

Key resources table S1

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Antibodies		
Purified anti-mouse CD16/32	Biolegend	101302
LIVE/DEAD™ Fixable Red Dead Cell Stain	Biolegend	423110
7-AAD Viability Staining Solution	Biolegend	420404
PE/Cyanine7 anti-mouse CD45	Biolegend	103114
PerCP/Cyanine5.5 anti-mouse CD45 Antibody	Biolegend	157208
APC/Cyanine7 anti-mouse CD45 Antibody	Biolegend	157618
Brilliant Viloet 605™ anti-mouse CD45 Antibody	Biolegend	103140
APC anti-mouse Ly-6G Antibody	Biolegend	127614
APC anti-mouse/human CD11b Antibody	Biolegend	101212
FITC anti-mouse CD170 (Siglec-F) Antibody	Biolegend	155504
PE anti-mouse Siglec-E Antibody	Biolegend	677103
APC anti-mouse CD182 (CXCR2) Antibody	Biolegend	149312
FITC Rat IgG2a, κ Isotype Ctrl Antibody	Biolegend	400505
PE anti-mouse Ly-6G Antibody	Biolegend	127608
PE/Cyanine7 anti-mouse/human CD11b Antibody	Biolegend	101216
FITC anti-mouse F4/80 Antibody	Biolegend	123108
PE anti-mouse NK-1.1 Antibody	Biolegend	156504
PE/Cyanine7 anti-mouse CD31 Antibody	Biolegend	102418

APC anti-mouse Ly-6C Antibody	Biolegend	128016
APC anti-mouse CD19 Antibody	Biolegend	152410
APC/Cyanine7 anti-mouse CD3 Antibody	Biolegend	100222
APC anti-mouse CD8b.2 Antibody	Biolegend	140410
Brilliant Viloet 510™ anti-mouse CD8b(Ly-3) Antibody	Biolegend	126631
PE anti-mouse CD25 Antibody	Biolegend	101903
FITC anti-mouse CD69 Antibody	Biolegend	104506
FITC anti-mouse CD62L Antibody	Biolegend	161212
PE anti-mouse CD274(B7-H, PD-L1) Antibody	Biolegend	124308
APC anti-mouse CD4 Antibody	Biolegend	100412
PE anti-mouse CD4 Antibody	Biolegend	100408
Brilliant Viloet 510™ anti-mouse CD3 Antibody	Biolegend	100234
Brilliant Viloet 421™ anti-mouse FOXP3 Antibody	Biolegend	126419
PerCP/Cyanine5.5 anti-house/mouse/rat CD278(ICOS)Antibody	Biolegend	313518
APC anti-mouse IFN- $\gamma$ Antibody	Biolegend	505810
PerCP/Cyanine5.5 anti-mouse IL-4 Antibody	Biolegend	504124
APC anti-mouse IL-17A Antibody	Biolegend	506916
Brilliant Viloet 421™ anti-mouse IL-10 Antibody	Biolegend	505022
Chemicals, peptides, and recombinant proteins		
Lipopolysaccharide	Sigma-Aldrich	L2880

Clophosome®-A-Clodronate Liposomes (Anionic)	FormuMax	F70101C-A
SB225002	Selleck	S7651
Mouse Siglec-F Antibody	R&D	MAB17061-100
Mouse IgG1 Isotype Control	R&D	MAB002
InVivoPlus anti-mouse IL-10R (CD210)	BioXCell	BP0050
InVivoPlus rat IgG1 isotype control, anti-horseradish peroxidase	BioXCell	BP0088
InVivoPlus anti-mouse Ly6G	BioXCell	BP0075-1
InVivoPlus rat IgG2a isotype control, anti-trinitrophenol	BioXCell	BP0089
InVivoMAb anti-mouse IL-2	BioXCell	BE0043-1
InVivoMAb rat IgG2a isotype control, anti-trinitrophenol	BioXCell	BE0089
Recombinant IL-33	Chamot	CM030-MP
Critical commercial assays		
Mouse CXCL1/KC Quantikine ELISA Kit	R&D	MKC00B-1
Mouse CXCL2/MIP-2 Quantikine ELISA Kit	R&D	MM200
Mouse IFN-gamma Quantikine ELISA Kit	R&D	MIF00-1
Mouse IL-10 Quantikine ELISA Kit	R&D	M1000B-1
EasySep™ Mouse T Cell I solution Kit	Stemcell	19851
Bacterial strains		
Escherichia coli	BioVector NTCC	ACCC01634

Key resources table S2

Primer list for the qPCR experiments.

Gene(s)	RT-qPCR Oligonucleotides
m-GAPDH-F	AGGTCGGTGTGAACGGATTTG
m-GAPDH-R	TGTAGACCATGTAGTTGAGGTCA
m-IL-6-F	TAGTCCTTCCTACCCCAATTTCC
m-IL-6-R	TTGGTCCTTAGCCACTCCTTC
m-IFN- $\gamma$ -F	ATGAACGCTACACACTGCATC
m-IFN- $\gamma$ -R	CCATCCTTTTGCCAGTTCCTC
m-TNF $\alpha$ -F	AAGCCTGTAGCCCACGTCGTA
m-TNF $\alpha$ -R	GGCACCAGTGTGGTTGTCTTTG
m-IL-10-F	GGTTGCCAAGCCTTATCGGA
m-IL-10-R	GGGAGAAATCGATGACAGC
m-CXCL1-F	CTGGGATTCACCTCAAGAACATC
m-CXCL1-R	CAGGGTCAAGGCAAGCCTC
m-CXCL2-F	CCAACCACCAGGCTACAGG
m-CXCL2-R	GCGTCACACTCAAGCTCTG
m-CXCL3-F	AGTGTGGCTATGACTTCCG
m-CXCL3-R	GAATTCACCTCAAGAACATCCA
m-CXCL5-F	TCCAGCTCGCCATTCATGC
m-CXCL5-R	TTGCGGCTATGACTGAGGAAG
m-CXCL8-F	TAAGTTCTTTAGCACTCCTTGG
m-CXCL8-R	TTCCTGATTTCTGCAGCTC

Figure S1 Indicators in the PICS mice model.

The PICS mice model was characterized by various indicators on the eighth day after cecal ligation and perforation (CLP).

**(A-B)** Proportion of macrophages and neutrophils in the spleens of PICS mice measured by flow cytometry.

**(C-F)** Proportion of neutrophils in the bone marrow and peripheral blood, and proportion of CD4<sup>+</sup> and CD8<sup>+</sup> T lymphocytes in the spleens of PICS mice assessed by flow cytometry.

