

1 *(Manuscript for Theranostics)*

2

3 **Illumination of targeting Nanotherapeutics with Precision Eyes: From Optical to**
4 **Radio-nanotheranostics**

5

6 *Maharajan Sivasubramanian^a, Yao-Chen Chuang^{a,b}, Chia-Hui Chu^a, Yu Hsia^a, Li-Jie*

7

*Lin^a, Leu-Wei Lo^a **

8

^a Institute of Biomedical Engineering and Nanomedicine, National Health Research

9

Institutes, Zhunan 35053, Taiwan

10

^bDepartment of Radiation Oncology, Taipei Medical University Hospital, Taipei

11

110301, Taiwan

12

13

14

15

16

17

18

19

20

21

22

23

24 **Corresponding author*

25 *Leu-Wei Lo, Ph.D.*

26 *Institute of Biomedical Engineering and Nanomedicine*

27 *National Health Research Institutes, Zhunan 35053, Taiwan*

28 *Email:*

29 *Tel: 886-37-246-166 ext. 37115*

30 *Fax: 886-37-586-440*

31 **Abstract**

32 Cancer continues to rank among the deadliest diseases globally, claiming numerous
33 lives due to its high mortality rates. Chemotherapy, a primary form of cancer treatment,
34 offers significant benefits but is hampered by drawbacks that compromise patients'
35 quality of life. Consequently, developing minimally invasive treatment alternatives
36 remains a formidable challenge. Among these, nanomaterial-based, light-activated
37 phototherapies—including photodynamic therapy, photothermal therapy, and
38 radiotherapy—emerge as promising options, providing precise spatial and temporal
39 control with reduced invasiveness. Advances in nanoscience and engineering have led
40 to the creation of nanoparticles (NPs) that integrate therapeutic and diagnostic
41 capabilities, known as theranostics, which enhance the effectiveness of clinical cancer
42 management. This review, summarizes recent advancements in nanotheranostics,
43 spanning optical to radio-based approaches. We discuss the roles of various
44 nanomaterials, such as upconversion NPs, gold NPs, nano-scintillators, and
45 mesoporous silica NPs, among others, highlighting their dual diagnostic and therapeutic
46 functionalities. Furthermore, NP-mediated induction of programmed cell death
47 mechanisms—ferroptosis, pyroptosis, and cuproptosis—underscoring their
48 significance in targeted cancer therapies and immune modulation were also explored.
49 Additionally, we discuss how NPs can activate the stimulator of interferon genes
50 pathway, amplify antitumor immunity and offer new avenues for improving cancer
51 treatment outcomes.

52

53 **Keywords:** Nanotheranostics, phototherapies, radiotherapy, radionuclides,
54 programmed cell death, immunomodulation and stimulator of interferon genes.

55

56

57

58

59

60

61

62

63

64

65

66

67

| | |
|----|---|
| 68 | Contents |
| 69 | Introduction |
| 70 | Functional photodynamic therapy (PDT) |
| 71 | NIR PDT |
| 72 | (UCNPs PDT) |
| 73 | TP PDT |
| 74 | X-ray PDT |
| 75 | TME modulating PDT |
| 76 | Image-guided PDT |
| 77 | Image-guided photothermal therapy (PTT) |
| 78 | Optical imaging based NIR-I imaging-guided PTT |
| 79 | Optical imaging based NIR-II imaging-guided PTT |
| 80 | Radionuclide nanotheranostics |
| 81 | Radionanotheranostics of α -and β -emitters |
| 82 | Radionanotheranostics of Cherenkov radiation |
| 83 | Comparison of proton therapy and intensity modulated radiotherapy |
| 84 | Advancing cancer immunotherapy with NPs |
| 85 | NPs combined with immune checkpoint blockade |
| 86 | NPs enabled immunogenic cell death |
| 87 | NPs enabled programmed cell death pathways |
| 88 | NPs enabled stimulator of interferon genes pathway |
| 89 | Conclusion and Outlook |
| 90 | Acknowledgements |
| 91 | References |
| 92 | |
| 93 | |

94 **Introduction**

95 Cancer is one of the leading causes of death worldwide, with mortality rates
96 continuing to increase according to the World Health Organization (WHO) estimates
97 [1]. However, systemic chemotherapy (CMT), a main clinical treatment modality for
98 cancer, is associated with setbacks, such as poor bioavailability and heterogeneous drug
99 distribution within tumors as well as drug resistance [2, 3]. Phototherapies, such as
100 photodynamic therapy (PDT) and photothermal therapy (PTT), have emerged as
101 alternatives primarily owing to the following reasons: 1) reduced toxicity and selective
102 activation for penetration into various tissues and 2) minimal invasiveness with
103 spatiotemporal precision and control of irradiation on the disease site. In PDT,
104 excitation of a photosensitizer (PS) using a suitable wavelength generates reactive
105 oxygen species (ROS) to selectively destroy the cancer cells [4-8]. However, classical
106 PDT with ultraviolet (UV) or visible (Vis) light-activated PSs are confined to surface-
107 accessible tumors due to poor tissue penetration. Near-infrared (NIR) light (NIR-I
108 (700–900 nm) and NIR-II (1000–1700 nm)) within the biological transparency window
109 provides deeper tissue penetration because of the minimal absorbance of endogenous
110 chromophores in the NIR range [9, 10]. Consequently, there have been significant
111 efforts to develop NIR light-activated PSs to extend the clinical applications of PDT.
112 NIR PDT can be classified into the following three types: 1) direct absorption of NIR
113 light by PS, 2) two-photon (TP) absorption by organic PSs and nanomaterials that have
114 a TP absorption cross-section for TP PDT [11, 12], and 3) upconversion nanoparticles
115 (UCNPs), in which nanomaterials absorb NIR light and emit fluorescence in the UV or
116 Vis wavelength range, thereby enabling the therapeutic potential of classical UV–Vis
117 PS to be realized indirectly. Furthermore, the emission characteristics of UCNPs can
118 be tailor-made according to PS absorption by carefully selecting the type and number
119 of ions doped [13-15]. Conversely, PTT is based on the activation of PS by a suitable
120 light to induce local heat to ablate cancer cells. Numerous NIR-induced photothermal
121 conversion nanomaterials have been shown to be excellent PTT agents [16-18].
122 Although the application of standalone NPs for either therapy or imaging has yielded
123 moderate success, recent developments in nanoscience have further prompted
124 researchers to produce multifunctional nanomaterials. This venture resulted in
125 nanotheranostics, an emerging paradigm that implements diagnostic and therapeutic
126 functions in a single nano-entity, and has found potential applications. For instance,
127 NP-mediated PTT/PDT combined with imaging modalities, such as computed
128 tomography (CT), optical imaging, magnetic resonance imaging (MRI), etc., have been
129 reported as useful cancer theranostics [19, 20]. Besides the image-guided PTT/PDT,
130 nuclear medicine, which involves the use of radioactive nuclides for therapy and
131 imaging (positron emission tomography (PET) and single-photon emission tomography

132 (SPECT)), has been well received and is rapidly advancing [21-23]. Recently,
133 Cherenkov radiation (CR) and Cherenkov luminescence have also been explored as
134 radiotheranostics. CR occurs when charged particles from radioactive decay traverse a
135 dielectric medium with a speed greater than the velocity of light luminesce in the UV–
136 blue spectrum, which in turn can be used to excite PSs to generate ROS [24, 25]. Using
137 nanotechnology, radiolabeled NPs can be designed and used for diagnostic and
138 therapeutic purposes when tagged with appropriate radionuclides [26-28].

139 The transition from optical to radioactive nanotheranostics represents a paradigm
140 shift from qualitative preclinical testing to quantitative clinical application. While
141 optical modalities such as NIR-I and NIR-II fluorescence offer high spatial resolution
142 and real-time intraoperative guidance, they remained inherently limited by tissue
143 penetration depth, where signal attenuation and autofluorescence impede deep-seated
144 tumor detection. The evolution toward radioactive modalities was primarily fueled by
145 three driving factors: First, the high energy of gamma rays and positrons ensures
146 minimal attenuation, allowing for whole-body imaging regardless of tissue depth.
147 Second, unlike optical signals, which are relative, radiotracers allow for absolute
148 quantification of injected dose per gram of tissue. Third, the chemical
149 interchangeability of diagnostic isotopes with therapeutic emitters provided a seamless
150 pathway for 'see-and-treat' protocols.

151 Optical nanotheranostics rely on externally applied light (e.g., visible/NIR) to
152 activate NP for imaging and therapy, with design focusing on optical absorption,
153 wavelength tuning, and energy conversion efficiency. In contrast, radio-based
154 nanotheranostics include radiotherapy (RT)/radiotheranostic strategies that utilize
155 radionuclides as internal energy sources for tumor imaging and therapeutic radiation
156 delivery, with design considerations centered on radionuclide stability, radiolabeling
157 efficiency, dosimetry control, and in vivo pharmacokinetics. We further explain that
158 these two modalities are complementary rather than isolated, as optical systems offer
159 precise spatiotemporal control but are limited by tissue penetration, whereas
160 radiological approaches overcome depth constraints and can also be integrated with
161 optical mechanisms in hybrid platforms such as scintillator-mediated systems.

162 Herein, we discuss the latest developments in nanotheranostics ranging from
163 “optical to radio” using various NP strategies by describing their design of systems,
164 operational mechanisms, and applications. NP-induced immunogenic cell death (ICD)
165 has emerged as a promising therapeutic approach [29, 30]. Cancer cells often develop
166 resistance mechanisms to evade programmed cell death (PCD), thereby reducing the
167 effectiveness of conventional treatments. In this context, nanotechnology provides
168 innovative solutions by providing targeted delivery systems and enabling precise

169 modulation of PCD pathways within the tumor microenvironment (TME). This review
170 also focuses on the role of NPs in promoting PCD in cancer cells such as ferroptosis,
171 cuproptosis, and pyroptosis to improve therapeutic outcomes while simultaneously
172 sparing normal tissues [31, 32]. We analyze the design, functionalization, and
173 interactions of NPs within cancer cells to induce PCD and activate innate immune
174 pathways [33, 34]. Moreover, this review discusses the therapeutic potential of
175 harnessing these pathways for improved cancer treatment.

176

177 **FUNCTIONAL PDT**

178 **NIR PDT**

179 Classical PDT in which a PS that is activated by UV–Vis light is used suffers from
180 poor efficacy due to limited penetration depth and inappropriate activation of the PS
181 [35]. A viable alternative to avoid these problems is the use of PSs that are responsive
182 to NIR light (NIR–I (700–900 nm) and NIR–II (1000–1700 nm)). In these biological
183 transparent windows, tissues have relatively low attenuation coefficients compared
184 with that in UV–Vis windows and thus improved penetration depth [36–38]. UCNPs
185 comprise an important method to deliver light into deep tissues for PDT. Lanthanide-
186 doped UCNPs, as an energy transducer, emit high-energy photons when excited with
187 lower-energy NIR photons [14, 39]. The merits of UCNPs include the following: 1)
188 they can realize the therapeutic potential of well-investigated and confirmed traditional
189 UV light–activated PSs and 2) they are endowed with exciting optical properties, such
190 as improved penetration depth, well–stabilized photochemical properties, and no
191 autofluorescence background [40]. Herein, we discuss the advancements of UCNP-
192 mediated deep PDT in the subsequent section.

193 A hydrogen peroxide-driven black TiO₂ mesoporous Janus nanomotor (HTiPC) was
194 designed to enhance tumor penetration and NIR-triggered PDT [41]. Comprising a
195 NIR-responsive H-TiO_{2-x} nanosphere and a catalase (CAT)-modified periodic
196 mesoporous organosilica (PMO) nanorod, the nanomotor autonomously navigates
197 through the TME by catalytically decomposing overexpressed H₂O₂ into oxygen and
198 water. This process not only propels the nanomotor but also alleviates tumor hypoxia,
199 improving ROS generation for PDT. The optimized HTiPC-ii nanomotor demonstrated
200 enhanced self-propulsion, increased cellular uptake, and deep tumor penetration,
201 leading to effective intracellular oxidation and apoptosis under 808 nm NIR irradiation.
202 In vivo studies confirmed superior tumor inhibition, highlighting the potential of these
203 TiO₂-based mesoporous nanomotors as a promising strategy for deep-tissue cancer
204 treatment.

205 NIR PS Secy7, a selenium-substituted heptamethine cyanine, was developed to
206 overcome the low singlet oxygen ($^1\text{O}_2$) generation in PDT [42]. By inducing an
207 intramolecular charge transfer effect and leveraging the heavy-atom effect of Se, Secy7
208 significantly narrows the energy gap (0.51 eV), enhances spin-orbit coupling (1.44
209 cm^{-1}), and achieves a high triplet state yield (61%), leading to an ultrahigh $^1\text{O}_2$ quantum
210 yield. Under low-power 850 nm irradiation, Secy7 demonstrated a ~ 24.5 -fold higher
211 $^1\text{O}_2$ generation than indocyanine green (ICG) and exhibited excellent phototoxicity with
212 minimal dark toxicity. In vivo studies confirmed its potent antitumor and antimetastatic
213 effect. A NIR photoactivatable nano-PROTAC (NAP) was developed for precise,
214 spatiotemporal control of protein degradation in tumors. NAP consists of a PROTAC
215 linked to an NIR PS via a $^1\text{O}_2$ -cleavable linker, forming a self-assembled
216 nanoformulation with initially silent proteolytic activity. Upon systemic administration,
217 NAP accumulates in tumors through the enhanced permeability and retention (EPR)
218 effect. NIR irradiation at the tumor site generates $^1\text{O}_2$, cleaving the linker to activate
219 PROTAC, which then degrades BRD4 and sensitizes cancer cells to PDT [43].

220 KD1@HPEG NP, a NIR-II PDT platform was designed for repeatable, precise, and
221 durable cancer treatment. This nanoplatform combines deep-tissue NIR-II fluorescence
222 imaging (FLI) with a Type I PS (KD1) that can generate oxygen-independent ROS.
223 KD1 was engineered to improve intersystem crossing, reduce energy loss, and
224 minimize unwanted photothermal heating. The hyaluronic acid-PEG coating improves
225 stability, provides intrinsic tumor targeting, and importantly avoid anti-PEG immune
226 responses, allowing repeated dosing without rapid clearance. Mechanistically, the
227 nanoplatform not only damages tumors through ROS but also triggers mitochondrial
228 dysfunction and ferroptosis-related pathways, for robust tumor killing [44].

229 NanoPcCu, a NIR-activated nano-photocatalyst, was designed to improve PDT in
230 hypoxic tumors. NanoPcCu can generate Type I reactive species such as $\text{O}_2^{\bullet-}$ and $\bullet\text{OH}$
231 even under low-oxygen conditions, making it much more effective in the hypoxic TME.
232 Under 685 nm light, the NPs also generate heat for PTT, so the treatment combines
233 PDT and PTT for robust tumor killing. In addition, NanoPcCu can perform catalytic
234 reactions, which generate more toxic radicals through Cu(II)/Cu(I) photoredox cycling,
235 further amplifying oxidative damage. The system also enables photoacoustic imaging
236 (PAI)-guided therapy, allowing tumor monitoring during treatment. Importantly,
237 NanoPcCu also promoted dendritic cell maturation and systemic antitumor immune
238 responses, that controls the growth of both primary and distant tumors [45].

239 A NIR-responsive “dark state photocage” nanoplatform that combines PDT and
240 CMT was designed and developed. The system stays largely inactive before irradiation,

241 but under 650–660 nm light, it undergoes efficient photolysis even at low power,
242 allowing deep-tissue activation with reduced off-target effects. When irradiated,
243 methylene blue (MB) phenothiazine/phenoxazine-based cages transition from ground
244 state to excited state, which promotes intersystem crossing and triplet-state formation,
245 allowing cargo release. In the representative system MB-Lv, NIR light triggers the
246 release of both STAT3 inhibitor (Lv) and MB, which produces $^1\text{O}_2$ for PDT. As a result,
247 enhanced PDT + CMT synergistic effect was observed, along with a clear fluorescence
248 turn-on signal for imaging-guided treatment [46]. An 808 nm NIR light-activatable
249 prodrug system for photoimmunotherapy of unresponsive tumors was developed.
250 BODIPY-based PS can be activated by 808 nm light, which penetrates tissue more
251 deeply and enables spatiotemporal drug release at the tumor site. In this system, R837
252 was linked to the NIR PS through a ROS-cleavable linker. Upon laser irradiation, the
253 NPs generated ROS and heat, which not only caused direct phototherapeutic tumor
254 killing but also cleaves the linker to release R837. This dual action promoted ICD,
255 release of tumor antigens and DAMPs, dendritic cell maturation, and stronger CD8⁺ T-
256 cell infiltration [47]

257 A mitochondria-targeting nanoplatfrom (NZ@TG) was developed to overcome the
258 shallow penetration and hypoxia limitations of conventional PDT by using 808 nm NIR
259 light. The system combines a lanthanide-doped nanocrystal with a phthalocyanine PS
260 and a hypoxia-activated prodrug, enabling dual action: localized oxidative damage in
261 mitochondria and hypoxia-triggered DNA damage. This synergistic effect induces
262 strong ICD, releasing DAMPs that activate systemic anti-tumor immunity, including
263 macrophage polarization toward the M1 phenotype and activation of CD8⁺ T cells.
264 Importantly, NIR-PDT achieved significant tumor suppression in both subcutaneous
265 and orthotopic liver tumor models, demonstrating effective treatment of deep-seated
266 tumors. It also generated long-term immune memory, reduced metastasis and
267 recurrence, and significantly enhanced the efficacy of immune checkpoint blockade
268 (ICB) therapy [48].

269 Studies have demonstrated that mild hyperthermia improves tumor oxygenation in
270 rodent, canine, and human tumors. The increase in the tumor oxygenation lasted for up
271 to 1–2 days after heating at mild temperatures (about 39–42 °C). The increase in
272 oxygenation occurs mainly due to enhanced tumor blood perfusion [49]. In one study,
273 semi-quantitative analysis of hypoxia-positive signals in tumor slices after mild NIR
274 PDT (~45 °C) treatment with DiR-hCe6-liposome showed a significant reduction in
275 tumor hypoxia from about 38% to ~12% [50]. PTT can trigger the upregulation of heat
276 shock proteins (HSPs), which protect cancer cells and reduce their sensitivity to heat-
277 induced apoptosis [51]. This limitation can be overcome by combining PTT with PDT.

278 In this strategy, the ROS generated during PDT can damage HSPs and directly kill
279 cancer cells, thereby enhancing the overall therapeutic effect [52]. Younis et al.
280 demonstrated that a nanopatform composed of AuNRs/MoS₂ loaded with ICG enables
281 simultaneous PDT and PTT under a single low-power NIR laser (0.2 W/cm²). The
282 system showed high photothermal conversion efficiency (PCE) (68.8%), raising the
283 temperature to ~60 °C within 5 min. The heat generated by the dual plasmonic NP
284 triggered the release of ~85% of ICG, leading to ¹O₂ production for PDT. As a result,
285 the combined PDT/PTT eradicated cancer cells and tumors in vitro and in vivo,
286 showing significantly higher antitumor efficacy than either PDT or PTT alone [53].

287 UCNPs PDT

288 UCNPs absorb low-energy NIR light and convert it into higher-energy UV or visible
289 light [54-56]. Because NIR light can penetrate deeper into tissues with less absorption
290 by biological components, UCNPs enable activation of light-responsive systems in
291 deeper regions [57]. Their lanthanide-doped structure allows efficient energy
292 conversion, making them useful as energy transducers to generate ROS and enhance
293 processes like PDT [58, 59].

294 The following study presents an advanced PDT system using core-multishell
295 UCNPs capable of emitting three distinct colors—red, green, and blue—when exposed
296 to different NIR wavelengths (1550 nm, 808 nm and 980 nm) [60]. These UCNPs are
297 integrated with PSs and nitric oxide (NO) donors to create a programmable "off-on"
298 nanopatform for precise and controlled phototherapy. By tuning the luminescent and
299 inert shell thicknesses, the UCNPs independently activate imaging, NO release, and
300 ROS generation, thus allowing for stepwise and targeted treatment. Remarkably, the
301 sequential release of NO before PDT reduces tumor hypoxia, thereby improving the
302 overall therapeutic efficacy (**Figure 1**). A study introduces stimuli-sensitive tumor-
303 targeted photodynamic NPs (STPNs) designed for deep tumor treatment, overcoming
304 challenges such as limited NIR tissue penetration and poor accumulation at target sites
305 [61]. These STPNs combine lanthanide-doped UCNPs for improved imaging and
306 photostability with the PS Purpurin 18 (Pu18), which exhibits persistent luminescence
307 (PL). Before intravenous administration, STPNs are preactivated by 980 nm NIR laser
308 irradiation, allowing them to accumulate at tumor sites and enter cells via HER2
309 receptors. Within the TME, STPNs disassemble, thereby enabling the UCNPs to trigger
310 the photoactivity of Pu18 and produce ROS for effective antitumor therapy.
311 Furthermore, STPNs provide diagnostic capabilities through MRI and intraoperative
312 NIR imaging due to their gadolinium content.

313 A cold-responsive nanoplatfrom (UCNPs@SiO₂-Ce6-HA) that integrates
314 cryotherapy with PDT for enhanced melanoma treatment was developed. The low-
315 temperature environment amplifies upconversion luminescence (2.45-fold) and ¹O₂
316 production (3.15-fold), significantly boosting PDT efficacy. The HA coating ensures
317 efficient transdermal delivery, leading to a remarkable 79% tumor growth inhibition in
318 melanoma-bearing mice, outperforming cryotherapy (17%) and PDT (55%) alone [62].
319 A recent study introduced a programmable NIR-controlled nanosystem (PB@UA) to
320 enhance PDT by overcoming tumor antioxidant defenses [63]. Using switchable
321 UCNPs, the system independently triggers berbamine (BBM) release at 980 nm and
322 activates a PS (ZnPc) at 808 nm. BBM inhibits antioxidant enzyme activity and disrupts
323 calcium ion regulation, making tumor cells more vulnerable to ROS induced apoptosis.
324 This sequential activation significantly improves PDT efficiency, achieving an 80.91%
325 tumor inhibition rate, surpassing PDT (31.78%) or BBM (11.29%) alone.
326 UCNPs@AgBiS₂ core-shell NPs were successfully synthesized using an ion exchange
327 reaction, enhancing PCE from 14.7% to 45% through cross-relaxation between Nd ions
328 and AgBiS₂ [64]. The optimal Nd ion doping (1%) in the inner core facilitated strong
329 upconversion emissions, which excited the AgBiS₂ shell to generate ROS for PDT.
330 These NPs demonstrated significant antitumor efficacy in vitro and in vivo under 808
331 nm laser irradiation by combining PTT and PDT.

332 Tumor-targeted upconversion nanospheres (ALUMSNs) that combine
333 upconversion PDT and PTT with multimodal imaging was developed by Palanikumar
334 et al. NaYF₄:Yb/Er core converts deeply penetrating NIR light into visible light, which
335 then activates Ce6 to generate ROS for PDT, while Bi₂Se₃ simultaneously generates
336 heat for PTT. The nanosystem was further designed with a mesoporous silica shell for
337 stable Ce6 loading, a lipid/PEG coating, and an ATRAM peptide that helps selective
338 uptake in the mildly acidic tumor environment. In addition, Gd in the core enables MRI,
339 and the system also supports thermal and fluorescence imaging, allowing treatment
340 monitoring [65].

341 Chu et al. developed UCNPs@AgBiS₂ core-shell NPs as an upconversion-based
342 PDT/PTT platform for cancer treatment. The Nd³⁺-sensitized upconversion core
343 converts 808 nm NIR light into higher-energy emission, which then activates the
344 AgBiS₂ shell to generate ROS for PDT. At the same time, energy interaction between
345 Nd ions and AgBiS₂ greatly improved the PCE from 14.7% to 45%, making the system
346 much stronger for PTT as well. 1% Nd in the inner core produces strong upconversion
347 emission and better therapeutic performance [66]. A smart upconversion-assisted
348 radiosensitizer (RS) platform (DH&UH NPs) that improves both tumor imaging and
349 RT was developed by Mo et al. The upconversion NPs (UH NPs) produce UV light

350 under 980 nm NIR irradiation, which remotely triggers in situ NP aggregation through
351 chemical bond formation with the downconversion partner (DH NPs). This aggregation
352 greatly prolongs tumor retention, allowing long-term NIR-II FLI for treatment
353 monitoring and tumor tracking. At the same time, the system carries diselenide-based
354 radiosensitizing groups, which generate strong oxidative products and increase ROS
355 under X-ray exposure, making cancer cells more sensitive to RT [67].

356 Wang et al. developed PURH, a smart upconversion-based PDT immunotherapy
357 nanoplatform that can be controlled by two different NIR light for precise cancer
358 treatment. Under 980 nm irradiation, the system activates rose bengal (RB) to generate
359 ROS, causing photodynamic tumor killing and ICD. Under 808 nm irradiation, the same
360 platform triggers the controlled release of CpG, which promotes dendritic cell
361 maturation and T-cell infiltration. The mesoporous silica shell stores RB, while
362 hyaluronic acid helps tumor targeting [68]. A carbon-based upconversion
363 nanocomposite was developed by Yang et al. to improve light utilization and
364 therapeutic performance under 980 nm irradiation. By coating mesoporous carbon NPs
365 with a lanthanide oxysulfide layer ($Y_2O_2S:Yb^{3+},Er^{3+}$), the system converts NIR light
366 into visible emission, which enhances photothermal efficiency (from ~59% to ~83%)
367 and supports upconversion-assisted photodynamic effects. This platform enables rapid
368 heating (~50 °C within 150 s) and efficient tumor ablation while also carrying drugs for
369 combined therapy with controlled release via a stimuli-responsive hydrogel. In vitro
370 and in vivo studies, including subcutaneous and ocular melanoma models, showed
371 strong tumor targeting, significant reduction in cancer cell activity, and marked
372 inhibition of tumor growth. The treatment activates tumor-suppressive pathways and
373 suppresses proliferation-related signaling, improving drug sensitivity [69].

374 **TP PDT**

375 TP PDT provides deep-tissue penetration by absorbing two low-energy NIR
376 photons simultaneously and emitting high-energy photons. TP PDT involves nonlinear
377 photon absorption, which allows three-dimensional selectivity to target tumor cells, a
378 critical feature that cannot be achieved by conventional single-photon absorption [11,
379 70]. Herein, we discuss various NP-based TP PS approaches for deep PDT. Using the
380 well-ordered mesoporous structure of MSN, Cheng et al. chemically attached a donor
381 TP antenna molecule in the silica framework and an acceptor PS molecule in the
382 nanochannels. This precise design allowed an efficient FRET-mediated energy transfer
383 rate of up to 93% to produce cytotoxic ROS that exerted improved PDT effects in a
384 breast cancer model [71].

385 This study shows the development of a novel PDT PS, BF₂DCz, an organic material
386 with room-temperature phosphorescence (RTP) characteristics [72]. Encapsulated
387 within a bovine serum albumin matrix, BF₂DCz-BSA demonstrates high
388 photoluminescence quantum yield ($47.7 \pm 3\%$), impressive intersystem crossing
389 efficiency ($\sim 90.3\%$), excellent biocompatibility, negligible dark toxicity, and superior
390 photostability (**Figure 2**). Using an NIR femtosecond laser, the material achieves
391 efficient ROS generation, enabling precise spatial and temporal therapeutic control. In
392 addition to demonstrating selective blood vessel closure and TP imaging in the mouse
393 brain vasculature, this study emphasizes the potential of BF₂DCz-BSA for deep-tissue
394 PDT, providing a promising approach for improving the outcomes of cancer therapy.
395 Emo/HSA NPs (E/H NPs), a highly efficient TP-PDT agent derived from emodin, a
396 natural anthraquinone, was developed and studied [73]. The NPs exhibited an
397 exceptional TP absorption cross-section ($4.02 \times 10^7 \text{ GM}$) and significant ¹O₂ quantum
398 yield (31.9%). In vitro, the E/H NPs exerted improved anticancer effects against MCF-
399 7 cells, whereas in vivo experiments revealed prolonged tumor retention and effective
400 tumor ablation at an ultra-low dosage (0.2 mg/kg) under 800 nm femtosecond laser
401 irradiation.

402 A novel nanohybrid combining thiolated chitosan-coated gold (Au) nanostars
403 (AuNS-TCS) and riboflavin-conjugated N,S-doped graphene quantum dots (Rf-N,S-
404 GQDs) for synergistic TP-PDT/PTT was synthesized [74]. Using a single low-power
405 pulsed laser (760 nm, 200 mW·cm⁻²), the nanohybrid leverages the spectral overlap
406 between the localized surface plasmon resonance (LSPR) of AuNS and the TP
407 absorption of Rf-N,S-GQDs, achieving simultaneous photodynamic and photothermal
408 effects. The thiolated chitosan coating improves colloidal stability and facilitates the
409 anchoring of Rf-N,S-GQDs, and the plasmonic effect significantly boosts ¹O₂
410 generation and photothermal performance. Compared with individual therapies, the
411 combined TP-PDT/PTT system exhibits superior phototoxicity and therapeutic
412 outcomes against 2D cell monolayers and 3D tumor spheroids, highlighting its potential
413 for efficient and simplified cancer treatment under single-laser activation. The first
414 development of metal-free NIR thermally activated delayed fluorescence NPs (NIR-
415 TADF NPs) as effective agents for TP PDT and imaging was presented in this study
416 [75]. These NPs exhibit excellent ¹O₂ generation, inherent TP excitation, and NIR
417 fluorescence emission, demonstrating remarkable biocompatibility and biosafety.
418 Validated in A549 tumor xenograft models, the NIR-TADF NPs exerted significant
419 antitumor effects and high precision in both single-photon and TP imaging and therapy.

420 A hypoxia-activated, novel mitochondria-localized iridium (III) endoperoxide
421 prodrug (2-O-IrAn) was designed for synergistic PDT and photoactivated CMT (PACT)

422 was developed [76]. Activated by NIR TP irradiation, 2-O-IrAn releases a cytotoxic
423 iridium (III) complex, $^1\text{O}_2$, and alkoxy radicals, demonstrating high phototoxicity in
424 hypoxic tumor cells and multicellular tumor spheroids at low concentrations.
425 Encapsulation in biotin-functionalized phospholipid NPs enhanced tumor selectivity
426 and pharmacological properties, enabling near-complete tumor eradication in a mouse
427 model after a single treatment. The limitations of Ru(II) polypyridine complexes in
428 PDT were addressed by coordinating them with graphitic carbon nitride (g-C₃N₄)
429 nanosheets to create an oxygen-self-sufficient TP PDT immunotherapy [77]. The
430 conjugates exhibit increased TPA, which is significantly stronger than that of molecular
431 Ru(II) complexes, and generate a robust ROS storm, even under hypoxic conditions,
432 by catalytically converting H₂O/H₂O₂ into O₂. Encapsulation with an amphiphilic
433 polymer improved the pharmacological properties, thereby enabling mitochondrial and
434 endoplasmic reticulum (ER) accumulation. Upon irradiation with a deeply penetrating
435 800 nm laser, the nanomaterial induced cell death via apoptosis, paraptosis, ferroptosis,
436 and ICD in both monolayer and multicellular tumor spheroids. In a melanoma-bearing
437 mouse model, it significantly inhibited tumor growth in primary tumors and activated
438 immune responses to secondary distant tumors, exhibiting its potential to treat hypoxic
439 solid tumors and metastatic cancer.

440 MeTTh, a high-performance TP PS for treating small residual glioblastoma (GBM)
441 was developed by Li et al. MeTTh was designed to have a large TP absorption cross-
442 section, strong ROS generation, and both Type I and Type II photodynamic activity,
443 making it useful even in the hypoxic GBM environment. After transferrin conjugation,
444 the modified NPs successfully targeted orthotopic GBM. It also enabled deep TP
445 imaging of brain structures up to 940 μm , allowing image-guided therapy [78]. A TP
446 theranostic nanoplatfrom based on mesoporous silica NPs (MTP-MSNs) for
447 simultaneous deep-tissue imaging and targeted cancer therapy was developed by Wu et
448 al. The NPs were engineered to produce multicolor TP fluorescence and were capped
449 with the cancer-targeting aptamer AS1411, which recognizes nucleolin overexpressed
450 on tumor cells. Once internalized by cancer cells, the aptamer gate opens the nanopores
451 and releases the loaded drug, enabling selective intracellular therapy [79]. Ke et al.
452 developed a biodegradable iridium (III)-based coordination polymer NPs (IrS NPs) to
453 improve PDT against cancer. These NPs remain stable in physiological conditions but
454 break apart in the tumor environment, where they reduce glutathione (GSH) levels and
455 release active iridium complexes. The NPs selectively accumulate in mitochondria,
456 making the treatment more precise at the subcellular level. Upon light activation, they
457 generate both $^1\text{O}_2$ and superoxide radicals, triggering cancer cell death through a
458 combination of apoptosis and ferroptosis rather than a single pathway [80].

