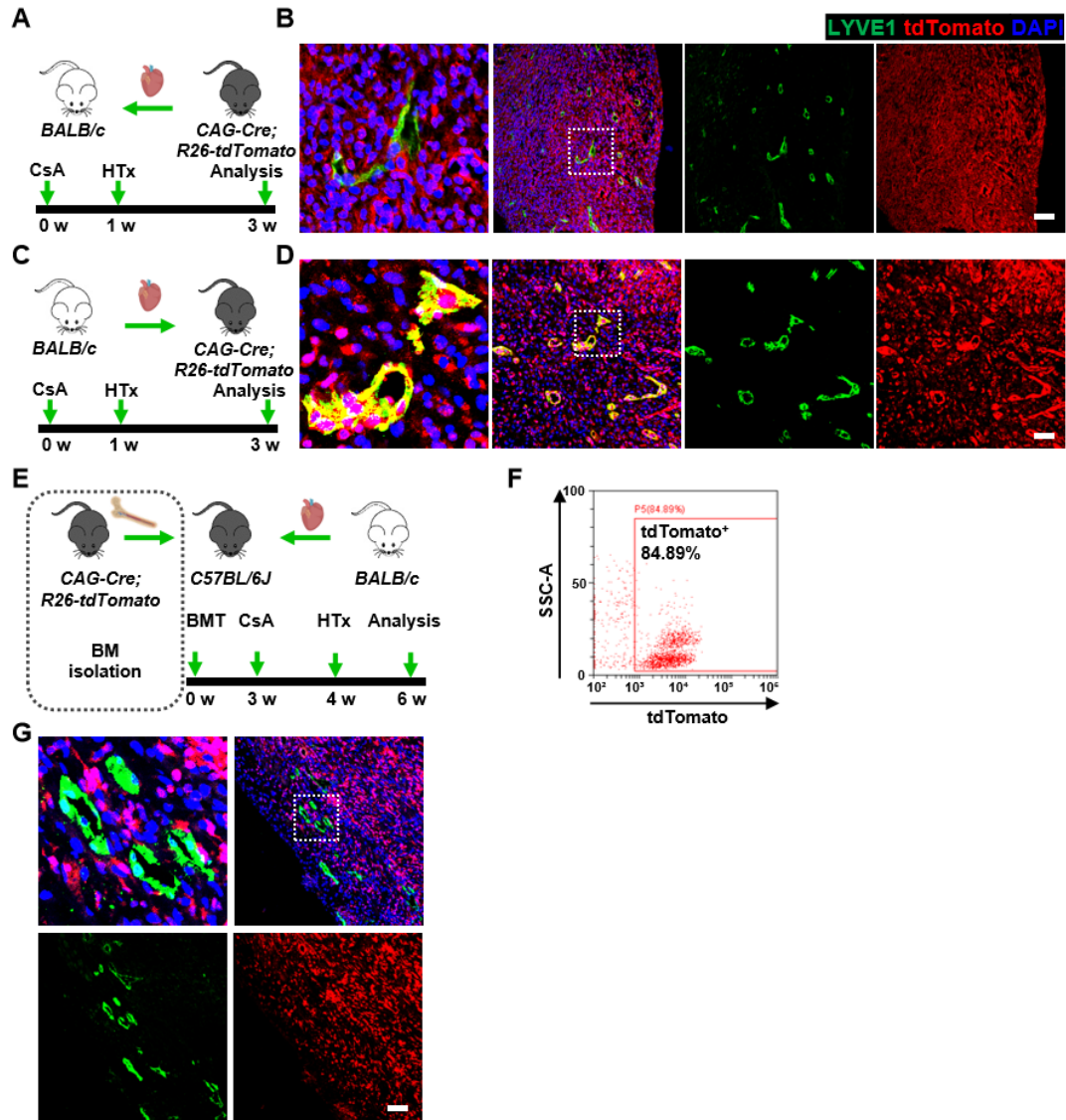


Figure S3. A, Schematic of heart transplantation model: Donor heart from Cag-Cre;R26-tdTomato mouse was transplanted into BALB/c recipient mouse. B, Immuno-staining of lymphatic vessels in allograft heart 2 weeks after transplantation which was described in (A). The images within the dashed box are displayed on the left. C, Schematic of heart transplantation model: Donor heart from BALB/c mouse was transplanted into Cag-Cre;R26-tdTomato recipient mouse. D, Immuno-staining of lymphatic vessels in allograft heart 2 weeks after transplantation which was described in (C). E, Schematic of heart transplantation model: Bone marrow (BM) of C57BL/6J recipient mice was replaced with the BM harvested from Cag-Cre;R26-tdTomato mice, subsequently the donor hearts from BALB/c mice were transplanted into C57BL/6J recipient mice. F, Quantification of tdTomato positive cells by flow cytometry in the reconstructed BM of recipient mice before heart transplantation which was described in (E). G, Immuno-staining of lymphatic vessels in allograft heart 2 weeks after transplantation which was described in (E). The image within the dashed box is displayed on the right. Scale bars, 100 μ m (B, D and G).

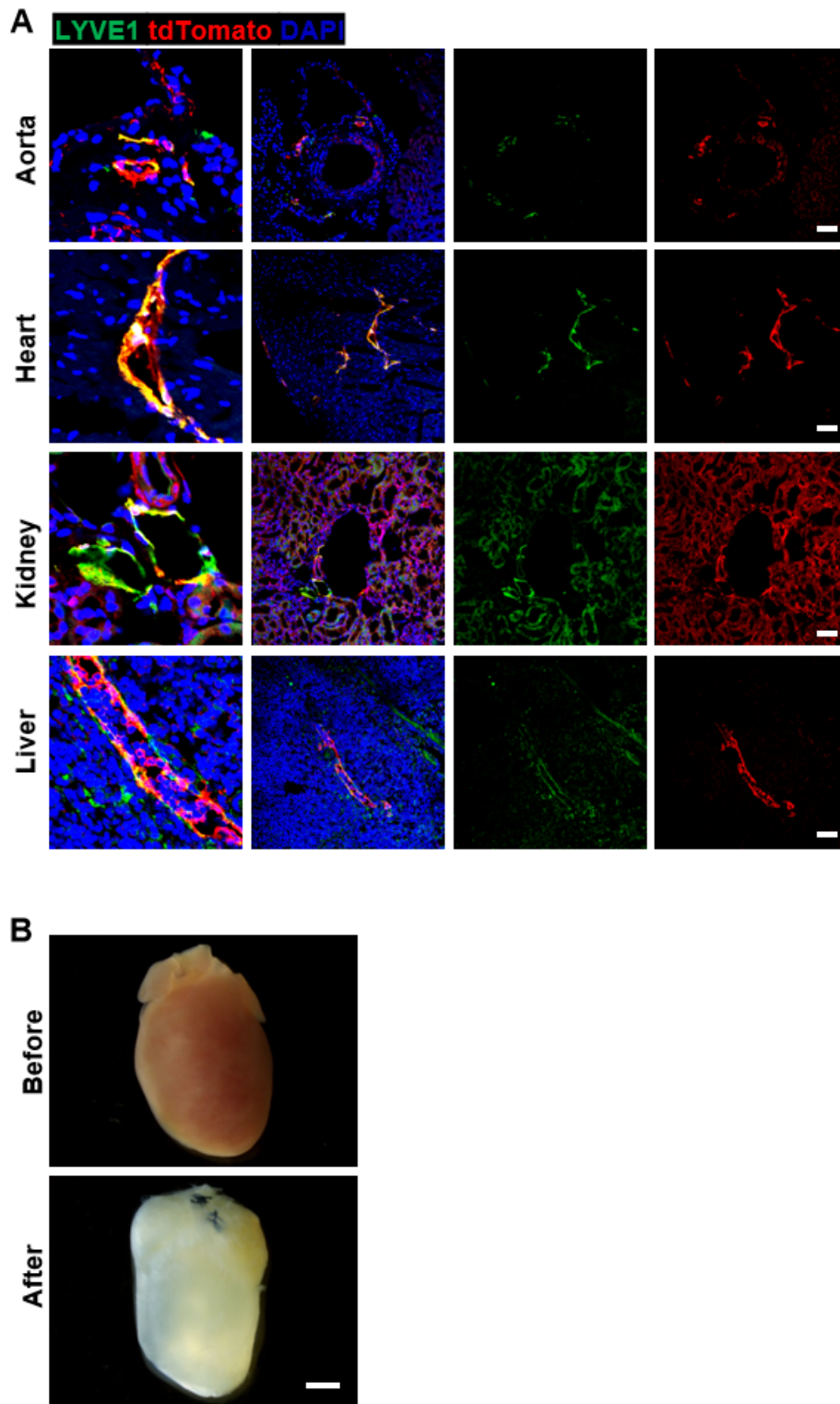


Figure S4. A, Immunofluorescence staining to verify the efficiency of tdTomato-labeled LYVE1+ cells in Lyve1-CreERT2;R26-tdTomato mice. B, Whole heart clearing with CUBIC. Scale bar: 100 μ m (A), 1 mm (H).