**(G-I)** Fluorescence intensity of PD-L1<sup>+</sup> CD4<sup>+</sup> T lymphocyte, PD-L1<sup>+</sup> CD8<sup>+</sup> T lymphocyte, and FoxP3<sup>+</sup> CD4<sup>+</sup> T lymphocyte in the spleens, as well as proportion of FoxP3<sup>+</sup> CD4<sup>+</sup> T lymphocytes, determined by flow cytometry.

**(J)** Expression of IL-10 in splenic CD45<sup>+</sup> immune cells of PICS mice detected by flow cytometry.

**(K)** Percentage calculated by subtracting the preoperative weight of mice from the weight at day 8 after CLP or sham surgery, then dividing by the preoperative weight of mice.

All indicators of the PICS group compared with the SHAM group and statistically analyzed by Student's *t* test. Data are presented as mean ± SEM. *n* = 4-10, \**P* < 0.05, \*\**P* < 0.01, \*\*\**P* < 0.001, \*\*\*\**P* < 0.0001.

Figure S2 Proportions of immune cells in the spleens.

**(A)** Flow cytometry used to detect the proportion of lymphocytes, monocytes, and granulocytes in the spleens of the mice.

**(B)** Flow cytometry used to detect the proportion of CD11b<sup>+</sup> immune cells in the spleen, peripheral blood, and bone marrow of the mice.

**(C-J)** Flow cytometry used to detect the proportion of B cells, NK cells, naive CD4<sup>+</sup>, CD8<sup>+</sup> T lymphocytes, Th1, Th2, Th17, and immature macrophages in the spleens of the mice. The results in the PICS and PICS+LPS 6 h groups were compared with the SHAM and SHAM+LPS 6 h groups, respectively, by one-way ANOVA followed by Dunnett's post hoc test. Data are presented as mean

± SEM. n = 3-10, *ns*  $P > 0.05$ , \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$ , \*\*\*\* $P < 0.0001$ .

Figure S3 Function of Treg cells in the spleen of PICS mice after secondary challenge.

**(A-B)** Proportions of Treg cells, ICOS<sup>+</sup> Treg, and fluorescence intensity of ICOS<sup>+</sup> Treg in the spleens were examined by flow cytometry.

**(C)** Efficiency of depleted Treg cells in the spleen of PICS mice.

**(D-F)** After Treg cells depletion, proportion of CD45<sup>+</sup> immune cells and expression of IL-10, IL-6, IFN- $\gamma$ , and TNF- $\alpha$  in the spleens of PICS+LPS 6 h and PICS+Anti-IL-10R+LPS 6 h mice were examined by flow cytometry and enzyme-linked immunosorbent assay.

**(G-H)** Proportion of CD4<sup>+</sup> and CD8<sup>+</sup> T lymphocytes in IL-10<sup>+</sup> CD45<sup>+</sup> immune cells and fluorescence intensity of TGF $\beta$ <sup>+</sup> CD45<sup>+</sup> immune cells were detected by flow cytometry.

One-way ANOVA followed by Dunnett's post hoc test was applied in **(A-B, G-H)**. Student's *t* test was applied in **(D-F)**. Data are presented as mean ± SEM. n = 3-7, *ns*  $P > 0.05$ , \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$ .

Figure S4 Expression of IL-10 in immune cells of the spleen.

**(A)** Expression of IL-10 in splenic neutrophils detected by flow cytometry.

**(B)** Relative mRNA expression of IL-6, IFN- $\gamma$ , and TNF- $\alpha$  in splenic neutrophils of PICS mice and PICS+LPS 6 h mice examined by qPCR.

**(C)** Efficiency of depleting neutrophils in PICS mice.

**(D)** Apparent changes observed in the spleen after depleting neutrophils in PICS mice.

**(E-G)** Expression of IL-10 in NK cells, CD3<sup>+</sup>, and CD4<sup>+</sup> T lymphocytes of the spleens examined by flow cytometry.

One-way ANOVA followed by Dunnett's post hoc test was applied in **(E-G)**. Student's *t* test was applied in **(B)**. Data are presented as mean ± SEM. n = 3-6, *ns*  $P > 0.05$ , \* $P < 0.05$ , \*\* $P < 0.01$ .

Figure S5 Effects of depleting neutrophils on the spleen.

**(A-E)** Expression of IL-10 in the spleen and proportions of CD45<sup>+</sup> immune cells, CD3<sup>+</sup>, CD4<sup>+</sup>, and CD8<sup>+</sup> T lymphocytes after neutrophils depletion.

**(F-H)** Splenic neutrophils and T lymphocytes from PICS mice were sorted by flow cytometry. Neutrophils were stimulated with LPS (1 µg/ml) in vitro for 6 h, and the supernatants were co-cultured with the T lymphocytes pre-incubated with Isotype or Anti-IL-10R for 6 h. Subsequently, T lymphocytes were stimulated with LPS for 6 h, and their function and activity were detected by flow cytometry. The relative mRNA expression of IFN-γ in T lymphocytes was examined by qPCR.

Student's *t* test was applied in **(A-H)**. Data are presented as mean ± SEM. n = 3-7, *ns* *P* > 0.05, \**P* < 0.05, \*\**P* < 0.01, \*\*\**P* < 0.001, \*\*\*\**P* < 0.0001.

Figure S6 Impact of the macrophages on T lymphocytes in the spleen.

**(A-B)** Proportion of splenic macrophages and expression of IL-10 in the splenic macrophages after secondary challenge, examined by flow cytometry.

**(C)** Relative mRNA expression of IL-10 in splenic macrophages detected by qPCR.

**(D)** Efficiency of macrophage depletion in PICS mice.

**(E-G)** After macrophage depletion, proportion of CD45<sup>+</sup> immune cells in PICS+LPS 6 h mice and PICS+Liposomes+LPS 6 h mice, and the activity of CD4<sup>+</sup> and CD8<sup>+</sup> T lymphocytes in the spleens were detected by flow cytometry.

One-way ANOVA followed by Dunnett's post hoc test was applied in **(A-C, F-G)**. Student's *t* test was applied in **(E)**. Data are presented as mean ± SEM. n = 3-6, *ns* *P* > 0.05, \**P* < 0.05, \*\**P* < 0.01.

Figure S7 Expression of CXCLs in the spleen, T lymphocytes and macrophages.