459 A sulfur-containing polymer PS for TP PDT was prepared by a simple catalyst-free
460 thiol–yne click reaction. Nanoplatfrom in this study were designed with a donor– π –
461 acceptor structure, which gave them strong TP absorption in the 700–800 nm range,
462 making them suitable for deeper tissue light activation. The introduction of sulfur
463 enhanced intersystem crossing and improved $^1\text{O}_2$ generation, while polymerization
464 further amplified this effect. In addition, the aggregation-induced emission unit helped
465 reduce energy loss and improve performance in the aggregated NP state. As a result,
466 the optimized NPs showed strong ROS production and effective cancer cell killing
467 under TP excitation [81].

468 X-ray PDT

469 Although fascinating, NIR-based deep PDT suffers from: 1) limited penetration
470 depth (up to 1.5 cm) and 2) difficulty in synthesizing of NIR PS with a broad energy
471 gap. X-rays, as an ionizing radiation with unlimited penetration depth and photon
472 energy ranging from keV to MeV, have been a mainstay in clinical imaging and
473 treatment of cancer, and could be used as an excitation source for deep PDT [82-84].
474 Nevertheless, direct activation of PSs by X-ray cannot be achieved, which has resulted
475 in the development of scintillating NP (SCNPs) [85] and persistent luminescent NP
476 (PLNPs) [86, 87]. X-ray excited PDT uses SCNPs to down-convert high-energy X-rays
477 into UV-Vis light to activate PSs, whereas X-ray excited PLNPs can activate PSs even
478 after the cessation of irradiation. Incorporation of heteroatoms (oxygen, nitrogen, and
479 sulfur) significantly enhanced X-ray energy harvesting, intersystem crossing, and
480 subsequent formation of ROS, particularly oxygen-independent $\bullet\text{OH}$ and $^1\text{O}_2$.
481 Encapsulating TBDCR in a crystalline PEG shell increased particle compactness and
482 further boosted ROS production. Remarkably, TBDCR NPs demonstrated strong NIR
483 fluorescence and potent anticancer efficacy against both standard HeLa and radio-
484 resistant HeLaR tumors in vitro and in vivo, establishing a new direction for purely
485 organic type I and II X-ray-excited PDT (X-PDT) nanoscintillators. $\text{NaYF}_4:\text{Tb}@\text{NaYF}_4$
486 core–shell NPs exhibiting enhanced PL were successfully synthesized for improved X-
487 PDT [88]. The optimized synthesis significantly enhanced X-ray excitation optical
488 luminescence and persistent luminescence by over 5.2 and 3.5 times, respectively.
489 Covalent conjugation of these NPs with the PS RB created an effective nanocoupling
490 system. A novel hyperfractionated irradiation strategy further amplified ROS
491 production, achieving 85% tumor inhibition in B16–F10 tumor-bearing mice at a low
492 irradiation dose (2 Gy) without notable side effects.

493 Although studies using SCNPs for X-ray PDT produced encouraging results,
494 substantial room remains for research in the following areas. First, improving SCNPs
495 parameters, such as size and luminescence properties, could promote efficient energy

496 transfer between SCNPs and PS to enhance ROS production. Second, targeting blood
497 vasculature rather than cancer cells is an attractive strategy to enhance tumor
498 accumulation. Third, validation of *in vivo* properties of NPs using orthotopic tumor
499 models is promising to minimize systemic toxicity. A novel purely organic
500 phosphorescent nanoscintillator was developed to achieve effective X-PDT without
501 relying on heavy-metal-containing inorganic agents [89]. These organic
502 nanoscintillators efficiently transferred energy to produce substantial $^1\text{O}_2$ under low-
503 dose X-ray irradiation (0.4 Gy), effectively combining the advantages of RT and PDT
504 (**Figure 3**). *In vivo* experiments demonstrated robust therapeutic performance against
505 deep-tissue tumors with minimal side effects, highlighting the potential for safe, low-
506 dose cancer treatment. A novel nanoplatform (Sc-D@Lip/Pt) was developed to
507 overcome limitations of PDT such as tumor hypoxia, insufficient tissue penetration,
508 and transient ROS generation. Incorporating a type I PS with core-shell scintillator NPs
509 enabled sustained production of less oxygen-dependent ROS ($\bullet\text{OH}$ and $\text{O}_2\bullet^-$) under X-
510 ray irradiation, continuing for over 4 h even after irradiation ceased [90]. Additionally,
511 a Pt(IV) prodrug was integrated and selectively converted into cisplatin within tumors,
512 enhancing chemotherapeutic and radiosensitizing effects. *In vitro* and *in vivo* studies
513 confirmed the platform's efficacy without aggravating hypoxia or increasing hypoxia-
514 related factors (HIF-1 α , VEGF), presenting a promising strategy for persistent type I
515 X-PDT.

516 In a study, low-dose X-PDT using purely organic phosphorescent nanoscintillators,
517 offering a safer alternative to traditional therapies that rely on heavy-metal-containing
518 agents was developed [91]. These nanoscintillators serve both as scintillators and PS,
519 effectively generating $^1\text{O}_2$ through energy transfer when exposed to X-rays. This
520 approach enhances deep-tissue cancer treatment by combining the strengths of PDT
521 and RT, while minimizing adverse effects on healthy tissues. The low dosage of 0.4 Gy
522 used in this therapy demonstrates significant efficacy in treating deep tumors,
523 highlighting its potential for broader applications. A novel UV PL material, particularly
524 Bi and Sb co-doped LaGaO₃, capable of ultra-long UV emission lasting over 2000 h
525 was developed [92]. These materials, activated by X-rays, exhibit high energy
526 collection and storage efficiency due to oxygen vacancies acting as energy traps within
527 the perovskite structure. The UV emission peaks at 372 nm, making these materials
528 highly effective for various applications, including X-ray imaging, phototherapy, and
529 photocatalysis. A key application is in PDT, where the LaGaO₃,Sb@g-C₃N₄ platform
530 shows excellent *in vitro* and *in vivo* results using low-dose X-ray irradiation (0.51 Gy),
531 overcoming traditional tissue penetration challenges while significantly reducing X-ray
532 dosage.

533 This study presents a non-invasive X-PDT strategy combined with real-time NIR-
534 II imaging for bladder cancer treatment. The system uses engineered nanotransducers
535 with lanthanide-doped nanoscintillators, which convert X-ray energy into visible light
536 to activate PSs for PDT, while simultaneously emitting NIR-II signals for deep-tissue
537 imaging. Tumor-targeting peptides help these nanotransducers selectively accumulate
538 in tumors, enabling efficient treatment. Under X-ray irradiation, this approach achieves
539 strong tumor regression, reduced recurrence, improved survival, and restoration of
540 immune balance. Importantly, NIR-II imaging allows continuous monitoring from
541 diagnosis to treatment and prognosis, and enables on-demand, fractionated therapy by
542 adjusting radiation dose based on imaging signals [93].

543 A pure organic aggregation-induced emission (AIE) nanoscintillator (TBDCR NPs)
544 was developed that can directly absorb X-rays and efficiently generate both type I •OH
545 and type II ROS without external energy converters. The incorporation of heteroatoms
546 improves X-ray absorption and promotes excited-state processes, enhancing ROS
547 production, while the AIE property and a rigid PEG shell further boost efficiency. This
548 system showed strong fluorescence, deep tumor penetration, and high therapeutic
549 performance, effectively suppressing both regular and radio-resistant tumors in vitro
550 and in vivo. Overall, this strategy introduces a new class of pure organic
551 nanoscintillators for efficient, hypoxia-tolerant X-PDT [94]. A biocompatible organic
552 nanoscintillator (BPT-HOF@PEG) for X-PDT was developed to treat hepatocellular
553 carcinoma (HCC). This nanoplateform uses a hydrogen-bonded organic framework that
554 acts as both a scintillator and PS, directly converting X-ray energy into ROS. Upon X-
555 ray irradiation, it produces strong oxidative damage to cell membranes and
556 mitochondria, while X-rays also cause direct DNA damage, leading to enhanced tumor
557 cell death. This dual action significantly improves treatment outcomes, achieving
558 strong tumor suppression in vivo. Mechanistically, the therapy promotes apoptosis and
559 inhibits tumor growth pathways such as NF- κ B, MAPK, and TNF signaling [95]. **Table**
560 **1** shows a list of deep tissue PDT nanosystems and their experimental parameters in
561 animal models.

562

563 **TME modulating PDT**

564 TME is highly dynamic and is mildly acidic (pH ~6.5–6.9) due to high lactate
565 production from altered cancer metabolism and poor blood supply. This acidity
566 increases with distance from blood vessels and is more pronounced in hypoxic regions.
567 Low pH promotes tumor growth by inducing abnormal vasculature, activating matrix-

568 degrading enzymes, and enhancing invasion. It also damages nearby normal cells while
569 tumor cells adapt, and drives immune suppression by polarizing macrophages toward a
570 tumor-supportive phenotype [96-99]. GSH levels are higher in tumor cells than in
571 normal tissues, typically in the millimolar range (about 5–10 mM), while normal cells
572 have lower levels (around 1–5 mM) [100, 101]. ROS, particularly hydrogen peroxide
573 (H_2O_2), are present at much higher levels in tumor tissues than in normal cells. Tumor
574 cells generate excess ROS through altered metabolism, mitochondrial activity, and
575 enzyme systems such as NADPH oxidase, with concentrations reaching up to ~100
576 μM —far higher than the low nanomolar levels in normal tissues [102]. This elevated
577 ROS environment supports tumor development, progression, and metastasis, and also
578 contributes to immune suppression by influencing macrophage behavior. At the same
579 time, cancer cells adapt to this oxidative stress and rely heavily on antioxidant systems
580 for survival [103]. Matrix metalloproteinases (MMPs) are zinc-dependent enzymes that
581 degrade extracellular matrix (ECM) components and regulate tissue structure and
582 signaling [104]. In normal tissues, their activity is very low and tightly controlled,
583 appearing mainly during physiological processes such as wound healing and
584 angiogenesis. In contrast, within the TME, MMP activity is significantly elevated due
585 to an imbalance between proteases and their inhibitors (TIMPs), leading to excessive
586 ECM breakdown. MMP-9 is strongly upregulated across many cancers compared to
587 normal tissues, whereas MMP-2 shows variable expression. Both enzymes degrade
588 type IV collagen in basement membranes, promoting tumor invasion and metastasis.
589 Overall, elevated MMP activity in the TME drives ECM breakdown and cancer
590 progression, unlike the tightly controlled activity in normal tissues [105, 106].

591 Hypoxia is a key feature of solid tumors, where oxygen levels are much lower than
592 in normal tissues due to rapid tumor growth and abnormal blood vessel formation.
593 While normal tissues typically have oxygen levels around 5% (≈ 38 mmHg), tumors
594 often show much lower values, sometimes dropping to 0.3–2.2% (≈ 2 –17 mmHg), with
595 even more severe oxygen depletion in tumor cores. This low-oxygen environment
596 arises from poor and irregular blood flow, which limits oxygen delivery and diffusion.
597 In response, tumor cells adapt through hypoxia-inducible factors (mainly HIF-1), which
598 regulate many genes involved in metabolism, angiogenesis, and survival. These
599 adaptations help cancer cells thrive under stress but also promote aggressive behavior,
600 metastasis, and resistance to PDT [107-109]. Hypoxia is a vital factor in the TME that
601 promotes metastatic progression, and hence leads to poor survival in several tumor
602 types. Alleviating hypoxia can be an attractive strategy for improving the diminished
603 PDT efficacy, for instance, hyperbaric oxygen therapy (HBOT) increases the amount
604 of oxygen a patient breathes by raising ambient pressure, which leads to more oxygen
605 dissolving in the blood plasma and reaching tissues [110]. In PDT, this can enhance

606 treatment by supplying more oxygen for $^1\text{O}_2$ production, as long as the oxygen can
607 effectively diffuse to the illuminated tumor region [111]. However, systemic
608 approaches like HBOT are not the only way to improve oxygen availability. An
609 alternative strategy is to generate oxygen directly within the tumor. Nanozymes, which
610 are nanomaterials with enzyme-like activity, can mimic catalase and convert
611 endogenous hydrogen peroxide (H_2O_2) into oxygen ($2 \text{H}_2\text{O}_2 \rightarrow 2 \text{H}_2\text{O} + \text{O}_2$). This
612 approach links oxygen production to the tumor's own redox environment, although its
613 effectiveness depends on the availability of H_2O_2 and the stability of the nanozyme in
614 biological conditions [112-114].

615 Another widely explored method involves oxygen carriers. Perfluorocarbons (PFCs)
616 can dissolve large amounts of oxygen physically, with oxygen loading and release
617 driven by partial pressure gradients [115-117]. Because of their high oxygen solubility,
618 PFC-based nanoemulsions or nanodroplets can act as efficient oxygen reservoirs for
619 PDT [118]. In contrast, hemoglobin-based systems rely on reversible oxygen binding,
620 similar to natural red blood cells, and release oxygen according to physiological
621 dissociation behavior, potentially enabling more controlled and biologically relevant
622 oxygen delivery [119, 120]. Finally, gas-filled microbubbles or nanobubbles provide a
623 distinct approach by storing oxygen in a gaseous core. These systems can be externally
624 triggered—most commonly using ultrasound—to release oxygen at a specific time and
625 location [121, 122]. This allows precise spatiotemporal control of oxygen delivery,
626 making them particularly useful for synchronizing oxygen supply with PDT irradiation
627 [123, 124]. This section reviews NP-based strategies that counter hypoxia to elevate the
628 effects of PDT.

629 HA, a glycosaminoglycan, is a principal component of ECM that participates in the
630 proliferation, invasion, and metastasis of cancer cells [125, 126]. Hyaluronidase
631 (HAase) is an enzyme that degrades HA, and has been used to improve the
632 permeability of anticancer drugs [127, 128]. A novel strategy adopted by Gong et al.
633 used HAase to alleviate hypoxia and improve the efficacy of PDT. Their results
634 demonstrated increased microvascular densities that amplified tumor perfusion to
635 relieve hypoxia. Consequently, improved tumor oxygenation and accumulation of
636 photoactive micelles by the improved EPR effect resulted in PDT-mediated tumor-
637 killing effects in a 4T1 tumor model [129]. PDT that leverages ROS faces challenges
638 in the hypoxic TME, where oxygen scarcity limits its efficacy. To address this issue,
639 two NO-releasing zinc(II) phthalocyanines, ZnPc-2NO and ZnPc-4NO, were
640 developed [130]. These compounds interact with intracellular GSH to release NO,
641 which inhibits mitochondrial respiration, reducing oxygen consumption and alleviating
642 hypoxia. Both conjugates generate ROS upon light irradiation, exhibiting cytotoxicity

643 under both normoxic and hypoxic conditions. Remarkably, ZnPc-2NO triggers ICD,
644 releasing damage-associated molecular patterns (DAMPs) that stimulate dendritic cells
645 (DC) maturation and anti-tumor immune responses. In vitro and in vivo experiments
646 have confirmed ZnPc-2NO-mediated PDT as an oxygen-efficient strategy that
647 suppresses tumor growth and improves anti-tumor immunity.

648 A novel photodriven nanozyme, Fe-TCPP-R848-PEG (Fe-MOF-RP) was
649 developed to address the immunosuppressive TME through a synergistic approach
650 combining photodynamic, chemodynamic, and immunotherapy [131]. Using Fe-TCPP
651 metal-organic frameworks (Fe-MOFs) as both catalytic cores and delivery vectors for
652 the immune agonist R848, the nanozyme enables precise tumor targeting and prolonged
653 circulation (**Figure 4**). It catalyzes the decomposition of hydrogen peroxide at tumor
654 sites to release oxygen, thereby alleviating hypoxia and improving PDT. Concurrently,
655 ferroptosis induced by the Fenton reaction promotes the release of tumor-associated
656 antigens (TAA), triggering ICD and robust anti-tumor immune responses. R848 further
657 stimulates DC maturation and reprograms tumor-associated macrophages (TAMs),
658 reshaping the TME. A recent study synthesized a carbon dot-based bifunctional
659 nanosystem, MnZ@Au, designed to improve PDT by addressing tumor hypoxia and
660 high glucose metabolism. MnZ@Au consists of Mn-doped carbon dots (Mn-CDs) as
661 the core, a ZIF-8 shell, and ultras-small AuNPs on the surface and acts as both a PS and
662 a cascading nanozyme with glucose oxidase (GOx)-and CAT-like reactivity [132]. It
663 catalyzes glucose consumption and hydrogen peroxide generation, which triggers
664 oxygen production to alleviate hypoxia and improve PDT efficacy. In vitro and in vivo
665 studies have demonstrated enhanced tumor penetration, improved ROS accumulation,
666 and significant tumor growth inhibition in breast cancer models. Furthermore,
667 MnZ@Au enabled PAI and PET imaging to monitor oxygen saturation and reduced
668 glucose uptake in tumors, respectively, thus validating its real-time catalytic activity
669 and therapeutic effects.

670 A novel light-responsive nanoplatform for targeted PDT in pancreatic ductal
671 adenocarcinoma (PDAC) was developed to address its dense desmoplastic and
672 immunosuppressive TME. The system uses semiconducting polymer NPs (PCP-NPs)
673 modified with midkine-specific nanobodies (MDK Nbs) to achieve precise delivery to
674 PDAC tissues enriched with MDK expression [133]. Upon light irradiation, the
675 nanoplatform generates ROS at the tumor site, inducing apoptosis and ICD via ER
676 stress and mitochondrial dysfunction. This process reprograms the TME by promoting
677 DC maturation, T-cell infiltration, and cytokine release. The integration of PDT with
678 programmed cell death protein 1 (PD-1) checkpoint blockade amplifies anti-tumor
679 efficacy, achieving maximal tumor growth inhibition and extended survival in mice.

680 TCPP-TER-Zn@RSV nanosheets (TZR NSs), a two-dimensional nanoplatform, were
681 designed to enhance the efficiency and immunogenicity of PDT while addressing the
682 immunosuppressive TME [134]. TZR NSs combine an ER-targeting PS with
683 resveratrol (RSV), which promotes autophagy and inhibits indoleamine-(2,3)-
684 dioxygenase (IND). Upon laser irradiation, TZR NSs generate ROS in the ER, inducing
685 oxidative stress. This process triggers ICD, releasing DAMPs and promoting DC
686 maturation. RSV further regulates T cell abundance, increasing the proportions of CD8⁺
687 and CD4⁺ T cells while reducing immunosuppressive Foxp3 regulatory T cells, thereby
688 reversing immunosuppressive TME and boosting anti-tumor immunity.

689 Dai et al. synthesized T-NPCe6-L-N, a tumor-targeted self-illuminating
690 supramolecular NP designed to address the limitations of conventional PDT [135]. By
691 integrating the PS Ce6, luminol, and NO donor SNAP into a self-assembled NP, this
692 system leverages high levels of tumor-specific H₂O₂ to trigger chemiluminescence
693 resonance energy transfer, enabling self-excited PDT without external light. The
694 selective activation of Ce6 in H₂O₂ and Fe³⁺-rich cancer cells ensures minimal side
695 effects on healthy tissues, whereas NO release alleviates tumor hypoxia and depletes
696 intracellular GSH levels, further amplifying the production of ROS. This dual
697 mechanism effectively suppresses subcutaneous, deep-seated, and metastatic tumors
698 while simultaneously inducing ICD for sustained anti-tumor immunity. Gao et al.
699 developed TDR848@FPB, a TME-responsive antigen-capturing nanoplatform
700 designed for systemic administration. A key point for TME modulation is that the
701 platform is selectively activated by peroxynitrite (ONOO⁻) in the tumor, where it
702 generates quinone methide traps that covalently capture TAA released after therapy.
703 Upon light irradiation, the AIE-based PS produces strong ¹O₂, induces ICD, and
704 releases R848, a TLR7/8 agonist, which helps turn the TME into a more pro-
705 inflammatory and immune-active state. Importantly, this system not only enhances
706 dendritic cell maturation and T-cell activation, but also actively reprograms suppressive
707 immune cells in the TME, including converting M2 macrophages toward the anti-tumor
708 M1 phenotype and shifting myeloid-derived suppressor cells toward antigen-presenting
709 behavior [136]. This study by Attar et al. presents a tumor-targeted self-illuminating
710 nanoplatform (T-NPCe6-L-N) that improves PDT by combining internal light
711 activation and TME modulation. Instead of relying on external light, the system uses
712 high tumor H₂O₂ levels to trigger chemiluminescence (via luminol), which activates the
713 PS (Ce6) through energy transfer, enabling effective PDT even in deep tumors. At the
714 same time, the release of NO plays a key role in TME modulation by reducing hypoxia
715 through vascular normalization and lowering intracellular GSH levels, which weakens
716 tumor antioxidant defenses and enhances ROS generation. This dual strategy allows
717 selective tumor killing, as activation mainly occurs in cancer cells with high H₂O₂ and

718 Fe³⁺ levels, minimizing damage to normal tissues. Additionally, the treatment induces
719 ICD, leading to sustained immune responses and long-term tumor suppression,
720 including effects on metastatic tumors [137].

721 Photodynamic “gel-bombs” (DCM@OPR) was designed to improve cancer therapy
722 by enhancing drug delivery and modulating the TME. These gel structures carry a PS
723 (Ce6), MnO₂ NPs, and doxorubicin within a calcium-crosslinked matrix. Upon light
724 irradiation, they generate ROS, producing explosive energy that breaks the gel into
725 small fragments, allowing deep penetration into tumor tissue through physical gaps and
726 receptor-mediated transcytosis. This mechanism ensures better distribution of
727 therapeutic agents inside tumors. Importantly, MnO₂ reacts with tumor H₂O₂ and acidity
728 to generate oxygen, helping to relieve hypoxia—a key TME barrier—and thereby
729 improving PDT efficiency. At the same time, released Mn²⁺ and doxorubicin can
730 activate immune pathways such as stimulator of interferon genes (STING), while
731 excess Ca²⁺ promotes tumor cell death [138]. Zhao et al. developed a TME-responsive
732 biomimetic nanoplatform (FBFO@HM@aOPN) designed to improve the combination
733 of PDT and immunotherapy for GBM by actively modulating the TME. After systemic
734 injection, the platform accumulates in the tumor and responds to the acidic TME, where
735 it promotes vascular normalization and ECM remodeling, enabling the NPs to penetrate
736 more effectively. At the same time, its nanozyme core generates oxygen to relieve
737 hypoxia, which enhances PDT efficiency. Importantly, the system also targets TAMs,
738 converting them from a pro-tumor state to an anti-tumor M1-like phenotype, while also
739 inducing immunogenic tumor cell death and ferroptosis, which increases neoantigen
740 release. These combined effects strengthen both innate and adaptive anti-tumor
741 immunity, making the tumor more responsive to immune checkpoint therapy such as
742 anti-PD1, and leading to stronger tumor suppression and reduced recurrence [139].

743 Xiaohui et al. synthesized a biomimetic nanosystem (G-IrC8) that combines
744 targeted PDT with TME modulation for stronger anticancer effects. The system was
745 designed by loading a flexible-chain iridium PS (IrC8) into giant plasma membrane
746 vesicles (GPMVs) derived from tumor cells, which improves biocompatibility and
747 homologous tumor targeting. Once inside tumor cells, IrC8 preferentially accumulates
748 in the mitochondria, where light activation triggers strong ROS production and efficient
749 tumor cell killing. Importantly, the treatment induces ICD, which reprograms the TME
750 by promoting antitumor immune activation [140]. A TME-responsive small molecule
751 (Ir-Fc) was designed to improve cancer treatment by combining PDT and ferroptosis
752 through effective TME modulation. The molecule contains an acid-sensitive imine
753 bond that breaks in the acidic lysosomal environment of tumors, releasing two active
754 parts: Ir-NH₂, which works as a strong PS producing type I and type II ROS, and Fc-

755 CHO, which drives Fenton reactions to generate highly toxic $\bullet\text{OH}$ and trigger
756 ferroptosis. Importantly, this process helps overcome two major TME barriers—high
757 GSH and hypoxia—because ferroptosis and Fenton chemistry consume GSH and also
758 contribute to oxygen generation, both of which enhance PDT efficiency. As a result, Ir-
759 Fc not only kills tumor cells directly but also reprograms the redox and oxygen balance
760 of the TME, creating a self-reinforcing therapeutic effect with strong anti-tumor activity
761 and good safety [141].

762 A hypoxia-responsive nanotheranostic (NanoPcN8O) was developed to address the
763 oxygen-deficient TME. This system was based on a hydrophilic phthalocyanine that
764 self-assembles into stable NPs and becomes activated specifically in hypoxic tumor
765 regions through bio-reduction, converting into an active form (NanoPcN8). This
766 transformation switches on type I photodynamic activity, generating oxygen-
767 independent radicals, along with a photothermal effect, enabling effective tumor killing
768 even under low oxygen conditions. Importantly, this design allows selective activation
769 within the hypoxic TME, as confirmed by targeted PAI, thereby minimizing damage to
770 normal tissues. The system shows significantly higher ROS generation under hypoxia
771 compared to conventional PSs and achieves strong tumor growth inhibition in vivo
772 [142]. A dual-mode nanotherapeutic system (HAP@BMPns) was developed to
773 overcome TME limitations and improve PDT outcomes. This system takes advantage
774 of the acidic TME, where the hydroxyapatite-based nanomaterial breaks down after
775 entering tumor cells, releasing Ca^{2+} and the PS. The increased intracellular Ca^{2+} causes
776 mitochondrial damage and triggers apoptosis, representing a biological effect, while
777 simultaneous activation of type I PDT under 800 nm irradiation generates oxygen-
778 independent reactive species for photochemical tumor killing. This coordinated
779 response effectively modulates the TME by exploiting its acidity and inducing
780 mitochondrial stress. As a result, the combined therapy shows much stronger tumor
781 inhibition, with significant reduction in cell survival and improved efficacy in both cell
782 and animal models [143].

783 A NIR-activated, heavy-atom-free PS system (CHL) that improves cancer treatment
784 through both phototherapy and TME modulation was recently developed. The PS Cy-
785 BF was packed into phospholipids and platelet exosome vesicles, which improved
786 tumor targeting and enabled 760 nm light-activated PTT plus type I PDT, even under
787 hypoxic conditions. After activation, CHL caused mitochondrial damage and ICD,
788 while also reducing tumor lactate production, an important metabolic factor in the TME.
789 This metabolic shift helped remodel the immune environment by decreasing regulatory
790 T cells (Tregs) and increasing CD8^+ T cells. To further overcome lactate-driven
791 immune suppression, lithium carbonate was added so that lactate could be repurposed

792 as an energy source for CD8⁺ T cells, further strengthening anti-tumor immunity [144].
793 A lactate-fueled self-acting PDT nanosystem for triple-negative breast cancer that
794 works by both killing tumor cells and modulating the TME was introduced. The
795 platform combines a PS (HPPH), luminol, and lactate oxidase (LOx) inside hollow
796 MnO₂ NPs coated with hyaluronic acid for tumor targeting. LOx first consumes excess
797 tumor lactate, converting it into pyruvate and H₂O₂, which helps reduce the harmful
798 lactate-rich, immunosuppressive TME. The generated H₂O₂ then triggers luminol
799 chemiluminescence, which activates HPPH without external light, allowing self-
800 illuminated PDT. At the same time, MnO₂ reacts in the acidic tumor environment to
801 generate oxygen, helping relieve hypoxia, a major barrier to PDT. Mn²⁺ formed during
802 this process further enhances the reaction and also enables MRI tracking [145].

803 **Image-guided PDT**

804 NIR FLI-guided PDT is promising for detecting and treating deep tumors, but its
805 effectiveness is often reduced by dye aggregation and the trade-off between
806 fluorescence and ROS production. To circumvent this, a pentamethine cyanine dye
807 (C5T) was engineered using a specially designed triphenylphosphine counterion with
808 an oligoethylene glycol chain. This design improves the dye's balance between
809 hydrophilic and hydrophobic properties and strengthens interactions within the system,
810 preventing aggregation. As a result, the modified NPs (C5T-Pco) show strong NIR-II
811 fluorescence and efficient type I ROS generation, both of which can be tuned by
812 changing the excitation wavelength. Compared to standard dyes, this system provides
813 deeper tissue imaging, higher ROS production, and better tumor targeting, including
814 mitochondrial localization [146].

815 A series of Au(I)-based complexes were designed by combining Au units with
816 special ligands, enabling a multifunctional system that integrates chemo/photo and
817 immunotherapy. Among the series of synthesized complexes, one optimized complex
818 showed strong tumor-specific imaging ability, including organelle-level targeting such
819 as the endoplasmic reticulum, allowing image-guided therapy with better precision. Its
820 anticancer effect originates from multiple coordinated actions, including inhibition of
821 thioredoxin reductase, high ROS generation, and induction of ICD, which activates
822 immune responses through stress signals and release of damage-associated molecules.
823 Both in vitro and in vivo studies confirmed that this system can effectively treat tumors
824 through image guidance, demonstrating strong theranostic potential [147].
825 A pentavalent bismuth-based nanoplatfrom (NaBiVO₃-PEG) for cancer treatment that
826 can generate ROS without requiring external light, oxygen, or hydrogen peroxide. In
827 the acidic tumor environment, the NPs undergo hydrolysis, leading to continuous
828 production of •OH and ¹O₂ through Bi(V) to Bi(III) conversion and lattice changes.

829 Concurrently, sodium ion release triggers pyroptosis and promotes strong immune
830 activation, enabling both direct tumor killing and systemic anti-tumor immunity,
831 including effects on distant tumors and metastasis. Importantly, after i.v. administration,
832 these NPs accumulate effectively at tumor sites and can be monitored in real time using
833 CT imaging, enabling image-guided therapy. The system also combines
834 immunotherapy and RT, showing strong tumor inhibition [148].

835 Treating deeply located GBM is difficult, especially when both accurate imaging
836 and effective therapy are needed at the same time. To address this, a tumor-targeted
837 europium hexaboride nanoplatform (EuB₆@RGD-K) was developed, where the RGD-
838 K peptide enables specific binding to $\alpha\beta 3$ receptors on GBM cells. This system acts
839 as a theranostic agent by combining MR imaging with NIR-II (1064 nm) and NIR-III
840 (1550 nm) light-triggered phototherapy. It allows clear tumor visualization and real-
841 time monitoring of treatment, enabling image-guided therapy with high precision. Upon
842 light activation, the NPs generate ROS and heat, producing combined PDT and PTT
843 effects. The long-wavelength NIR light enables deeper tissue penetration and non-
844 invasive treatment, reducing damage to surrounding brain tissue. In animal studies, this
845 approach significantly improved survival, especially with NIR-III irradiation. Overall,
846 this platform demonstrates a powerful image-guided strategy for precise diagnosis,
847 monitoring, and treatment of aggressive brain tumors [149]. A theranostic nanoplatform
848 (CDSP NPs) was developed by combining carbon dots with a paclitaxel prodrug. These
849 NPs provide strong NIR afterglow imaging, deep tissue penetration, and high signal
850 clarity, enabling real-time, image-guided surgical navigation for precise tumor removal.
851 In addition to imaging, CDSP NPs deliver combination therapy by generating ROS for
852 PDT and releasing paclitaxel for CMT, leading to enhanced tumor suppression [150].
853 A smart dual-activatable nano-immunomodulator (DIR NPs) was designed for cancer
854 treatment using NIR-II FLI-guided photodynamic immunotherapy. The NPs remain
855 dormant during bodily circulation but become activated in the TME under 808 nm laser
856 irradiation. Upon activation, they release a PS (DIR) and R848. The released DIR binds
857 to tumor-associated proteins, which restores strong NIR-II fluorescence for clear tumor
858 imaging and enhances ROS production for effective PDT. The released R848 boosts
859 immune responses by promoting dendritic cell activation and subsequent T-cell-
860 mediated tumor killing. This combined effect not only destroys primary tumors but also
861 suppresses distant tumors and metastasis. [151].