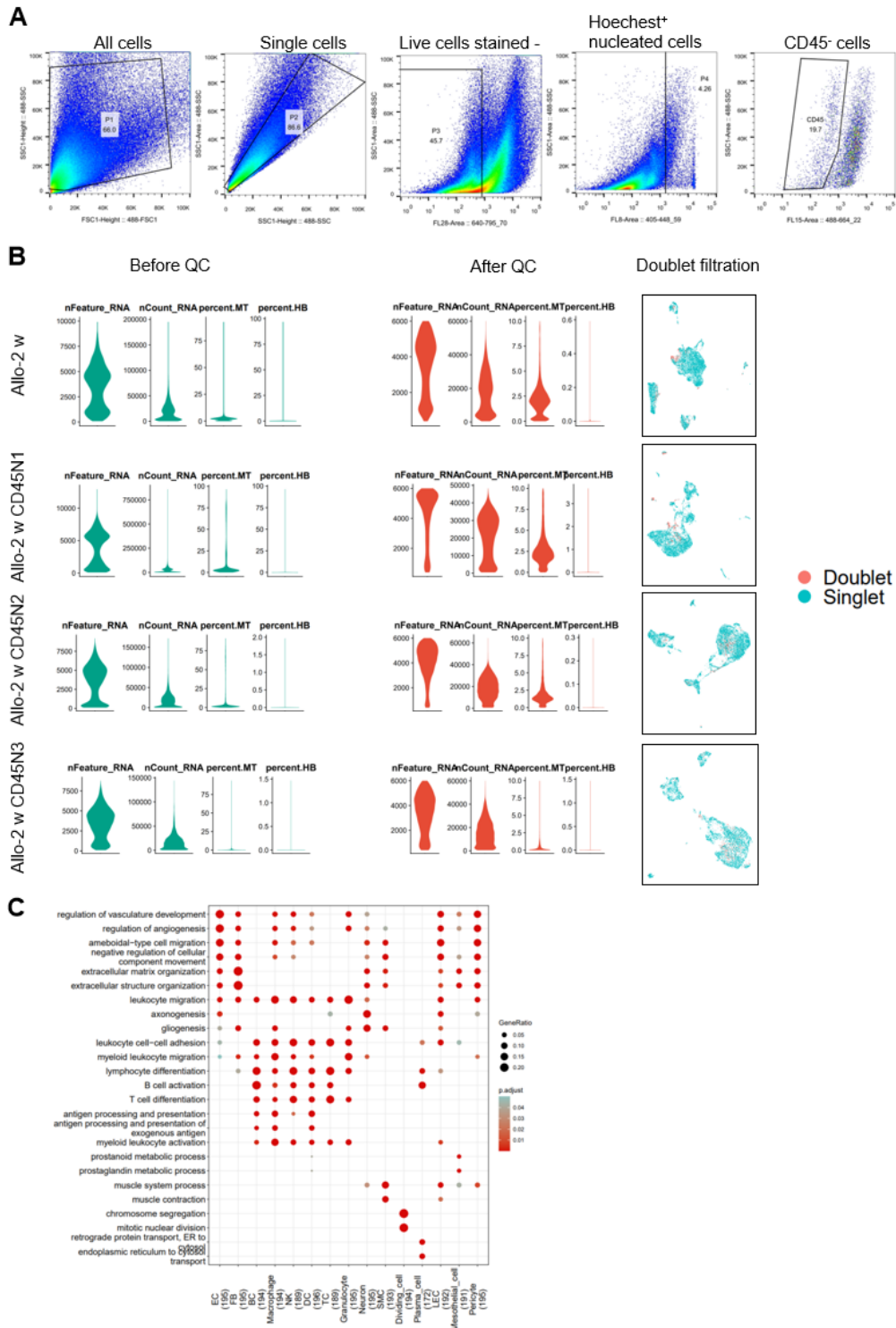


Figure S5. A, Flow cytometry of sorting strategy. B, Quantity control of single-cell RNA sequencing data. C, Dot plot shows the GO enrichment terms of major cell types.

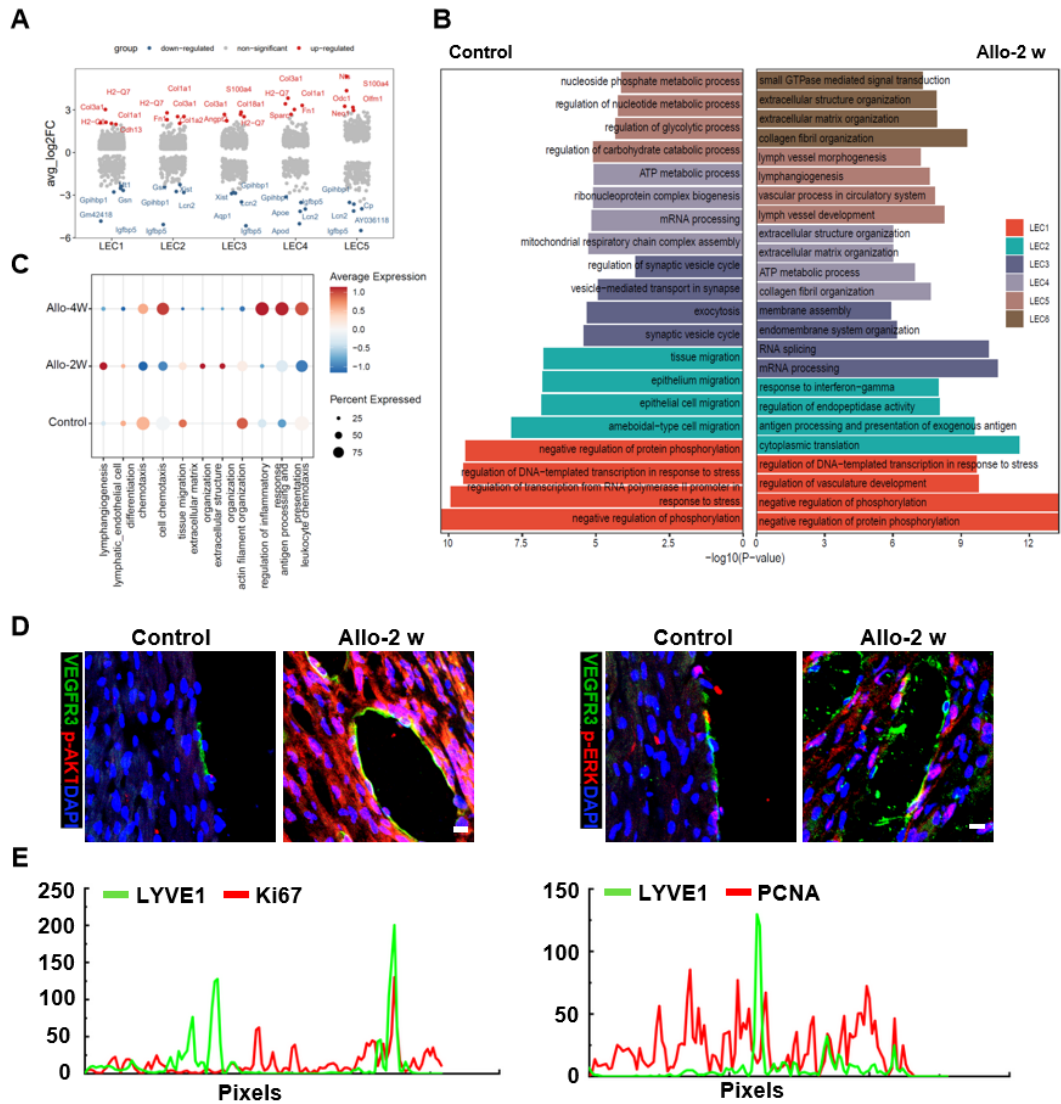


Figure S6. **A**, Volcano plot shows DEGs between two groups. **B**, Bar plot shows GO enriched in LEC sub-clusters from two groups. **C**, Dot plot shows GO enrichment of DEGs in different groups. **D**, Immuno-staining of p-AKT (left) and p-ERK (right) in LECs from two groups. **E**, Image J analysis of co-immuno staining of Ki67 (left) and PCNA (right) with LYVE1 in the cardiac allograft group. Scale bars, 25 μ m (D).

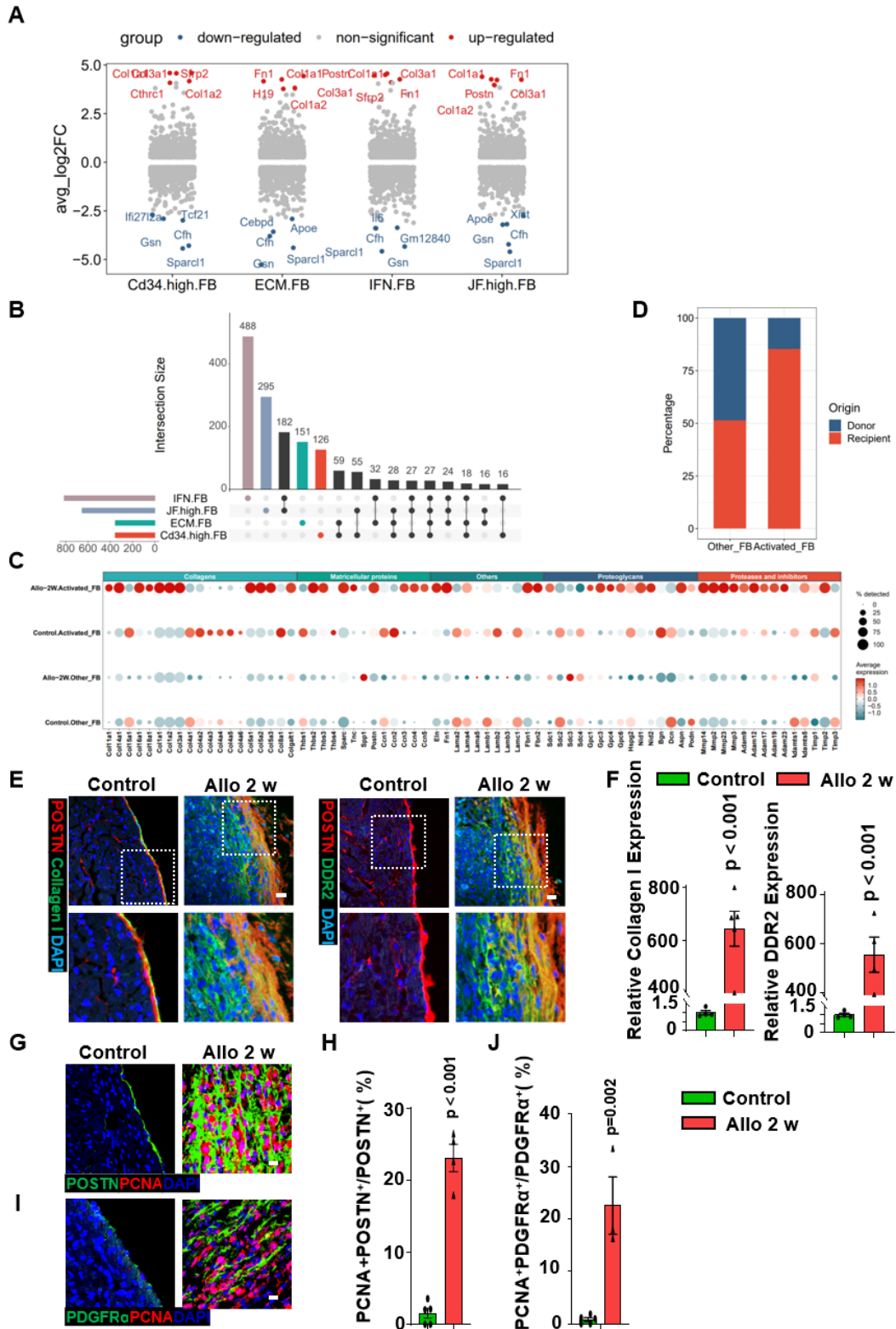


Figure S7. **A**, Volcano plot shows differentially expressed genes (DEGs) between allo-2 w and control group, genes in red represents up-regulated expression in allo-2 w group, genes in blue represents down-regulated expression in control group. **B**, Upset plot shows gene ontology (GO) terms enriched in each sub-cluster. The GO terms were enriched according to top 200 DEGs of each sub-cluster, and the terms with p value < 0.01 were used for further analysis. **C**, Dot plot