**(A-E)** Expression of CXCL1, CXCL2, CXCL3, CXCL5, and CXCL8 in splenic T lymphocytes of SHAM, PICS, and PICS+LPS 6 h mice.

**(F-I)** mRNA expression of CXCL1, CXCL2, CXCL3, and CXCL5 in the spleens

of SHAM, PICS, and PICS+LPS 6 h mice, detected by qPCR.

**(J)** mRNA expression of CXCL2 in splenic macrophages of SHAM, PICS, and PICS+LPS 6 h mice, detected by qPCR.

One-way ANOVA followed by Dunnett's post hoc test was applied in **(A-J)**. Data are presented as mean  $\pm$  SEM.  $n = 3-6$ ,  $ns P > 0.05$ ,  $*P < 0.05$ ,  $**P < 0.01$ ,  $****P < 0.0001$

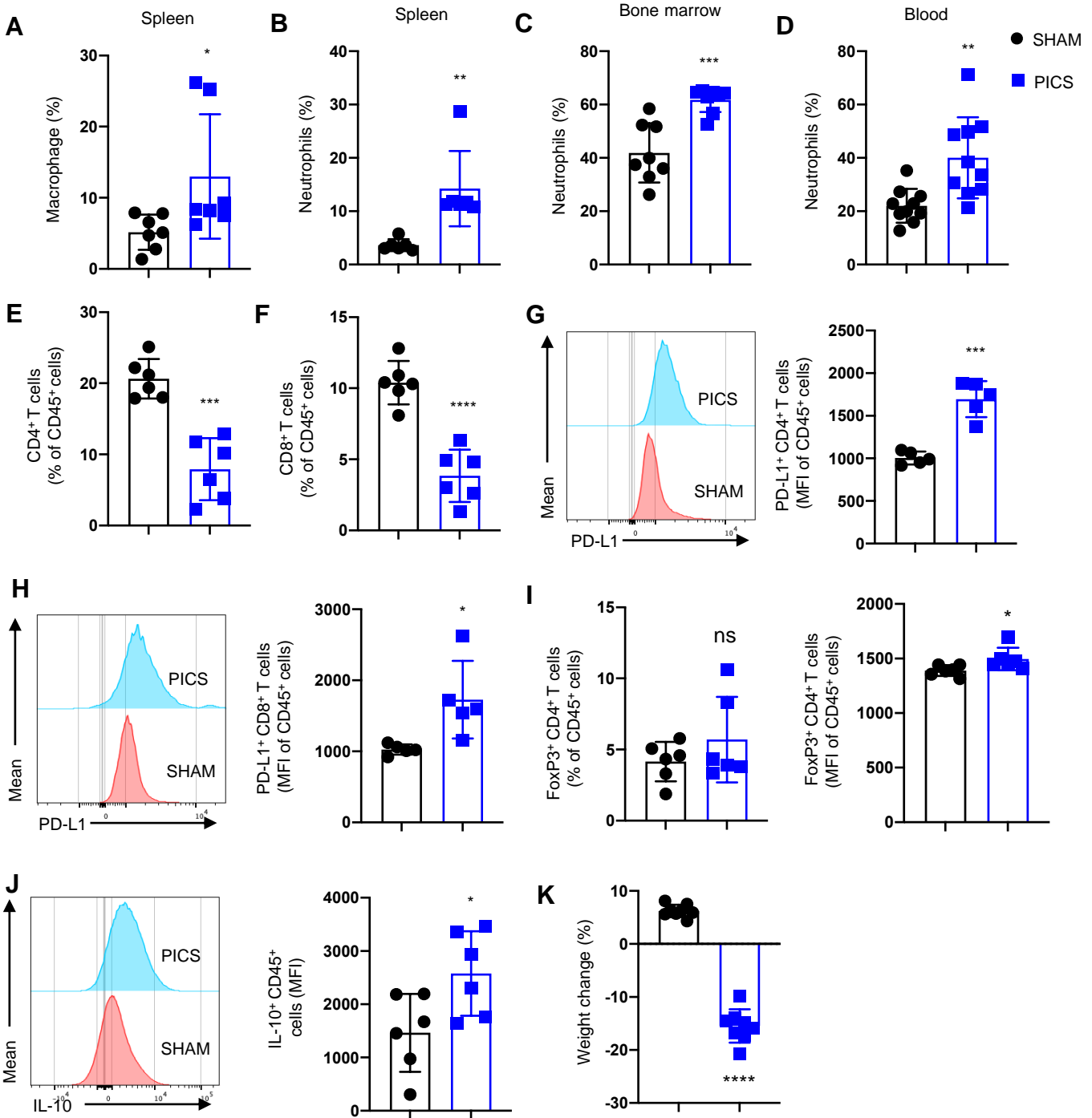
Figure S8 Effects on immune cells with expanded eosinophils after secondary challenge.