862 A NIR-II phototheranostic platform based on a mitochondria-targeting moiety
863 (MYM) that combines imaging and therapy for GBM treatment was designed and
864 developed. To improve delivery, MYM was loaded into exosomes and modified with
865 the iRGD peptide, allowing it to cross the blood–brain barrier and selectively

866 accumulate in tumor. The system enables multimodal NIR-II imaging, providing real-
867 time visualization of tumor location and treatment progress, supporting precise image-
868 guided therapy. Upon 808 nm laser irradiation, MYM@iRGD-Exo exhibits
869 phototherapeutic effects, leading to strong tumor inhibition and also enhances immune
870 responses by promoting T-cell infiltration and activating immune-related pathways, as
871 confirmed by RNA analysis [152]. This study presents a smart nanoplatform (CDZP
872 NPs) for breast cancer that combines sensitive miR-21 detection with enhanced PDT in
873 a single system. Based on a ZIF-8 MOF, the NPs carry both an imaging module and a
874 therapeutic module and release their components in response to the acidic tumor
875 environment. The system uses a unique signal amplification strategy to detect very low
876 levels of miR-21 with high sensitivity, enabling accurate tumor identification and real-
877 time monitoring for image-guided therapy. The released Zn^{2+} activates DNzyme that
878 suppresses GPX4, reducing the cell's ability to remove ROS, thereby boosting PDT
879 effectiveness. [153].

880 To overcome the drug leakage issues from cell membrane coated NP delivery systems,
881 Zhang et al. developed an electrostatically stabilized-light activated membrane delivery
882 platform [Hm]@NPs for pancreatic cancer therapy. [Hm]@NPs was synthesized by the
883 loading of AIE PS into a positively charged polymer with thioketal bonds vulnerable to
884 ROS and coated is with red blood cell and pancreatic cancer cell membranes. When
885 irradiated with white light, the highly stable polymer generated ROS by PDT effects,
886 which disintegrates both ROS-responsive polymer and hybrid membrane, enabling
887 drug release. In a pancreatic tumor model, [Hm]@NPs preferentially accumulated in
888 the tumor and suppressed tumor growth by FLI-guided PDT [154]. Conventional
889 fluorescence probes for early diagnosis of pancreatic cancer suffer from poor
890 penetration and accuracy. Zhu et al. developed an enzyme-activatable high contrast
891 fluorescence probe using AIE fluorophore (QM) amphiphilic peptides QM-HSP-CPP.
892 It primarily consists of QM, a hydrophobic peptide vulnerable to the enzyme cathepsin
893 E (CTSE) and a cell-penetrating peptide. CPP has the ability to modulate the molecular
894 dispersion properties that masks the fluorescence of QM in bodily circulation and due
895 to CTSE activity in the tumor tissues emanates strong fluorescence. QM-HSP-CPP
896 achieved intraoperative diagnosis of human PC sections, tracking PC in heterotopic
897 nude mouse models with high specificity and long-term tracking ability [155].
898 Conventional PDT systems cannot produce type I and type II ROS simultaneously due
899 to their oxygen dependence. Wang et al. synthesized AIE PS Pys-QM-TT with optimal
900 electronic properties capable of generating both type I and II ROS with simultaneous
901 FLI. Notably, the strong donor- π -acceptor (D- π -A) structure of Pys-QM-TT enhances
902 the ICT effect, which promotes emission in the NIR region. In addition, this structure
903 lowers the singlet-triplet energy gap, aiding the system generate ROS more efficiently.

904 In addition, the pyridinium salt group helps the excited PS transfer electrons more easily,
905 which further increases the production of type I ROS. In vivo, Pys-QM-TT
906 demonstrated the capability as image-guided PDT agent for cancer [156].

907 **Image-guided PTT**

908 In contrast to PDT, which has experienced almost 50 years of development, PTT,
909 which uses heat generated from laser irradiation of light-absorbing agents, has only
910 recently been confirmed to achieve promising photothermal ablation (PTA) of tumors.
911 Nevertheless, there have been considerable attempts to explore PTT nanomaterials,
912 with significant achievements obtained in recent years. To date, a large number of
913 photothermal nanomaterials have been explored for PTT and can be classified into the
914 following four categories: 1) noble metal-based materials (e.g., Au) [157, 158]; 2)
915 transition metal-based materials (e.g., Cu_{2-x}S , FeS, MoO_x and WS₂) [159-162]; 3)
916 carbon-based materials (e.g., carbon nanotubes, graphene, and fullerenes) [163-165],
917 and 4) organic nanomaterials (porphyrin, polypyrrole, and semiconducting polymer)
918 [166, 167]. Au nanorods exhibit much stronger NIR absorption and scattering than other
919 Au nanostructures due to their shape and aspect ratio. Smaller nanorods are efficient
920 light absorbers, while larger ones enhance scattering. Unlike spherical particles, they
921 enable uniform heat generation across the entire structure. This makes them highly
922 effective for photothermal applications [168-170]. Carbon-based nanomaterials arise
923 from the versatile bonding ability of carbon (sp , sp^2 , sp^3), leading to a wide range of
924 structures such as carbon dots, fullerenes, graphite, carbon nanotubes (CNTs), and
925 graphene derivatives [17, 171, 172]. These materials are attractive for photothermal
926 applications due to their strong light absorption, chemical stability, low density, and
927 cost-effectiveness. Their heat generation originates from excitation and relaxation of π -
928 electrons, enabling efficient light-to-heat conversion across a broad spectrum. Optical
929 and thermal properties depend on factors like size, shape, layer number, and doping, as
930 well as fabrication strategies (top-down or bottom-up). Porous and hierarchical designs
931 further enhance light absorption by reducing reflection and promoting multiple light
932 scattering. In biomedical applications, especially cancer therapy, CNTs and graphene-
933 based materials are widely explored as photothermal agents, often combined with drug
934 delivery, gene therapy, or catalytic functions to improve therapeutic outcomes [173,
935 174]. Palladium-based nanomaterials, especially ultrathin nanosheets, are promising
936 photothermal agents due to strong NIR absorption, high heat conversion, and good
937 stability. Their size and shape can be tuned to optimize optical properties [175, 176].
938 These features make them useful for cancer treatment and imaging applications [177].

939 Furthermore, phototherapy can be effectively combined with optical imaging
940 diagnostic technologies, such as FLI and PAI to realize the integration of tumor

941 diagnosis and treatment, which will significantly improve the efficiency and accuracy
942 of tumor therapy. This section highlights and discusses the recent progress in optical
943 imaging-guided PTT nanomaterials in biomedical applications, primarily comprising
944 recent developments in different modalities of imaging-guided PTT, further
945 advancements of NP-based PTT, and the potential of PTT in clinical applications in the
946 near future.

947 **Optical imaging-guided PTT**

948 With the continuous development of optical imaging technology, the biological
949 imaging window has gradually expanded from the visible (Vis) light region (400–700
950 nm) to the first NIR region (NIR-I, 700–900 nm) and the second NIR region (NIR-II,
951 1000–1700 nm). When working in the NIR window, NIR light imaging can provide
952 superior performance in spatiotemporal resolution, signal-to-noise ratio, and imaging
953 depth than Vis light imaging. In the past few years, nanomaterial-based NIR-I imaging
954 agents have been widely used in clinical settings, including clinical trials and animal
955 experiments [178, 179]. Recent studies on NIR-II have attracted considerable attention,
956 due to the lower tissue absorption/scattering and autofluorescence effects and improved
957 resolution and deeper tissue visualization compared with that of NIR-I imaging, which
958 is expected to be used in the clinic in the future. Here, we introduce the latest progress
959 in the development of NIR-I and II optical imaging agents, including FLI and PAI,
960 which may help readers grasp the trends in development and act as a possible guideline
961 for the future of optical imaging agents.

962 **NIR-I window**

963 With significant efforts dedicated to developing FLI, NIR FLI agents are
964 particularly useful in research and clinical applications, where operators can directly
965 explore the kinetics of drug movement or identify the intraoperative tumor margin.
966 However, NIR-FLI agents with the “always-on” mode, with high background signals
967 inevitably result in “false positive” signals in the complex and dynamic biological
968 milieu. Currently, activatable or smart’ FLI agents, capable of changing signals upon
969 interaction with targets of interest, can significantly reduce the background
970 fluorescence, while simultaneously meeting the requirements of absorption to generate
971 heat for PTT. Du et al. reported an “on-off” GSH-activatable photothermal agent biotin-
972 cystamine-Cys-Lys (Cypate)-CB, which was fabricated by biotin, cypate, 2-
973 cyanobenzothiazole (CBT) and disulfidelinked cysteine (Cys). In the presence of GSH,
974 the disulfide bond of biotin-cystamine-Cys-Lys (Cypate)-CB is reduced to initiate the
975 CBT-Cys click condensation reaction (intramolecular quenching) and self-assembly
976 (intermolecular quenching) into NPs, resulting in strong quenching of their

977 fluorescence due to the FRET effect. Meanwhile, photothermal efficacy of the cyprine
978 fluorophore increases significantly and its PTT efficiency on cancer cells and tumors
979 improves significantly [180]. In addition to the “on-off” mode, activatable FLI
980 characterized by the imaging signals “off/on” switch is another design strategy [181].
981 Huang et al. designed a smart “off-on” system of CyPT–AuNRs nanohybrids, which
982 combined AuNRs, heptamethine cyanine dye (CyPT), and GSH sensitive linker of
983 thiodiglycolic acid [182]. Firstly, the system would “sense” the tumor by providing NIR
984 fluorescence, simultaneously exhibiting quenching in normal cells, because the
985 fluorescent CyPT is released from the CyPT-AuNRs due to the stimulation of GSH.
986 Moreover, in this system, the free AuNRs recovered the photoconverted thermal effect
987 and can be excited by a single 808 nm laser and generate PTT and PDT effects
988 simultaneously.

989 Regarding fluorescence-guided phototherapy, it is well understood that light
990 emission and photosensitization are competing processes. Because most conventional
991 organic dyes with strong intermolecular π – π stacking, exhibit strong competition from
992 nonradiative pathways when their molecules are used as solid films or in the aggregate
993 state. Therefore, the aggregation-caused quenching (ACQ) effect severely restrains
994 their application in nanomaterial systems, because they are often used as aggregates
995 rather than in discrete states. This problem can be resolved by enhancing the AIE.
996 Compared with the planar structure of ACQ molecules, AIE agents generally have rotor
997 structures, which restricts their molecular movements in the aggregate state and avoids
998 nonradiative decay [183, 184]. Recently, Yang et al. designed an NIR AIE-active
999 probe with a donor-acceptor-donor (D–A–D) structure, T-BDP, by the conjugation of
1000 boron-dipyrromethene (BODIPY) and TPA. The AIE effect is attributed to the
1001 nonplanar configuration and three freely rotatable benzene rings. Accordingly, the
1002 amphiphilic T-BDP could spontaneously self-assemble into NPs (T-BDP NPs), which
1003 exhibited good water solubility and strong fluorescence in the NIR region with a
1004 maximum peak at around 724 nm. Furthermore, the self-assembled AIE agent T-BDP
1005 NP possessed robust photothermal conversion efficiencies (PCEs) of 50.9%. Under
1006 irradiation with a single 635-nm laser, the T-BDP NPs could generate ROS and heat
1007 simultaneously, facilitating the ablation of cancer cells through photodynamic and
1008 photothermal effects [185]. More recently, Tian et al. designed Ce6-conjugated
1009 AuNPs that demonstrated significantly improved one- and two-photon excitation fluorescence,
1010 $^1\text{O}_2$ generation, and photothermal effects. The optimum 1PE and 2PE fluorescence
1011 intensities of aggregated Ce6-AuNPs improved by 24.2- and 47.0-fold compared with
1012 those of pre-quenched Ce6-AuNPs and by 3.9- and 8.5-fold compared with that of free
1013 Ce6. Because the $^1\text{O}_2$ generation efficiency of aggregated Ce6-AuNPs is 13.4- and 2.3-
1014 times better than that of pre-quenched Ce6-AuNPs and free Ce6, respectively. The

1015 aggregated Ce6-AuNPs exhibited excellent PDT and PTT performance under 660-nm
1016 lasers (194 mW/cm²) applied below the skin tolerance threshold [184].

1017 In addition to approaches that depend on fluorescence responses, photoacoustic (PA)
1018 techniques have been incorporated into the design of precise biomedical imaging
1019 systems. During the therapeutic process of PTT, contrast agents or tissues that interact
1020 with input light would convert some of the optical energy into heat, and then ultrasound
1021 waves would be generated due to the heat-induced thermoelastic expansion. Thus, PAI,
1022 which detects phonons instead of photons after light excitation, makes PA and PTT an
1023 ideal pair for seamlessly and synergistically combining into theranostics [186]. Among
1024 photothermal agents, noble metal plasmonic nanostructures, such as Au nanomaterials,
1025 are well known to possess strong LSPR and improved PCE at the NIR light region.
1026 With numerous photothermal clinical trials currently in progress (e.g., NCT00848042
1027 and NCT01679470), the use of Au nanomaterials in PTT has established the foundation
1028 for the development of clinical applications in the near future. They also provide good
1029 chemical stability and interesting physical properties, making them powerful agents in
1030 imaging-guided PTT. For instance, El-Sayed et al. described the simulation prediction
1031 of the absorption efficiency of various Au materials. They estimated that Au
1032 nanostructures have a low luminescence quantum yield and exhibit absorption
1033 efficiency that confers large light-to-heat conversion efficiencies for PTT [187].
1034 Accordingly, Chen et al. designed biodegradable Au nanovesicle (BGV) assemblies
1035 composed of poly(ethylene glycol)-b-poly(ϵ -caprolactone) (PEG-b-PCL)-tethered
1036 AuNPs, which demonstrated LSPR absorbance in the range of 650–800 nm. The PCE
1037 was high, reaching 37%, which can be attributed to the high absorption-to-extinction
1038 ratio of small AuNPs that convert optical energy into heat with high efficiency. Results
1039 showed that tumors on mice were completely eliminated after NIR laser irradiation.
1040 Remarkably, noble metal-based nanomaterials can also act as PAI agents, whereas
1041 contrast-enhanced PAI can be used to guide and monitor therapies [188, 189].

1042 In addition to Au nanomaterials, semiconductor nanomaterials, such as
1043 chalcogenides, as well as metal oxides, are considered the most promising candidates
1044 for PAI-guided PTT. Researchers have developed several chalcogenides (e.g., Cu_xS_y,
1045 [190, 191], and Ce-doped MoO_x [192]), all of which exhibited strong LSPR in the NIR
1046 region. Therefore, porphyrin-based PTT constitutes a promising method for highly
1047 selective cancer treatment and monitoring. Furthermore, other researchers have
1048 developed efficient heating agents by constructing organic dye-based NPs. For instance,
1049 ICG [193] and IR dyes [194] have been used to combine with encapsulating polymer
1050 or protein for PTT. Among them, ICG is a Food and Drug Administration (FDA)-
1051 approved drug for a number of clinical imaging applications [195]. Tong et al.

1052 developed an improved ICG-mediated PTT platform by loading ICG into porphyrin-
1053 based covalent organic framework NPs (ICG@COF NPs). The porous COF structure
1054 allows efficient ICG loading and enhances its photostability and light absorption,
1055 resulting in a high PCE (~56.7%) under 808 nm irradiation. The composite showed
1056 stronger heat generation and better performance in killing cancer cells. Both in vitro
1057 and in vivo experiments confirmed that ICG@COF NPs can effectively ablate tumors,
1058 with near-complete tumor regression observed in 4T1 tumor-bearing mice, while
1059 maintaining good biocompatibility and minimal toxicity [196]. Several research groups
1060 have reported the use of conjugated polymers, including polyaniline [197] and
1061 polypyrrole-based NPs [198] for PTT due to their high PCE, excellent photostability,
1062 and good biocompatibility. Owing to the synthetic methods, polymer NPs could be
1063 designed to possess a configurable backbone and side chains, which endows them with
1064 tunable photophysical properties. In 2013, Levi-Polyachenko et al. designed a nano-
1065 PCPDTBT for PTT applications *in vitro* under NIR irradiation. Their pioneering work
1066 demonstrated that a donor-acceptor (D-A) structure endowed conjugated polymer NPs
1067 with a low band gap and ensured strong absorbance in the NIR window [199].
1068 Furthermore, a recent work by Pu et al. presented alternating D–A semiconducting
1069 polymers containing diketopyrrolopyrrole (DPP) moieties as the acceptor groups. Their
1070 study provided evidence of how to finely tune bandgaps by copolymerizing DPP with
1071 different electron-donating monomers.

1072 A Nd-Yb co-doped nanomaterial, termed water-heating NIR NPs, were developed
1073 for NIR imaging-guided PTT by leveraging the strong absorption of water at 1.0
1074 μm [200]. The incorporation of Tm ions extended the NIR lifetime, enabling precise
1075 tumor visualization and targeted heating in deep tissues. In a GBM multiforme (GBM)
1076 mice model, anti-CD133-conjugated NPs (Ab-NPs) improved tumor targeting by 2.4
1077 times compared to unconjugated NPs, leading to 78.9% tumor volume reduction under
1078 808 nm continuous-wave laser irradiation. Systematic in vitro and in vivo studies
1079 confirmed effective brain tumor localization, deep-tissue imaging, and PTT,
1080 demonstrating the potential of these multifunctional NPs for intracranial cancer
1081 treatment with minimal toxicity. Fe²⁺-chelated PFC-encapsulated polyepinephrine
1082 (PFC@PEPP-Fe) nanoshells were designed as a theranostic platform for dual-mode
1083 imaging and synergistic cancer therapy [201]. Structurally, the PFC core functions as
1084 an ultrasound (US) contrast agent and oxygen carrier, enhancing chemodynamic
1085 therapy (CDT) by self-supplying O₂ and H₂O₂, while the PEPP-Fe shell enables
1086 fluorescence (FL) imaging and PTT under NIR light. The NPs exhibit tumor-specific
1087 US imaging via microbubble generation and cancer-selective “OFF-ON” FL signals
1088 due to Fe²⁺ chelation. NP-mediated PTT can be optimized to prevent metastasis by
1089 precisely tuning photothermal temperature [202]. A theranostic nanoplatform,

1090 GNDs@gelatin, was developed to selectively activate NIR-II photoacoustic image
1091 (PA)-guided PTT through MMP-triggered in situ assembly. Unlike conventional
1092 "always-on" PTT agents, GNDs@gelatin offers tumor-specific activation, precise PA-
1093 based treatment planning, and efficient dual-NIR (808 nm and 1064 nm) energy
1094 harnessing. Notably, transcriptome analysis revealed that high PTT temperatures (>55
1095 °C) induced metastasis-related pathways, whereas moderate temperatures (43–50 °C)
1096 effectively ablated tumors without metastasis (**Figure 5**). This "moderate-is-better"
1097 approach demonstrated improved long-term survival in glioma models, highlighting
1098 GNDs@gelatin as a promising strategy for metastasis-free, tumor-targeted PTT. A
1099 multifunctional NIR-triggered phototheranostic agent, TTNH, was developed for dual-
1100 modal imaging and synergistic type-I PDT and PTT in hypoxic tumors. By integrating
1101 an electron-deficient 2-cyanothiazole unit, TTNH achieves efficient intersystem
1102 crossing, enabling superoxide radical ($O_2^{\bullet-}$) generation under 808 nm laser irradiation.
1103 TTNH NPs (TTNH NPs) exhibit strong NIR-II fluorescence (QY = 2.08%), excellent
1104 PCE (51.8%), and enhanced PAI capabilities. The NPs demonstrate long tumor
1105 retention (>120 h) and induce potent mitochondrial damage, promoting apoptosis in
1106 both normoxic and hypoxic conditions [203].

1107 A multifunctional image-guided phototherapy nanoplatform that integrates
1108 fluorescence and photoacoustic (PA) imaging with therapy for precise cancer treatment
1109 was developed by Kang et al. The system is built using a near-infrared AIEgen
1110 combined with a hypoxia-responsive paclitaxel (PTX) prodrug, and further coated with
1111 a macrophage cell membrane to enhance tumor targeting. The strong optical properties
1112 of the AIEgen enable high-quality fluorescence and PA imaging, allowing accurate
1113 tumor localization and real-time guidance during treatment. Upon light irradiation, the
1114 platform generates ROS, inducing PDT and ICD, while also consuming oxygen and
1115 increasing tumor hypoxia. This hypoxic condition then accelerates the release of the
1116 PTX prodrug, creating a self-enhancing therapeutic cycle. As a result, the combined
1117 effects of imaging-guided therapy, PDT, and CMT trigger strong immune responses,
1118 effectively suppressing both primary and distant tumors [204]. A TME- responsive,
1119 image-guided PTT strategy for HCC was developed by Zeng et al. It identifies
1120 ATP6V0C, a proton pump subunit, as a key driver of tumor acidity, which is closely
1121 linked to tumor growth, metastasis, and poor prognosis. To monitor this, a pH-
1122 responsive ratiometric photoacoustic sensor (PPS) was developed, enabling real-time,
1123 high-resolution imaging of tumor acidity during cancer initiation, progression, and
1124 metastasis. This imaging approach allows precise visualization of tumor boundaries and
1125 dynamic pH changes, improving early diagnosis and disease monitoring. Importantly,
1126 PPS also shows enhanced photothermal effects in acidic conditions, and when
1127 combined with a proton pump inhibitor, it achieves synergistic tumor suppression [205].

1128 Shi et al. presented a NIR FLI-guided PTT strategy using a newly designed
1129 semiconducting polymer, PF2. By introducing fluorine into the polymer structure, the
1130 researchers improved both FLI ability and heat generation, overcoming the usual trade-
1131 off between imaging and photothermal performance. Compared with its chlorinated
1132 counterpart, PF2 NPs showed stronger NIR fluorescence, a high PCE of 69.8%, and
1133 better molecular properties such as improved planarity and light absorption. These
1134 features allowed clear tumor imaging and efficient tumor heating under 808 nm laser
1135 irradiation. In 4T1 tumor-bearing mice model, PF2 NPs achieved strong tumor
1136 suppression (about 80% inhibition) with good biocompatibility and safety [206]. Zhao
1137 et al. developed a multifunctional, image-guided PTT nanoplatfrom, called ASNP NPs,
1138 to improve anti-tumor immunity. The system combines a NIR photothermal agent with
1139 sodium nitroprusside (SNP) inside heat-sensitive liposomes. Under NIR laser
1140 irradiation, ASNP NPs generate heat for PTT and simultaneously release SNP, which
1141 reacts with GSH to produce NO and Fe²⁺. The Fe²⁺ further reacts with H₂O₂ to
1142 generate •OH, promoting ferroptosis. This three-way treatment-PTT, NO gas therapy,
1143 and ferroptosis—greatly increases the release of DAMPs, which helps reprogram the
1144 tumor immune microenvironment and activate stronger tumor-killing immunity. In
1145 addition, the NPs show strong fluorescence, allowing better imaging guidance during
1146 treatment. In a 4T1 breast tumor model, this strategy significantly suppressed tumor
1147 growth [207].

1148 **NIR-II window**

1149 As well documented over the past several decades that most photoinduced
1150 theranostics operate in the NIR-I window, numerous photoinduced agents in the
1151 traditional NIR window have been extensively investigated in both animal and clinical
1152 applications. Nonetheless, there remains much room for improvement. In particular,
1153 endogenous optical species, including Hb and deoxyhemoglobin, exhibit light
1154 absorption in the NIR-I region and, combined with light scattering within the same
1155 spectral region, have contributed to significant attenuation of excitation light, which in
1156 turn has led to a decrease in imaging sensitivity, spatial resolution, and tissue
1157 penetration depth. To further increase tissue penetration depth, techniques using longer
1158 wavelengths have also been developed to overcome the limitation of optical imaging
1159 depth. In 2009, Smith et al. proposed the second “transparent window” with a longer
1160 wavelength from 1000 to 1700 nm, which is also known as the NIR-II window [208].
1161 Compared with that in the NIR-I window, optical imaging in the NIR-II window can
1162 yield significantly deeper penetration depth, with good temporal and spatial resolutions
1163 and minimized autofluorescence [209]. Using a single component, Ag₂S QDs with a
1164 high fluorescence quantum yield (15%) in the NIR-II range have been reported to detect

1165 tumors with a high signal-to-background ratio through passive tumor targeting [210].
1166 Subsequently, both *in vitro* and *in vivo* NIR-II imaging results were achieved using
1167 Ag₂S QDs, which were found to be attractive NIR-II fluorescence probes with good
1168 biocompatibility and high quantum efficiency. In addition to the individual NIR-II FLI
1169 function, Ag₂S QDs can exhibit photothermal characteristics, which have been used for
1170 image-guided PTT [211-213]. For instance, Liu et al. used the reversal of ACQ strategy
1171 to design a pH-responsive photothermal NP for FLI-guided PTT. In their work, the
1172 stimulation-responsive assembled Ag₂S vesicle (Ag₂S Ve) was proposed by the self-
1173 assembly of Ag₂S QDs coated with pH-sensitive copolymer thiolated polystyrene-co-
1174 poly(4-vinylpyridine). The Ag₂S Ve demonstrated strong fluorescence quenching in the
1175 second NIR-II region. Triggered by the acidic environment, transformation of Ag₂S
1176 QDs from aggregated to disaggregated states promptly mediated the quenched NIR-II
1177 fluorescence from “off” to “on.” Due to these properties, the theranostic Ag₂S Ve can
1178 be specifically activated in acidic tumor tissues, whereas it remains nonfunctional in
1179 normal tissues. Besides the above-mentioned types of photothermal agents, imaging-
1180 guided PTT was successfully realized in tumor-bearing mice using several different
1181 types of NIR-II absorbers, including semiconducting polymer NPs [214], lanthanide
1182 NPs [215], Au nanostructure [216], silicon oxide NPs (H-SiO_x NPs) [217], semimetal
1183 NPs [218], metal carbide [219], etc. All these systems demonstrated efficient anticancer
1184 ability. Especially, semiconducting polymer NPs are not only capable of photothermal
1185 heating and fluorescence/PA imaging of tumors but also possess good biocompatibility
1186 and biodegradability, thus providing great promise for translational medicine.

1187 A novel approach to tumor treatment using genetically engineered *Escherichia coli*
1188 MG1655 (MG1655-M) that biosynthesize melanin NPs was developed [220]. These
1189 bacteria target hypoxic tumor regions, where they effectively colonize and deliver
1190 melanin. This delivery can be tracked in real-time using PAI, allowing precise timing
1191 for PTT. Upon exposure to an 808-nm laser, the melanin induces localized heating,
1192 resulting in significant tumor ablation (**Figure 6**). Moreover, the PTT triggers strong
1193 anti-tumor immune responses, creating long-term immune memory that helps prevent
1194 tumor metastasis and recurrence. This integrated platform provides image-guided,
1195 tumor targeting therapy with combined photothermal and immune treatment, presenting
1196 a promising strategy for future cancer therapies. A novel hydrogen-based cancer
1197 therapy using diameter and shape-controlled magnesium (Mg) NPs, including
1198 hexagonal nanosheets, nanoflowers, and small NPs, were investigated. Among them,
1199 pH-sensitive polymer-coated Mg nanoflowers (MgNF@PEG/PMMVP) demonstrate
1200 significant potential in NIR-II PAI and bubble-enhanced ultrasound imaging (USI) for
1201 cancer treatment [221]. These nanoflowers, approximately 100 nm in size, have a
1202 unique structure that allows strong NIR-II absorption, making them effective imaging

1203 contrast agents. When introduced into the acidic tumor environment, the polymer shell
1204 disassembles, causing a reaction between Mg and water that generates hydrogen (H₂)
1205 bubbles. These bubbles improve ultrasound imaging signals, induce cavitation that
1206 ruptures lysosomes, and disrupt cellular energy metabolism, leading to oxidative stress
1207 and cancer cell death. This drug-free method effectively combines bubble burst-like
1208 action with H₂ therapy, resulting in significant tumor inhibition with minimal side
1209 effects and good biosafety. The approach holds promise for clinical applications due to
1210 its selective targeting and non-invasive nature.

1211 However, unsatisfactory imaging depth and signal-to-background ratio remain
1212 intrinsic issues for clinical application. A few recent studies have explored the use of
1213 lasers in optical subwindows such as 1300–1400 nm (termed the NIR-IIa window) and
1214 1500–1700 nm (termed as the NIR-IIb window) for the better performance of FLI. For
1215 instance, Wu et al. reported a PEG-stabilized copper sulfide NP (CuS NP) that
1216 possesses a broad absorption range, and was used as a PTT agent and excited by either
1217 808- or 1275-nm laser. This multifunctional CuS NP not only serves as a contrast agent
1218 for NIR-IIa PAI but also functions as a photothermal agent for PTT. Moreover, the
1219 1275-nm laser possesses deeper tissue penetration capability than the 808-nm laser. The
1220 authors also demonstrated that the temperature increments in the CuS-PEG NP solution
1221 irradiated by the 1275-nm laser at depths of 2, 5, 10, 15, and 20 mm were 10.5-, 9.1-,
1222 6.5-, 6.2-, and 4.8-fold higher than those observed with the 808-nm laser [222]. Besides,
1223 semiconducting polymer NPs (SPNs) have been widely used as NIR-II PAI or PTT.
1224 Zhang et al. used diketopyrrolopyrrole and benzobisthiadiazole (BBTD) as two
1225 acceptors and constructed D-A1-D-A2 semiconducting polymer NPs for NIR-IIa PAI-
1226 guided chemo-photothermal combination therapy of cancers. BBTD exhibits desperate
1227 electron-withdrawing capability in D-A polymers and decrease the bandgap to drive the
1228 absorption spectra to the longer wavelength. Therefore, their established two-acceptor
1229 semiconducting polymers (SP1-3) with the strongest electron deficient acceptor not
1230 only exhibited a high photothermal conversion efficiency (60%) at 1064 nm, but also
1231 as an excellent PAI agent with the strong photoacoustic signal at 1280 nm [223]. Due
1232 to the lack of endogenous fluorescent molecules emitting beyond 1400 nm, the NIR-
1233 IIb window provides near-zero autofluorescence and negligible scattering, thus
1234 providing an optimal imaging platform for *in vivo* research [224, 225]. To date, there
1235 have been several efforts toward the design and synthesis of nanomaterials with longer
1236 and brighter NIR-IIb imaging. For instance, Huang et al. designed a quantum-dot-based
1237 nanoprobe with broad NIR absorption and emission tailing beyond 1500 nm and used
1238 it for imaging-guided PTT [226]. A novel small-molecule NIR-IIb dye IT-TQF with a
1239 D–A–D structure was synthesized by Cheng et al. The IT-TQF NP-based NIR-IIb
1240 imaging exhibited high spatial resolution and high tissue penetration depth, and the

1241 extended imaging wavelengths improved the tumor signal-to-background ratio to 9.42
1242 in orthotopic osteosarcoma models. Importantly, the IT-TQF NPs displayed high PTT
1243 efficacy (PCE: 47%) for effective tumor treatment in mice [227]. Recently, Wang et al.
1244 reported a TME-responsive hollowed virus-bionic MnO₂ nanoshell (HvMnO₂@QDs-
1245 IR1061), which loaded IR1061 in the cavity and anchored QDs (PbS@CdS) on the
1246 surface. The NIR-IIb fluorescent signal in the HvMnO₂@QDs-IR1061 can be
1247 completely quenched in normal tissues via absorption competition-induced emission
1248 mechanism. Subsequently, the pH of the TME triggered NIR-IIb fluorescence recovery
1249 and exerted efficient NIR-II photothermal effect for lymphatic metastasis under NIR-
1250 IIb fluorescent imaging navigation [228]. Despite the large number of strategies used
1251 for realizing NIR-IIb imaging-guided PTT, the major absorption peaks are concentrated
1252 in the NIR-I region. In general, high excitation power or exposure time is required in
1253 the tail NIR-II fluorescence bioimaging. Small organic molecules generally display low
1254 fluorescence brightness in water due to the lipid-soluble characteristics. Moreover, the
1255 major peaks of absorption/emission rarely reach beyond 1300 nm. Therefore, the
1256 development of high-brightness and long-wavelength emissive fluorophores remain a
1257 major research challenge.