shows the expression of genes associated with ECM metabolism in two fibroblast sub-populations. **D**, Bar plot shows the proportions of donor-derived and recipient derived cells in two fibroblast sub-populations. **E**, **F**, Immuno-staining (**E**) and quantification (**F**) of Collagen I and DDR2 expression in control group and allograft group. The images within the dashed box are displayed in the second row. The area of POSTN+Collagen I+ or POSTN+DDR2+ in control group as the criterion (1.0); Number of mice in group (Collagen I): n = 4 (control), n = 5 (Allo-2 w); Number of mice in group (DDR2): n = 4; unpaired t-test. **G**, **H**, Immunostaining (**G**) and quantification (**H**) of proliferation marker (PCNA) in cardiac fibroblasts (POSTN+) of two groups; Number of mice in each group: n = 5 (control), n = 4 (Allo-2 w), unpaired t-test. **I**, **J**, Immunostaining (**I**) and quantification (**J**) of proliferation marker (PCNA) in cardiac fibroblasts (PDGFR α +) of two groups; Number of mice in each group: n = 5 (control), n = 3 (Allo-2 w), unpaired t-test. Scale bars, 100 μ m (E, G, I).

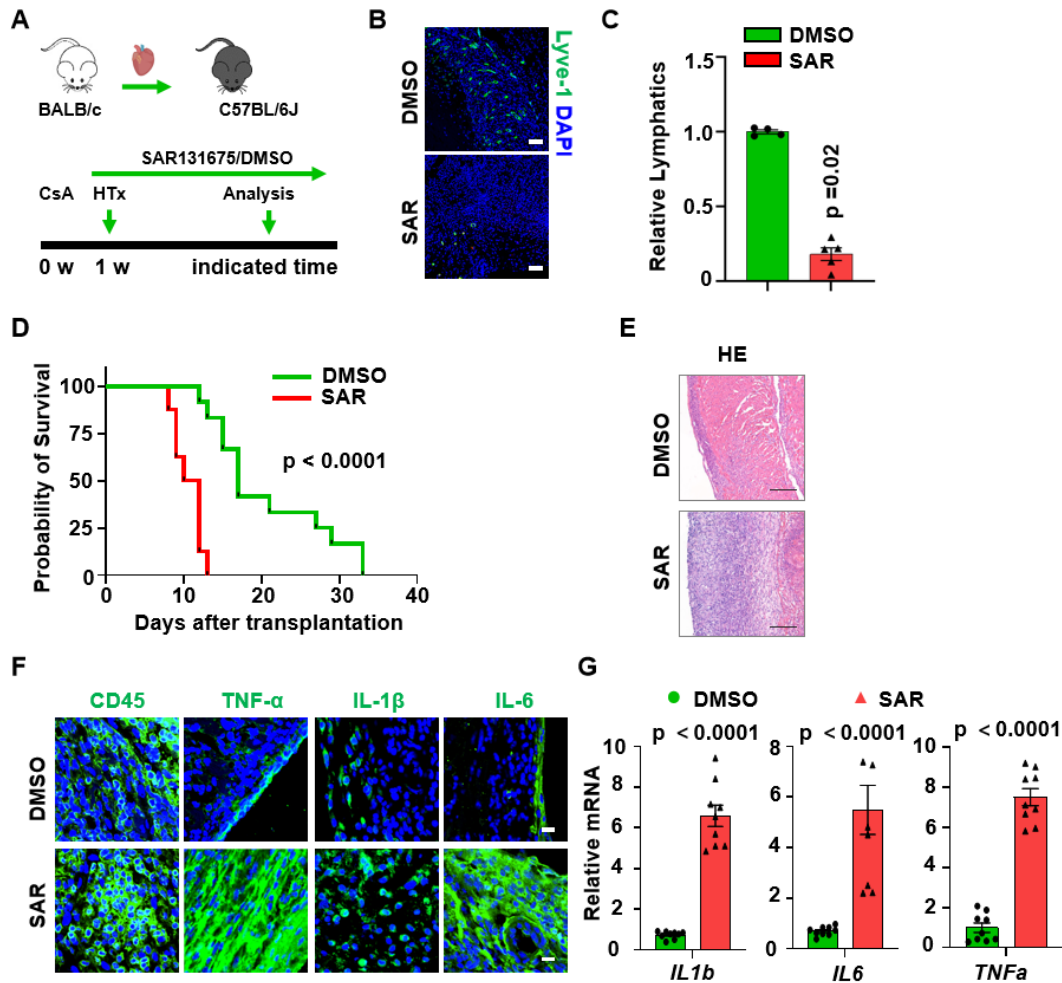


Figure S8. A, Sketch of the experimental design for lymphatic vessel growth inhibition after heart transplantation: VEGFR3 inhibitor (SAR131675) was used to suppress lymphatic vessel formation (SAR group), while DMSOI solution (5% DMSO in 30% PEG 300+5% Tween 80+ddH2O solution) was used as control group. B, C, Immunostaining (B) and quantification (C) of lymphatic vessels of cardiac allografts in two groups after 12 days of transplantation; Number of mice in each group: n = 4 (DMSO), n = 5 (SAR); unpaired t-test. D, Survival time of cardiac allografts in two groups; Number of mice in each group: n = 12 (DMSO), n = 8 (SAR); Gehan-Breslow-Wilcoxon test. E, HE staining of cardiac allograft slices in two groups. F, Immunostaining assay of CD45, TNF- α , IL-1 β and IL-6 expression in cardiac allografts from two groups. G, qPCR quantification of IL-1b, IL-6 and TNFa in cardiac allografts of two groups, mRNA level in the DMSO group as the criterion (1.0); Number of mice in each group (IL1b, TNFa): n = 9; Number of mice in group (IL6): n = 9 (DMSO), n = 7 (SAR); unpaired t-test. Scale bars, 50 μ m (B), 200 μ m (E), 25 μ m (F).

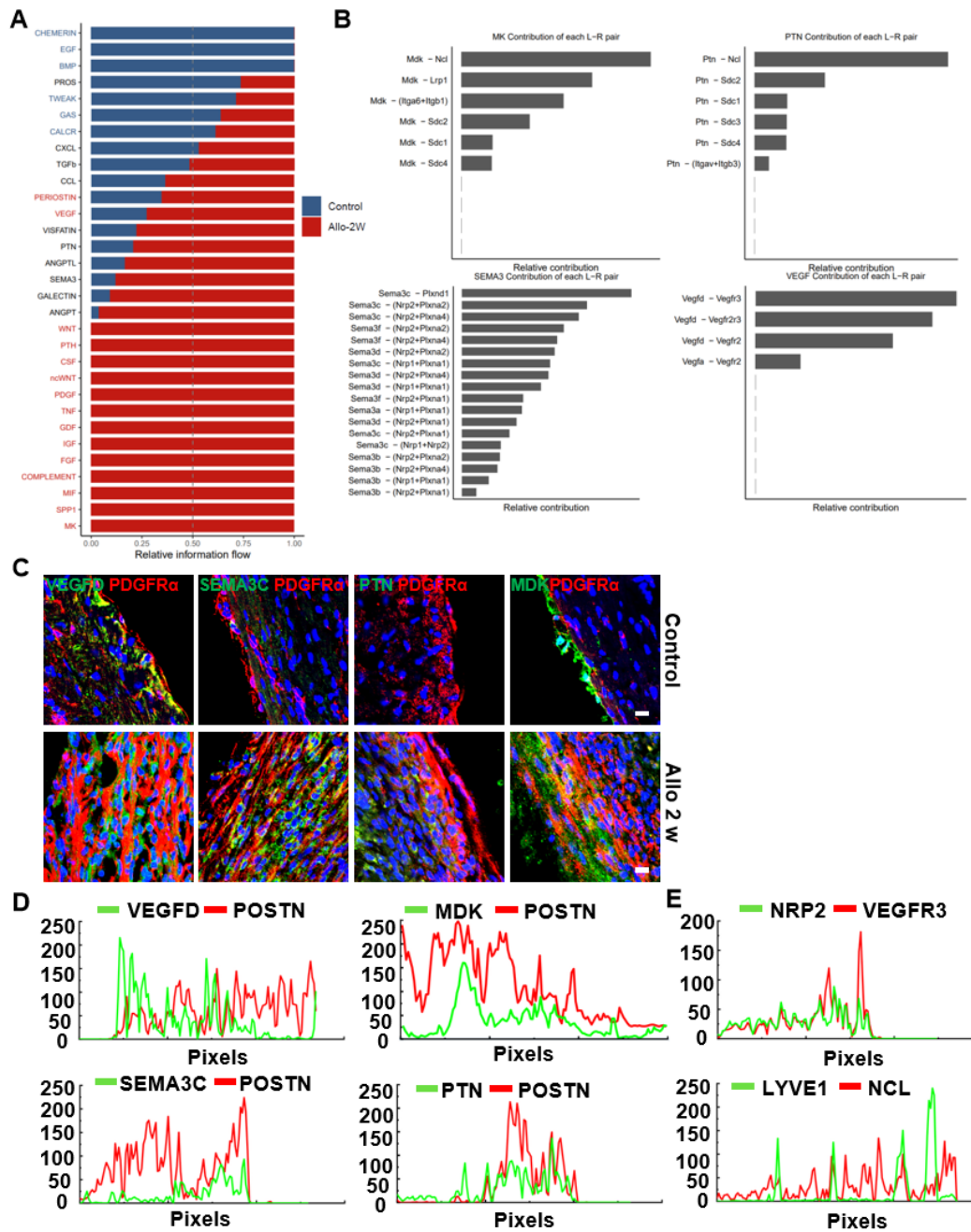


Figure S9. **A**, Bar plot shows the rank of cell-cell communication patterns in two groups. **B**, Bar plot shows the contribution of ligand-receptor pairs in selected signaling patterns. **C**, Immuno-staining of ligands in PDGFR α + fibroblasts of the cardiac allografts from two groups. **D**, Image J analysis of co-immuno staining of potential ligands (VEGFD, MDK, SEMA3C and PTN) with POSTN in allograft group. **E**, Image J analysis of co-immuno staining of NRP2 with VEGFR3, and NCL with LYVE1 in allograft group. Scale bars, 25 μ m (C).