**(A)** Proportion of Siglec-F<sup>-</sup> neutrophils after secondary challenge detected by flow cytometry.

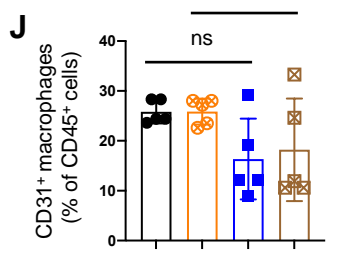
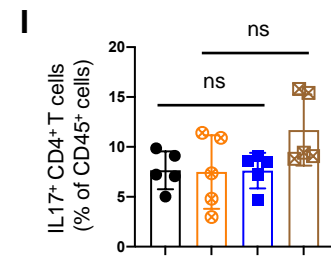
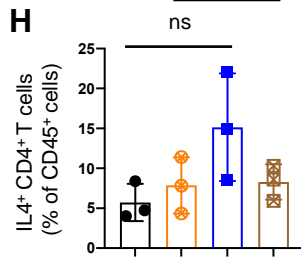
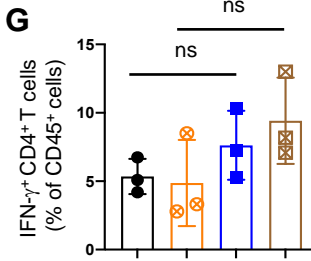
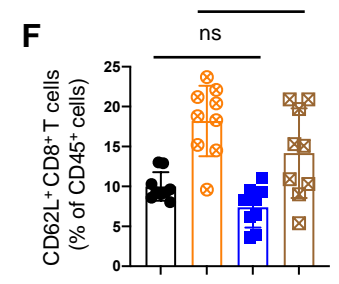
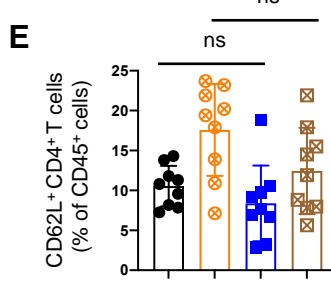
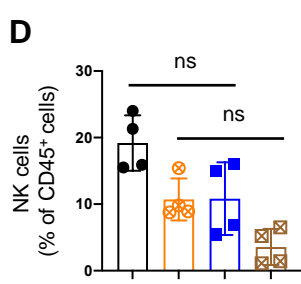
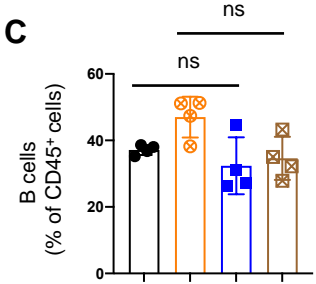
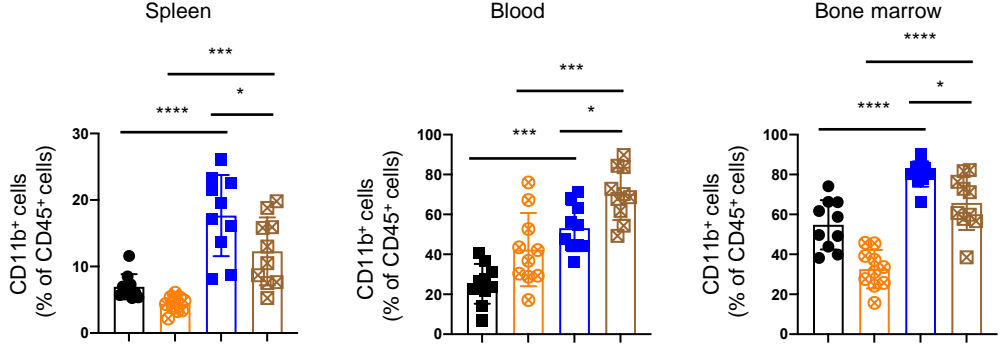
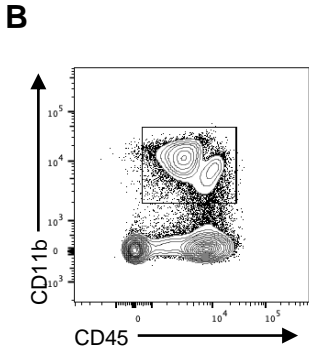
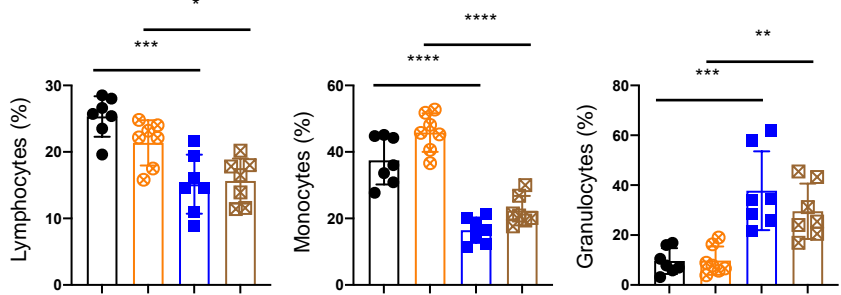
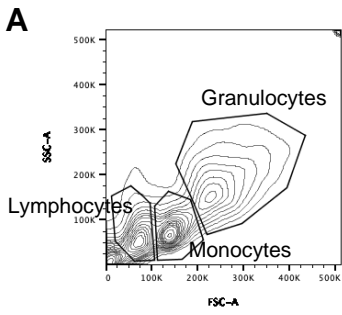
**(B-H)** After intraperitoneally injecting IL-33 to expand eosinophils of in PICS mice, the proportions of Siglec-F<sup>-</sup> neutrophils, neutrophils, CD11b<sup>+</sup> cells, CD45<sup>+</sup> immune cells, CD3<sup>+</sup>, CD4<sup>+</sup>, and CD8<sup>+</sup> T lymphocytes in PICS+LPS 6 h mice and in PICS+IL-33+LPS 6 h mice were examined by flow cytometry.