1258 NP-based PTT has been extensively investigated recently, and the development of
1259 depth-independent imaging-guided multifunction nanoplatfoms has been well
1260 documented in several reviews. Being realistic, the maximum optical penetration depth
1261 achievable in human tissues is within the range of a few centimeters. Therefore,
1262 therapeutic light-triggered PTT is incompatible with depth-limitless imaging
1263 technology for preclinical applications and clinical translation. Instead, several research
1264 groups intend to explore novel light delivery systems to circumvent the major limiting
1265 issue of deep-tissue treatments. NP-based photothermal treatment of deep-seated
1266 tumors is still achievable if an optical fiber delivery is used in combination with depth-
1267 limitless imaging-guided techniques. Recently, Ma et al. [229] introduced transvascular
1268 interventional PTT (Ti-PTT), a minimally invasive approach that integrates
1269 endovascular delivery with PTT for precise tumor treatment. A small microcatheter (1.8
1270 Fr) equipped with an ultrafine optical fiber (100 μm) was used to simultaneously deliver
1271 ICG-based photothermal agents and 808 nm laser irradiation directly through blood
1272 vessels. Two formulations were investigated: ICG solution for direct photothermal
1273 heating and ICG–ethiodized oil (ICG–EO) emulsion, which improves retention and acts
1274 as a photothermal embolic agent. Under laser activation, localized heat induces
1275 embolization, preventing blood supply and causing tissue necrosis even beyond the
1276 normal light penetration depth. The ICG–EO formulation showed better retention and
1277 more effective embolization than free ICG. In animal models, Ti-PTT achieved precise
1278 vascular occlusion and strong therapeutic effects, and its parameters can be adjusted

1279 based on vascular structure. Huang et al. developed a catheter-based iPTT system that
1280 combines real-time imaging and therapy using a single 1310 nm light source for precise
1281 treatment of orthotopic colorectal cancer (CRC). A miniaturized catheter (0.9 mm) with
1282 an optical fiber enables simultaneous optical coherence tomography imaging and PTT,
1283 allowing accurate tumor localization and controlled heat delivery. Folate-modified
1284 Bi/Bi₂S₃ NPs act as both imaging enhancers and photothermal agents, showing strong
1285 NIR-II absorption, scattering signals, and efficient heat generation due to localized
1286 surface plasmon resonance, along with effective tumor targeting. In vitro and in vivo
1287 studies demonstrated effective tumor ablation, high imaging contrast, and minimal
1288 toxicity, confirming good biosafety [230]. To date, AuNP-based functionalized stents
1289 have been introduced for the local treatment of cancer cells or tissue hyperplasia
1290 adjacent to stented nonvascular luminal organs [231-233]. For instance, Hu et al.
1291 developed a localized PTA therapy for obstructive rectal cancer, which operates
1292 through AuNP-coated stent with NIR irradiation. Aided by the excellent NIR
1293 absorption properties of branched AuNPs, the nanotherapeutic stents displayed
1294 effective suppression of tumor growth after stent placement by heat-induced tumor
1295 necrosis. In the same year, Cho et al. used the same concept in a rat esophagus model.
1296 Their results demonstrated that the AuNP-coated stent-mediated local PTT protocol
1297 could be used to treat not only granulation tissue formation but also tumor ingrowth or
1298 overgrowth through the stent meshes. Although further preclinical studies are required
1299 to investigate the efficacy and safety of localized heat treatment, the developed nano-
1300 functionalized SEMS and localized PTA therapy should be promising for clinical
1301 applications.

1302 A multifunctional bionic nanoplatform (Au@MnO₂@PM; AMP) was developed
1303 for multimodal imaging and NIR-II PTT of lung cancer [234]. The system integrates
1304 Au nano-bipyramids (Au NBPs) for photothermal conversion and MnO₂ for enhanced
1305 MRI, with a homologous cancer cell membrane coating for precise tumor targeting.
1306 AMP enables CT, photothermal imaging (PTI), and MRI, while achieving high
1307 photothermal conversion efficiency (52.07%) under 1064 nm irradiation. In vitro and
1308 in vivo studies confirmed effective tumor ablation with no significant systemic toxicity.
1309 The tumor-homing ability and microenvironment-responsive MnO₂ release further
1310 enhance imaging and therapeutic efficacy, highlighting AMP as a promising
1311 nanotheranostic for precise cancer diagnosis and treatment. A next-generation
1312 photothermal agent, Stealth NanoBomb (SNB), was developed to enhance mild-
1313 temperature (<45 °C) PTT by inhibiting HSPs expression in cancer cells. SNB consists
1314 of self-assembled small molecular NIR-II AIEgens and a CO carrier polymer
1315 (PLGA(CO)) coated with PEG-lipid, ensuring stability and safe circulation for
1316 intravenous administration [235]. Upon tumor accumulation, SNB responds to the

1317 overexpressed H₂O₂ in the TME, releasing CO gas to suppress HSPs and enhance PTT
1318 efficacy. Compared to previous nanobomb generations, SNB offers improved safety,
1319 stability, and targeted CO gas/drug co-delivery, representing a promising strategy for
1320 more effective, low-toxicity cancer therapy. A novel small-molecule phototheranostic
1321 agent, CY-1234, was developed for NIR-II PAI-guided PTT with excellent
1322 biocompatibility and tumor-targeting capabilities [236]. CY-1234, featuring an
1323 extended π -conjugation structure, exhibits strong NIR-II absorption at 1234 nm and is
1324 nanoencapsulated (CY-1234 NPs) for aqueous solubility and in vivo stability. These
1325 NPs achieve high photothermal conversion efficiency (76.01%) under 1064 nm laser
1326 irradiation, leading to efficient tumor ablation with a 97% apoptosis rate in HeLa cells.
1327 In vivo studies confirmed strong PA signals, effective tumor inhibition, and no systemic
1328 toxicity. A bismuth-doped iron selenide (BFS) NPs platform was developed to enhance
1329 CDT and second NIR-II PTT for CRC. BFS NPs exhibit high photothermal conversion
1330 efficiency (31.9%) and accelerate the Fenton reaction under NIR irradiation, increasing
1331 ROS generation and improving CDT efficacy [237]. Additionally, bismuth doping
1332 enables T₂-weighted MRI and CT imaging, providing precise guidance for NIR-II PTT.
1333 The synergistic CDT/PTT approach effectively inhibits tumor growth while
1334 minimizing side effects.

1335 As discussed earlier, a large number of nanomaterials have been developed as NIR-
1336 absorbing photothermal agents to serve as heat carriers for local hyperthermia. Local
1337 hyperthermia in tumors can not only directly eradicate cancer cells but also exert
1338 additive effects that improve the effect of conventional PTT. For instance, PTT has
1339 been combined with CMT to compensate for each other's limitations, e.g., poly(N-
1340 isopropylacrylamide) (PNIPAM), a well-known thermally responsive polymer in
1341 which temperature-dependent swelling and shrink phase transition behavior can drive
1342 actuated spatiotemporal drug release under NIR irradiation. Moreover, several research
1343 groups constructed numerous photo-responsive components for synergistic PTT/CMT,
1344 including PNIPAM derivative copolymer, poly(ϵ -caprolactone), and poly(vinyl alcohol)
1345 [238-240]. In addition to the spatiotemporal- and dose-controlled delivery of CMT for
1346 improved therapeutic efficacy and reduced systemic side effects, thermal-sensitive
1347 materials were successfully realized in a biodegradable nanoplatform to accelerate NPs
1348 elimination from the living body. Specifically, the subunit thermal-sensitive polymer-
1349 tethered NPs self-assemble into a larger formulation. With laser irradiation, when the
1350 thermal-sensitive polymer exhibits structural damage or change, the building blocks
1351 dissociate into their original sizes, thereby facilitating their elimination from the body
1352 [241, 242]. The degradation and clearance process of self-assembled formulation
1353 through the hepatobiliary and renal route was demonstrated, encouraging the translation
1354 of NP-based PTT to clinical applications [243, 244].

1355 In addition to the direct efficacy of NP-based PTT, some studies have focused on
1356 the indirect influence of NP-based hyperthermia to tackle the surrounding TME
1357 sequentially and then the core tumor cells [245]. Conversely, in terms of tumor vessels,
1358 local hyperthermia could be exploited for increasing the vascular permeability to result
1359 in better extravasation (100 nm) of NPs when the heating temperature range is 40 °C–
1360 42 °C. In contrast, the effect of PTT also influences tumor stroma in which temperature
1361 increase affects the collagen architecture and then remodels the ECM. For instance, the
1362 research groups of Bhatia and Gazeau confirmed that PTT may serve as a nano-heater
1363 to induce localized denaturation of collagen, especially in the study of Gazeau, through
1364 local denaturation of adjacent tissue. In particular, on collagen fibers, light-exposed
1365 multiwalled carbon nanotubes induce remodeling and softening of the TME
1366 concomitant to tumor regression [246, 247]. Remarkably, the structural change of TME
1367 after local heating, which favored further penetration of NPs and drug, has the potential
1368 to improve the efficacy of current treatments [248]. **Table 2** shows the summary of NIR
1369 I and II image-guided PTT nanosystems.

1370 Jiang et al. developed an image-guided PTT nanomotor for treating superficial
1371 tumors through peritumoral subcutaneous injection. The nanomotor consists of a
1372 polydopamine (PDA)-coated spherical core loaded with ICG, allowing it to convert
1373 NIR-I light into both heat and NIR-II fluorescence. Under laser irradiation, the
1374 generated heat not only enables tumor-killing PTT, but also creates a thermophoretic
1375 driving force that actively pushes the nanomotor through subcutaneous tissue toward
1376 the tumor. At the same time, the NIR-II fluorescence provides real-time imaging
1377 guidance, allowing the movement of the nanomotor to be tracked and directed
1378 accurately. The heat also helps reduce tissue barriers by loosening fat and connective
1379 tissue, improving delivery efficiency [249]. Shi et al. reported a NIR FLI-guided PTT
1380 platform based on a newly designed photothermal agent, 3TPA. The molecule was
1381 engineered using a CF₃-BODIPY core with triphenylamine (TPA) rotors, which
1382 enhances light absorption and promotes efficient heat generation through strong
1383 nonradiative decay, resulting in a high photothermal conversion efficiency (~57%).
1384 When encapsulated into a biocompatible polymer (DSPE-mPEG), the resulting 3TPA
1385 NPs (3TPA NPs) show improved tumor targeting, stability, and photostability
1386 compared to conventional dyes. Importantly, these NPs enable dual NIR-I/NIR-II
1387 fluorescence imaging, allowing precise visualization and guidance during therapy.
1388 Under 808 nm laser irradiation, 3TPA NPs rapidly raise tumor temperature to effective
1389 levels for ablation, leading to near-complete tumor elimination with no recurrence in
1390 animal models, while maintaining good biosafety[250]. Patnaik et al. developed a Au-
1391 coated solid lipid NP (Au-SLN) as a promising image-guided PTT agent for breast
1392 cancer. The NPs showed very high photothermal conversion efficiency (about 80%),

1393 When exposed to laser, Au-SLNs rapidly raised tumor temperature to above 60 °C,
1394 enabling effective thermal ablation. Importantly, treatment response was monitored
1395 using multiple non-invasive imaging methods, including infrared thermography for
1396 real-time heat tracking, bioluminescence imaging (BLI) and NIR fluorescence for
1397 tumor activity, and microCT for anatomical and volumetric assessment. This
1398 multimodal imaging approach allowed precise evaluation of both treatment
1399 effectiveness and healing [251].

1400 A smart image-guided phototherapy platform, called BOD-D was developed by Sun
1401 et al., which can switch treatment modes during therapy to improve tumor killing.
1402 Initially, BOD-D works as a NIR-I FLI-guided PDT agent, producing $^1\text{O}_2$ to destroy
1403 tumor cells. As treatment continues and the tumor becomes more hypoxic, PDT
1404 becomes less effective, so light activation triggers BOD-D to release NO and convert
1405 into BOD-T, a photothermal agent with NIR-II fluorescence. This allows the therapy
1406 to shift from NIR-I-guided PDT to NIR-II-guided PTT. Importantly, the released NO
1407 also acts as gas therapy and helps sensitize both PDT and PTT, making the overall
1408 treatment stronger [252]. Chen et al. introduced a novel image-guided NIR-II PTT
1409 based photoimmunotherapy strategy using a membrane-anchoring small molecule,
1410 CBT-3, that directly disrupts cancer cell membranes. The molecule was designed to
1411 attach to tumor cell membranes and, under NIR-II laser irradiation (1064 nm), generates
1412 a mild photothermal effect (~ 43 °C) that rapidly induces plasma membrane rupture and
1413 necrotic cell death. This approach offers strong spatiotemporal control and works
1414 efficiently at relatively low doses. Importantly, the induced cell damage triggers ICD
1415 and activates inflammatory pathways, reshaping the TME and enhancing immune cell
1416 infiltration. As a result, both local tumor destruction and systemic anti-tumor immune
1417 responses are achieved [253]. Chen et al. designed and developed a TME-responsive,
1418 degradable phototheranostic nanoplatform, CX@PSS, for NIR-II FLI-guided PTT. The
1419 system uses a GSH-sensitive polyurethane carrier loaded with a multifunctional NIR-
1420 II cyanine PS (CX), allowing it to specifically degrade in the GSH-rich TME, which
1421 may help reduce long-term toxicity. Under 808 nm laser irradiation, CX@PSS provides
1422 high-resolution NIR-II FLI for precise tumor visualization while simultaneously
1423 generating heat for PTT and ROS for PDT. Importantly, this combined phototherapy
1424 also induces ICD, which releases tumor-associated antigens and activates anti-tumor
1425 immunity. Both cell and animal studies showed strong tumor suppression, good
1426 biosafety, and promising potential to reduce tumor metastasis and recurrence [254].

1427 **Design principles of PTT agents for optimal performance**

1428 The photothermal effect in plasmonic NPs is primarily governed by factors that
1429 determine how efficiently absorbed light is converted into heat. A key parameter is the

1430 LSPR, which occurs when the wavelength of incident light matches the natural
1431 oscillation frequency of electrons in the NPs, resulting in strong light absorption and
1432 efficient heat generation. The efficiency of this process is further influenced by NPs
1433 characteristics such as size, shape, composition, and the dielectric properties of the
1434 surrounding environment [17, 18]. For example, smaller particles often favor
1435 absorption-dominated processes, while anisotropic structures such as nanorods can
1436 enhance photothermal conversion through stronger plasmonic oscillations. In addition,
1437 optical parameters including the absorption cross-section and the ratio of absorption to
1438 total extinction determine the level of incident energy is converted into heat rather than
1439 scattered [255]. For biomedical applications, the excitation wavelength is particularly
1440 important. Early photothermal approaches were limited because visible light is strongly
1441 absorbed and scattered by biological tissues, restricting treatment mainly to superficial
1442 tumors and causing unwanted heating of nearby healthy tissue. This limitation has been
1443 largely addressed by using NIR light within the biological windows ($\approx 700\text{--}980$ nm and
1444 $\approx 1000\text{--}1400$ nm), where tissue absorption and scattering are lower, allowing deeper
1445 light penetration and more efficient activation of NPs within tumors. By localizing heat-
1446 generating NPs in tumor tissues and using appropriate laser wavelengths, stronger and
1447 more selective heating can be achieved with lower laser power, reducing damage to
1448 surrounding healthy tissue [256, 257]. Although light penetration in human tissue is
1449 still limited to a few centimeters, deeper tumors can be treated by combining NPs
1450 delivery with optical fiber or endoscopic illumination, enabling localized photothermal
1451 heating at the tumor site.

1452

1453 **Radionuclide theranostics**

1454 Currently, PDT and PTT have demonstrated the potential for effective cancer
1455 therapy in preclinical studies. Specifically, an external beam is used to irradiate the
1456 tumor site to achieve PDT and PTT. A critical challenge that must be overcome in
1457 clinical development is the penetration depth. Currently, RIT is a mature technology
1458 for the clinical treatment of cancer by self-emitted radiation. Targeted delivery of
1459 radionuclides to the tumor remains a limitation of RIT. Radioisotope-related
1460 theranostics, such as α - and β -emitters, as well as CR and proton therapy, will be
1461 discussed in the following section.

1462 **Radionanotheranostics of α - and β -emitters**

1463 The high linear energy transfer and the short path length of α -emitters enable them
1464 to destroy local tumor cells. These properties are suitable for the treatment of small cell
1465 clusters and micrometastases. Targeted alpha particle therapy (TAPT) provides more

1466 precise tumor cell killing and reduces damage to the surrounding normal tissues. An α -
1467 emitting radionuclide, such as ^{213}Bi [258, 259] and ^{149}Tb [260], is labeled with a
1468 monoclonal antibody or targeting ligand to achieve TAPT.

1469 Targeted alpha therapy, a potent modality in nuclear medicine, uses nuclides that
1470 emit alpha particles to exert localized therapeutic effects. Nevertheless, radionuclides
1471 such as ^{223}Ra and ^{225}Ac decay into radioactive progeny, which may leave the original
1472 vector and cause irradiation in non-target organs. A previous study investigated the fate
1473 of $^{211}\text{Pb}/^{211}\text{Bi}$ progeny from ^{223}Ra surface-labeled TiO_2 NPs *in vitro* and in animal
1474 model [261]. Stability studies demonstrated that up to 40% of ^{211}Bi and 25% of ^{211}Pb
1475 progeny were released within 48 h under static conditions, with biodistribution data
1476 confirming these findings *in vivo*. The ^{211}Pb activity was still detectable in blood and
1477 kidneys, supporting the theory that daughter radionuclides from alpha decay can
1478 migrate to non-target organs, thus increasing radiation exposure. Although some studies
1479 suggest that progeny remain in target tissues once internalized, this is not always the
1480 case, especially considering factors such as lesion size and tumor perfusion. Therefore,
1481 it is crucial to analyze the fate of recoils from alpha decay cascades when introducing
1482 new radiopharmaceuticals. Further studies are required to quantify progeny release and
1483 migration, although preliminary findings support the hypothesis that significant
1484 portions of daughter progeny can be transported to nontargeted organs through the
1485 bloodstream. Auger emitters, with their high LET that causes clustered DNA damage
1486 and efficient cell death, are ideal for targeting metastasized tumors. Despite their
1487 potential, few radiopharmaceuticals using Auger emitters exist due to the need for these
1488 electrons to reach critical cell components such as the nucleus. One study combined the
1489 Auger emitter ^{125}I with ultrasmall AuNPs to create a novel radiopharmaceutical [262].
1490 These ^{125}I -labeled NPs accumulated in the cell nucleus, demonstrating high tumor-
1491 killing efficiency in 2D and 3D models. These findings suggest that ultrasmall NPs,
1492 which naturally accumulate in the nucleus, could improve targeted radionuclide therapy,
1493 especially when combined with tumor targeting agents. This new radiopharmaceutical
1494 approach could provide significant advancements in cancer treatment.

1495 Internal α -therapy is effective for micrometastatic diseases, with ^{224}Ra being a
1496 promising candidate due to its multiple α -particle emissions and a half-life of 3.6 days
1497 [263]. However, the use of ^{224}Ra has been limited to bone-seeking applications because
1498 it cannot be stably bound to targeting molecules. A previous study explored calcium
1499 carbonate microparticles as carriers for ^{224}Ra , aimed at treating cancers in cavitary
1500 regions such as peritoneal carcinomatosis. The ^{224}Ra -labeled CaCO_3 microparticles
1501 demonstrated high labeling efficiencies and retention of both ^{224}Ra and its daughter
1502 nuclide ^{212}Pb for up to 1 week *in vitro*. Biodistribution studies in mice revealed that the

1503 radioactivity predominantly remained in the peritoneal cavity, with reduced skeletal
1504 uptake of ^{224}Ra at higher microparticle doses. These results suggest that ^{224}Ra -labeled
1505 CaCO_3 microparticles could be a promising new method for localized α -therapy in
1506 cavitory cancers. PCa, the second most common cancer in men, often progresses to
1507 metastatic castration-resistant prostate cancer (mCRPCa), necessitating new targeted
1508 therapies.

1509 Hydroxyapatite and TiO_2 NPs are promising candidates for carrying medicinal
1510 radionuclides due to their large surface area, radiation stability, and low toxicity. In one
1511 study, they were radiolabeled with $^{99\text{m}}\text{Tc}$ for diagnostics and ^{223}Ra for therapy using
1512 two methods, surface labeling and intrinsic incorporation. Both methods achieved
1513 $>94\%$ labeling yields. Stability tests in various biological media revealed that ^{223}Ra -
1514 labeled TiO_2 NPs had the best stability, with $<6\%$ activity release over 59 h and $<3\%$
1515 over 55 days [264]. Both $^{99\text{m}}\text{Tc}$ -labeled NPs demonstrated approximately 20% activity
1516 release in short-term tests. These data suggest that although HAp is better for local
1517 applications and controlled release, TiO_2 is more suitable for systemic applications due
1518 to its superior long-term stability. Surface modifications could further improve the
1519 stability of these radiolabeled NPs. An injectable brachytherapy hydrogel based on Fe-
1520 tannin NPs was developed, allowing stable and chelator-free radiolabeling of clinically
1521 used radionuclides (^{131}I , ^{90}Y , ^{177}Lu , ^{225}Ac). Crosslinking FeTA NPs with 4-arm PEG-
1522 SH created a biocompatible hydrogel capable of prolonged tumor retention and real-
1523 time in vivo monitoring via MRI signals from embedded ferric ions. In a CT26 colon
1524 cancer model, the ^{225}Ac -labeled hydrogel effectively eradicated local tumors and, when
1525 combined with immunoadjuvant imiquimod and anti-PD-L1 therapy, significantly
1526 inhibited metastatic tumor growth by stimulating robust anti-tumor immune responses
1527 without adverse effects [265].

1528 A multifunctional nanoprobe ($^{177}\text{Lu}\text{-Fe}_3\text{O}_4\text{@HA/DBCO}$) was developed to enhance
1529 bladder-preserving treatment of bladder cancer by integrating mucosal penetration,
1530 bioorthogonal targeting, and internal irradiation therapy [266]. Iron oxide NPs coated
1531 with HA and dibenzocyclooctyne effectively penetrated mucosa and selectively
1532 accumulated in azide-expressing cancer cells via bioorthogonal click chemistry (**Figure**
1533 **7**). Enhanced MRI distinguished non-muscle-invasive from muscle-invasive bladder
1534 cancers (MIBC), while internal irradiation by ^{177}Lu significantly reduced tumor size.

1535 A multifunctional black phosphorus-based nanosheet (BPNS) system was
1536 developed that integrated Cu^{2+} to achieve both rapid degradation and enhanced
1537 photothermal stability [267]. This BP@Cu structure exhibited superior PTT
1538 performance, enabled CDT by generating $\bullet\text{OH}$, and facilitated real-time PET imaging
1539 using $^{64}\text{Cu}^{2+}$. The approach demonstrated effective tumor-targeting capability and

1540 introduced a versatile nanoplatform for PET-guided, CDT-enhanced photothermal
1541 cancer therapy. A polymersome-based nanocarrier was developed to modulate innate
1542 immunity by efficiently targeting splenic red pulp myeloid cells, significantly reducing
1543 tumor growth in a mice melanoma model [268]. Utilizing ^{89}Zr -radiolabeling and PET
1544 imaging, large spherical polymersomes rapidly accumulated in the spleen, effectively
1545 delivering β -glucan to hematopoietic organs. Preclinical biodistribution studies in non-
1546 human primates confirmed splenic targeting and biocompatibility, demonstrating their
1547 translational potential for cancer immunotherapy. GSH-modified GNCs labelled with
1548 radionuclides ($^{99\text{m}}\text{Tc}@Au$ NCs and $^{177}\text{Lu}@Au$ NCs) were synthesized via a simple
1549 chelation strategy, significantly enhancing internal radionuclide therapy (RNT)
1550 efficacy. While $^{99\text{m}}\text{Tc}@Au$ NCs enabled both imaging and radiosensitization, the
1551 therapeutic radionuclide $^{177}\text{Lu}@Au$ NCs effectively induced ICD, activating DC and
1552 synergizing with anti-PD-L1 checkpoint blockade to suppress distant and
1553 spontaneously metastatic tumors [269]. Radiolabeled, urease-powered mesoporous
1554 silica nanobots were evaluated in an orthotopic mice model of bladder cancer,
1555 demonstrating significantly enhanced tumor accumulation and penetration. Using PET
1556 imaging with ^{131}I , nanobots achieved approximately eightfold higher tumor localization,
1557 validated by ex vivo optical imaging methods [270]. This increased accumulation
1558 enabled effective radionuclide therapy, reducing tumor size by roughly 90% at notably
1559 lower therapeutic doses.

1560 Wang et al. developed a radionuclide-assisted chemodynamic therapy platform,
1561 ^{125}I -MIL-88B(Fe), to improve cancer treatment in pancreatic tumors. The system
1562 consists of iron-containing metal-organic framework NPs that can concentrate
1563 hydrogen peroxide and promote the Fenton reaction to generate highly toxic $\bullet\text{OH}$. The
1564 attached radioiodine-125 continuously produces hydrated electrons in the tumor
1565 environment, which help convert Fe^{3+} to Fe^{2+} . By accelerating this iron redox cycling,
1566 the NPs produce more ROS and achieve stronger tumor cell killing than conventional
1567 CDT alone. Both cell and animal studies confirmed enhanced anti-tumor effects, mainly
1568 through ROS-triggered activation of the MAPK/p53 apoptosis pathway [271].

1569 A new radioembolic agent, ^{131}I -labeled methacrylated gelatin microspheres (^{131}I -
1570 GMs) were developed for transcatheter arterial radioembolization (TARE) in HCC.
1571 These particles are biodegradable, elastic, and highly uniform in size, which is
1572 important for safe and effective arterial delivery. They were synthesized using a
1573 microfluidic method that allows precise size control, making them potentially useful
1574 for personalized radionuclide therapy. In animal studies, the Ms showed good
1575 radioactive stability, remained well retained in the hepatic artery and tumor tissue, and
1576 significantly slowed liver tumor progression [272]. Jia et al. developed a

1577 multifunctional NPs system, ^{131}I -BaGdF₅@PDA-CDDP, to combine transarterial
1578 chemoembolization (TACE) and TARE for improved treatment of HCC. The NPs carry
1579 cisplatin for CMT and iodine-131 for radionuclide therapy, allowing simultaneous local
1580 PTT and CMT directly inside the tumor after injection through the hepatic artery. This
1581 is important because RNT can help kill residual tumor cells that often survive after
1582 embolization and contribute to recurrence or metastasis. The BaGdF₅ core also provides
1583 CT/MRI capability, while ^{131}I enable SPECT imaging, allowing real-time monitoring
1584 of NP distribution and treatment delivery in vivo. In animal models, this combined
1585 approach reduced tumor metabolism and increased tumor cell apoptosis more
1586 effectively than single treatments alone [273].

1587 Shao et al. developed ^{177}Lu -labeled porous granular hydrogels as a new RNT
1588 platform for liver tumors. These injectable particles combine embolization and local
1589 internal radiation, while also allowing SPECT imaging for treatment tracking.
1590 Compared with conventional radioactive Ms, they showed stable radionuclide loading,
1591 controllable size, good tumor retention, and reduced reflux into non-target tissues. In
1592 animal models, they achieved strong tumor inhibition with good safety, highlighting
1593 their promise for precise intra-arterial brachytherapy [274]. Xiao et al. developed
1594 ^{177}Lu -labeled polymeric Ms for RNT of liver cancer through image-guided intra-arterial
1595 brachytherapy. ^{177}Lu provides both local radiation treatment and SPECT imaging,
1596 allowing real-time tracking after delivery. The Ms showed stable radionuclide loading,
1597 uniform tumor embolization, strong anti-tumor effects, and good safety without off-
1598 target embolization [275].

1599 Li et al. developed a multifunctional biodegradable microsphere (^{131}I -ICT/R848-
1600 MS) for HCC that combines transarterial radioembolization with CMT and immune
1601 activation. The RNT component, iodine-131, is important because it delivers local
1602 radiation to kill tumor cells and induce ICD, while also enabling SPECT/CT imaging
1603 to monitor microsphere retention and distribution after treatment. The Ms also carry
1604 icaritin, which further promotes tumor cell death, and R848, an immune stimulator that
1605 enhances dendritic cell activation and strengthens T-cell-mediated anti-tumor
1606 immunity. In tumor models, the Ms showed strong tumor-specific retention, robust
1607 immune activation, and improved anti-tumor effects without relapse [276]. This study
1608 by Zhao et al. highlights a promising RNT strategy using ^{177}Lu -LNC1004, a fibroblast
1609 activation protein (FAP)-targeted radiopharmaceutical designed to deliver radiation
1610 directly to tumors and remain there for longer periods. In preclinical models, ^{177}Lu -
1611 LNC1004 showed strong tumor uptake and anti-tumor activity, however it also
1612 temporarily increased PD-L1 expression, suggesting that tumors may activate immune
1613 escape pathways after radiation. To address this, it was combined with anti-PD-L1

1614 immunotherapy, which resulted in complete tumor eradication and long-lasting
1615 immune memory in mice. Mechanistic analyses showed that this combination reshaped
1616 the TME by enhancing CD8⁺ T-cell activity, increasing M1 macrophages, improving
1617 immune communication, and broadening T-cell receptor diversity. Early clinical data
1618 also suggested that ¹⁷⁷Lu-LNC1004 is safe, well-tolerated, and potentially effective in
1619 patients with advanced FAP-positive cancers [277].

1620 Zhang et al. developed a novel RNT strategy using a ²²³Ra/Ba single-atom
1621 nanozyme to induce tumor cell senescence and enhance anticancer immunity. The
1622 radionuclide ²²³Ra was delivered efficiently using a barium-based nanozyme carrier,
1623 which also has enzyme-like catalytic activity that increases ROS production, leading to
1624 tumor cell damage and senescence. However, senescent cells can promote tumor
1625 recurrence, the treatment was combined with anti-PD-L1 immunotherapy inducing
1626 senescence with radionuclide therapy, then eliminating those cells through immune
1627 activation. This combination significantly suppressed both primary and distant tumors
1628 while improving immune response [278]. Pei et al. inactivated *Salmonella* and labeled
1629 it with iodine-131 (¹³¹I-VNP) as a tumor-targeting carrier for RNT. The bacteria helped
1630 retain radioiodine in tumors, improve internal radiation delivery, and activate both
1631 innate and adaptive immunity. Importantly, treatment stimulated the innate immune
1632 pathway, promoted dendritic cell maturation, and enhanced T-cell responses. When
1633 combined with anti-PD-L1 therapy, it not only controlled primary tumors but also
1634 suppressed distant tumors and helped prevent recurrence, showing strong potential for
1635 radio-immunotherapy [279].

1636 ATP-responsive nanoassembly, USINAs (¹³¹I-aPD-L1), was developed by Shen et
1637 al. to improve RNT by combining ferroptosis, radiopharmaceutical therapy, and
1638 immunotherapy in one platform. This system exhibited good stability in blood and then
1639 selectively disintegrated in the ATP-rich TME, releasing ultrasmall iron NPs and ¹³¹I-
1640 labeled anti-PD-L1. The iron NPs induce ferroptosis, while the ¹³¹I component delivers
1641 targeted radiation and also blocks PD-L1, creating a dual radioimmunotherapy effect.
1642 Importantly, RNT and ferroptosis reinforced each other: radiation increased ROS and
1643 sensitized tumors to ferroptosis, while ferroptosis made tumor cells more vulnerable to
1644 radiation. This combination also triggered strong ICD, promoted dendritic cell
1645 maturation, enhanced T-cell responses, and supported SPECT imaging for treatment
1646 monitoring [280].

1647 **Radionanotheranostics of Cerenkov radiation (CR)**

1648 Currently, PET, SPECT, and CT are general translational imaging systems to
1649 perform radio nanotheranostics in clinical settings. The high-energy radionuclides

1650 release three signals, viz., gamma energy for imaging systems and redshifted emission
1651 and blue light for optical imaging and PDT [281]. One of the signals, blue light,
1652 constitutes a novel avenue for basic and applied science by combining advances in
1653 nanotechnology and materials science.

1654 CR, also termed Cherenkov luminescence light (CLI), is an optical light originating
1655 from energetic radionuclides, especially β -emitters. In general, CR can be used to
1656 initiate photodegradation of compounds and generate fluorescent emission for imaging
1657 [282]. CR signals are tissue-depth-dependent and entirely restricted by background
1658 light. To improve the intensity of CR signals, radiopharmaceutical-excited FLI [283],
1659 secondary Cherenkov-induced FLI [284], Cherenkov radiance energy transfer (CRET)
1660 [285], radioluminescence imaging [286], and radionuclide energy transfer [287] have
1661 been validated. A novel therapeutic approach was developed to improve the
1662 effectiveness of CR-PDT and minimize its side effects using ^{131}I -labeled ALA-loaded
1663 EMs (^{131}I -EM@ALA) [288]. This multifunctional platform was designed to target
1664 tumors through the EPR effect and homologous targeting, where the abundant
1665 mitochondria would convert ALA into the active PS PpIX, generating ROS under CR
1666 stimulation. The combination of ^{131}I RT and CR-PDT was anticipated to exert strong
1667 anti-tumor effects with minimal adverse effects due to the low accumulation of PpIX
1668 in normal tissues. Results showed that a single injection of ^{131}I -EM@ALA significantly
1669 inhibited tumor growth in subcutaneous tumor-bearing mice, and the strategy
1670 effectively reduced treatment side effects compared with that using PpIX directly. This
1671 study presents a promising multitherapeutic strategy for anti-tumor treatment,
1672 combining RT and CR-PDT without the need for an external light source.