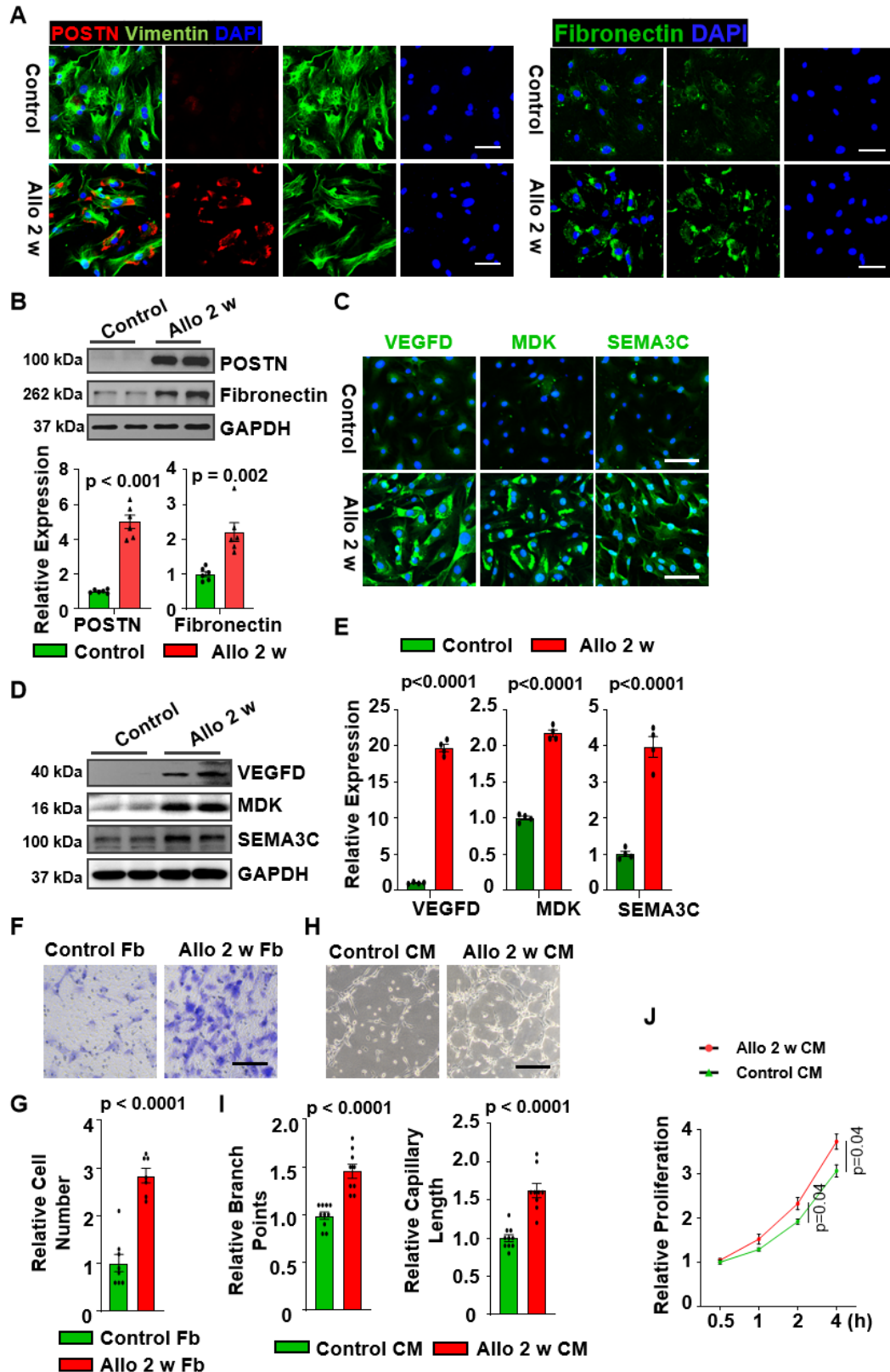


Figure S10. **A**, Immunostaining of POSTN, Vimentin (left panel) and Fibronectin (right panel) in fibroblasts isolated from normal hearts and allograft hearts after 2 weeks of transplantation. **B**, Western blotting assay (up panel) and quantification (down panel) of POSTN and Fibronectin expression in control fibroblasts and allograft fibroblasts, the protein expression in the control

group as the criterion (1.0); Number of mice in each group: n = 6, unpaired t-test. **C**, Immunostaining of ligands (VEGFD, MDK and SEMA3C) in control fibroblasts and allograft fibroblasts. **D, E**, Western blotting assay (**D**) and quantification (**E**) of VEGFD, MDK and SEMA3C in the fibroblasts from two groups, the protein expression in the control group as the criterion (1.0); Number of mice in each group: n = 4, unpaired t-test. **F, G**, Transwell assay (**F**) and quantification (**G**) of LECs co-cultured with fibroblasts from two groups, cell number in the control group as the criterion (1.0); Replication number in each group (from four heart isolation): n = 8 (Control Fb), n = 7 (Allo 2 w Fb) , unpaired t-test. **H, I**, Tube formation assay (**H**) and quantification (**I**) of LECs cultured with fibroblasts conditioned medium of two groups, the branch points and the capillary length in the control group as the criterion (1.0); Replication number in each group (from four heart isolation): n = 10 (Control CM), n = 9 (Allo 2 w CM), unpaired t-test. **J**, CCK8 assay of LECs cultured with fibroblasts conditioned medium of two groups; Replication number in each group (from four heart isolation): n = 6, unpaired t-test. Scale bars, 25 μm (A, C), 50 μm (F), and 200 μm (H).

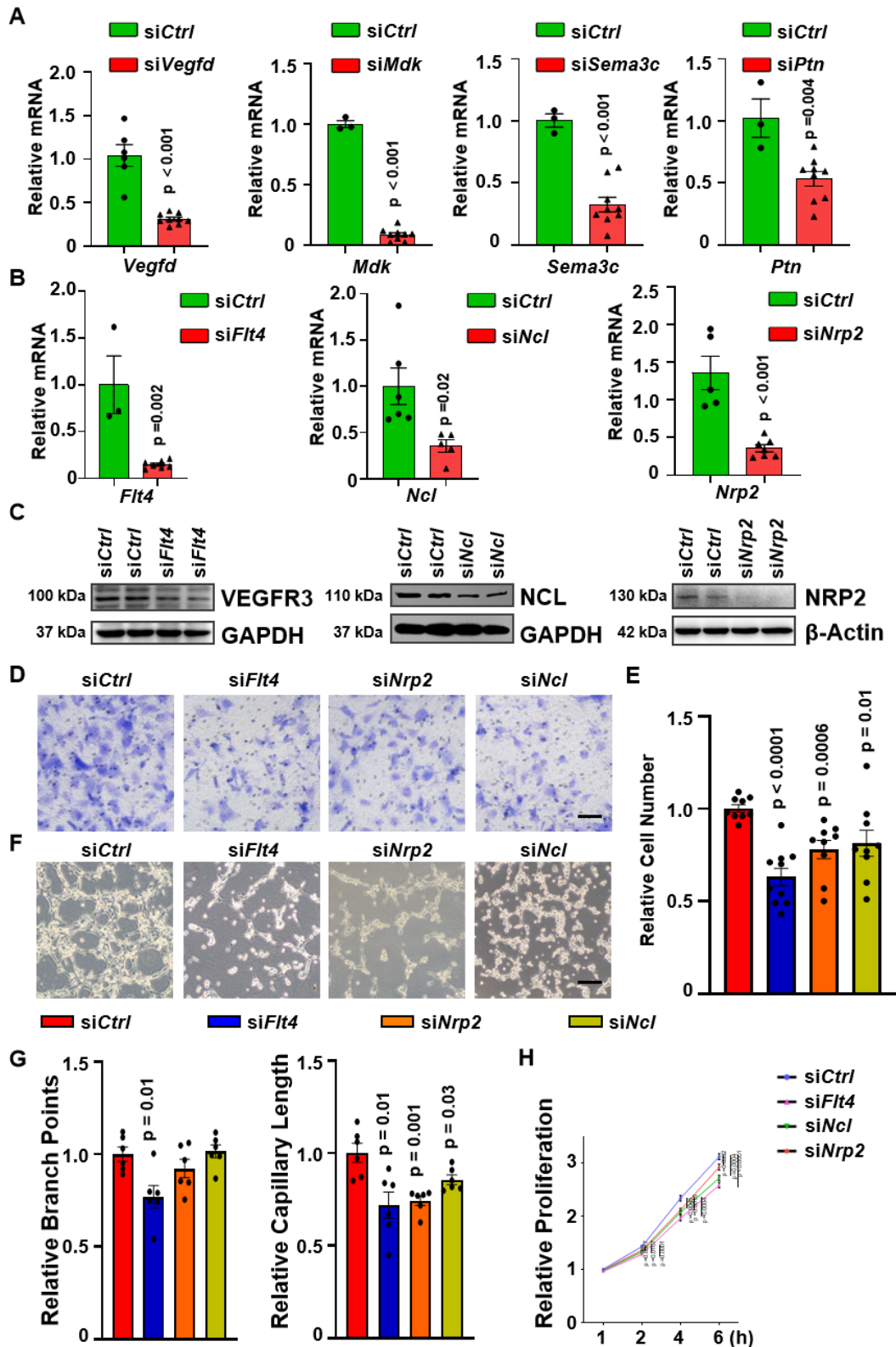


Figure S11. A, qPCR quantification of *Vegfd*, *Mdk*, *Sema3c* and *Ptn* in the transplanted heart fibroblasts treated with siRNA, mRNA expression in the control group as the criterion (1.0); Cells were isolated from four heart isolation, replication number in group (*Vegfd*): n = 6 (siCtrl), n = 9 (siVegfd); group (*Mdk/Sema3c/Ptn*): n = 3 (siCtrl), n = 9 (siMdk/siSema3c/siPtn); unpaired t-test.