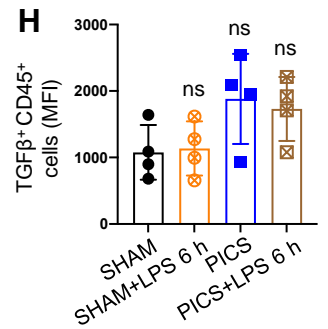
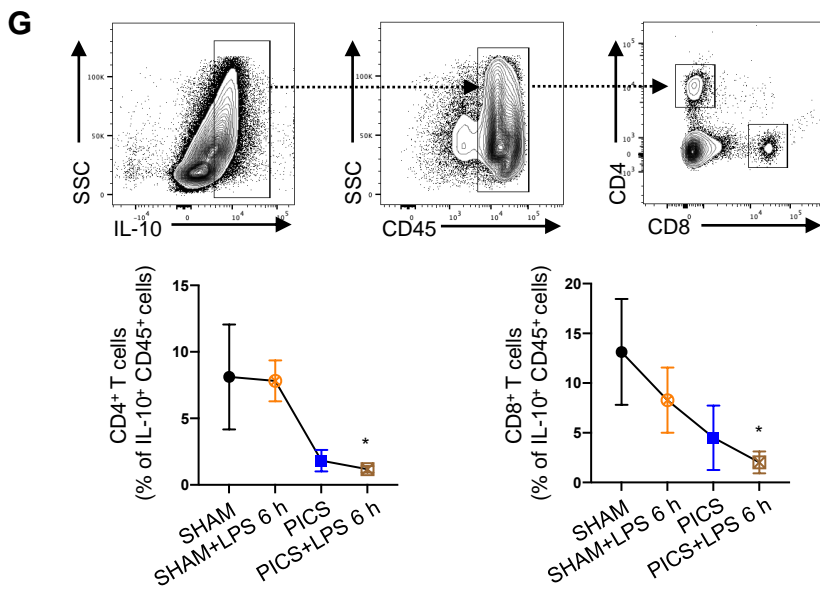
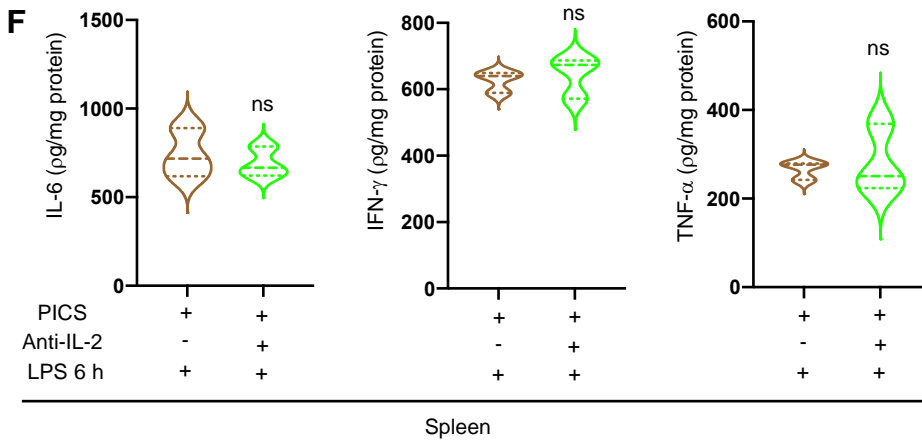
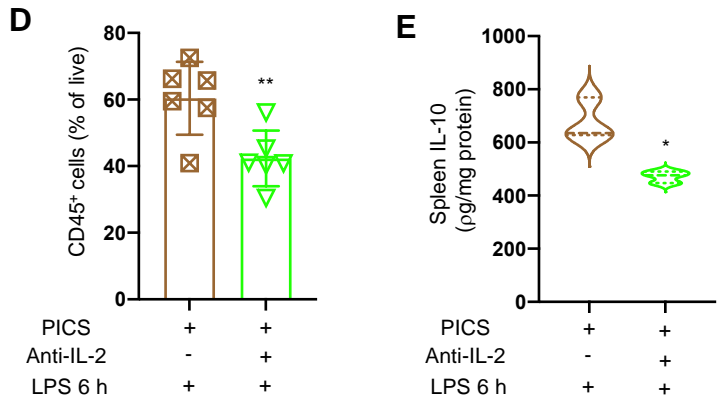
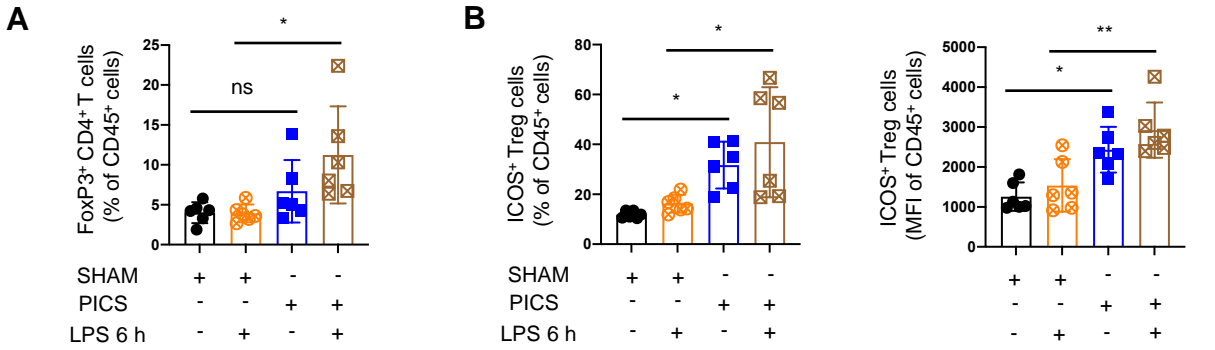
One-way ANOVA followed by Dunnett's post hoc test was applied in **(A)**. Student's *t* test was applied in **(B-H)**. Data are presented as mean  $\pm$  SEM.  $n = 4-6$ ,  $ns P > 0.05$ ,  $*P < 0.05$ ,  $**P < 0.01$ ,  $***P < 0.001$ .