1673 The therapeutic efficacy of traditional PDT is often limited by the shallow
1674 penetration of external light. To address this problem, CR has been investigated as an
1675 alternative; however, existing type I PSs for CR-induced PDT (CRIT) have significant
1676 limitations. An innovative approach by engineering $\text{NH}_2\text{-Ti}_3\text{O}_2$ nanocluster-derived
1677 ultrasmall nano-PSs (TDPs) using dopamine ligands was designed and developed by
1678 Li et al. [289]. These ligands improved water solubility, photocatalytic properties, and
1679 tumor targeting through dopamine receptor binding on cancer cells. Under CR
1680 irradiation, TDPs effectively generate $\bullet\text{OH}$ by separating electron-hole pairs, resulting
1681 in improved type I PDT and augmented CDT. The use of CR not only promotes CRIT
1682 but also improves CDT, demonstrating significant *in vitro* and *in vivo* anti-tumor
1683 properties with minimal side effects. Another study by Guo et al. integrates
1684 nanotechnology with nuclear medicine, thereby advancing CR-induced combined
1685 therapy and nanocatalytic medicine. The combination of Ce6@GEV and ^{18}F -FDG
1686 significantly reduced cell viability from 88.02% to 23.79%, demonstrating effective

1687 CL-induced PDT [290]. The Ce6@GEV + ¹⁸F-FDG treatment group achieved the
1688 highest tumor inhibition rate of 58.02% and the longest survival rate of 35 days (40%)
1689 compared with other groups. This approach, by combining Ce6 and ¹⁸F-FDG, aims to
1690 address the limitations of traditional PDT and may extend its application to various
1691 tumors. In addition, NPs were combined with CR to improve diagnosis and theranostics.
1692 CR-induced PDT can kill cancer cells, and the therapeutic types can be divided into the
1693 following three approaches: 1) free radionuclides combined with drug-loaded NPs, 2)
1694 radionuclide-modified NP-doped PS, and 3) radionuclides combined with NPs. First,
1695 CR generated from ¹⁸F radionuclides activate tumor-surrounding photosensitive drugs
1696 to induce cell death [291] (**Figure 8**). Moreover, chemotherapeutics transforms to
1697 radiotherapeutics for the precise treatment of disseminated cancer. The strategy rescues
1698 abandoned photosensitive drugs with poor therapeutic outcomes into precision
1699 phototherapeutics.

1700 Goel et al. developed a multifunctional core–satellite nanostructure by combining
1701 copper sulfide (CuS) NPs with [⁸⁹Zr]-labeled hollow mesoporous silica nanoshells
1702 filled with porphyrin molecules to improve cancer imaging and therapy [292]. It enables
1703 simultaneous tetra-modal imaging (PET, fluorescence, Cherenkov luminescence, and
1704 CR energy transfer) for accurate tumor detection and multimodal image-guided therapy.
1705 The synergy between CuS-mediated PTT and porphyrin-mediated PDT resulted in
1706 complete tumor eradication within a day, with no recurrence. The study emphasizes a
1707 versatile approach to creating high-performance core–satellite nanohybrids, which can
1708 be tailored for various imaging and therapeutic applications. Cherenkov imaging is
1709 advancing the use of radiotracers and therapeutic agents in nuclear imaging by shifting
1710 Cherenkov light to the red-light region using europium oxide NPs. Traditional CR is
1711 limited by its UV/blue emission, which is poorly penetrative *in vivo*. To address this
1712 problem, Zhang et al. developed ultrasmall Eu₂O₃ NPs with improved brightness
1713 through a down-conversion technique involving europium oxide and trimethylamine
1714 N-oxide [293]. These NPs, functionalized with polyethylene glycol and radiolabeled
1715 for intravenous injection, enabled more effective cancer imaging in mice, including
1716 lymph node and tumor visualization. The high density of emitters and efficient
1717 fluorescence of europium make it ideal for improving Cherenkov light and achieving
1718 sensitive, multimodal imaging. This approach not only improves the *in vivo* imaging
1719 capabilities of europium-based NPs but also lays the groundwork for further
1720 development of smaller, biocompatible particles for potential clinical applications and
1721 advanced diagnostic technologies.

1722 Radionuclides have distinct patterns of emission for imaging and RT. Some CRIT
1723 involves radiation emission from α - and β -emitters, as well as a contrast agent for

1724 SPECT or PET imaging, and can serve as radiotheranostics for the precise treatment of
1725 cancer. Furthermore, the low range and high energy of α -emitters can treat lung tumor
1726 colonies. β -emitters can treat a wide range of cancers, such as hepatocarcinoma and
1727 ovarian and head and neck cancers. CR is an optical light generated from energetic
1728 radionuclides and restricted by tissue depth. The participation of NPs can improve CR
1729 application from diagnosis to theranostics. In addition to treating solid tumors, CR-
1730 combined NPs have the potential to treat disseminated cancers.

1731 Su et al. introduced a strategy using CR from the clinical radiotracer [^{68}Ga]Ga-FAPI
1732 as an internal light source to activate TiO_2 NPs for CR-mediated PDT (CR-PDT). Since
1733 [^{68}Ga]Ga-FAPI specifically targets CAFs, it enables localized CR generation within
1734 tumors. The activated NPs produce ROS that damage both tumor cells and CAFs,
1735 reduce collagen formation, and affecting the ECM structure. This remodeling relieves
1736 vessel compression, improves drug penetration, and enhances CMT delivery.
1737 Additionally, CR-PDT consumes oxygen, increasing tumor hypoxia, which further
1738 activates hypoxia-sensitive drugs for greater therapeutic effect [294]. Zhu et al.
1739 developed a biohybrid system made of the probiotic bacterium *E. coli* Nissle 1917
1740 loaded with an aggregation-induced emission PS. After administration, these
1741 biohybrids preferentially accumulate in tumors with the tumor-seeking
1742 radiopharmaceutical ^{18}F -FDG, whose CR activates the PS directly inside the tumor.
1743 This internal activation triggers CR-PDT, leading to tumor cell damage and ICD.
1744 Importantly, the bacteria also act as immune stimulators, improving DC maturation and
1745 $\text{CD}8^+$ T-cell activation. As a result, the combination of CR-PDT and immune activation
1746 produced much stronger tumor inhibition and longer survival than CR-PDT alone,
1747 while maintaining good safety [295].

1748 A tumor-selective CR-PDT system (^{89}Zr -ALA-Liposome-ART) that uses ^{89}Zr as an
1749 internal CR source to activate protoporphyrin IX (PpIX) inside tumor cells was
1750 developed by Liu et al. The acidic tumor environment triggers ALA release, while the
1751 conversion of PpIX to heme is further used to activate artemisinin. This design
1752 improves the selectivity and effectiveness of CR-based therapy, leading to strong tumor
1753 inhibition and enhanced anti-tumor immunity. When combined with anti-PD-L1, it also
1754 helped suppress tumor recurrence [296]. Jo et al. developed a precise Cerenkov
1755 luminescence energy transfer-based PDT for HER2-positive cancers by combining two
1756 clinically used components: 5-aminolevulinic acid (5-ALA) and ^{64}Cu -DOTA-
1757 trastuzumab. After administration, 5-ALA is selectively converted inside cancer cells
1758 into the PS protoporphyrin IX (PpIX), while ^{64}Cu -DOTA-trastuzumab specifically
1759 accumulates in HER2-expressing tumors. When both localize in the same tumor,
1760 Cerenkov light emitted by ^{64}Cu excites PpIX directly inside the cancer, producing a

1761 highly tumor-selective PDT effect [297]. Rosenkrans et al. developed semiconducting
1762 polymer NPs (SPNs) to improve CR-induced therapy (CRIT). Since CR is a weak but
1763 internally generated light source, the SPNs were designed to efficiently capture and
1764 amplify it, then transfer the energy to a PS for stronger PDT effects. The NPs also
1765 showed good tumor uptake on PET and optical imaging, supporting both therapy and
1766 imaging. [298]. Herein, we list α -emitters, β -emitters, and CR to compare their
1767 characteristics (**Table 3**).

1768 **Comparison of proton therapy and intensity-modulated radiation therapy** 1769 **(IMRT)**

1770 RT is commonly applied to cancerous tumors due to its ability to control cell growth.
1771 Radionuclides that emit gamma rays or positrons can provide diagnostic information
1772 and identify the functions of specific organs and are also used for tumor therapy by
1773 systemic or targeted circulation. Compared with radionuclide-associated
1774 nanotheranostics, proton therapy is another method of external beam RT that utilizes
1775 ionizing radiation in clinical settings [299, 300]. Proton therapy represents a type of
1776 radiation that uses a particle, the proton, to deliver radiation while minimizing the dose
1777 to nearby organs. Protons have mass and a positive elementary charge, and with high
1778 momentum to a specific depth that depends on both the initial speed of the protons and
1779 the density of tissue. At that specific depth, protons rapidly decelerate, and this rapid
1780 deceleration deposits the dose with a steep falloff phenomenon, known as the Bragg
1781 peak [301]. These charged particles damage the DNA of cells, ultimately killing the
1782 cells by blocking reproduction. Cancerous cells are particularly vulnerable to attacks
1783 on DNA because of their high rate of division and reduced ability to repair DNA
1784 damage. The physical processes produced by protons that generate positron emitters
1785 can be monitored during or after irradiation by PET. Furthermore, CR is used to
1786 measure the nuclear cross-sections of protons, and research suggests that the method is
1787 convenient and widely applicable for high-precision proton therapy [302]. To improve
1788 the effect of proton therapy, metallic NPs could be used to augment efficacy through
1789 the particle-induced X-ray emission effect. The metallic NPs generate secondary
1790 electrons and characteristic X-ray to cause tumor cell death after proton irradiation. In
1791 addition to therapeutic improvement, secondary radiation also exhibited high contrast
1792 in a multispectral imaging system [303]. In clinical settings, the latest generation of
1793 proton treatment methods is known as intensity-modulated proton therapy (IMPT),
1794 which adjusts the precision, depth, and intensity of a proton beam to complex spider-
1795 like tumors, simultaneously avoiding healthy tissue. Compared with RT, IMPT
1796 significantly reduces the dose in surrounding normal tissues [304]. Although proton
1797 therapy is a good method to treat patients with cancers, there are three major challenges,

1798 viz., 1) its limited availability, which may delay or preclude treatment for patients who
1799 require expedited treatment, 2) the high cost of proton therapy, i.e., some insurance
1800 companies may not approve payment of this treatment, and 3) inapplicability to patients
1801 with disseminated tumors. Because of these challenges, other methods of radiation
1802 therapy are more widely utilized.

1803 Proton therapy combined with bismuth NPs (Bi NPs) was evaluated for enhanced
1804 treatment of deeply-seated, unresectable tumors [305]. Bi NPs synthesized via pulsed
1805 laser ablation in liquids and coated with Pluronic-F127 polymer showed good colloidal
1806 stability and moderate toxicity below 25 $\mu\text{g/mL}$. At the Bragg peak, proton irradiation
1807 significantly increased apoptosis, disrupted cell membranes, reduced clonogenic
1808 activity up to 97%, and notably suppressed primary tumor growth by 60% and
1809 metastatic spread in S37 sarcoma-bearing mice, demonstrating substantial promise for
1810 improved cancer RT. Proton-based RT combined with noble metal NPs was evaluated
1811 to determine factors influencing ROS generation. Ligand-free colloidal platinum (Pt)
1812 and gold (Au) NPs, synthesized without organic stabilizers to avoid interference, were
1813 irradiated, generated ROS, which depend mainly on total particle surface area rather
1814 than mass or size [306]. Pt NPs consistently exhibited higher ROS yields compared to
1815 Au and Au-Pt alloy NPs, emphasizing the significant role of surface atom chemistry in
1816 ROS generation. These findings clarified that surface characteristics of noble metal NPs
1817 substantially influence proton-induced ROS production.

1818 Simulations evaluating GNPs as RSs in proton therapy revealed that ROS generated
1819 upon irradiation diffused several hundred nanometres away, maximizing enhancement
1820 around 50 nm from the NPs, particularly towards the proton source [307]. Although
1821 proximity between NPs significantly elevated local doses, it also reduced overall
1822 reactive species yield by up to 60% due to increased absorption. These findings
1823 emphasized balancing dispersed NP distributions to enhance total radiolysis yield with
1824 clustered arrangements strategically positioned near biological targets, guiding the
1825 optimal use of GNPs in proton therapy (**Figure 9**). A recent study evaluated GNPs as
1826 RSs in proton beam therapy (PBT) by investigating their ability to enhance ROS
1827 generation in tumor cells irradiated with a 230 MeV proton beam at the spread-out
1828 Bragg peak. Proton irradiation triggered electron emission from GNPs, leading to
1829 increased ROS production and severe mitochondrial and cytoskeletal damage within 48
1830 h post-irradiation, resulting in a radiosensitization enhancement factor of 1.24 at 30%
1831 cell survival after 8 days [308].

1832 Lee et al. developed a three-dimensional (3D) spheroid-based high-throughput
1833 screening platform to evaluate how head and neck cancer cells respond to PBT and drug
1834 combinations. Using two cancer cell lines, the system allowed precise delivery of

1835 different proton doses and testing of multiple drugs at the same time. The results
1836 showed that both proton radiation and certain drugs reduced tumor cell viability in a
1837 dose-dependent manner. Importantly, combination therapy with olaparib showed a
1838 strong synergistic effect [309]. Tudor et al. showed that iron oxide NPs loaded with
1839 doxorubicin can sensitize chondrosarcoma cells to proton therapy, and can overcome
1840 chemo- and radioresistance. The combination reduced cell survival, increased DNA
1841 damage, and was more effective with low-LET protons than with high-LET protons. It
1842 also caused measurable changes in cell and nuclear features, suggesting possible
1843 markers for treatment response [310]. Silva et al. showed that porphyrin-coated Au NPs
1844 can improve treatment of triple-negative breast cancer by combining PDT with RT.
1845 After light activation, they increased ROS and $^1\text{O}_2$, disrupted mitochondrial function,
1846 inhibited thioredoxin reductase, and enhanced cancer cell killing. The combination
1847 reduced tumor cell survival more effectively than radiation alone, with proton therapy
1848 showing the strongest effect [311].

1849 IMRT is an advanced mode of high-precision photon RT that uses computer-
1850 controlled linear accelerators to safely and painlessly deliver precise radiation doses to
1851 a malignant tumor or specific areas within the tumor. IMRT allows for the radiation
1852 dose to conform more precisely to the three-dimensional shape of the tumor by
1853 modulating or controlling the intensity of the radiation beam in multiple small volumes.
1854 IMRT delivers the same photons to treat a tumor but with the potential to lower the
1855 high doses of radiation received by healthy structures. Currently, IMRT is being
1856 employed most extensively to treat PCa, the head and neck, and the central nervous
1857 system. IMRT has also been used in limited situations to treat breast, thyroid, lung, as
1858 well as in gastrointestinal, gynecologic malignancies, and certain types of sarcomas.
1859 IMRT may also be beneficial for treating pediatric malignancies [312]. The
1860 radiotherapeutic management of cancers is rapidly changing, with IMRT now being the
1861 standard of care. Moreover, AuNPs are used to increase the therapeutic effect of IMRT
1862 and improve X-ray imaging in brain and PCa [312, 313]. Metallic NPs are agents that
1863 can potentially improve diagnostic imaging and IMRT and selectively improve RT
1864 effectiveness. In clinics, the challenges of IMRT include disseminated tumor treatment
1865 and that a larger range of normal organs receive a low dose of radiation.

1866 A clinical trial evaluated NBTXR3 (Hafnium oxide NPs) in combination with
1867 external beam radiotherapy in patients with advanced soft tissue sarcoma prior to
1868 surgical intervention. Of twenty-two patients who finished therapy were monitored for
1869 a duration of up to 40 months. Tumors diminished by approximately 40% at elevated
1870 dosages. Results indicated that NBTXR3 was safe, effective in diminishing tumor size,
1871 and potentially suitable for preoperative use in advanced sarcoma patients. Preoperative

1872 chemo-RT, a standard treatment for locally advanced rectal cancer, was administered
1873 in conjunction with PEP503 (NBTXR3), which has demonstrated the ability to maintain
1874 therapeutic efficacy at diminished radiation doses [315]. A prospective, single-arm
1875 Phase Ib/II clinical trial evaluated the safety and efficacy of PEP503 administered via
1876 intratumoral injections in conjunction with concurrent chemoradiotherapy in patients
1877 without metastatic disease. In Phase Ib, dose escalation established the appropriate
1878 dosing, whereas Phase II assessed anti-tumor response rates. This groundbreaking study
1879 confirmed PEP503's ability to safely enhance the effectiveness of radiation in advanced
1880 rectal cancer treatment. The Act.In.Sarc trial tested NBTXR3, combined with RT in
1881 patients with locally advanced sarcoma [316]. Results showed the formulation
1882 significantly improved complete tumor removal and did not increase serious side effects
1883 compared to radiation alone. Treated patients had fewer serious adverse events and
1884 reported better quality of life scores. NBTXR3 was tested for the first time in a patient
1885 with non-operable PDAC. The NPs were safely injected into the tumor using an
1886 endoscope, followed by RT (45 Gy delivered in 15 fractions) [317]. The treatment
1887 caused no immediate side effects, and imaging showed effective local tumor targeting.
1888 These initial results indicated that endoscopic delivery of NBTXR3 with radiation was
1889 safe and feasible, offering a potential new approach for treating patients who cannot
1890 undergo surgery. A phase I study tested the safety of the radioenhancer NBTXR3 in
1891 patients with advanced tumors, given by injection at increasing doses followed by RT.
1892 NBTXR3 was safe, causing no serious side effects, and patients tolerated it well, even
1893 at higher doses. The study did not reach a maximum tolerated dose, and early signs
1894 suggested that NBTXR3 could effectively slow tumor growth. These results supported
1895 further testing of NBTXR3 at a recommended dose combined with RT [318].

1896 Chajon et al. evaluated NBTXR3, a radioenhancer injected directly into liver tumors,
1897 in combination with stereotactic body RT for HCC and liver metastases. NBTXR3 was
1898 designed to improve the effect of radiation specifically within the tumor. In this trial,
1899 the treatment showed a manageable safety profile, with no dose-limiting toxicities and
1900 no treatment-related deaths. Most serious side effects were not directly linked to
1901 NBTXR3, and the recommended dose was established at 42% of the gross tumor
1902 volume. Early results also showed encouraging anti-tumor activity, with objective
1903 responses seen in about half of treated lesions in both liver cancer and metastatic cases
1904 [319]. We summarize proton therapy and IMRT with NPs for theranostics in **Table 4**.
1905 Some NPs participate in proton therapy and IMRT to improve localized therapeutic
1906 efficacy in preclinical studies.

1907

1908 **Advancing cancer immunotherapy with NPs**

1909 NP based phototherapies, including PDT and PTT, provide several advantages,
1910 such as minimal invasion, high efficacy, and minimal side effects, compared with
1911 traditional anti-tumor therapies. PDT and PTT ablate tumors through ROS and heat.
1912 Moreover, the consequent tumor cell necrosis or apoptosis after PDT or PTT provides
1913 a potential antigen pool to induce an immune response [320, 321]. Compared with other
1914 anti-tumor therapies, such as PDT, PTT, and CMT, the target in cancer immunotherapy
1915 is not tumor cells directly but rather the body's immune system. The tactics of cancer
1916 immunotherapy include triggering the natural capacity of the immune system to attack
1917 tumor cells. Studies in cancer immunology and cancer immunotherapy, including
1918 artificial antigen-presenting cell-based therapy [322], and cell-based therapies, such as
1919 DC, adoptive T-cell transfer, and checkpoint blockade therapy [323-325], have
1920 suggested promising clinical applications in the past decade, especially for checkpoint
1921 blockade therapy. For instance, in 2018, a few Nobel laureates established a whole new
1922 strategy for cancer therapy. James P. Allison investigated a known protein, termed
1923 cytotoxic T-lymphocyte-associated antigen 4 (CTLA-4) [326], and Tasuku Honjo
1924 found a protein, called PD-1 [327], on immune cells. Both these proteins operate as a
1925 brake on the immune system, but with a different mechanism. They demonstrated a
1926 different operation by releasing these brakes on the immune system, which could trigger
1927 immune cells to attack tumors [328]. Such improvement has provided various
1928 combined treatments, such as PTT and immunotherapy. PTT not only ablates the tumor
1929 by hyperthermia-induced cell apoptosis and necrosis but also triggers immunotherapy
1930 to treat both primary and metastatic tumors by killing tumor cells, releasing TAA and
1931 then inducing a systemic anti-tumor immune response. In fact, this method is already
1932 being used in preclinical and clinical studies [329].

1933 **NPs combined with ICB**

1934 Ionizing RT is a functional tool for local tumor treatment, but it is restricted to
1935 symptom mitigation. Recently, a crucial approach to improve cancer treatment has been
1936 to combine RT with checkpoint blockade to induce not only a local tumor treatment
1937 effect but also systemic regression of metastatic disease, known as the abscopal effect
1938 [330]. Further research has demonstrated that when anti-CTLA-4 antibody is combined
1939 with RT for tumor treatment, fractionated, rather than a single-dose, RT induces higher
1940 efficacy of the abscopal effect [331]. For successful checkpoint blockade therapy
1941 against cancer, it is necessary to reduce immune-related cytotoxicity. Targeted delivery
1942 of checkpoint inhibitors to specific locations is also critical. Numerous NPs have been
1943 investigated to provide controlled release to the tumor site. Several preclinical studies
1944 have shown that NP-based targeted delivery of anti-CTLA-4 antibody augments tumor
1945 destruction efficacy and reduces cytotoxicity. Nevertheless, there are still no clinical

1946 studies to estimate the NP-mediated delivery of checkpoint inhibitors combined with
1947 RT. NP-mediated ICB treatment combined with RT may provide another opportunity
1948 to achieve optimal anti-tumor efficacy with specific targeting and decreased
1949 cytotoxicity [332].

1950 In traditional cancer CMT, the most crucial problem is tumor metastasis due to its
1951 significant effects and restricted efficacy [333]. A probable option is to trigger the
1952 immune system to treat metastatic tumors. NPs combined with immunoadjuvants not
1953 only induce an innate immune response but also increase immune cell infiltration in the
1954 TME to release inflammatory cytokines, such as IL-6, IL-12, IL-1 β , TNF- α , and IFN-
1955 γ [334]. PTT-induced hyperthermia can inhibit tumor growth and metastasis by
1956 upregulating the expression of HSPs [335]. NPs-based PTT can achieve a higher
1957 concentration of particle accumulation in the tumor site because of active or passive
1958 targeting and kill tumors via a thermal effect. Cell debris and TAA are released and
1959 further activate the immune system to kill metastatic tumors. Nevertheless, a major
1960 challenge exists not only in PTT but also in immunotherapy. Specifically, tumor cells
1961 are tolerant to the immune system under certain conditions. Some anti-tumor negative
1962 immune regulatory signals, such as CD47 signal regulatory protein- α (SIRP α) on
1963 macrophages [336], CTLA-4 on regulatory T cells [337], and PD-1 on T cells [338],
1964 block the immunotherapy of immune cells, which causes treatment failure. Therefore,
1965 PTT combined with immunotherapy can improve treatment efficacy. For instance,
1966 Chen et al. developed a combined adjuvant NPs-mediated PTT with checkpoint
1967 blockade immunotherapy. ICG, a photothermal agent, and a toll-like receptor seven
1968 agonist (R837), were loaded in PLGA, which, when combined with anti-CTLA-4
1969 exerted a markedly suppressive effect not only on primary tumors but also on distant
1970 tumors. Furthermore, this combined treatment resulted in long-term survival by
1971 inducing the memory of T cells, thus inhibiting tumor recurrence. Anti-PD-L1
1972 combined with AuNS-mediated PTT was found to be an effective treatment for primary
1973 tumors and distant tumors via heat ablation-induced immune response in a bladder
1974 cancer animal model [339]. In recent years, a large number of clinical trials have
1975 demonstrated that combined treatment with anti-CTLA-4 and anti-PD-1 can achieve
1976 better treatment efficacy than monotherapy of anti-CTLA-4 or anti-PD-1 in sarcoma
1977 and melanoma [340, 341].

1978 PDT that kills cancer cells using PSs to generate ROS under irradiation is a valid
1979 therapeutic modality that provides minimal invasiveness [6]. PDT can induce anti-
1980 tumor immune responses by producing tumor-derived protein antigens from dying
1981 tumor cells. Nevertheless, traditional PSs used in PDT have limited tissue penetration
1982 depth because of the short excitation wavelength. Although PDT may induce immune

1983 responses, such effects are generally not sufficiently powerful for inhibiting tumor
1984 growth after PDT [342]. PDT causes Treg cell infiltration, which causes overexpression
1985 of PD-L1 to inhibit tumor-killing by immune cells, such as T lymphocytes [343].
1986 Therefore, combination with ICB therapy to improve the anti-tumor immune response
1987 of PDT constitutes a better strategy.

1988 A multifunctional NP system (PIC), composed of polylysine, iron oxide, and CpG,
1989 was designed to enhance the in-situ vaccine effect of radiation therapy (RT) and
1990 improve tumor response to ICB. RT alone has limited efficacy in optimizing tumor
1991 antigen presentation and modulating the tumor immune microenvironment, but the
1992 addition of PIC increases antigen presentation, shifts TAMs towards an M1-dominant
1993 phenotype, and stimulates a type I interferon response. In immunologically “cold”
1994 murine tumor models, the combination of RT, PIC, and ICB significantly improves
1995 tumor control, extends survival, and induces tumor-specific immune memory. This
1996 approach offers a scalable and readily translatable strategy to transform tumors into
1997 sites of adaptive immune activation, enhancing responsiveness to ICB and potentially
1998 other immunotherapies. Given the broad applicability of RT and PIC across solid tumor
1999 types, further preclinical and clinical investigations are warranted to evaluate their
2000 potential in metastatic cancer treatment [344]. A nanomaterial-based approach to
2001 enhance PDT and anti-tumor immunity using a 3-bromopyruvate (BrP)-anchored
2002 nanoscale metal–organic layer (BrP@MOL) [345]. By inhibiting mitochondrial
2003 respiration and glycolysis, BrP@MOL reprograms tumor metabolism, increasing
2004 oxygen availability and reducing lactate production, thereby alleviating hypoxia and
2005 immunosuppression in the TME. This metabolic modulation enhances ROS generation
2006 during PDT, leading to over 90% tumor growth inhibition, with 40% of treated mice
2007 achieving complete tumor regression and resistance to re-challenge, as well as
2008 prevention of lung metastasis. When combined with α PD-L1, the therapeutic efficacy
2009 further improves, resulting in >98% tumor inhibition and complete tumor clearance in
2010 80% of mice.

2011 A novel therapeutic approach for oral squamous cell carcinoma (OSCC) was
2012 developed by integrating NIR-II PTT with systemic immune activation. Using
2013 polymer-based NPs with strong electron donor-acceptor structures, this strategy
2014 enhances photothermal conversion, stability, and biocompatibility, enabling deep
2015 tumor ablation with minimal off-target effects. RNA sequencing reveals that PTT-
2016 treated cancer cells upregulate apoptosis-related and antigen-presenting pathways,
2017 leading to ICD, increased TAA release, and activation of DAMPs. This, in turn,
2018 stimulates dendritic cell activation and an adaptive immune response, effectively
2019 targeting both primary tumors and lymph node metastases. Compared to conventional

2020 ICG dyes, PNPs demonstrate superior photostability and photothermal efficiency,
2021 further amplifying the immunogenic response. The study underscores the potential of
2022 NIR-II photothermal immunotherapy as a paradigm shift in OSCC treatment, offering
2023 a promising strategy for tumor eradication and metastasis prevention. [346].

2024 **NPs enabled ICD**

2025 ICD is a regulated form of cell death that has the potential to elicit antigen-specific
2026 immune response. There are several determinants for ICD to occur that includes cellular
2027 stress, cell death, antigenicity, and adjuvanticity. Briefly, unregulated, accidental cell
2028 death cannot be immunogenic, dying cancer cells must express antigens to be
2029 recognized and taken up antigen-presenting cells of adaptive immune response such as
2030 DC or macrophages. To boost the adaptive immune response, ICD requires robust
2031 adjuvants such as HMGB1, extracellular ATP and translocation of calreticulin (CRT)
2032 from ER to plasma membrane. NP-based ROS-inducing therapies are well known to
2033 induce ICD through oxidative stress or proteotoxic stress and are summarized in this
2034 section [347].

2035 Triple negative breast cancer (TNBC) often resists immunotherapy due to its
2036 immunologically unresponsive microenvironment. Wang et al. developed a gas
2037 nanoadjuvant using a virus-mimicking hollow mesoporous silica doped with
2038 tetrasulfide, which co-encapsulates AIE-active luminogen and manganese carbonyl for
2039 the immunotherapy of breast tumors [348]. This nanosystem releases hydrogen sulfide
2040 and carbon monoxide in response to tumor-specific conditions, enhancing PDT and
2041 activating an innate immune pathway. Upon NIR laser irradiation, the AIEgen-
2042 mediated therapy causes mitochondrial damage and mitochondrial DNA leakage,
2043 which, along with the released CO and Mn^{2+} , activates an innate immune pathway. This
2044 strategy effectively improves the photoimmunotherapy of poorly immunogenic TNBC,
2045 resulting in significant tumor inhibition, and metastasis elimination in female mice. In
2046 this study, Turubanova et al. investigated two novel porphyrazine-based PSs, pz I and
2047 pz III, for their ability to induce ICD [349]. Both PSs, at an optimal light dose of 20
2048 J/cm², effectively induced cell death in cancer cells, with pz I localizing in the Golgi
2049 apparatus and lysosomes and pz III in the ER and lysosomes. Cell death induced by pz
2050 I-PDT was inhibited by an apoptosis inhibitor but not by ferroptosis or necroptosis
2051 inhibitors, while pz III-PDT-induced cell death was affected by both apoptosis and
2052 necroptosis inhibitors. Both PSs led to the release of key DAMPs, which triggered the
2053 maturation of DC in vitro and provided protective immunity in an animal model.

2054 The following study by Li et al. presents a dual-targeting nanosystem designed for
2055 effective PDT and PTT combined with immunotherapy by targeting the ER. The

2056 nanosystem, comprising ER-targeting pardaxin (FAL) peptide-modified ICG
2057 conjugated hollow Au nanospheres (FAL-ICG-HAuNS) and an oxygen-releasing
2058 hemoglobin (Hb) liposome (FAL-Hb lipo), addresses hypoxia and enhances treatment
2059 efficacy [350]. The ER-targeting approach induces significant ER stress and CRT
2060 exposure on the cell surface under NIR light, which triggers immune reaction leading
2061 to effective tumor growth inhibition and prolonged survival in animal models. Yang et
2062 al. developed a smart nanovesicle platform (pRNVs/HPPH/IND) to enhance cancer
2063 immunotherapy, combining pH-responsive nanovesicles with a PS (HPPH) and an IND
2064 [351]. These nanovesicles not only deliver therapeutic agents but also induce ICD by
2065 exposing CRT, enhancing anti-tumor effects. In a B16F10 melanoma model, this
2066 system achieved significant tumor reduction and abscopal effects through $^1\text{O}_2$
2067 generation, improved dendritic cell recruitment, and modulation of the TME by IND,
2068 which boosts CD8^+ T-cell development.

2069 To enhance RT-induced immunity, a biomineralization strategy was developed to
2070 synthesize $\alpha\text{PD-L1@MnO}_2$ NPs, which encapsulate ICB and reprogram the TME [352].
2071 The acidic TME triggers the release of Mn^{2+} , activating an innate immune pathway to
2072 promote dendritic cell maturation and type I interferon production, enhancing tumor-
2073 specific immunity. Simultaneously, the released αPDL1 blocks PDL1-PD1 interactions,
2074 increasing cytotoxic T lymphocyte (CTL) infiltration and boosting systemic anti-tumor
2075 responses. This strategy effectively inhibits primary and metastatic tumors by reversing
2076 the immunosuppressive TME, promoting CD8^+ T cell infiltration, and triggering a
2077 robust abscopal effect.