B, qPCR quantification of Flt4, Ncl and Nrp2 in mouse LECs treated with siRNA, mRNA expression in the control group as the criterion (1.0); Replication number in group (Flt4): n = 3 (siCtrl), n = 7 (siFlt4); group (Ncl): n = 6 (siCtrl), n = 5 (siNcl); group (Nrp2): n = 5 (siCtrl), n = 7 (siNrp2); unpaired t-test. **C**, Western blotting assay of Flt4, Ncl and Nrp2 in mouse LECs treated with siRNA. **D**, **E**, Transwell assay (**D**) and quantification (**E**) of LECs (treated with siRNA) co-cultured with allograft fibroblasts, cell number in the control group as the criterion (1.0); Number in each group: n = 9 (siCtrl), n = 10 (siFlt4), n = 9 (siNrp2), n = 9 (siNcl), one-way ANOVA test. **F**, **G**, Tube formation assay (**F**) and quantification (**G**) of LECs (treated with siRNA) cultured with allograft fibroblasts conditioned medium, the branch points and the capillary length in the control group as the criterion (1.0); n = 6, one-way ANOVA test. **H**, CCK8 assay of LECs (treated with siRNA) cultured with allograft fibroblasts conditioned medium; n = 6, one-way ANOVA test. Scale bars, 50 μ m (D), and 200 μ m (F).

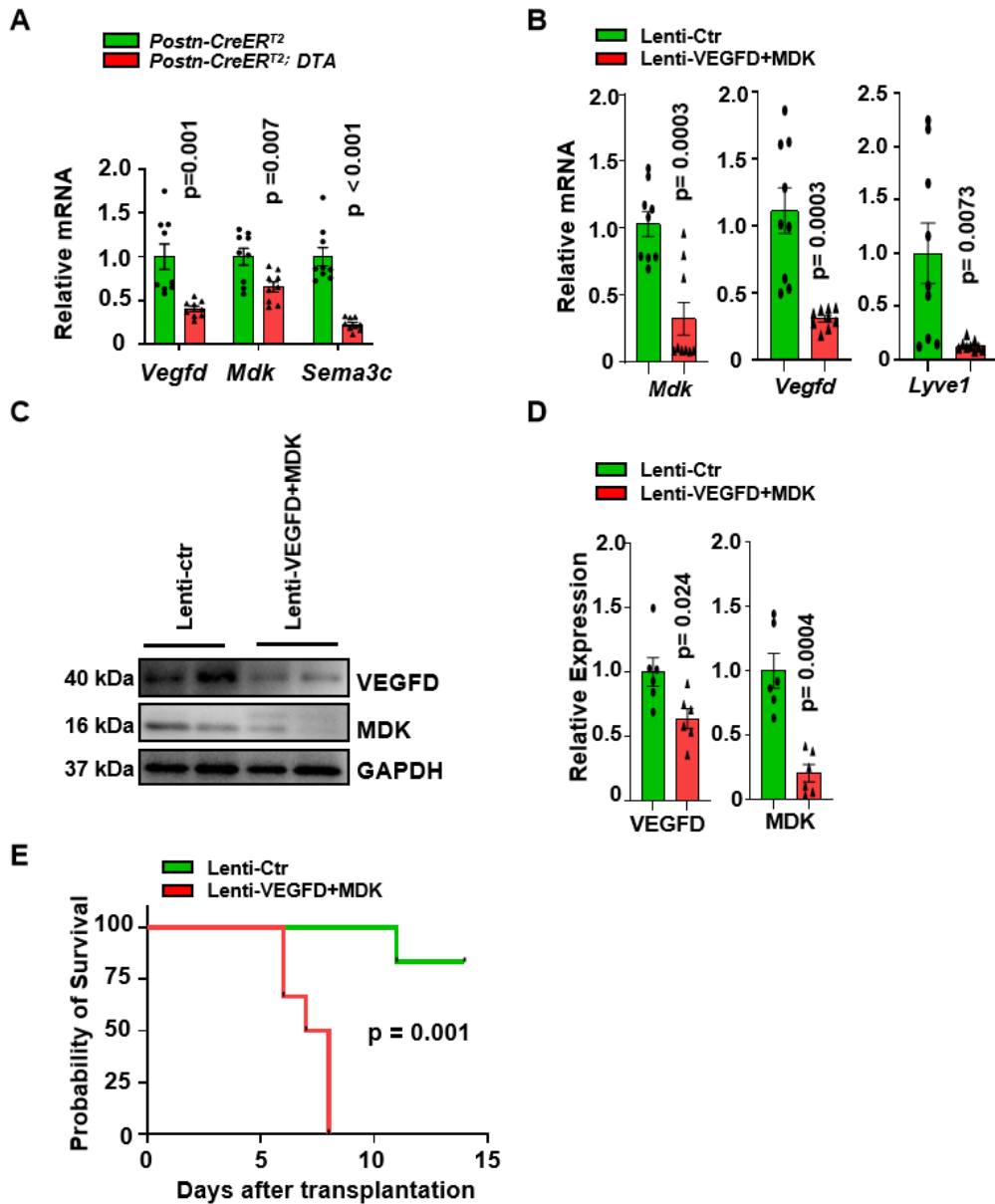


Figure S12. A, mRNA expression of secreted fibroblast ligand genes (*Vegfd*, *Mdk* and *Sema3c*) in the hearts from the POSTN⁺ cell depletion group (*Postn-CreERT2*;DTA) and the control group (*Postn-CreERT2*), mRNA expression in the control group as the criterion (1.0); Number of mice in each group: n = 9, unpaired t-test. B, qPCR quantification of *Vegfd*, *Mdk* and *Lyve1* in the cardiac allografts of the ligands-knockdown group and the control group, mRNA expression in the control group as the criterion (1.0); Number of mice in each group: n = 9, unpaired t-test. C, D, Western blotting assay and quantification of VEGFD and MDK expression in ligands-knockdown group and the control group, protein expression in the control group as the criterion (1.0); Number of mice in each group: n = 6, unpaired t-test. E, Survival time of cardiac allografts in VEGFD/MDK-knockdown group and the control group; Number of mice in each group: n = 6, Gehan-Breslow-Wilcoxon test.

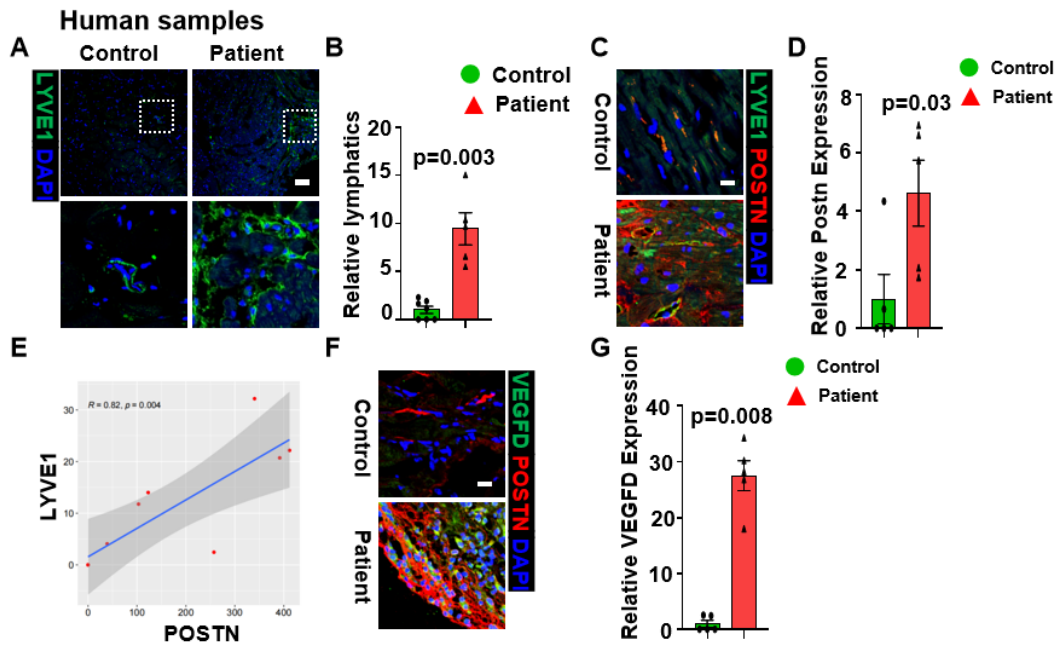


Figure S13. A, B, Immunostaining (A) and quantification (B) of cardiac lymphatic vessels in heart tissue of healthy people (control group) and the patients undergone the heart transplant operation, the lymphatic vessel density of the control group as the criterion (1.0); $n = 7$ (Control), $n = 5$ (Patient), unpaired t-test. C, D, Immunostaining (C) and quantification (D) of activated fibroblasts in two human groups, the area of POSTN+ expression in control group as the criterion (1.0); $n = 5$, unpaired t-test. E, Pearson correlation coefficient of POSTN and LYVE1 in human transplanted heart; $n = 5$. F, G, Immunostaining (F) and quantification (G) of VEGFD secreted by activated fibroblasts in two human groups, the area of POSTN+VEGFD+ expression in control group as the criterion (1.0); $n = 5$, unpaired t-test. Scale bars, 100 μm (A), 25 μm (C,F).