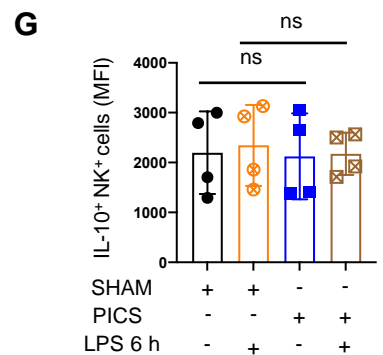
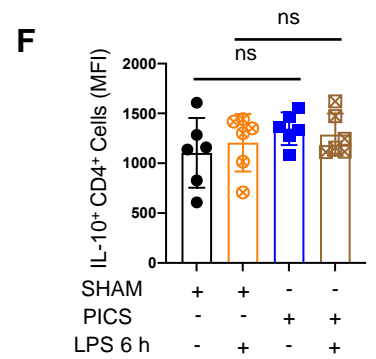
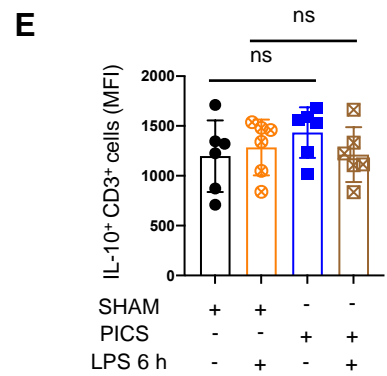
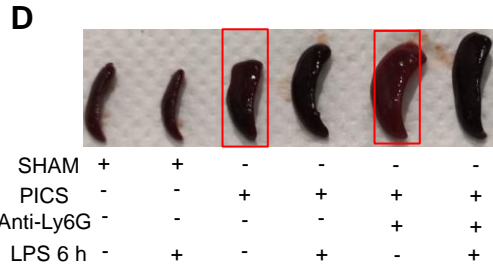
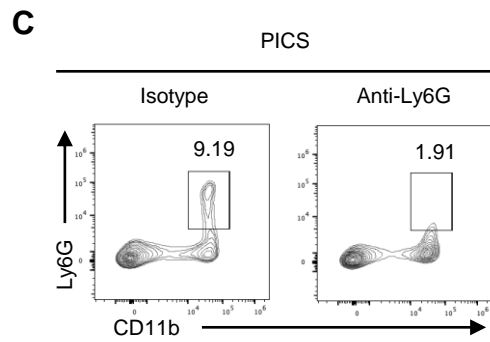
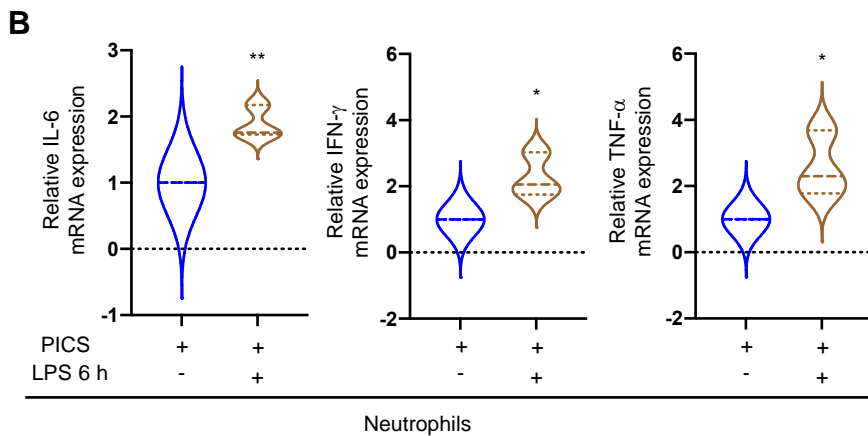
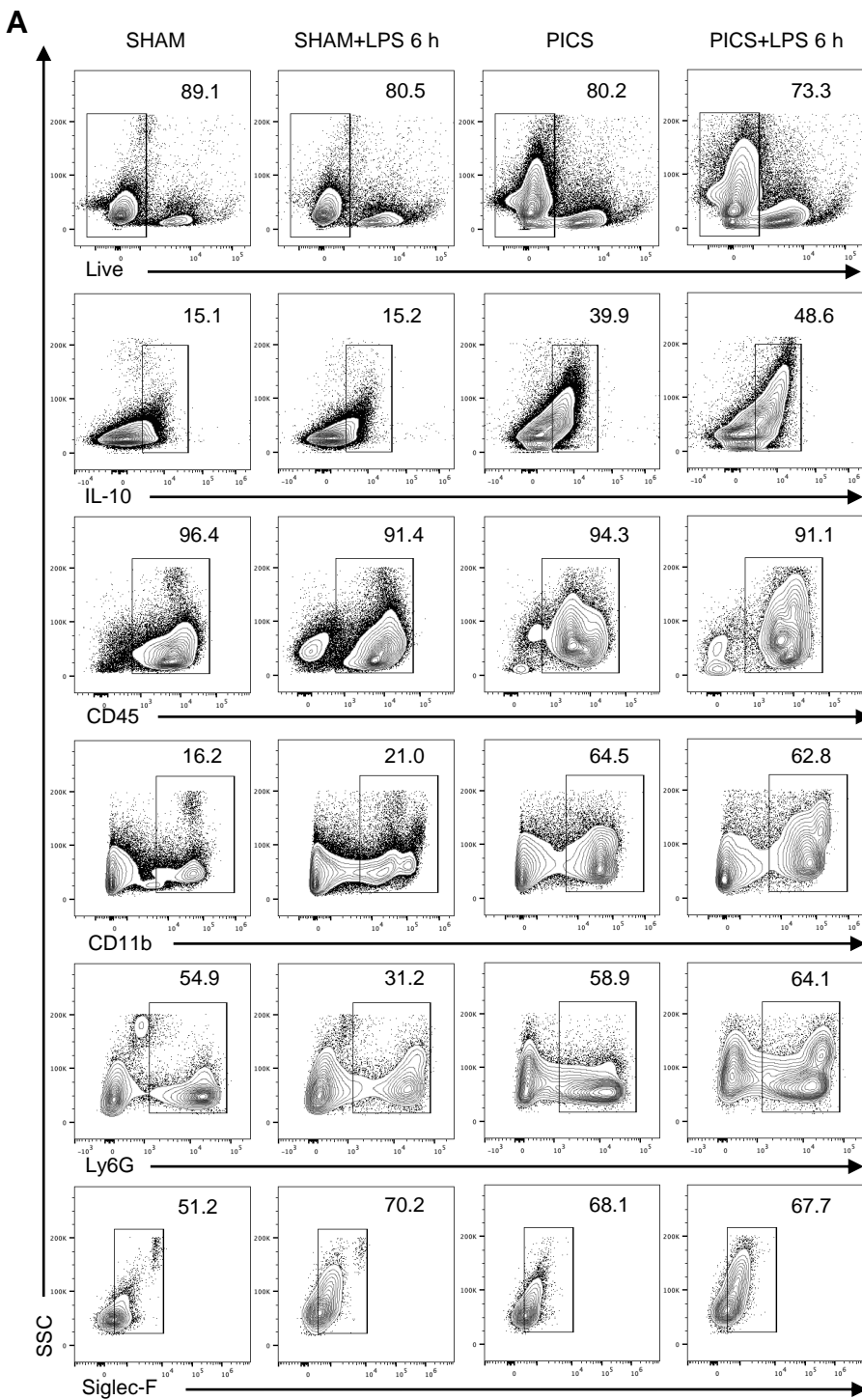