2078 RT has the potential to induce ICD and in situ vaccination (ISV) to activate systemic
2079 anti-tumor immunity, but its effectiveness is limited by insufficient X-ray deposition
2080 and an immunosuppressive TME. Nanoscale coordination NPs (AmGd-NPs) were
2081 developed by combining high-Z gadolinium (Gd) for radiosensitization and the CD73
2082 inhibitor AmPCP to improve the immune response towards tumors [353]. AmGd-NPs
2083 enhance RT by generating ROS to promote ICD, while gradually releasing AmPCP to
2084 inhibit CD73 activity. This prevents the conversion of extracellular ATP to adenosine,
2085 creating a proinflammatory microenvironment that drives DC maturation and primes
2086 CD8^+ T cell-dependent immune responses. When combined with RT, AmGd-NPs
2087 induced potent ISV, inhibited both primary and metastatic tumor growth, and prolonged
2088 survival in mice models. This effect was further enhanced by ICB, demonstrating the
2089 potential of AmGd-NPs as a promising adjuvant for RT in cancer immunotherapy. To
2090 improve the systemic anti-tumor immune response by RT, gadolinium-hemin-based
2091 nanoscale coordination polymers (H@Gd-NCPs) were developed as multifunctional
2092 RSs [354]. These NPs combine high-Z gadolinium for enhanced X-ray deposition and

2093 Hemin for GSH depletion and peroxidase-like catalytic activity, amplifying radiation-
2094 induced oxidative stress and ICD. H@Gd-NCPs also utilize tumor-overexpressed H₂O₂
2095 to produce ROS, further enhancing their radiosensitization effects. Additionally, the
2096 system serves as a MRI contrast agent and can potentiate ICB therapy, strengthening
2097 systemic anti-tumor immunity against primary, distant, and metastatic tumors.

2098 RT-induced abscopal effects are rare in clinical settings due to tumor hypoxia-
2099 related radioresistance, insufficient immune stimulation, and an immunosuppressive
2100 TME. As an attempt to overcome this, a RT-immunomodulated nanoplatform
2101 (THUNDER) was developed to synergize with RT by co-encapsulating TPZ and
2102 imiquimod [355]. THUNDER targets both hypoxic and normoxic tumor cells,
2103 enhancing TAA release and promoting DC maturation. Under RT, hypoxic tumor cells
2104 are precisely destroyed by hypoxia-activated TPZ, while normoxic cell death further
2105 aggravates hypoxia, amplifying TPZ activation and TAA generation. Simultaneously,
2106 R837 enhances immune stimulation in tumors and the spleen, leading to robust systemic
2107 immunity and immune memory. In murine models, the combination of THUNDER, RT,
2108 and ICB effectively suppresses tumor metastasis and recurrence, demonstrating
2109 significant potential for radioimmunotherapy. ISV with intratumorally injected
2110 adjuvants offers a promising approach to enhancing the abscopal effects of RT, but its
2111 clinical efficacy is limited by insufficient antigen availability from RT-induced ICD.
2112 To address this, a novel nanoadjuvant (FMC) was developed, comprising CpG-
2113 modified Fe₃O₄ NPs with maleimide residues designed to capture TAA via sulfhydryl
2114 groups. FMC maximizes antigen bioavailability and immune activation, while its
2115 magnetic properties enable tumor visualization through MRI (**Figure 10**). Combined
2116 with ICB, FMC-based ISV reverses T-cell exhaustion, promotes DC maturation,
2117 expands CD8⁺ T-cells, and reduces suppressive immune cells, effectively suppressing
2118 local and distant tumors. Mechanistic studies reveal that FMC disrupts redox
2119 homeostasis and alters amino acid metabolism in the TME, inducing ferroptosis and
2120 enhancing immune responses [356].

2121 **NPs enabled PCD pathways**

2122 **Ferroptosis**

2123 Ferroptosis is a form of PCD that differs from pyroptosis, apoptosis, and necroptosis,
2124 characterized by iron-dependent lipid peroxidation [357, 358]. Key parameters that help
2125 us understand ferroptosis include iron homeostasis, ROS biology, immunogenicity,
2126 amino acid and lipid metabolism, all of which are intricately linked in the process of
2127 ferroptosis. Type I ferroptosis involves the inhibition of cystine antiporter system Xc⁻
2128 and type II involves the direct inhibition or degradation of glutathione peroxidase 4

2129 (GPX4). NP mediated ROS based therapies that orchestrate ferroptosis are summarized
2130 below [359, 360].

2131 For gastric cancer (GC) treatment, targeting the TME has demonstrated promise.
2132 Improving the efficacy of PDT using di-iodinated IR780 (Icy7) increases ROS
2133 production, thereby inducing ICD and triggering neutrophil ferroptosis (**Figure 11**). A
2134 novel nanodrug, LLI, combining a ferroptosis inhibitor (liproxstatin-1) and Icy7, has
2135 demonstrated significant inhibition of tumor growth. This approach restores T-cell
2136 function by reducing oxidized lipids and boosts anti-tumor immunity when combined
2137 with ICB therapy. The synergistic treatment effectively decreased immunosuppression
2138 in the TME, promoted DC maturation, increased CD8⁺ T-cell infiltration, and exhibited
2139 strong systemic anti-tumor responses, including abscopal effects, emphasizing its
2140 potential for GC immunotherapy [361].

2141 To mitigate hypoxia, researchers developed TPEQM-DMA, an organic NIR-II PS
2142 with strong type I phototherapeutic efficacy [362]. TPEQM-DMA emits in the NIR-II
2143 region (>1000 nm) and produces superoxide anions and •OH through a low-O₂-
2144 dependent process under white light, specifically targeting the mitochondria of cancer
2145 cells. This process disrupts cellular redox balance, causes mitochondrial dysfunction,
2146 and increases lethal peroxidized lipid levels, resulting in apoptosis and ferroptosis by
2147 the downregulation of GPX4. Encapsulating TPEQM-DMA in NPs improved its
2148 pharmacological properties, allowing for effective tumor suppression and NIR-II FLI-
2149 guided PDT *in vivo*. A core-shell-structured Cu₂O@Mn₃Cu₃O₈ (CMCO) nanozyme has
2150 been developed to serve as a TME-activated copper ionophore for safe and efficient
2151 cuproptosis in CRC therapy [363]. The Mn₃Cu₃O₈ shell protects normal tissues from
2152 Cu₂O toxicity and improves enzyme-mimicking activities, which deplete GSH levels
2153 and generate O₂, favoring cuproptosis. Furthermore, the Fenton-like reaction from Mn
2154 ion release and GSH elimination triggers ferroptosis by the downregulation of GPX4,
2155 increasing lipid peroxidation, ROS production and boosts cuproptosis. Mild
2156 photothermal effects further improve these enzyme-mimicking activities. CMCO
2157 nanozymes exhibit strong catalytic activity, biocompatibility with normal cells, and
2158 significant tumor inhibition *in vivo*. ICD through apoptosis or necroptosis improves
2159 cancer treatment by triggering anti-tumor immunity; however, tumor resistance to these
2160 processes hinders effectiveness. A new photothermal NP (TPA-NDTA NP) has been
2161 designed to address this issue, improving ferroptosis induced ICD through excited-state
2162 intramolecular motion [364]. These NPs, exhibiting strong photothermal conversion
2163 and stability, were tested in 4T1 cancer cells and mouse models with poor immunogenic
2164 tumors, where significantly improved therapeutic effects through ferroptosis were

2165 observed. That study emphasizes the potential of TPA-NDTA NPs in boosting the
2166 immunogenicity of ferroptosis.

2167 A pH-responsive nanomedicine (DP-HBN/RA) was designed to act as a RS by
2168 amplifying ferroptosis and stimulating the immune system. This nanomedicine uses
2169 hollow Bi₂Se₃ NPs co-loaded with RSL3 and the STING agonist diABZi, modified with
2170 DSPE-PEoz for targeted delivery in acidic TME. It sensitizes tumors to X-rays,
2171 generating ROS that induce lipid peroxidation and ferroptosis. RSL3 further intensifies
2172 ferroptosis by inactivating GPX4, overcoming RT resistance. Concurrently, the DNA
2173 damage and fragments generated by DP-HBN/RA activate the cGAS-STING pathway.
2174 DiABZi amplifies this response by increasing the sensitivity of cGAS to DNA
2175 fragments, increasing immune reaction towards tumor [365]. Nasopharyngeal
2176 carcinoma, a head and neck malignancy, often exhibits resistance to RT, resulting in
2177 poor prognosis and treatment failure. One study identified circADARB1, a circular
2178 RNA significantly upregulated in NPC, as a key driver of RT resistance by inhibiting
2179 ferroptosis through the stabilization of SLC7A11 and GPX4. To overcome this
2180 resistance, Fe@Pdots-siRNA nanomaterials were developed that combine siRNA-
2181 targeting circADARB1 with iron ions, effectively suppressing circADARB1
2182 expression, increasing ferroptosis via lipid peroxidation, and improving radiosensitivity
2183 in NPC cells. This nanocomposite not only demonstrated improved tumor targeting
2184 efficiency and anti-tumor activity *in vitro* and *in vivo* but also confirmed its biosafety
2185 and potential for clinical applications [366].

2186 The clinical application of ferroptosis-inducing drugs remains limited because of
2187 safety concerns. Recent research emphasizes the synergistic potential of combining X-
2188 ray irradiation with HA-based NPs containing iron (FHA-NPs), which effectively
2189 induce ferroptosis by lipid peroxidation. FHA-NPs are internalized via CD44 receptors
2190 overexpressed in cancer cells and degraded in lysosomes, thereby triggering lipid
2191 peroxidation and ferroptotic cell death. Simultaneously, X-ray irradiation induces DNA
2192 damage, with Monte Carlo simulations confirming improved dose delivery in the TME.
2193 Both *in vitro* and *in vivo* studies have demonstrated significantly improved tumor
2194 control through this combination therapy, providing valuable insights into the
2195 mechanisms and therapeutic potential of ferroptosis in cancer treatment [367].

2196 **Cuproptosis**

2197 Cuproptosis is distinct from other forms of oxidative stress-related cell death, such
2198 as apoptosis and ferroptosis. It is characterized by mitochondrial stress resulting from
2199 the accumulation of lipoylated mitochondrial proteins and the depletion of Fe–S cluster
2200 proteins [368-370]. CMT for breast cancer often fails due to drug resistance, driven by

2201 endogenous and exogenous factors. Cuproptosis, a newly identified form of PCD
2202 triggered by copper ion accumulation, provides an alternative approach. However,
2203 achieving effective intracellular Cu ion levels is challenging; therefore, researchers
2204 have developed PEG@Cu₂O-ES, a nanocomposite combining elesclomol and cuprous
2205 oxide [371]. This composite can efficiently enter breast cancer cells, releasing ES and
2206 Cu₂O, which is further accelerated by the NIR-II region PTT effects. Cu₂O releases
2207 substantial Cu ions that participate in the TCA cycle, inducing cuproptosis by the
2208 downregulation of DLAT, FDX1, and LIAS proteins. The PTT-enhanced Fenton-like
2209 reactions generate ROS, inhibiting the ATP-Cu pump, amplifying cuproptosis. In
2210 addition, ES promotes Cu entry from the TME into cancer cells. *In vivo* studies have
2211 demonstrated that PEG@Cu₂O-ES not only exerted strong anti-tumor effects and
2212 reprogrammed the TME but also increased sensitivity to α PD-1 immunotherapy.
2213 Cuproptosis faces challenges in minimizing Cu ion release in normal tissues and
2214 maximizing its therapeutic effect at cancer sites. A photothermally triggered
2215 nanoplatform (Au@MSN-Cu/PEG/DSF) was developed for on-demand delivery and
2216 synergistic therapy [372]. This nanoplatform releases disulfiram to chelate Cu²⁺ and
2217 form cytotoxic CuET, inducing apoptosis and promoting mitochondrial protein
2218 aggregation, resulting in cuproptosis evidenced by the loss of DLAT and LIAS proteins.
2219 Combined with PTT, this approach effectively kills tumor cells and inhibits tumor
2220 growth by up to 80.1%. The multifunctional platform ensures controlled payload
2221 release at tumor sites, favoring instant intratumoral reactions upon NIR laser irradiation,
2222 with negligible effects on healthy tissues.

2223 Single-atom nanozymes with atomically dispersed active sites are better
2224 alternatives to natural enzymes; however, their full potential remains to be studied. A
2225 facile synthesis method was developed for sputtered SAzymes, creating copper-based
2226 SAzymes (CuSA) with a unique planar Cu-C₃ configuration. By improving tumor
2227 targeting through a bio-orthogonal approach, the engineered CuSACO demonstrates
2228 minimal off-target toxicity and improved multienzyme activities, generating a ROS
2229 storm for effective tumor destruction. CuSACO also releases Cu ions in the presence
2230 of GSH, inducing cuproptosis by the loss of LIAS, and oligomerization of lipoylated
2231 DLAT and improving tumor treatment efficacy. Furthermore, the photothermal
2232 properties of CuSACO enable precise PTT, significantly inhibiting orthotopic breast
2233 tumors, gliomas, and lung metastasis through combined enzyme-like catalysis,
2234 cuproptosis, PTT, and immunotherapy [373]. Residual tumors that develop after RT
2235 often acquire resistance to radiation, resulting in recurrence, metastasis, and challenges
2236 in re-irradiation. FDX1 and LIAS, key regulators of cuproptosis, are upregulated in
2237 these tumors, increasing their sensitivity to this form of cell death. To exploit this
2238 vulnerability, an interesting radiosensitization strategy was developed using copper-

2239 containing nanocapsule-like polyoxometalates, which release copper ions in a
2240 controlled manner upon X-ray irradiation. The released Cu^+ ions trigger cuproptosis by
2241 binding to lipoylated proteins in the TCA cycle, resulting in Fe–S cluster protein loss
2242 and protein aggregation. This approach effectively reverses radiation resistance,
2243 improves tumor immunogenicity, and activates a robust abscopal effect, achieving
2244 significant therapeutic efficacy in both local and metastatic tumors in radioresistant and
2245 reirradiation murine models [374]. Urchin-like $\text{Cu}_2\text{-xSe}$ hollow nanospheres enriched
2246 with copper vacancies were developed for high-performance plasmonic-thermoelectric
2247 catalytic therapy [375] (**Figure 12**). These nanostructures exhibit strong plasmonic
2248 absorption and a good PCE (67%) under 1064-nm laser irradiation, generating a
2249 localized temperature gradient that activates thermoelectric catalysis. This process
2250 improves peroxidase- and CAT-like activities, producing ROS to induce apoptosis in
2251 cancer cells. Moreover, the accumulation of copper ions within cells triggers
2252 cuproptosis via the loss of FDX1 and LIAS proteins, and promote DLAT
2253 oligomerization, generating a synergistic therapeutic effect. Density functional theory
2254 calculations reveal that the VCu in $\text{Cu}_2\text{-xSe}$ HNSs improve carrier concentration,
2255 electrical conductivity, and enzymatic activities, further increasing therapeutic
2256 efficiency. In tumor-bearing mice, this approach demonstrated significant anti-tumor
2257 efficacy, using dual pathways—membrane potential reduction and mitochondrial
2258 dysfunction—within a single nanostructure. This strategy emphasizes the potential of
2259 energy-converting nanomedicines for effective cancer therapy.

2260 A X-ray-activated HfO_2 -based RS ($\text{ES@HM-HfO}_2\text{:Cu}$) designed to enhance tumor
2261 RT while minimizing toxicity to normal tissues [376]. This nanocapsule system
2262 incorporates Cu ions doped into the shell and elesclomol (ES), within its mesoporous
2263 structure. Upon X-ray irradiation, Cu ions are specifically released at the tumor site,
2264 triggering cuproptosis, this dual sensitization strategy significantly enhances
2265 radiosensitivity, leading to a 77.9% tumor inhibition rate. The Cu ions generated during
2266 irradiation amplify proteotoxic stress by promoting mitochondrial lipoylated protein
2267 aggregation and DLAT accumulation, further amplifying tumor cell death. Compared
2268 to conventional HfO_2 sensitizers, $\text{ES@HM-HfO}_2\text{:Cu}$ demonstrates better efficacy and
2269 safety, providing a strong rationale for clinical applications.

2270 **Pyroptosis**

2271 Pyroptosis is a type of RCD triggered by disturbances in innate immunity.
2272 Pyroptosis is characterized by its dependence on inflammatory caspases (CASP) for its
2273 execution. This form of cell death can be initiated by various inflammatory or apoptotic
2274 caspases, such as CASP1, murine CASP11, human CASP4 and CASP5, and CASP3.
2275 Pyroptosis begins with the assembly of an inflammasome, a multiprotein complex that

2276 includes a sensor, the adaptor ASC, and CASP1. This complex facilitates the activation
2277 of CASP1, which then processes its substrates, including pro-IL-1 β and pro-IL-18, into
2278 their active forms, and GSDMD, which forms pores in the plasma membrane [377, 378].

2279 A novel nanoplatform integrating AIE luminogen (AIEgen) and a pyroptosis
2280 inducer into hypoxia-responsive covalent organic frameworks (COFs) was developed
2281 for improved image-guided cancer immunotherapy [379]. By synthesizing and
2282 comparing donor-acceptor type AIEgens, it was found that incorporating two acceptor
2283 units provided the longest response wavelength and most effective PDT. The
2284 constructed COF-based nanoplatform facilitates hypoxia-triggered COF degradation,
2285 improving fluorescence and PDT properties and accelerating the release of the
2286 pyroptosis drug. This system enables precise tumor imaging and effective tumor
2287 inhibition through synergistic PDT and pyroptosis by the upregulation of cleaved
2288 caspase-3 and the cleavage of GSDME into GSDME-N, leveraging exceptional ROS
2289 production and controlled drug release. In 4T1 tumor-bearing mice, the nanoagent
2290 effectively inhibited primary and distant tumor growth, demonstrating its potential as a
2291 self-synergistic cancer immunotherapy approach. The absence of tumor antigens, a
2292 challenge in ICB therapy, can be addressed by inducing tumor-specific pyroptosis. To
2293 achieve this, an ROS/GSH dual-responsive nanoprodrug loaded with paclitaxel and the
2294 PS purpurin 18 (P18) was designed [380]. This chemo-PDT system responds to the
2295 high ROS/GSH levels in the TME, enabling controlled drug release and inducing
2296 pyroptosis through increased levels of GSDME-N. Pyroptotic tumor cells release
2297 DAMPs, increasing ICB efficiency, initiating adaptive immunity, and preventing tumor
2298 recurrence. MCPP exhibits high tumor retention and penetration, promoting DC
2299 maturation and T-cell expansion and generating immunological memory.

2300 Golgi apparatus plays a vital role in NLRP3 activation. A Golgi apparatus-targeted
2301 photodynamic strategy was developed using self-assembled ChS-Ce6 nanovesicles to
2302 induce NLRP3 activation precisely [381]. This approach significantly upregulated
2303 NLRP3, resulting in an inflammatory response that improved innate immunity and
2304 triggered downstream caspase-1-dependent pyroptosis. This process promotes ICD, DC
2305 maturation, and robust anti-tumor immunity and inhibits distant cancer development
2306 (**Figure 13**). To improve pyroptosis-mediated PDT, Z1, an NIR-II PS with a
2307 thiopyrylium framework and high benzene ring count, was developed. Z1 exhibits
2308 superior type I ROS generation, and avid mitochondrial-targeting capabilities.
2309 Encapsulated into biocompatible NPs (Z1 NPs), this system demonstrates precise tumor
2310 targeting, guided by NIR-II FLI and PAI. Upon 808-nm laser irradiation, Z1 NPs induce
2311 mitochondrial dysfunction, triggering both pyroptosis by the upregulation of cleaved
2312 caspase-3 and GSDME-N and apoptosis to effectively suppress tumor growth. *In vivo*

2313 studies confirmed the high therapeutic efficacy of Z1 NPs with minimal side effects
2314 [382]. Macrophage-based biohybrid microrobots (IDN@MC) present a promising
2315 strategy for breast cancer treatment by inducing targeted pyroptosis and improving anti-
2316 tumor immunity [383]. These microrobots combine macrophages with pH-sensitive
2317 decitabine-loaded MOFs (DZNPs) and fluorescent PS, thus enabling photothermal
2318 conversion, fluorescence navigation, targeted drug delivery, and controlled release. The
2319 microrobots accumulate at tumor sites, where DZNPs release decitabine to polarize
2320 macrophages into the M1 phenotype for tumor cell phagocytosis and direct delivery of
2321 therapeutic agents. Under laser irradiation, the microrobots induce pyroptosis via the
2322 upregulation of cleaved caspase-3 and GSDME-N, releasing inflammatory factors,
2323 promoting DC maturation, T-cell expansion, and immune microenvironment
2324 modulation. This robust immune activation inhibits tumor growth, prevents metastasis,
2325 and primes adaptive immunity.

2326 A novel nanocomposite RS, DAC@O-HONs, composed of ultrasmall HfO₂ NPs
2327 loaded with decitabine to transform radiation-induced apoptosis into pyroptosis for
2328 treating TNBC was designed and developed [384]. The HfO₂ NPs improve tumor
2329 targeting through efficient osmosis and retention along with generating ROS under
2330 ionizing radiation to induce apoptosis. Simultaneously, DAC reverses GSDME
2331 silencing by inhibiting DNA methyltransferase, converting apoptosis into pyroptosis.
2332 This process releases TAA and mounts immune reaction towards tumor to inhibit tumor
2333 metastasis and recurrence.

2334 While initial nanomedicine research focused primarily on the induction of apoptosis,
2335 contemporary evidence suggests that advanced nanosystems often elicit a hybrid cell
2336 death landscape. For instance, while ROS-generating NPs may trigger rapid necrosis,
2337 the same system at a modulated concentration might initiate ferroptosis via GPX4
2338 inhibition or pyroptosis through NLRP3 inflammasome activation. The emerging
2339 paradigm of cuproptosis specifically induced by copper-ionophores and copper-based
2340 NPs further complicates this hierarchy by targeting mitochondrial respiration (**Table 6**).
2341 Understanding this link is essential, as the simultaneous activation of multiple RCDs
2342 by a single NP platform can prevent the compensatory survival mechanisms typically
2343 employed by heterogeneous tumor populations.

2344

2345 **NP promoting STING pathway**

2346 Recently the significance of the cyclic GMP-AMP synthase (cGAS)–STING
2347 pathway in activating anti-tumor immunity and transforming immunologically inactive
2348 ("cold") tumors into responsive ("hot") tumors has inspired researchers. Cytosolic

2349 double-stranded DNA (dsDNA), typically associated with damage, is detected by
2350 cGAS, leading to the production of cyclic GMP-AMP (cGAMP) [385, 386]. cGAMP
2351 acts as a second messenger to activate STING, prompting the release of type I
2352 interferons (IFNs), crucial for stimulating both innate and adaptive immune responses.
2353 Beyond its established role in microbial defense, recent studies highlight the importance
2354 of the cGAS–STING pathway in cancer progression, genomic instability, and shaping
2355 the TME. Therapeutically targeting STING with agonists has demonstrated promising
2356 preclinical anti-cancer efficacy, highlighting its potential in cancer immunotherapy
2357 strategies. Here, we introduce the design and applications of nanomedicine that could
2358 either deliver cGAS-STING activators or possess intrinsic capability to induce STING
2359 [387, 388].

2360 One study introduced novel bimetallic nanopolymers (TA-Fe/Mn-OVA@MB NPs)
2361 that combine PTT and PDT to target tumors [389]. These nanopolymers, formed by
2362 assembling tannic acid, bimetals, and ovalbumin, demonstrate excellent dispersity and
2363 biocompatibility. Under specific laser irradiation, they effectively destroy tumor cells
2364 and promote the release of tumor antigens. Furthermore, they improve the immune
2365 response by activating the cGAS-STING pathway, resulting in the maturation of DC
2366 and the activation of CTL, thus providing a potent strategy for combining localized
2367 phototherapy with robust anti-tumor immune activation. A bioactive injectable
2368 hydrogel (BG-Mngel), composed of manganese-doped bioactive glass and sodium
2369 alginate, was developed as a versatile platform for simultaneous anti-tumor
2370 immunotherapy and wound healing. The Mn^{2+} in BG-Mn activates the cGAS-STING
2371 pathway, eliciting a robust immune response, whereas its photothermal properties
2372 improve NP uptake in tumor cells and facilitate STING activation under NIR light. This
2373 synergistically inhibits tumor growth and prevents recurrence, particularly when
2374 combined with the α -PD-1, which strengthens long-term immune memory. Moreover,
2375 BG-Mngel promotes angiogenesis and tissue regeneration by releasing bioactive ions
2376 such as SiO_4^{4-} , Ca^{2+} , and Mn^{2+} , enabling rapid wound healing in melanoma models
2377 [390]. The cGAS-STING pathway plays a vital role in tumor immunotherapy; however,
2378 the lack of tumor-specific targeting by STING agonists has limited its clinical
2379 application. A photodynamic polymer was designed to encapsulate a cationic Pt (II)
2380 agent (56MESS), forming NPs (NPPDT-56MESS) [391]. These NPs accumulate at
2381 tumor sites and, upon 808-nm laser irradiation, generate ROS that disintegrate the NPs
2382 and release 56MESS. The ROS and 56MESS synergistically damage DNA and
2383 mitochondria, which activates the cGAS-STING pathway to induce robust anti-tumor
2384 immune responses. In mouse models of uveal melanoma, NPPDT-56MESS
2385 demonstrated effective tumor targeting, reduced systemic toxicity, and induced
2386 systemic anti-tumor immunity, including immune memory, which prevented tumor

2387 recurrence and metastasis. Researchers have developed a 2D nanoplatform,
2388 cGAMP/MOL, where cGAMP is conjugated to a nanoscale MOL [392]. This design
2389 improves both STING activation and radiosensitization within tumors. Upon X-ray
2390 irradiation, cGAMP/MOL induces ICD and improves the maturation of APCs, resulting
2391 in a “hot” tumor immune environment rich in inflammatory cytokines and type I IFNs.
2392 This environment fosters local innate and adaptive immune responses. When combined
2393 with ICB, cGAMP/MOL overcomes immune suppression, reinvigorates T cells, and
2394 bridges innate and adaptive immunity, resulting in systemic anti-tumor effects and
2395 tumor regression.

2396 This study introduces an innovative NP-based immunotherapy designed to target
2397 pulmonary APCs and improve anticancer responses against lung metastases. In mouse
2398 models, inhaling phosphatidylserine-coated liposomes loaded with the cGAMP allows
2399 for rapid distribution across the lungs, where the NPs are rapidly absorbed by APCs
2400 without triggering harmful immune responses [393]. The formulation is optimized for
2401 efficient cytosolic release of cGAMP, which activates STING signaling and promotes
2402 the production of type I IFNs in APCs. This activation transforms the TME into a
2403 proinflammatory state across multiple lung metastases. In addition, when fractionated
2404 radiation is applied to one of the tumor-bearing lungs, synergy was established with the
2405 NP-cGAMP, resulting in a robust systemic anticancer immune response, even when
2406 exposed to repeated tumor challenges.

2407 An ECM-degrading nanoagonist (dNac) was designed for targeted breast cancer
2408 therapy. The dNac contains FeS₂ NPs that serve as NIR-II photothermal converters and
2409 Fenton catalysts, along with a STING agonist and an ECM-degrading enzyme [394].
2410 Mild heat generated by the dNac after irradiation improves the Fenton reaction,
2411 inducing tumor cell death, ICD, and STING activation. Robust immune reactions
2412 effectively inhibited both primary and distant tumors in, along with suppressing liver
2413 and lung metastasis. A smart nanosystem (PBM) was developed based on Prussian blue
2414 NPs doped with Mn(III). This system improves the photothermal ablation-induced
2415 activation of the cGAS-STING pathway by generating cytosolic DNA damage and
2416 continuously releasing Mn(II) in response to the GSH-rich environment of the tumor
2417 [395]. The PBM nanosystem not only improves local tumor ablation but also triggers a
2418 strong systemic immune response, effectively reducing the risk of tumor recurrence and
2419 metastasis.

2420 This study presents a novel approach for treating TNBC by combining PTT with
2421 Mn²⁺ to activate the cGAS-STING pathway. The researchers developed a manganese-
2422 enriched nanoplatform that integrates Mn-doped Prussian blue NPs with MnOx [396].
2423 This platform leverages Mn²⁺ to amplify the cGAS-STING pathway and uses

2424 hyperthermia and ROS to elicit immune response against tumor. Experiments using
2425 4T1-bearing mice demonstrated that this treatment effectively suppressed both local
2426 and distant tumors by promoting innate and adaptive immunity (**Figure 14**). In another
2427 study, a multifunctional bionic nanoplatform was designed to improve the treatment of
2428 PCa by combining PDT, CMT, and immunotherapy. The nanoplatform incorporates the
2429 AIE PS TPAQ-Py-PF6 and PTX, in a macrophage cell membrane for active targeting
2430 [397]. Upon laser irradiation, the platform generates ROS that cause DNA damage,
2431 triggering ICD and activating the cGAS-STING pathway to mount a robust systemic
2432 immune response. The combination of PDT and CMT not only improves the anti-tumor
2433 effects but also transforms the TME from “cold” to “hot,” boosting cytotoxic T-cell
2434 infiltration and establishing lasting immune memory.

2435 A polymeric metal-organic framework (PMOF) NPs was designed to improve
2436 cancer immunotherapy by combining PDT with STING activation [398]. The porous
2437 structure of PMOF NPs, could accommodate a STING agonist (SR-717) and coated
2438 with a PEG shell. Upon intravenous injection and tumor targeting, light irradiation at
2439 the tumor site triggers the production of ROS cleaves thioketal bonds within the PMOF
2440 releasing SR-717, which activates the STING pathway. This dual action not only
2441 reverses the immunosuppressive TME but also improves anti-tumor immunity,
2442 effectively suppressing both primary and metastatic tumors. A manganese-doped
2443 layered double hydroxide (Mn-LDH) nanoplatform was developed using a
2444 hydrothermal method to reprogram the TME and potentiate anti-tumor immunity. Mn-
2445 LDH neutralizes protons and consumes hydrogen peroxide to produce oxygen,
2446 generating a positive feedback loop that releases Mn^{2+} and Mg^{2+} [399]. These ions
2447 activate the cGAS-STING pathway and sustain $CD8^+$ T-cell cytotoxicity, triggering a
2448 cascade of innate and adaptive immune responses. Locally administered Mn-LDH
2449 transformed the TME into an immune-supportive state, forming a network of mature
2450 DC, M1 macrophages, and cytotoxic and helper T cells, thereby effectively inhibiting
2451 tumor growth and metastasis in breast and colon tumor models. Furthermore, its
2452 photothermal conversion capability improved the therapeutic efficacy in large tumor
2453 models with a single administration and irradiation. To mitigate hypoxia-associated
2454 radioresistance in rectal cancer (RC), researchers have developed a catalytic RS,
2455 DMPtNPS, combined with a STING agonist, to improve RC radiosensitivity [400]. The
2456 combination of DMPtNPS@cGAMP, RT, and ICB resulted in a durable complete
2457 response at the primary tumor site and an improved abscopal effect on distant tumors
2458 in a bilateral tumor model.

2459 A nanotheranostic platform combining PDT and epigenetic therapy to improve
2460 cancer treatment by inducing pyroptosis and activating the cGAS-STING pathway in a

2461 light-controlled manner was developed. The system involves two types of NPs, viz.,
2462 NP1, which is ROS-sensitive and loaded with the PS TBE, and NP2, which contains
2463 the epigenetic drug decitabine. NP2 reverses promoter methylation, restoring STING
2464 and gasdermin E (GSDME) expression, whereas NP1 generates excessive ROS under
2465 808-nm laser exposure, causing mitochondrial damage and the release of DNA
2466 fragments. This stimulates the cGAS-STING pathway and activates caspase-3 to cleave
2467 GSDME into its pore-forming fragment, initiating pyroptosis[401]. A strong anti-tumor
2468 immune response is then triggered by the release of inflammatory cytokines, which
2469 enhance T-cell-mediated immunity and long-term immunological memory.
2470 Additionally, the system uses NIR fluorescence bioimaging to provide TME regulation
2471 and guided photoimmunotherapy. SAPTN was designed and developed to improve
2472 cancer immunotherapy by targeting the STING pathway. The SAPTN is engineered
2473 with NIR-II properties for precise imaging and PAI-guided therapy. It delivers the
2474 STING agonist diABZIs in a sustained manner within the tumor's ECM, ensuring
2475 durable drug activity. When combined with mild PTT, which triggers neutrophilic
2476 inflammation, SAPTN improves tumorous retention and stimulates a robust and long-
2477 lasting immune response. This strategy achieved tumor-free survival for at least two
2478 months in preclinical studies, when nearly 100% of mice with breast cancers had their
2479 tumors completely eliminated. When given locally, SAPTN also had an abscopal
2480 impact, slowing the growth of distant tumors. Additionally, NIR-II imaging makes it
2481 possible to clearly identify the tumor margin for improved treatment guidance.[402]. A
2482 MOF-based nanoagonist (DZ@A7) for precise tumor-specific activation of the cGAS-
2483 STING pathway and improved photodynamic–metalloimmunotherapy was developed
2484 [403]. Inside the cancer cells, when exposed to NIR irradiation, DZ@A7 performs
2485 several functions such as i) generates mitochondrial-targeted ROS, ii) releases oxidized
2486 mitochondrial DNA (mtDNA) into the cytoplasm, activating the cGAS-STING.
2487 Simultaneously, hypoxia-responsive drugs inhibit DNA repair, resulting in increased
2488 cytosolic damaged DNA, further boosting immune activation. Furthermore, NIR-
2489 accelerated zinc ion overloading improves cGAS enzymatic activity, amplifying
2490 STING activation. The combination of these effects promotes DC maturation and
2491 cytotoxic T-cell infiltration, causing the eradication of primary tumors and establishing
2492 long-term anti-tumor immunity to prevent metastasis. **Table 5** shows NP mediated cell
2493 death pathways and innate immune system activation.