Figure S1

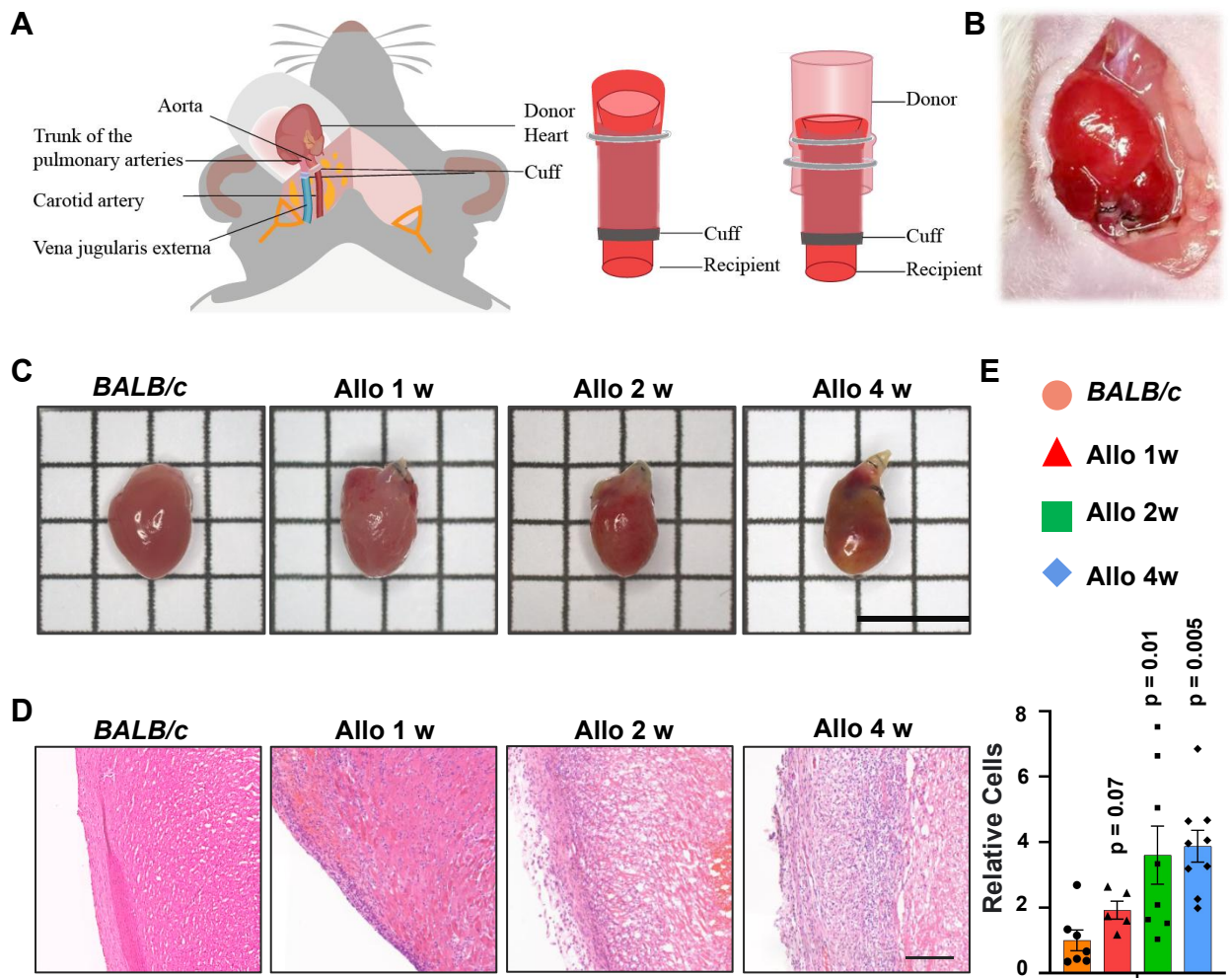


Figure S2

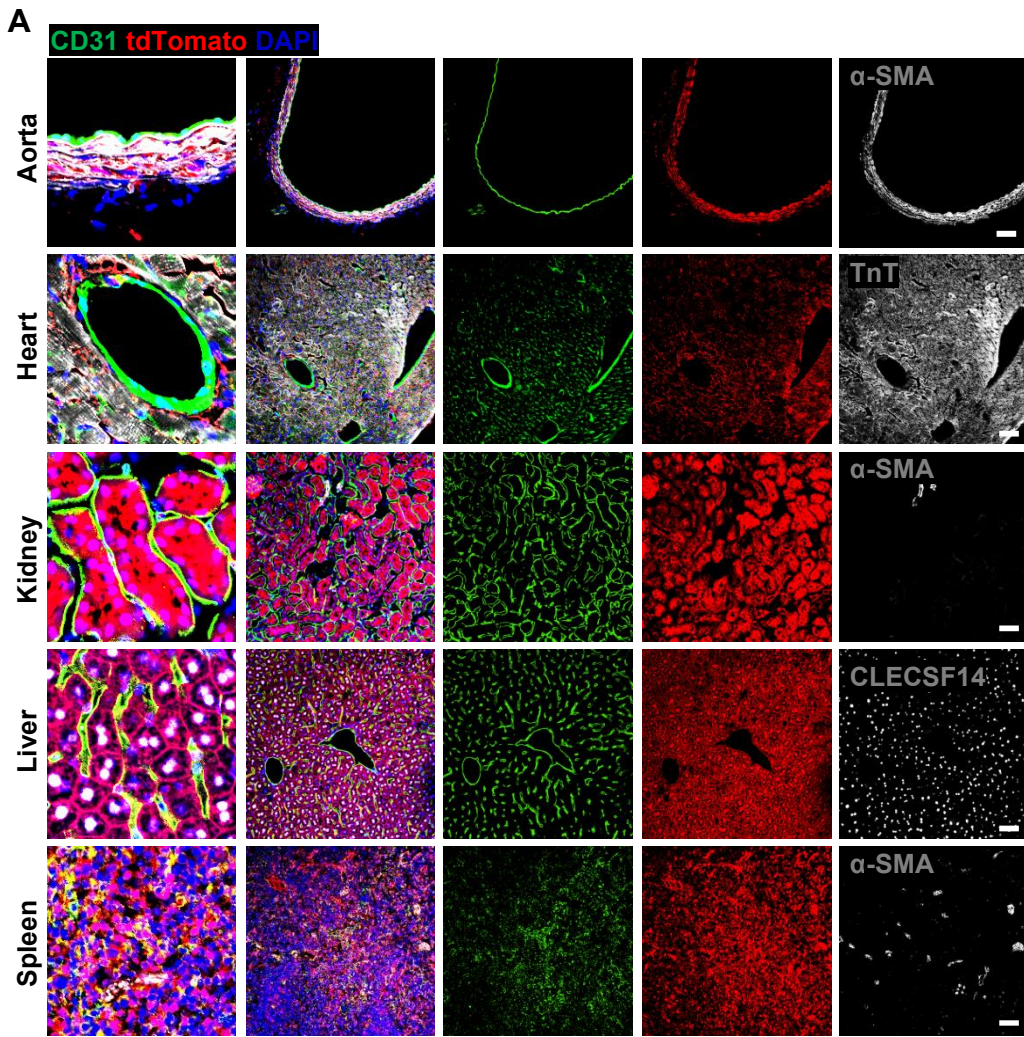


Figure S3

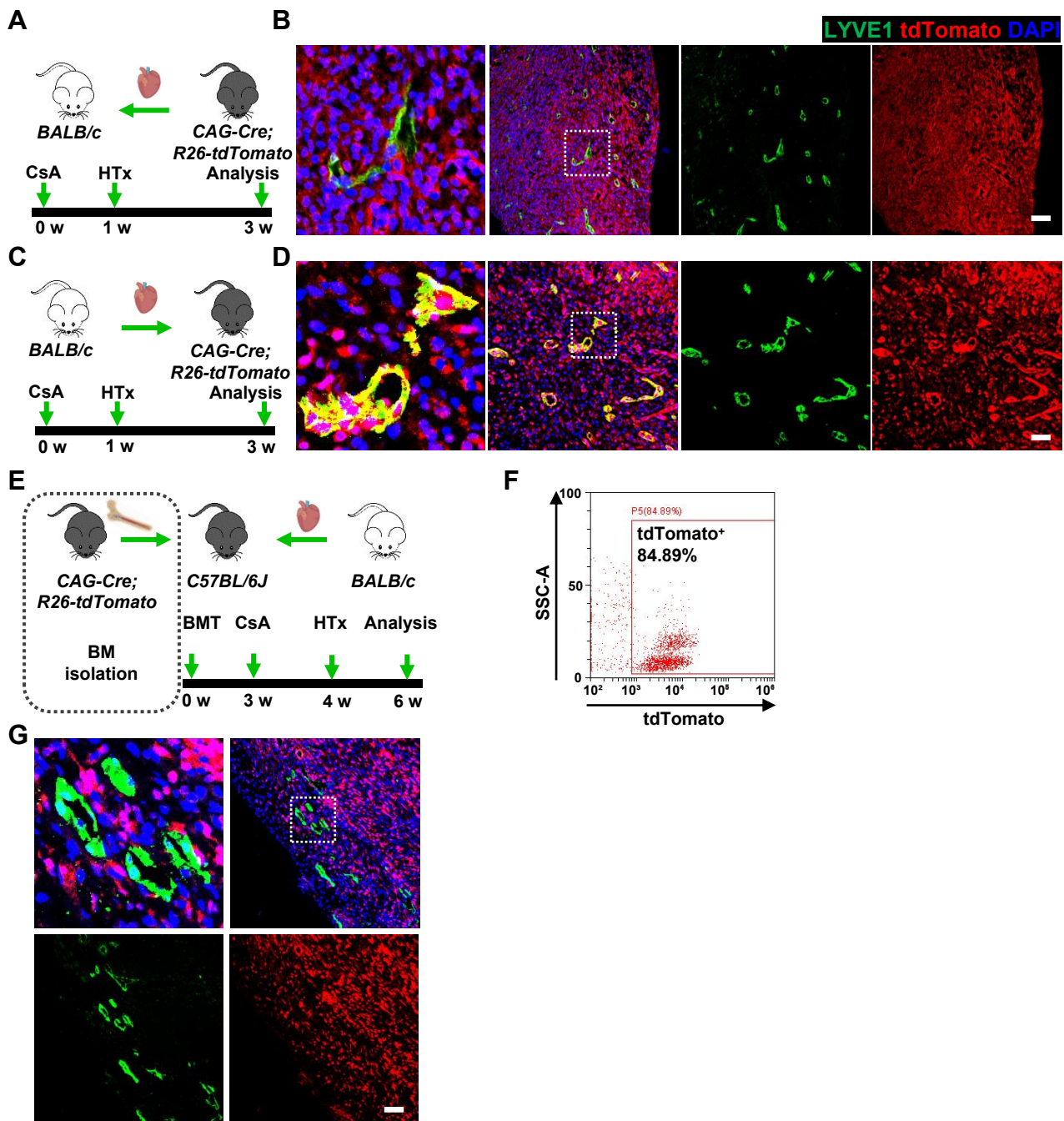
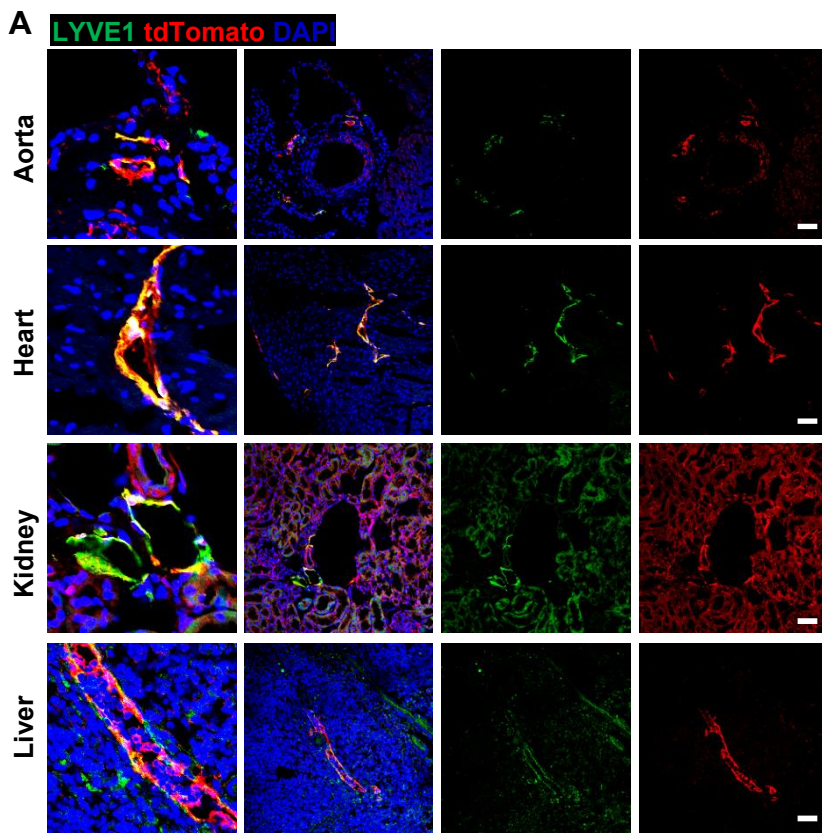