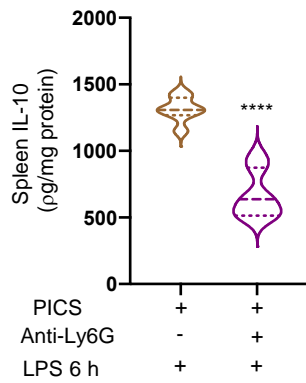
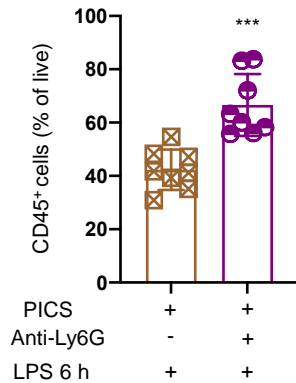
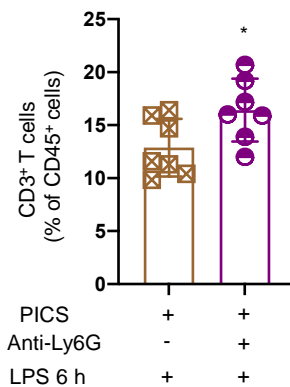
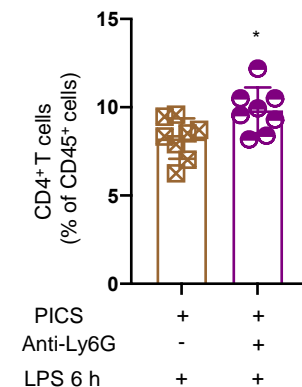
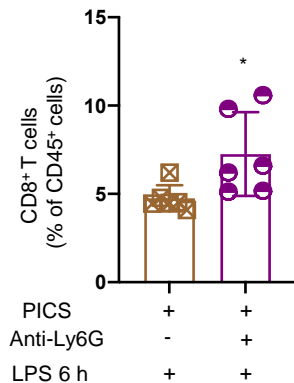
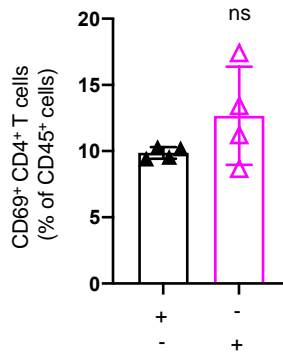
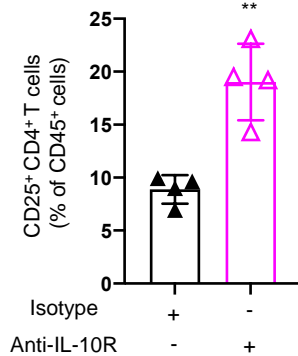
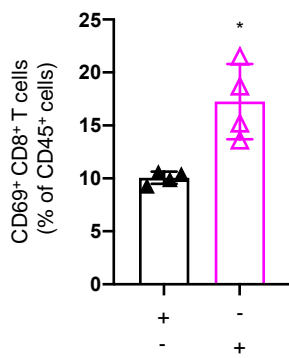
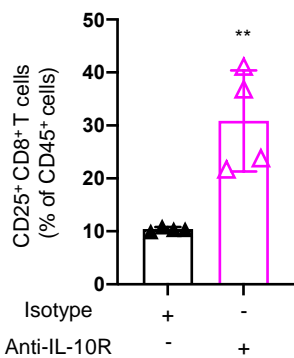
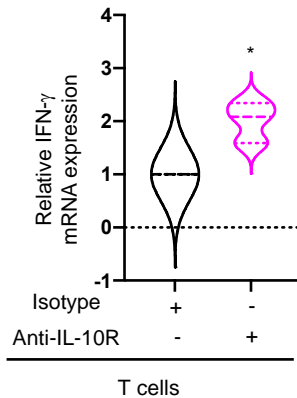


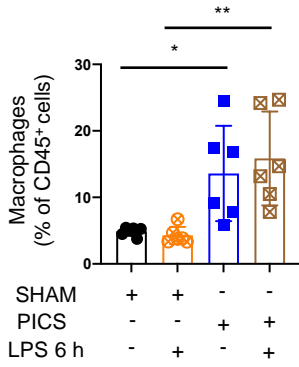
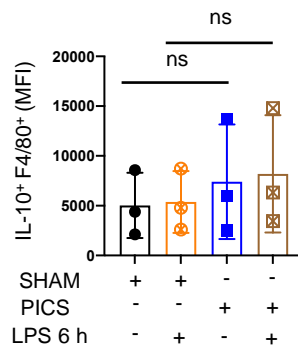
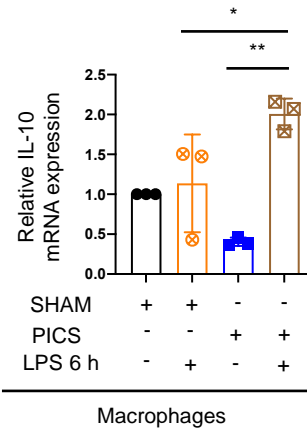
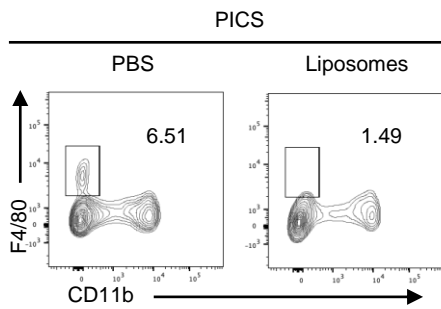
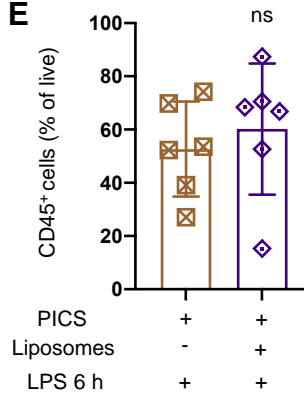
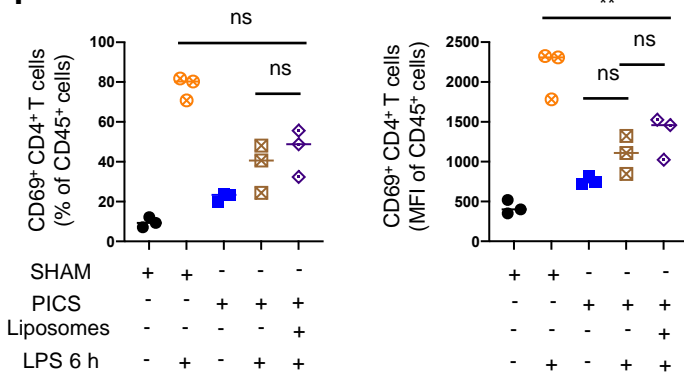
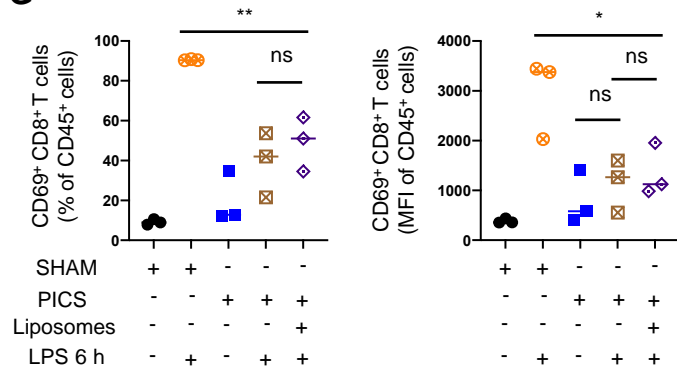
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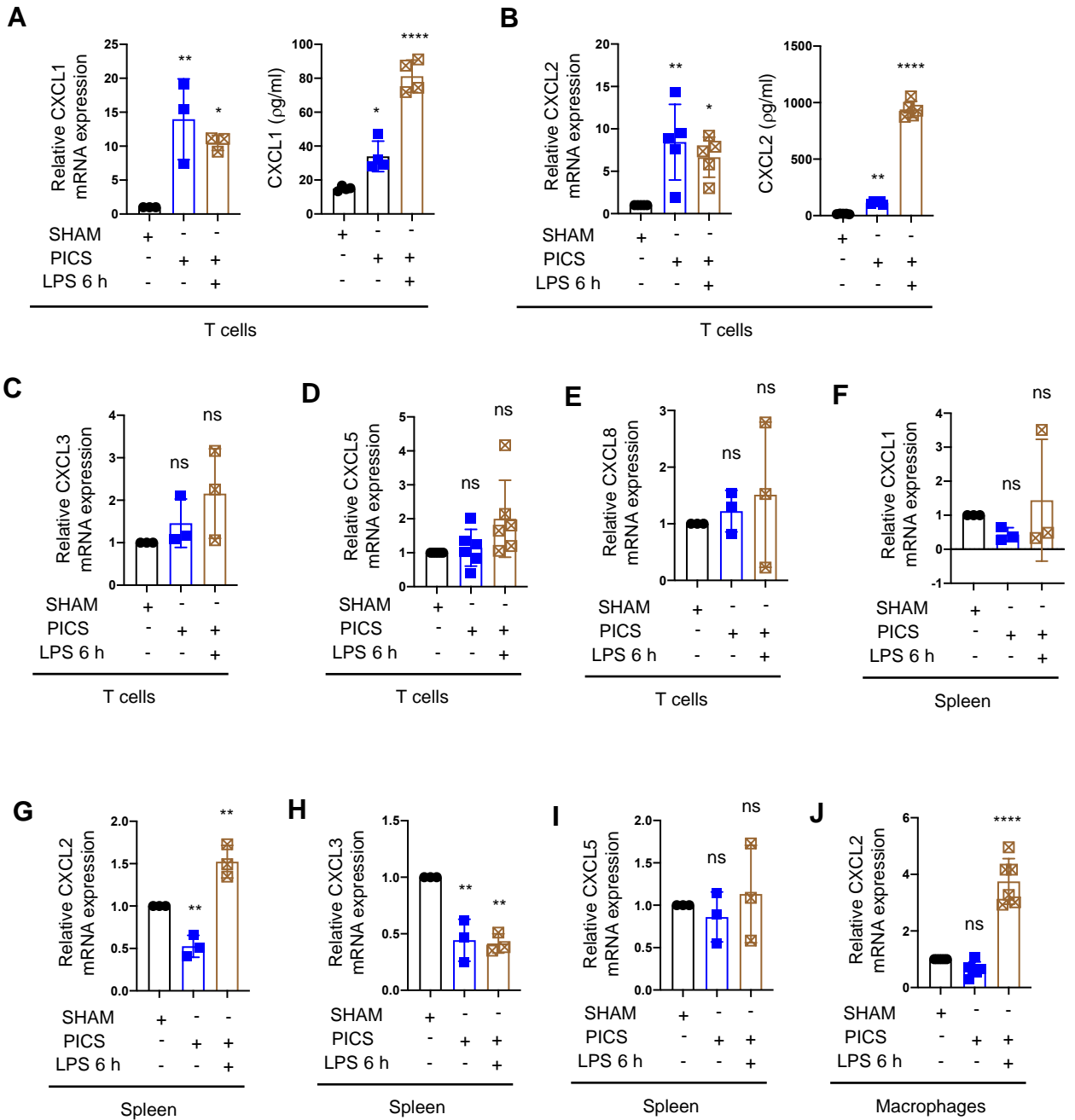