2494 **Toxicity of NPs**

2495 Nanotoxicity can be ascribed to their rate of elimination. Glomerular endothelial
2496 cells (GECs) is a part of RES, which is integral to the glomerular filtration mechanism.
2497 GECs contain numerous fenestrae with diameters ranging from 60–80 nm, which

2498 facilitates rapid exchange of macromolecules between the vascular and extravascular
2499 spaces [404, 405]. Inorganic NPs with a size less than 6 nm and organic NPs with size
2500 cutoff 8-10 nm are eliminated from the body through renal mechanism [406, 407], while
2501 large sized NPs is trapped in the GECs base membrane [408]. NPs can also be
2502 eliminated through bodily secretions such as saliva, breast milk and sweat, but this
2503 warrants large-scale studies for better understanding [409-411]. NPs that do not
2504 undergo renal elimination obliges to hepatobiliary excretion, which is size and charge
2505 dependent. As demonstrated in a study, both MSN with more charge (+34.4 mV) and
2506 less charge (-17.6 mV) are accumulated in the liver, former undergoes hepatobiliary
2507 excretion into gastrointestinal tract while latter remains sequestered [412].

2508 Metabolism of NPs strongly depends on their organic and inorganic composition,
2509 among them, inorganic NPs show a high propensity to liver accumulation causing
2510 toxicity [413]. Inorganic NPs tend to degrade and release constitutive ions in the acidic
2511 lysosome compartment, which react with biomolecules, altering their biological
2512 activity [414]. Whereas organic counterparts, including lipid, polymeric NPs, show
2513 biotransformation rates, the resulting products may be more reactive or present low
2514 clearance rates. In addition, biotransformation rates are correlated to drug release
2515 profile. Accelerated biodegradation triggers untimely drug release, while slow
2516 degradation can lead to prolonged tissue accumulation and potential toxicity. Orally
2517 administered TiO₂ NPs interact with the gut microbiota affecting gut-associated
2518 metabolism, altering several metabolites and metabolic pathways. In particular, the
2519 levels of metabolites such as N-acetylhistamine and glycerophosphocholine increased,
2520 while others including L-histidine and L-ornithine decreased. This may lead to
2521 enhanced lipopolysaccharide production, which subsequently triggers oxidative stress
2522 and inflammatory responses in the intestine [415]. In vitro studies using TM-4 Sertoli
2523 cells and GC2-spd spermatocyte cells demonstrated that ZnO NPs can penetrate cells,
2524 disrupt cell membranes, and generate oxidative stress, that activates the ROS-ERK-
2525 TNF- α signaling pathway, leading to impairment of the blood-testis barrier by reducing
2526 junction proteins in Sertoli cells. In germ cells, ROS generation causes DNA damage,
2527 reduced DNA repair capacity, and S-phase cell cycle arrest, contributing to genotoxic
2528 effects [416]. Small amorphous silica NPs (~70 nm) can trigger strong coagulation
2529 responses after systemic exposure. In mice, i.v. injection caused platelet depletion, liver
2530 injury, and fatal coagulation reaction. This effect occurs through activation of the
2531 intrinsic coagulation pathway via interaction with coagulation factor XII, which is
2532 enhanced by the large surface area of smaller NPs [417].

2533

2534 **Conclusions and outlook**

2535 This review has comprehensively summarized the development of nanomaterials
2536 for precise nanotheranostics from optical to radioactive modalities. Imaging-guided
2537 photoinduced treatment has achieved enormous advancements in recent decades and
2538 plays an increasingly vital role in the global war against cancer. Photo-based therapies
2539 for cancer that exert minimal side effects and high specificity can effectively decrease
2540 side effects. Considering the tissue penetration limitation of PDT and PTT, tremendous
2541 attempts have been devoted to exploring deep light delivery techniques for treating
2542 orthotopic tumors inside patients. Very recently, compared with conventional NIR-I
2543 imaging, imaging in the NIR-II region has demonstrated superior performance to that
2544 in the NIR-I region due to its higher spatial resolution, deeper penetration depth, and
2545 lower optical absorption and scattering from biological substrates with minimal tissue
2546 autofluorescence. With this strong motivation, several novel NIR-II imaging systems
2547 have been applied to assist surgeons in precision surgery.

2548 Moreover, due to the high tissue-permeability of X-ray radiation and interstitial
2549 fiberoptic light sources, several investigations on orthotopic tumor models were
2550 performed within the past 2 years. Consequently, clinical applications of X-ray PDT or
2551 interventional PTT are reasonable in the near future. The incorporation of imaging
2552 techniques into RT has the potential to allow dose painting with minimal toxicity in the
2553 surrounding normal tissue and maximum therapeutic dose in localized tumors.
2554 Currently, NPs, such as gadolinium-based NPs (AGuIX) and HfO₂ NPs crystalline NPs
2555 (NBTXR3), in conjunction with RT are being used in clinical trials in imaging-guided
2556 RT. In particular, NBTXR3 and HfO₂ NPs, combined with IMRT, for the treatment of
2557 PCa are under Phase I/II investigation (NCT02805894). Furthermore, the clinical
2558 applications of IMPT, as well as IMRT, are rapidly increasing for the treatment of
2559 various cancers. Although IMPT has achieved encouraging results and cost-
2560 effectiveness in several cancers, additional clinical data are necessary. Apart from
2561 extrinsic RT, another methodology used to achieve deeper tissue penetration is the
2562 utilization of radionuclides as an internal activation light source. In 2015, Achilefu et
2563 al. demonstrated that tumors could be eliminated through CR-triggered NPs. To date,
2564 CLI, which combines nuclear and optical imaging, has been used in intraoperative FLI-
2565 guided surgery, including breast cancer, PCa, gastrointestinal cancer, and metastatic
2566 lymph nodes. Furthermore, NPs and microparticle-based imaging-guided theranostic
2567 agents have entered clinical oncological applications. For instance, liver
2568 radioembolization has achieved successful clinical trials. To date, ⁹⁰Y Ms are approved
2569 in the European Union, Asia, and the USA for the treatment of unresectable liver tumors,
2570 and more than 40,000 treatments have been performed using ⁹⁰Y-labeled Ms in hepatic
2571 solid tumors. Numerous novel particles are currently under development for next-
2572 generation imaging-guided theranostic agents, and successful clinical cases will

2573 accumulate with time. Nanotechnology has greatly advanced imaging-guided treatment
2574 of deep-seated tumors, improving both tumor detection and targeted therapy. Emerging
2575 imaging systems such as total-body PET and MARS-CT are enhancing sensitivity,
2576 quantification, and scan speed, which may allow earlier and more accurate diagnosis.
2577 On the therapeutic side, photoresponsive and radioactive NPs have shown strong
2578 potential for highly specific tumor ablation.

2579 NPs have emerged as powerful tools due to their ability to interact with cellular
2580 components and modulate various biochemical pathways. We emphasized their role in
2581 inducing specific cell death mechanisms such as ferroptosis, pyroptosis, and
2582 cuproptosis, each characterized by distinct molecular processes orchestrated by PDT,
2583 PTT, and RT. These cell death modalities instigated by either oxidative or
2584 mitochondrial proteotoxic stress have the potential to induce an immune response by
2585 promoting the release of tumor cell-specific antigens and cause ICD that could improve
2586 cancer immunotherapies. NPs have also emerged as potent modulators of the innate
2587 immune system, and are designed to overcome the delivery issues of STING agonists
2588 such as low bioavailability, poor in vivo circulation times and modest uptake by the
2589 tumor cells due to their negative charge [418-420]. To overcome the challenges
2590 associated with the i.v. delivery of STING agonists, Dosta et al. conjugated STING
2591 agonist CDN to poly(β -amino ester) (CDN NPs) through a cathepsin-sensitive linkage.
2592 The synthesized polymer-drug conjugate showed improved drug loading, good serum
2593 stability and low drug dose, which was sufficient to exhibit robust anti-tumor
2594 therapeutic response across various syngeneic tumor models and elicit immune memory.
2595 Tumor acts as a drug depot for CD NPs and releases them to nearby immune cells to
2596 activate STING and inhibit tumor growth [421]. Another interesting approach is
2597 leveraging the potential of Mn^{2+} ions, which can activate monomeric cGAS even
2598 without dsDNA and does not require cGAS oligomerization. Mn^{2+} and cytosolic
2599 dsDNA together enforce each other's binding to cGAS, markedly lowering the
2600 activation threshold of the STING pathway. Mn^{2+} increases the enzymatic activity of
2601 dsDNA-bound cGAS, leading to higher production of the second messenger cGAMP.
2602 Mn^{2+} can also trigger phosphorylation of TBK1 and p65 independently of STING and,
2603 together with STING agonists, promotes enhanceosome formation and stronger IFN- β
2604 production [422-424]. In a recent study, blackphosphorous nanosheets were modified
2605 with Mn^{2+} /CpG oligodeoxynucleotides (BPNS@ Mn^{2+} /CpG) through coordination
2606 bond. In TME, Mn^{2+} /CpG were released in a pH responsive manner activating immune
2607 cells to prime cytotoxic and helper T cells via Toll-like receptor 9 and STING pathways.

2608 Phototherapy combined chemodynamic therapy of tumor by BPNS@Mn²⁺/CpG
2609 induced ICD that not only suppressed the primary tumor but also inhibited the distant
2610 tumor growth [425]. A polymer based universal STING mimic (uniSTING) was
2611 developed that can self-assemble into active STING moieties and trigger STING
2612 signaling independent of endogenous STING expression. When delivered as mRNA
2613 through lipid NPs, uniSTING effectively activated the TBK1–IRF3–IFN-I pathway in
2614 both tumor cells and dendritic cells. It also promoted dendritic cell maturation and
2615 stronger CD8⁺ T-cell responses, leading to significant tumor suppression in several
2616 cancer models [426].

2617 A plethora of NPs have been synthesized, optimized, and demonstrated potential in
2618 preclinical levels. Nevertheless, their clinical translation is severely hampered by both
2619 immune and physiological barriers. Inside the body, based on the surface composition,
2620 NPs are covered by serum proteins known as opsonin, which are readily recognized
2621 and cleared from circulation by mononuclear phagocytic systems or reticuloendothelial
2622 systems. The transport of NPs to tumors occurs through inter-endothelial gaps in tumor
2623 blood vessels and retained by a poor lymphatic system, termed as the EPR effect.
2624 Nevertheless, the existence of the EPR effect is heterogeneous and dependent on tumor
2625 types. A relatively novel mechanism for NP entry into tumors, termed active
2626 transcytosis, has been proposed using Au NPs. This process involves NPs transport
2627 through endothelial cells and overexpression of transport pathway–related genes. Once
2628 inside the tumor, the NPs are retained through interactions with TME components, such
2629 as TAMs and ECM, rather than depending on poor lymphatic drainage. Furthermore,
2630 extensive investigations are required to validate the mechanism using soft NPs and
2631 various tumor models. Active tumor targeting involves engaging receptors expressed
2632 on tumors through ligand-modified NPs. The expression level of receptors and ligand
2633 density on NPs plays a vital role in cancer cell–specific uptake of NPs. To accelerate
2634 the clinical translation of NPs, regulatory bodies such as the FDA USA and the
2635 European Medical Agency have issued guidelines to ensure the safety, efficacy, and
2636 quality of NPs and emphasize the characterization of physicochemical properties,
2637 pharmacokinetics, and potential toxicological effects; however, current frameworks
2638 face challenges in addressing the complexities of nanoscale materials. Advances in
2639 computational tools, particularly AI-based platforms such as AlphaFold and Rosetta,
2640 have transformed the landscape by enabling precise protein structure prediction and
2641 novel protein design. These innovations provide critical insights into protein–ligand
2642 interactions and facilitate the development of targeted nanomedicine therapies with

2643 optimized safety and efficacy. The use of human-relevant experimental systems,
2644 including patient-derived organoids, 3D tumor models, and organ-on-chip platforms,
2645 can provide more realistic biological conditions than conventional 2D cell culture or
2646 murine models [427-429]. Improved animal models, such as orthotopic tumors,
2647 genetically engineered mouse models and patient-derived xenografts, can better mimic
2648 disease complexity and drug responses [430-432]. Improving patients' quality of life
2649 can expand the number of people willing to receive treatment, which also boosts the
2650 practical and commercial value of a therapy. For example, treatments that provide better
2651 effectiveness, cause fewer side effects, require fewer doses, and can be given through
2652 convenient routes of administration are more satisfactory to patients. These factors can
2653 improve patient adherence to therapy and ultimately allow a larger patient population
2654 to benefit from the treatment.

2655 Toxicity of the NPs is a crucial parameter impeding their clinical translation,
2656 profound accumulation of NPs in undesired vital organs is a major contributor to
2657 toxicity. NP often accumulates in organs such as the liver, lungs, kidneys, spleen, and
2658 bones. This is mainly because these organs contain cells of the reticuloendothelial
2659 system that capture and remove foreign bodies from the blood [433]. The extent of this
2660 accumulation depends on the NPs properties, including their size, surface charge,
2661 stiffness, and other physicochemical characteristics [434]. PEGylation is commonly
2662 used to give NPs a "stealth" property. It reduces opsonization by proteins in the
2663 bloodstream and prevents the particle aggregation [435, 436]. In addition,
2664 nanomedicines should be screened using human blood or serum complement assays,
2665 cytokine profiling, and more predictive models such as the porcine CARPA model,
2666 which has been widely used to study acute cardiopulmonary infusion reactions [437-
2667 439]. From the industry point of view, NPs bulk production is challenging due to their
2668 complex design, composition, and multistep synthesis that increases the production cost
2669 and time. Preclinical purification methods, such as ultrafiltration and evaporation, are
2670 often not efficient during large-scale production and may leave behind toxic solvent
2671 residues [440, 441]. By incorporating such tools into regulatory processes, the
2672 evaluation of nanomedicine interactions and off-target effects can be significantly
2673 improved, ultimately enhancing the therapeutic outcomes and advancing regulatory
2674 science.

2675

2676 **Abbreviations:** PDT, photodynamic therapy; PTT, photothermal therapy; RT,
2677 radiotherapy; NPs, nanoparticles; UCNP, upconversion nanoparticle; AuNP, gold
2678 nanoparticle; MSN, mesoporous silica nanoparticle; WHO, World Health Organization;
2679 PS, photosensitizer; ROS, reactive oxygen species; UV, ultraviolet; NIR, near infrared;
2680 FRET, fluorescence resonance energy transfer; Vis, visible; CT, computed tomography;
2681 MRI, magnetic resonance imaging; RIT, radionuclide therapy; CR, Cerenkov radiation;
2682 FA, folic acid; HA, hyaluronic acid; DOX, doxorubicin; MDR, multidrug resistance;
2683 AuNR, gold nanorod; Ce6, chlorin e6; ADR, adriamycin resistance; TP, two-photon;
2684 RB, rose bengal; NGO, nanographene oxide; AuNEs, gold nanoechinus; TME, tumor
2685 microenvironment; MB, methylene blue; HIF-1 α , hypoxia inducible factor-1 α ; PC,
2686 perfluorocarbon; APC, albumin-coated PC nanodroplets; RBC, red blood cells; TPZ,
2687 tirapazamine; HAdase, hyaluronidase; SCNP, scintillating nanoparticle; PLNP,
2688 persistent luminescent nanoparticles; MOL, metal organic layers; LSPR, localized
2689 surface plasmon resonance; PCE, photothermal conversion efficiency; BGV,
2690 biodegradable gold nanovesicles; DPP, diketopyrrolopyrrole; SWCNT, single-walled
2691 carbon nanotubes; ND, nanodots; HSA, human serum albumin; IPTT, interventional
2692 photothermal therapy; TNP, theranostic nanoparticles; PNIPAM, poly(N-
2693 isopropylacrylamide); PET, positron emission tomography; LET, linear energy transfer;
2694 BBB, blood brain barrier; SPECT, single photon emission computed tomography; ACE,
2695 artificial compound eye; CLI, Cherenkov luminescence light; REFI,
2696 radiopharmaceutical-excited fluorescence imaging; SCIFI, Cerenkov-induced
2697 fluorescence imaging; RET, radionuclide energy transfer; PIXE, particle-induced x-ray
2698 emission, IMPT, intensity-modulated proton therapy; IMRT, intensity-modulated
2699 radiation therapy; ICG, indocyanine green; NCP, nanoscale coordination polymer;
2700 CSiNP, cationic silica nanoparticle; cGAMP, cyclic GMP-AMP cGAMP; STING,
2701 stimulator of interferon genes.

2702

2703 **Acknowledgements**

2704 This study was supported by the grants BN-110-PP-04~BN-113-PP-04, BN-114-
2705 PP-04 from National Health Research Institutes of Taiwan, and 114-2221-E-400-002-
2706 MY3 from the National Science and Technology Council of Taiwan. The authors
2707 confirm that no artificial intelligence (AI) tools were used in this manuscript.

2708

2709 **Contribution**

2710 **Maharajan Sivasubramanian:** Writing original draft for the section “Functional
2711 photodynamic therapy”– review & editing. **Yao-Chen Chuang:** Writing original draft
2712 for the section “Image-guided photothermal therapy”– review & editing. **Chia-Hui Chu,**
2713 **Li-Jie Lin and Maharajan Sivasubramanian:-** Writing original draft for the section
2714 “Radionuclide nanotheranostics”, **Yu Hsia and Maharajan Sivasubramanian:**
2715 Writing original draft for the section “Advancing cancer immunotherapy with NPs”.
2716 **Leu-Wei Lo:** Conceptualization, Writing – review, editing and supervision-original
2717 draft,

2718

2719 **Declaration of competing Interests**

2720 The authors declare that they have no known competing financial interests or personal
2721 relationships that could have appeared to influence the work reported in this paper.

2722

2723

2724

2725

2726

2727

2728

2729

2730

2731

2732

2733

2734

2735

2736

2737

Table 1. Deep tissue PDT nanosystems and their experimental parameters.

| Energy modes | NPs (Size, zeta potential and morphology) | Treatment condition, λ power | Biological experiment | Ref. |
|--------------|---|--|---|------|
| Upconversion | UCNPs@mSiO ₂ -ZnPc-RBS, ~200 nm, hexagonal shape | 1550, 808 nm and 980 nm; 1 W·cm ⁻² | 4T1 subcutaneous xenograft (i.t.), tumor inhibitory effect | [60] |
| | NaGdF ₄ :Yb Tm@CaF ₂ :Eu@shell, ~150 nm, spherical shape | 980 nm; 2 W·cm ⁻² | GBC orthotopic xenograft (i.v.), tumor inhibitory effect | [61] |
| | PB@VA, 100.37 ± 4.05 nm, ~-30 mV | 980&808 nm; 1 W·cm ⁻² | HeLa subcutaneous xenograft (i.t.), tumor inhibitory effect | [63] |
| TP | Emo/HSA NPs (E/H NPs), 315 ± 158.1 nm, spherical shape | 800 nm; 2 W·cm ⁻² | MCF7 subcutaneous xenograft (i.v.), tumor inhibitory effect | [73] |
| | NIR-TADF NPs, ~140 nm, -46.1 mV, spherical shape | 800 nm; 2 W·cm ⁻² | A549 subcutaneous xenograft (i.v.), tumor inhibitory effect | [75] |
| | DSPE-PEG-Biotin@2-O-IrAn, ~116 nm, spherical shape | 750 nm, 50 mW·cm ⁻² | A549 subcutaneous xenograft (i.v.), tumor inhibitory effect | [76] |
| X-PDT | ITC NPS, 47.2 ± 0.8 nm, -1.0 mV, spherical shape | 0.4 Gy; 50 kV | 4T1 subcutaneous xenograft (i.t. and i.v.), tumor inhibitory effect | [89] |
| | Sc-D@Lip/Pt, 73 nm, hexagonal shape | 2 Gy | 4T1 subcutaneous xenograft (i.v.), tumor inhibitory effect | [90] |
| | NSs-RB-F/cRGD, 274.2 nm, -40.3 mV, Hexagonal shape | 0.025 Gy | NMIBC (intravesical fusion), tumor inhibitory effect | [93] |

2739

2740

2741

Table 2. NIR-I and II image-guided PTT nanosystems.

| Energy modes | Imaging technique | NPs (Size, zeta potential and morphology) | Exposer conditions for PTT | Biological experiment | Ref |
|--------------|-------------------|--|--|--|-------|
| NIR-I | FL | Water heating NIR NP, 18.5 ± 1.7 nm, +29.1 mV, spherical shape | 808 nm; 1.5 W·cm ⁻² | GBM orthotopic model (i.v), tumor inhibitory effect | [200] |
| | FL/US | PFC@PEPP-Fe, ~180 nm, +35.6 mV, spherical shape | 808 nm; 1 W·cm ⁻² | 4T1 subcutaneous xenograft (i.v), tumor inhibitory effect | [201] |
| | PA | GNDs@gelatin, 120.4 ± 26.6 nm, Multibranched spherical shape | 808 nm and 1160 nm; 1 W·cm ⁻² | C6 subcutaneous xenograft (i.v), tumor inhibitory effect | [202] |
| | FL/PA | Organic TTNH, 90.3 nm, -19.89 ± 2.93 mV | 808 nm; 1 W·cm ⁻² | CT26 orthotopic breast cancer xenograft (i.v), tumor inhibitory effect | [203] |
| | CT/MRI/PTI | Au@MnO ₂ @PM 246.12 nm, -27.10 mV, core/shell shape | 1064 nm; 1 W·cm ⁻² | A549 subcutaneous xenograft (i.v), tumor inhibitory effect | [234] |
| NIR-II | FL | SNB, 190 nm, -180 ± 0.3 mV, spherical shape | 808 nm; 1.5 W·cm ⁻² | 4T1 subcutaneous xenograft (i.v), tumor inhibitory effect | [235] |
| | PA | CY-1234, 55 nm, spherical shape | 1064 nm; 1 W·cm ⁻² | 4T1 subcutaneous xenograft (i.v), tumor inhibitory effect | [236] |
| | MRI/CT | BFS, 9 nm, -17.08 ± 2.13 mV, spherical shape | 1064 nm; 1 W·cm ⁻² | 4T1 subcutaneous xenograft (i.v), tumor inhibitory effect | [237] |
| | | | | HT29 subcutaneous xenograft (i.v), tumor inhibitory effect | |

2743

2744

2745

2746

2747

2748

2749

2750

2751

2752

2753

2754

2755

2756

2757

2758

2759

2760

2761

2762

2763

2764

2765

2766

2767

2768

Table 3. Radionuclides loaded NPs for radionuclide therapy.

| Therapy | Energy (keV) | Half-life | Emission type | NPs-Isotopes (Size, zeta potential and morphology) | Diseases | Ref. |
|---------------------|--------------|-------------------|----------------------|---|--|-------|
| Radionuclide | 192 | 8.02 to 8.04 days | β and γ | ^{131}I -ICT/R848-MS 48.59 \pm 7.2 μm , spherical shape | Orthotopic HCC, (hepatic artery injection) tumor inhibitory effect | [276] |
| | 5.79 MeV | 11.435 days | α | ^{223}Ra /Ba SAE 310 NM, -13 \pm 1.11 mV, nanosheets | Subcutaneous LLC tumor (i.t.) tumor inhibitory effect | [278] |
| | 192 | 8.02 to 8.04 days | β and γ | ^{131}I -VNP+ α PD-L1 Nano and micro sized bacterial vectors | Subcutaneous CT-26 tumor (i.t.), tumor inhibitory effect | [279] |
| | 192 | 8.02 to 8.04 days | β and γ | USINAS (^{131}I - α PD-L1) 139.31 \pm 24.89 nm, spherical shape | Subcutaneous 4T-1 tumor (i.v.), tumor inhibitory effect | [280] |
| Cherenkov radiation | 897 | 78.4 h | Blue light | ^{89}Zr -aEuNP@PEG \sim 11 nm, square plate | Subcutaneous CT-26 tumor imaging (i.v.), | [293] |
| | 511 | 67.6 to 68 min | Blue light | TiO ₂ NP-[^{68}Ga]Ga-FAPI+TPZ 60 nm, spherical shape | Subcutaneous Panc-1 tumor (i.v.), tumor inhibitory effect | [294] |
| | 633 | 109.8 min | Blue light | | | [295] |
| | 897 | 78.4 h | Blue light | ECN@TTVP- ^{18}F -FDG 2067.5 \pm 59.3nm, -28.5 \pm 0.5 mV, bacteria biohybrid 89ZrALA-liposome-ART | Subcutaneous CT-26 tumor imaging (i.v.), tumor inhibitory effect | [296] |

136.5 ± 6.5 nm,
spherical shape

Subcutane
ous 4T-1
tumor
(i.v.),
tumor
inhibitory
effect

2770

2771

2772

2773

2774

2775

2776

2777

2778

2779

2780

2781

2782

2783

2784

2785

2786

2787

2788

2789

2790

2791

2792

Table 4. Therapeutic NPs used in IMRT and proton therapy.

| Therapy | Dose | NPs (Size, zeta potential and morphology) | Diseases | Ref. |
|---------|-----------------------------|--|--|-------|
| Proton | 0.5, 1, 2 and 4 Gy | IONP _{DOX} 369.1, 20.9 mV, spherical shape | SW1353 3D spheroids, chemo and radio sensitization effects were observed | [310] |
| | 2 Gy | Au 25.12 ± 0.79 nm, -18.55 ± 0.15 Mv, spherical shape | MDA-MB 231 3D spheroids, proton therapy effectively reduced cell viability | [311] |
| IMRT | 50 Gy in 25 fractions | NBTXR3 Hafnium oxide 50 nm, -50mV | Rectal cancer patients, protocol of a phase Ib/II study to investigate the safety profile, dose-limiting toxicity and anti-tumor activity of (NBTXR3) RT+Chemo | [315] |
| | 50 Gy | NBTXR3 Hafnium oxide 50 nm, -50mV | Locally advanced soft tissue sarcoma patients, NBTXR3 did not negatively affect safety or quality of life and long-term safety | [316] |
| | 45 Gy in 15 daily fractions | NBTXR3 Hafnium oxide 50 nm, -50mV | Pancreatic ductal adenocarcinoma patients, initial feasibility of local endoscopic delivery of NBTXR3 activated by radiation | [317] |

2794
2795
2796
2797
2798
2799
2800
2801
2802
2803
2804
2805
2806
2807
2808
2809
2810
2811
2812
2813
2814
2815
2816
2817
2818
2819

| | |
|--|---|
| | therapy for patients with pancreatic cancer |
|--|---|

Table 5. NPs mediated cell death pathways and innate immune system activation.

| Cell death pathways | NPs (Size, zeta potential and morphology) | Therapy | Therapeutic conditions | Mechanism | In vivo cancer therapy | Ref |
|---------------------|--|---------|--------------------------------------|------------------------------------|---|-------|
| Ferroptosis | TPEQM-DMA 50 nm, spherical shape | PDT | White light; 100 mW·cm ⁻² | GPX4 downregulation | CT26 subcutaneous xenograft (i.t.), tumor inhibitory effect | [362] |
| | TPA-NDTA-NP 132 nm, -16.9 mV, spherical shape | PTT | 808 nm; 0.3 W·cm ⁻² | GPX4 downregulation | Bilateral 4T1 subcutaneous xenograft (i.t. and i.v.), tumor inhibitory effect | [364] |
| | DP-HBN/RA 34.26 nm, +21 mV, hexagonal shape | RT | 6 Gy | SLC7A1 1/GPX4 downregulation | 4T1 subcutaneous xenograft (i.v.), strong inhibitory effect | [365] |
| Cuproptosis | Au@MSN-Cu/PEG/DSF ~250 nm, ~20 mV, core/shell shape | PTT | 808 nm; 1 W·cm ⁻² | DLAT, LIAS and NPL4 downregulation | 4T1 subcutaneous xenograft (i.v.), tumor inhibitory effect | [372] |
| | CuSACO 104 ± 9.5 nm, -21.5 ± 29 mV, nanocapsule shape | PTT | 1064 nm; 0.4 W·cm ⁻² | LIAS downregulation | Orthotopic GL261 glioma xenograft (i.v.), tumor inhibitory effect | [373] |

| | | | | | | |
|------------|--|---------|------------------------------------|--------------------------------------|--|-----------|
| | ES@HM-HfO ₂ :Cu 100 ± 5 nm, hollow spherical nanocage | RT | 6 Gy (2Gy × 3) | Fe-S cluster proteins downregulation | 4T1 subcutaneous xenograft (i.t.), significant tumor inhibitory effect | [376] |
| Pyroptosis | ChS-Ce6 74.4 ± 10.2 nm, -22.3 ± 3.6 mV, vesicular shape | PDT | 660 nm; 500 mW·cm ⁻² | GSDMD | LLC spinal metastasis xenograft (i.v.), strong tumor inhibitory effect | [381] |
| | Metal organic framework (IDN@MC) 130 nm, 24 ± 1.27 mV, spherical shape | PTT | 808 nm; 1 W·cm ⁻² | GSDME | 4T1 breast cancer lung metastasis xenograft (i.v.), tumor lung metastasis inhibition | [383] |
| | DAC@O-HONS 4.5 nm, spherical shape | RT | 6 Gy | GSDME | 4T1 orthotopic xenograft (i.t.), strong tumor inhibitory effect | [384] |
| STING | BG-Mn ^{gel} hydrogel | PT T | 808 nm; 1 W·cm ⁻² | pSTING, pTBK1 and pIRF3 | B16F10 subcutaneous xenograft (i.t.), tumor inhibitory effect | [390] |
| | NP PDT-56 MESS 109.4 nm, -13.9 mV, spherical shape | PD T | 808 nm; 1 W·cm ⁻² | pSTING, pTBK1 and pIRF3 | Melanoma orthotopic xenograft (i.v.), tumor inhibitory effect | [391] |

| | | | | | |
|--|----|----------|--------------------|---|-----------|
| Phosphatidyl serine- liposome loaded cGAMP 118.8 nm, -40.7 mV, spherical shape | RT | 8 Gy × 3 | pTBK1 and pIRF3 | 4T1 breast cancer lung metastasis xenograft (inhalation), strong tumor inhibitory effect | [393] |
|--|----|----------|--------------------|---|-----------|

2820

2821

2822

2823

2824

2825

2826

2827

2828

2829

2830

2831

2832

2833

2834

2835

2836

2837

2838

2839

2840

2841

Table 6. Comparison of various cell death pathways and their representative

2842

nanosystem.