Figure S4



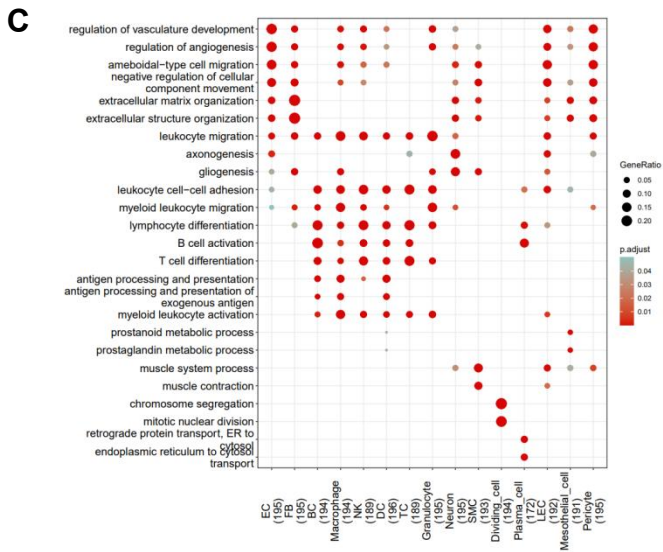
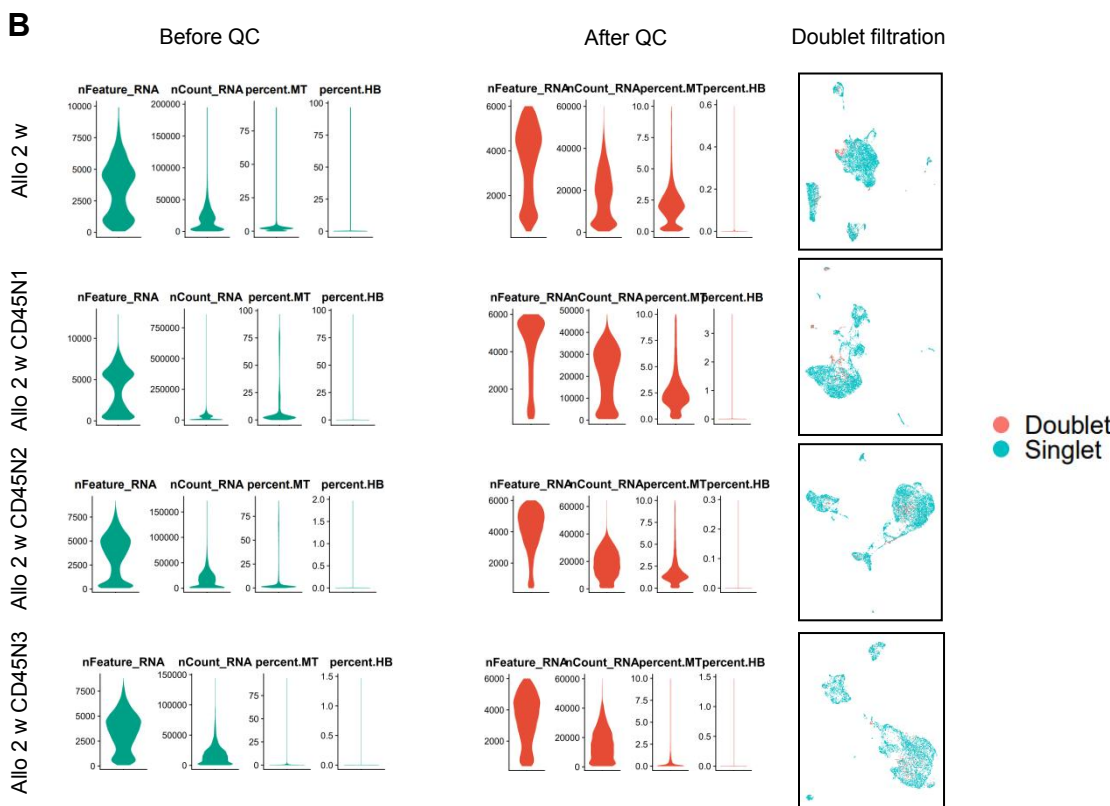
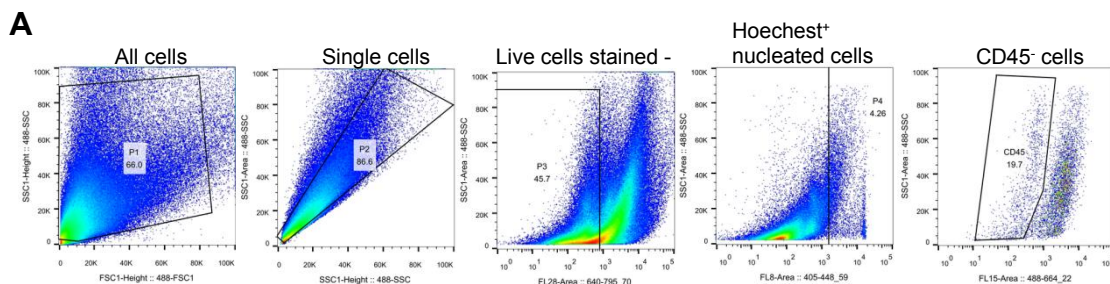
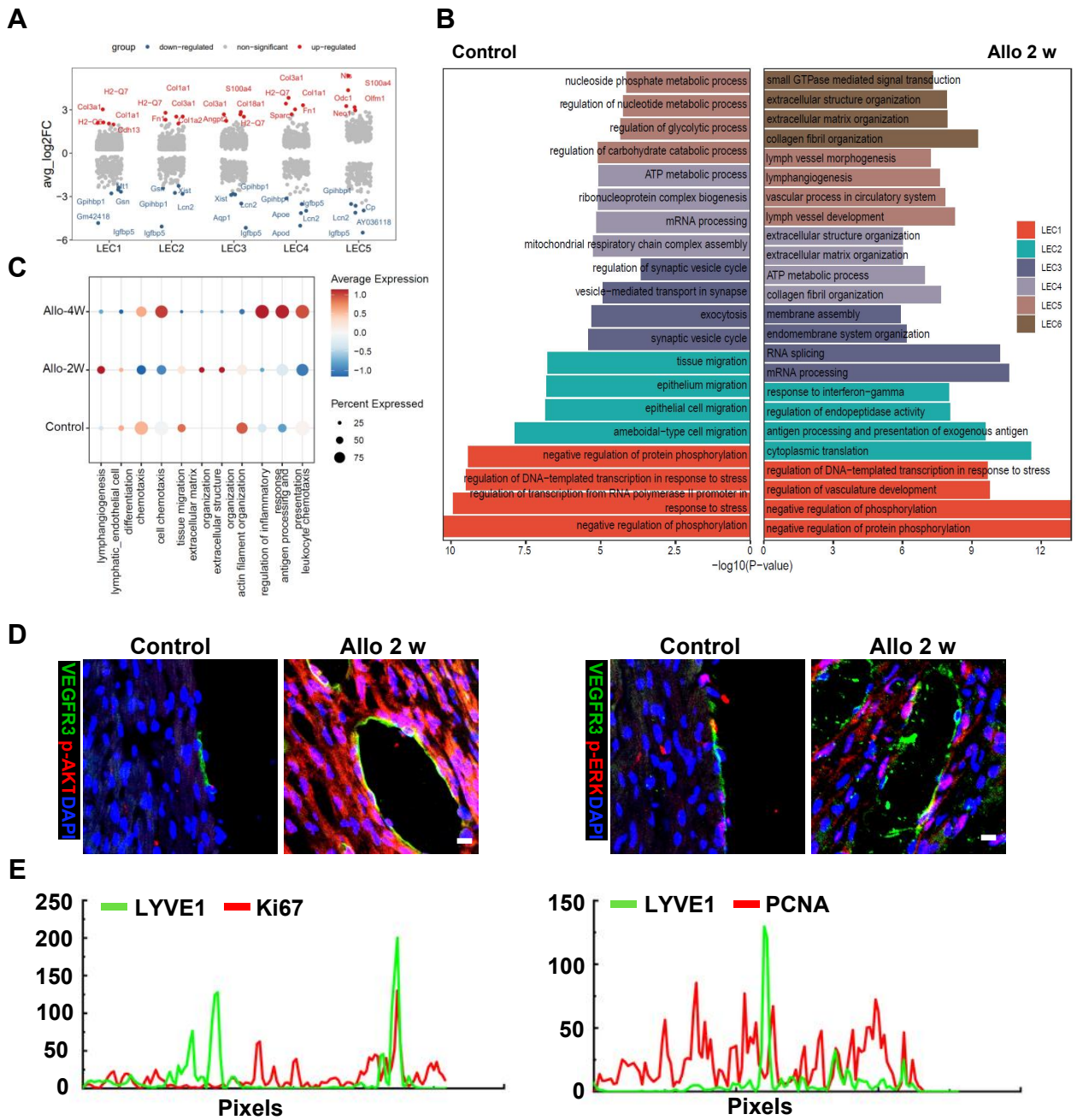