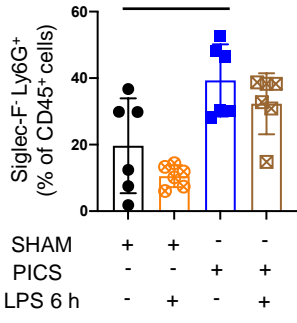
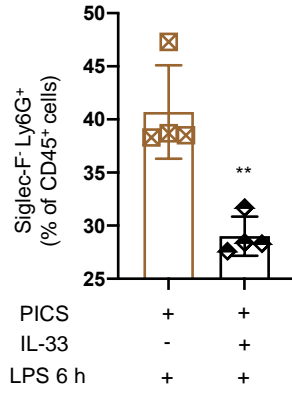
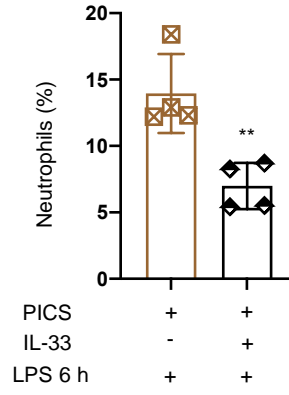
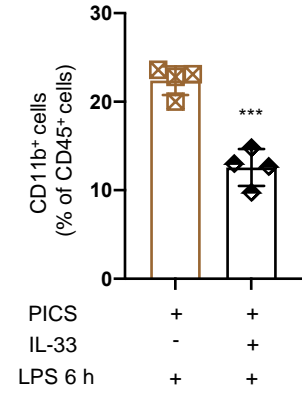
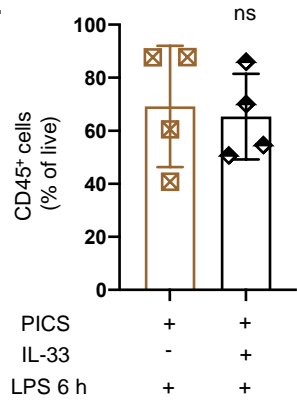
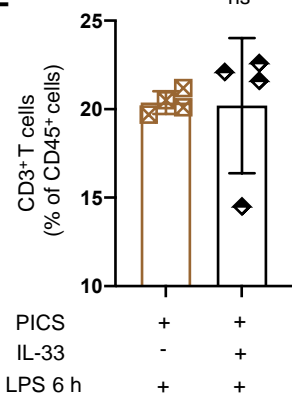
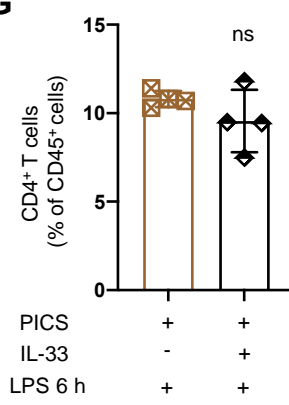




**A****B****C****D****E****F****G****H**

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