Figure S6



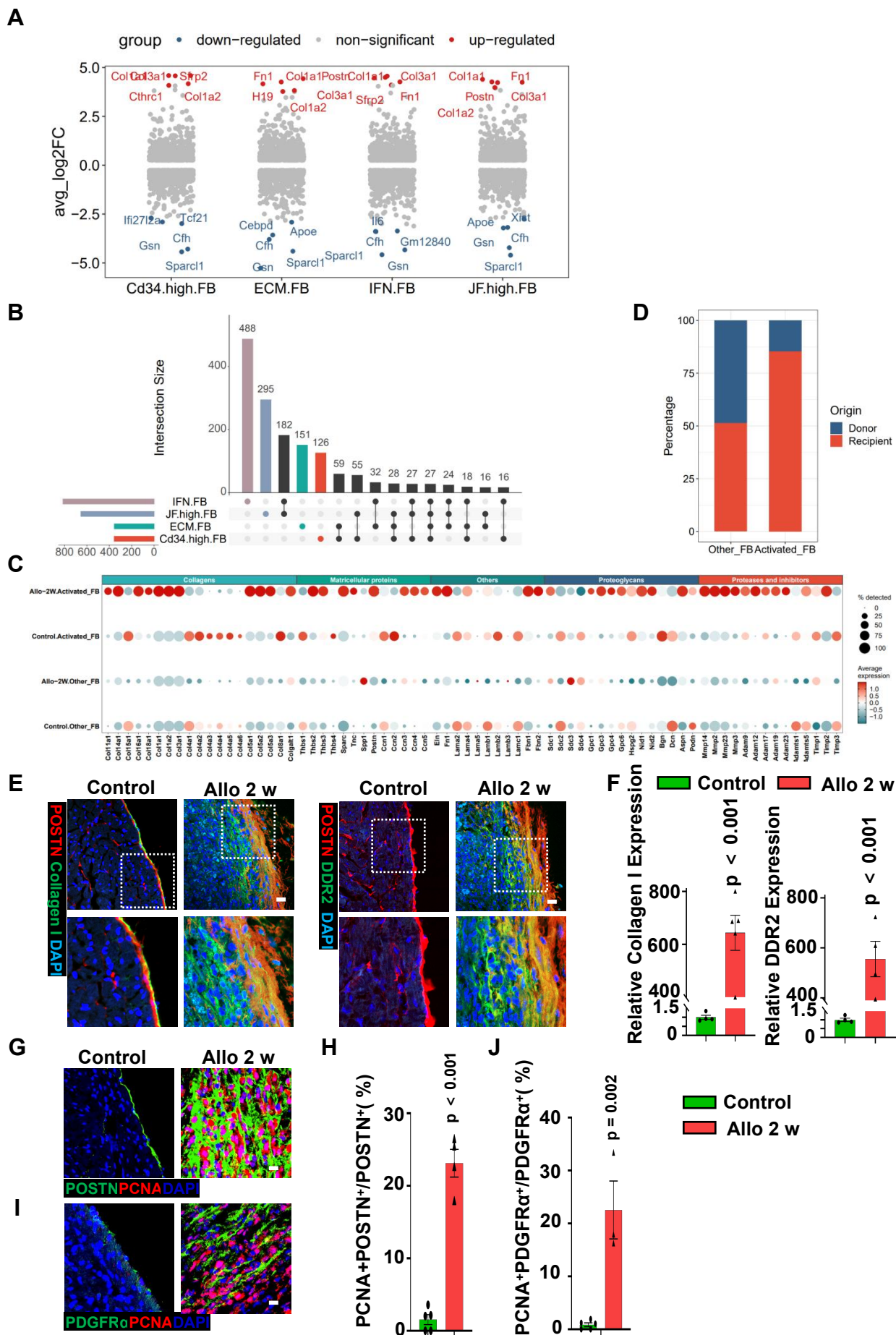


Figure S8

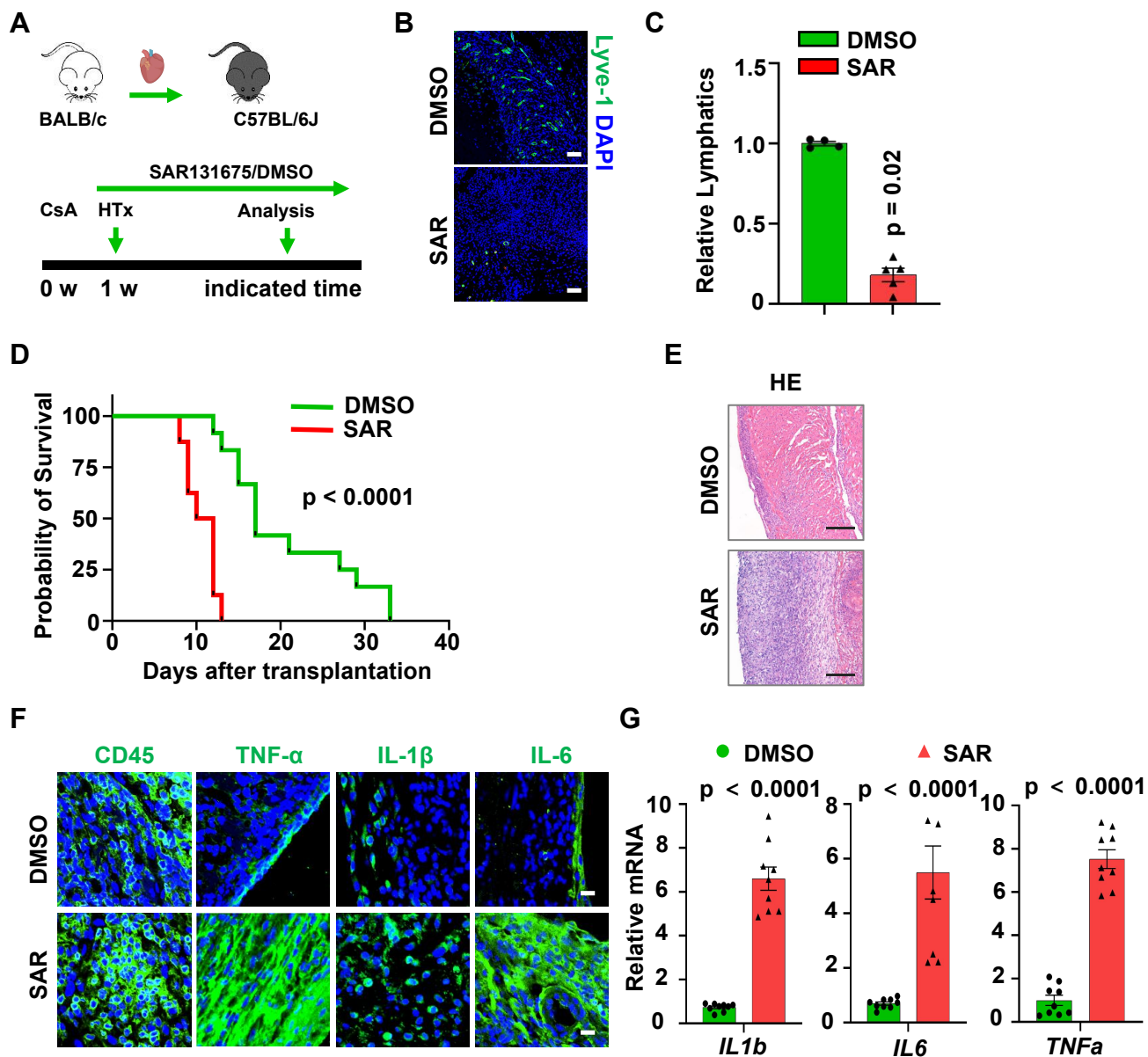


Figure S9