2843

| Cell Death pathway | Primary inducer | Molecular mechanism | Morphological features | Representative nanosystems |
|---------------------------|---|--|--|---|
| Apoptosis | Caspase activation | Caspase-3/7/9, Cytochrome c, Annexin V+ | Cell shrinkage, chromatin condensation, apoptotic bodies | Drug-loaded polymeric NPs, Gold NPs, TiO ₂ NPs |
| Necrosis | Severe ATP depletion, membrane mechanical or thermal damage | HMGB1 release, random DNA fragmentation | Cell swelling, membrane rupture, organelle breakdown | PTT NPs, Magnetic hyperthermia |
| Ferroptosis | Iron-dependent lipid peroxidation; GPX4 inhibition | Lipid ROS, GPX4 downregulation, SLC7A11 downregulation | Mitochondrial shrinkage, membrane rupture | Iron oxide NPs, Fe-doped MOFs |
| Pyroptosis | Inflammasome activation; Gasdermin-mediated pore formation | GSDMD/E cleavage, NLRP3, IL-1 β | Cell swelling, large bubbles (pores), membrane rupture | NLRP3-activating silica NPs, LDH-coated NPs |
| Cuproptosis | Cu-induced lipoylated protein aggregation; TCA cycle stress | Lipoylated DLAT/DLST, FDX1, HSP70 | Mitochondrial proteotoxic stress, membrane damage | Copper-sulfide NPs, Cu-doped MOFs, Cu-ionophores |

2844

2845

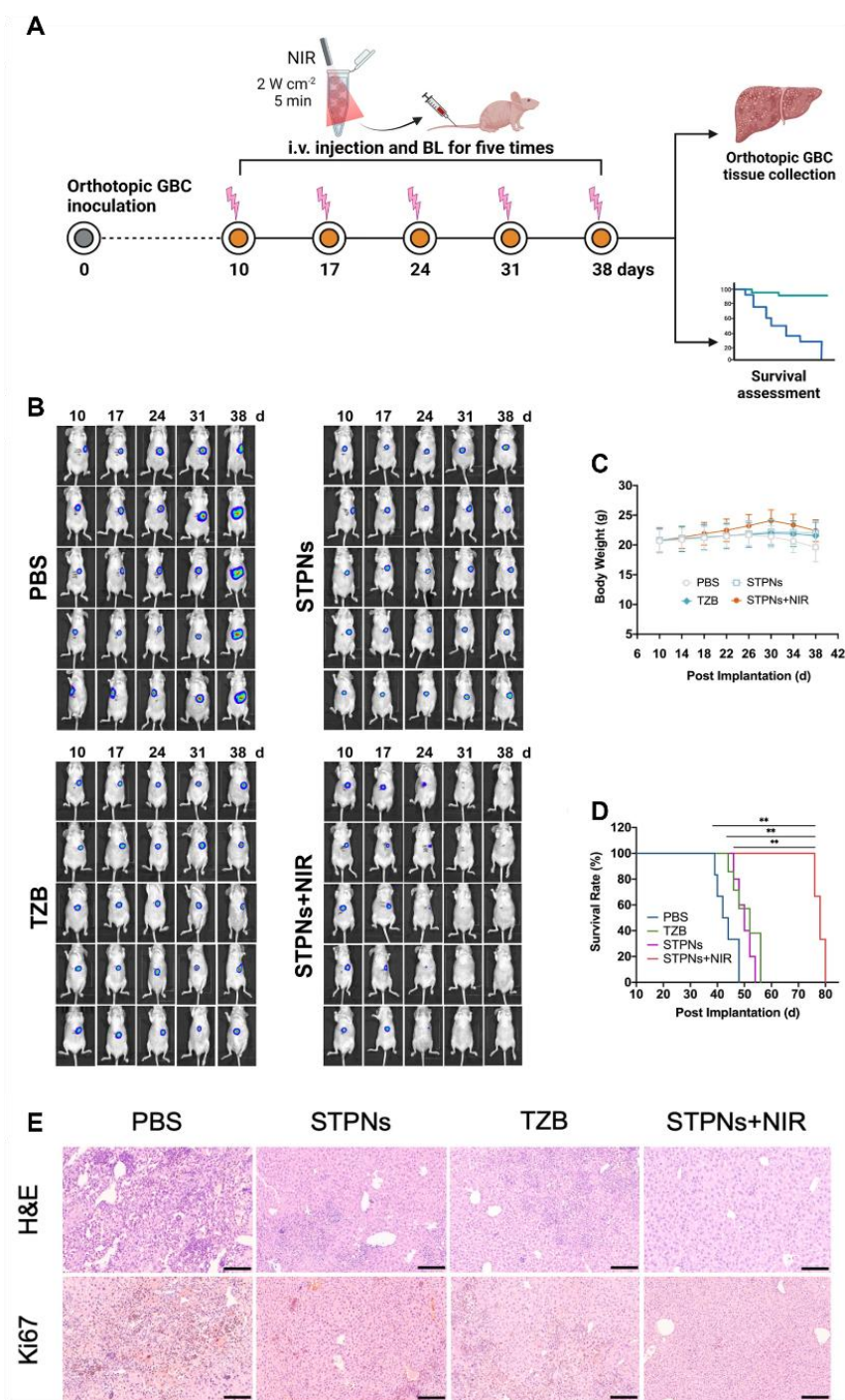
2846

2847

2848

2849

2850 **Figures**



2851

2852

2853 **Figure 1 (A)** schematic illustration of orthotopic GBC inoculation and treatment, **(B)**
2854 in vivo bioluminescence images of orthotopic GBC-bearing mice treated with various
2855 formulations irradiated by a 980 nm laser (2.0 W cm^{-2} , 5 min), **(C)** body weight changes,

2856 **(D)** Kaplan–Meier survival curves, and **(E)** H&E and Ki67 staining (scale bar = 250
2857 μm). Adapted with permission from [61], Copyright 2023 Springer Nature.

2858

2859

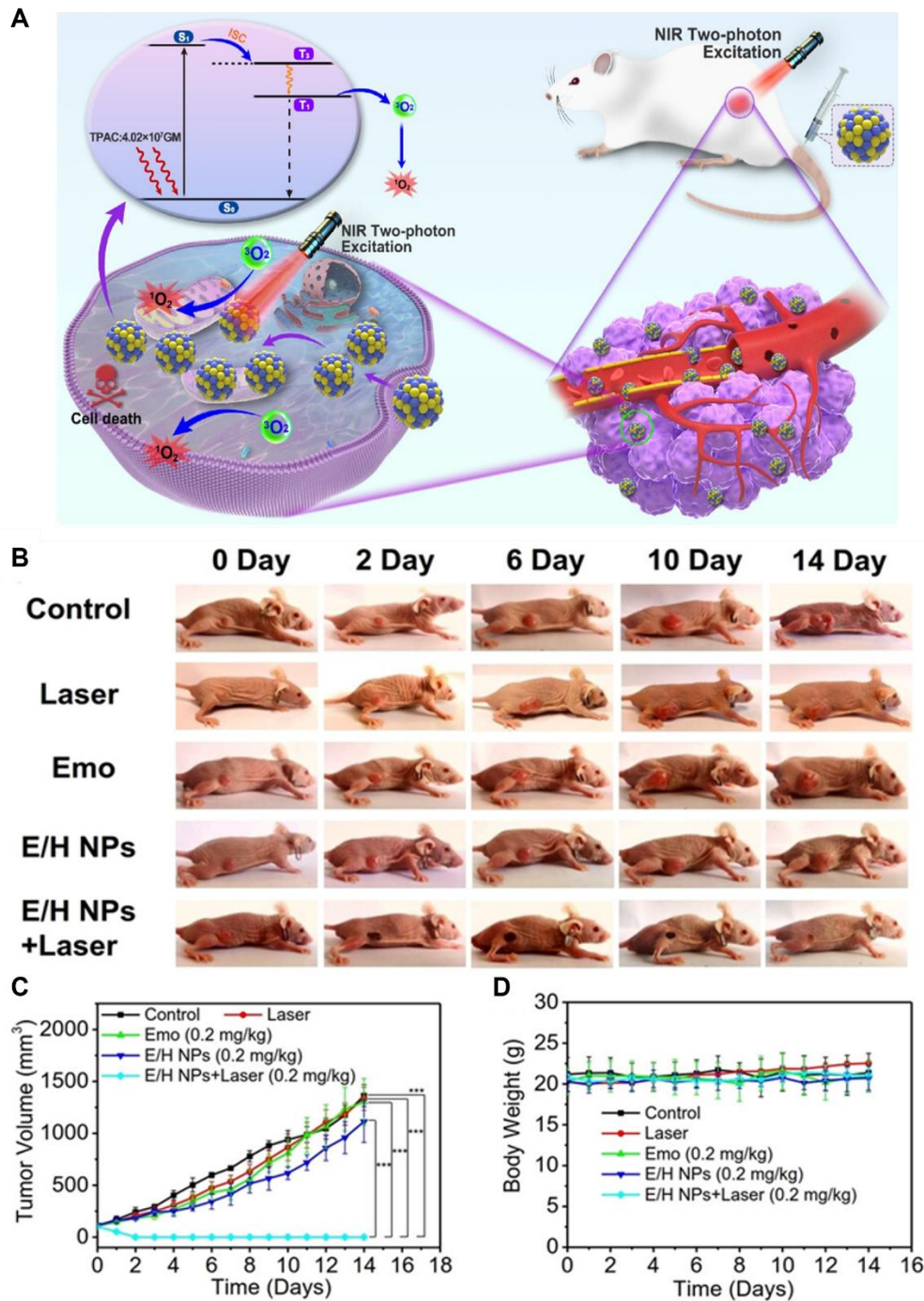
2860

2861

2862

2863

2864

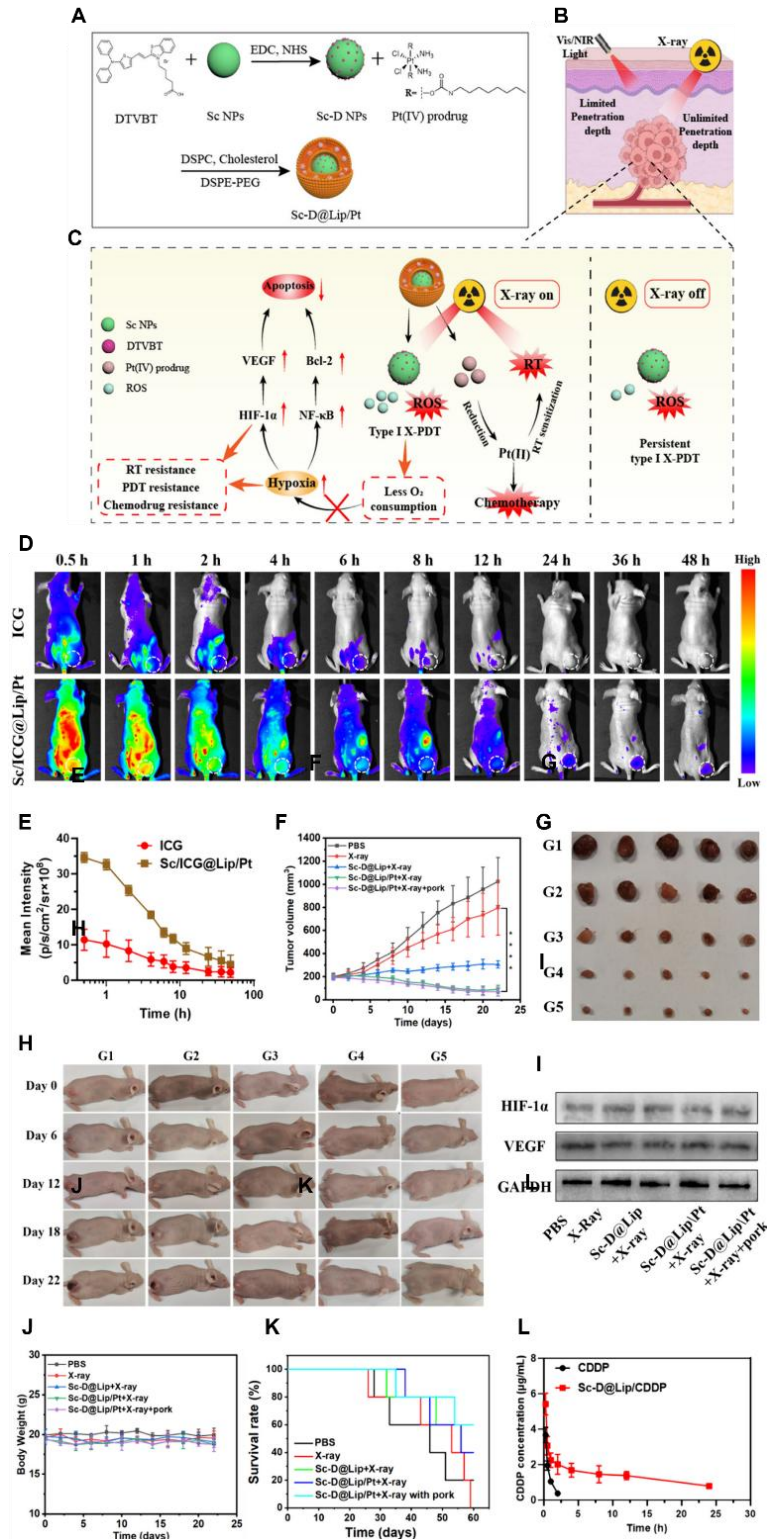


2865

2866

2867 **Figure 2 (A)** Schematic illustration of E/H NPs for efficient anti-cancer TPE-PDT, **(B)**
 2868 photographs of tumor-bearing mice at various time points after treatment, **(C)** tumor
 2869 volume changes, and **(D)** body weight changes. Adapted with permission from [73],
 2870 Copyright 2023 John Wiley and Sons .

2871



2872

2873 **Figure 3** Schematic illustration of the (A) design and (B, C) mechanism of X-ray
 2874 activated persistent type I PDT nanoplatform, (D) In vivo biodistribution of free ICG
 2875 and ICG-loaded NPs at various time points (white circles indicate tumors), (E)
 2876 Quantitative analysis of fluorescence intensity in tumor regions at different times, (F)
 2877 Tumor growth curves for each treatment group, (G) Photographs of tumors from

2878 different treatment groups, **(H)** Representative images of mice bearing 4T1 tumors after
2879 treatments at different time points, **(I)** Western blot analysis of HIF-1 α and VEGF
2880 expression in 4T1 tumors after treatments, **(J)** Body weight changes of tumor-bearing
2881 mice in each group, **(K)** Survival curves of treated mice, **(L)** Blood circulation profiles
2882 of CDDP and Sc-D@Lip/CDDP measured by HPLC following intravenous injection.
2883 Adapted with permission from [90], Copyright 2025 American Chemical Society.

2884

2885

2886

2887

2888

2889

2890

2891

2892

2893

2894

2895

2896

2897

2898

2899

2900

2901

2902

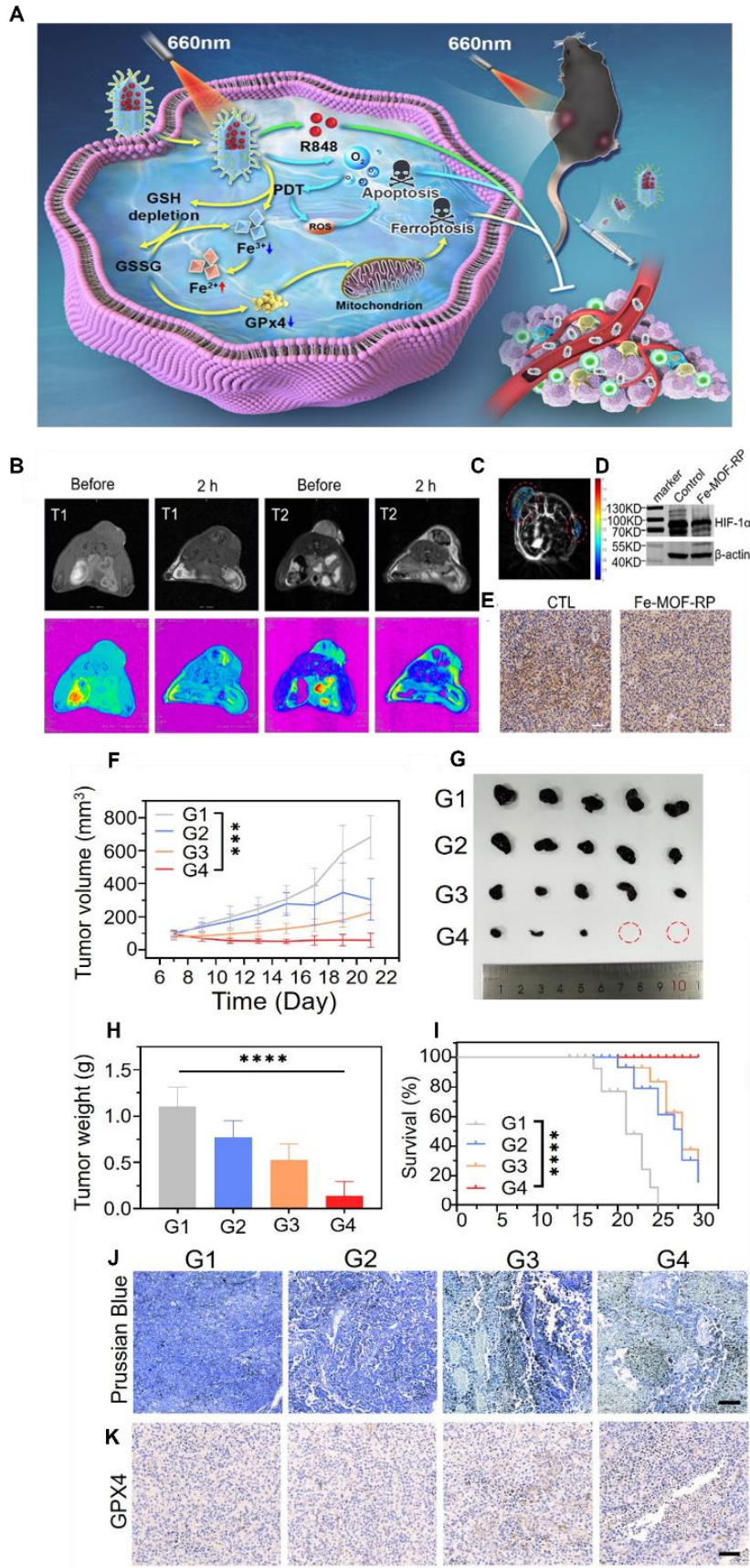
2903

2904

2905

2906

2907



2908

2909

2910 **Figure 4 (A)**Schematic Illustration of mechanism of the light-driven Fe-MOF-RP for
2911 efficient tumor therapy, **(B)** MRI imaging of tumor before and after the injection of Fe-
2912 MOF-RP, **(C)** Blood oxygen saturation measured by photoacoustic tomography after
2913 intratumoral injection of Fe-MOF-RP into the left tumor, **(D)** Western blot analysis,
2914 **(E)** immuno-staining of HIF-1 α in the tumor. In vivo anti-tumor effects of Fe-MOF-
2915 RP, **(F)** tumor growth curves of different groups, **(G)** photographs of isolated tumors,
2916 **(H)** tumor weights after treatment, **(I)** survival rates, **(J)** iron content in tumor tissues
2917 assessed by Prussian blue staining, and **(K)** GPX4 expression in tumor tissues evaluated
2918 by immunohistochemistry. Adapted with permision from [131], Copyright 2024
2919 American Chemical Society.

2920

2921

2922

2923

2924

2925

2926

2927

2928

2929

2930

2931

2932

2933

2934

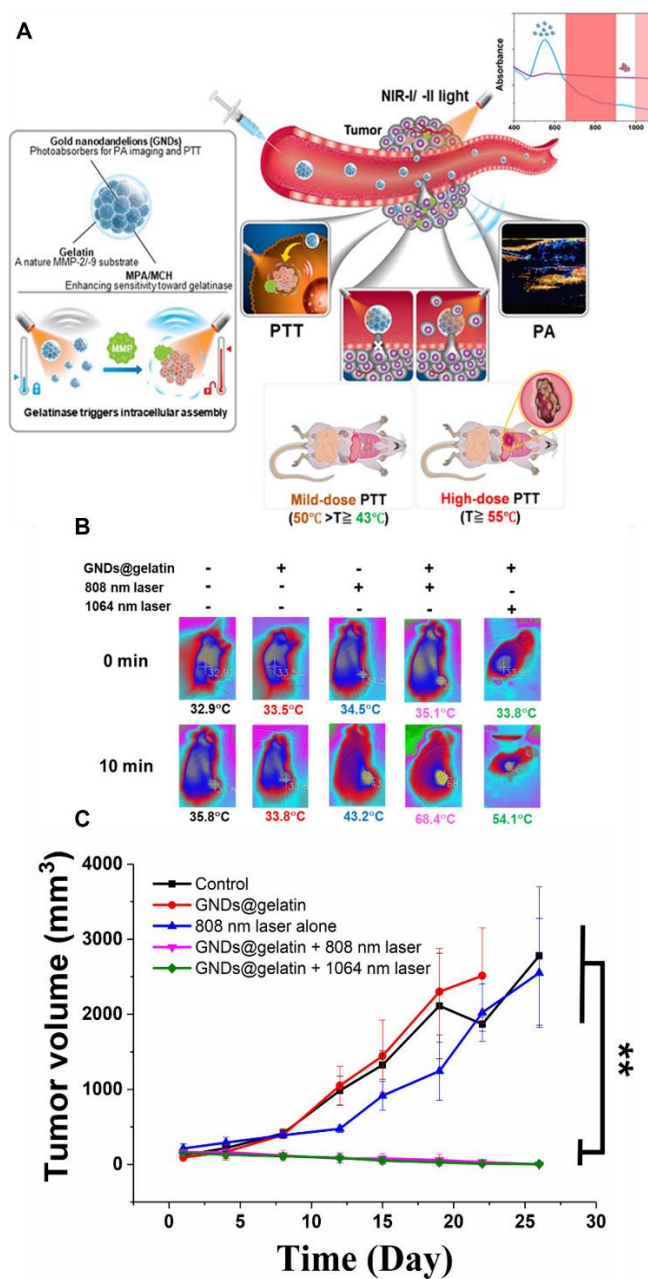
2935

2936

2937

2938

2939
 2940
 2941
 2942
 2943

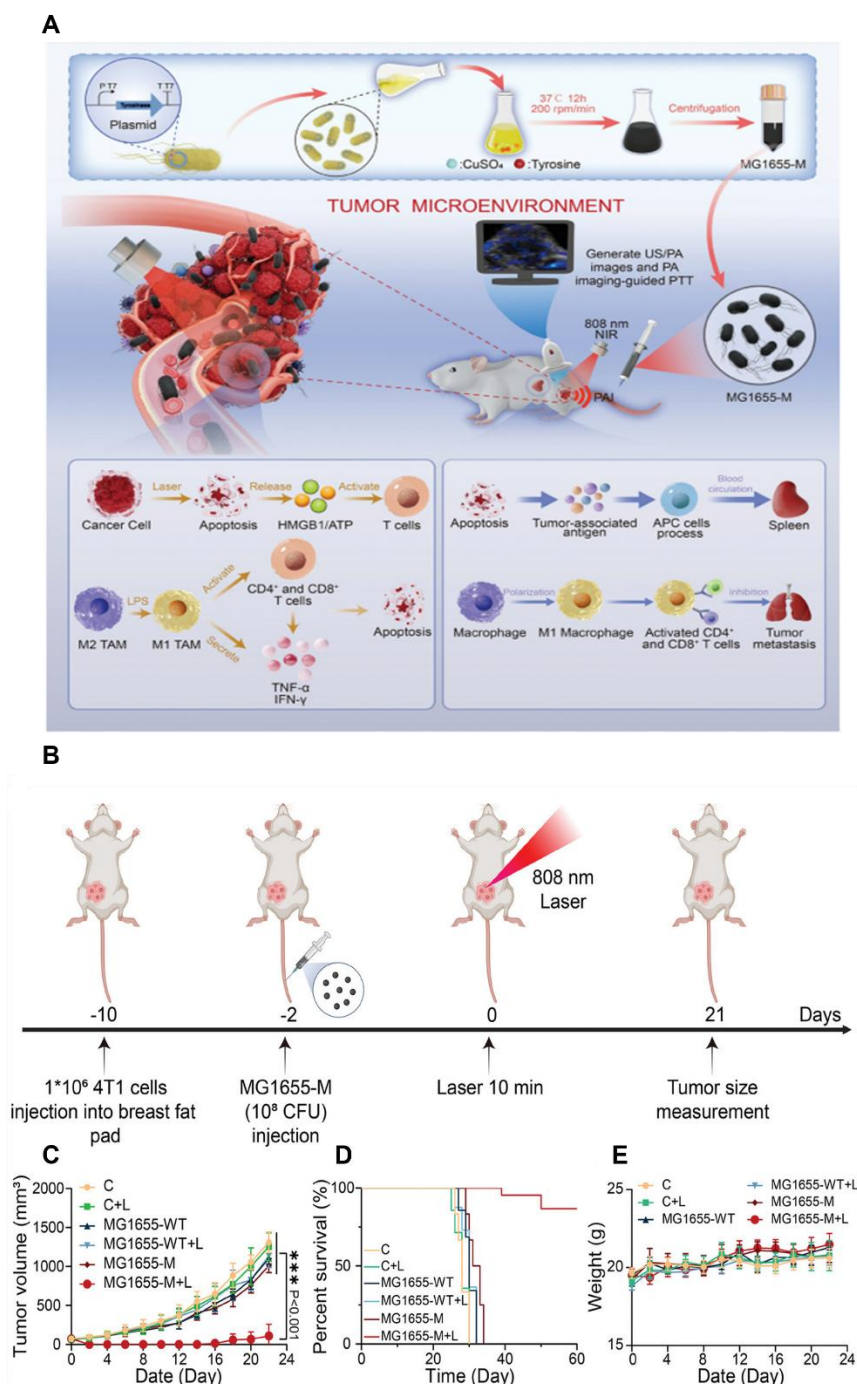


2944

2945 **Figure 5 (A)** Schematic of TME-responsive GNDs@gelatin and PAI-guided PTT and
 2946 metastasis modulation. In vivo NIR-I and NIR-II PTT against tumors, **(B)**
 2947 representative IR thermal images of C6 tumor-bearing mice injected with

2948 GNDs@gelatin and irradiated with an 808 nm ($1 \text{ W} \cdot \text{cm}^{-2}$) or 1064 nm ($1 \text{ W} \cdot \text{cm}^{-2}$) laser
 2949 at different time points, and (C) tumor growth curves after various treatments over 30
 2950 days. Adapted with permission from [202], Copyright 2024 American Chemical
 2951 Society.

2952



2953

2954 **Figure 6 (A)** Schematic of MG1655-M for photoacoustic image-guided photothermal
 2955 therapy and its anti-tumor mechanisms, **(B)** schematic of MG1655-M-mediated
 2956 photothermal immunotherapy in the 4T1 primary tumor model, **(C)** tumor growth

2957 curves in mice after different treatments, **(D)** survival curves of mice in each group, and
2958 **(E)** body weight curves of mice receiving different treatments. Adapted with
2959 permission from [220], Copyright 2024 John Wiley and Sons.

2960

2961

2962

2963

2964

2965

2966

2967

2968

2969

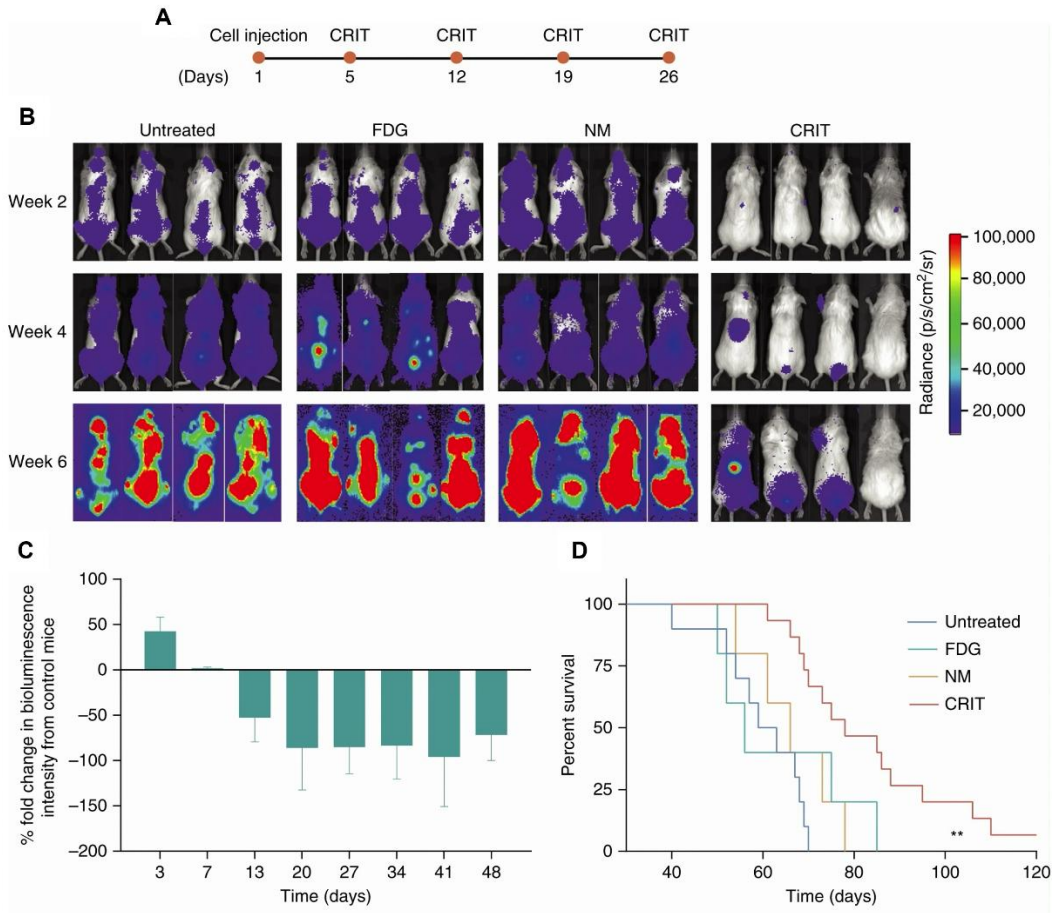
2970

2971

2972

2973

3002



3003

3004

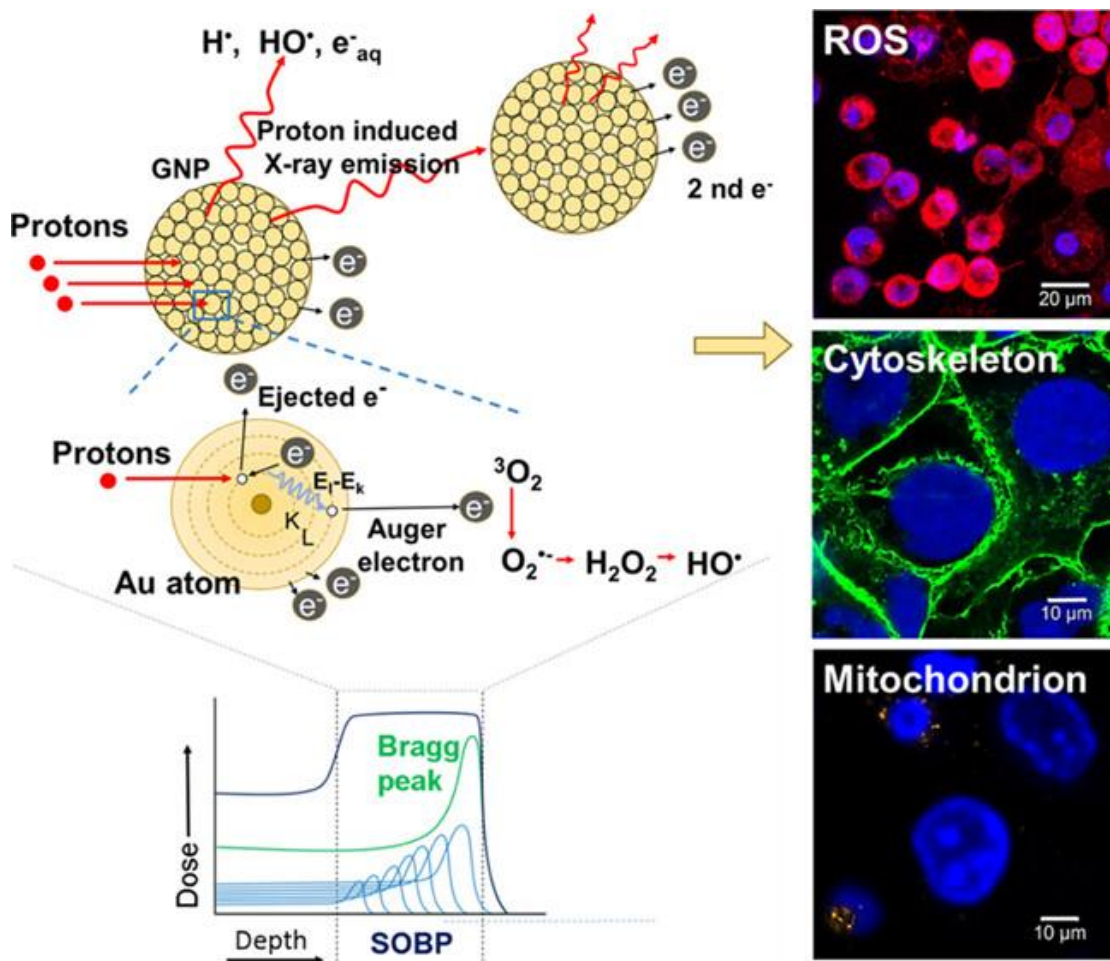
3005 **Figure 8** Response of multiple myeloma to CRIT. **(A)** Treatment timeline, **(B)** BLI of
3006 representative multiple myeloma-bearing mice under various treatments, **(C)** changes
3007 in bioluminescence intensity following treatments relative to untreated control, **(D)**
3008 survival comparison showing twofold increase in survival for treated mice versus
3009 controls. Adapted with permission from [291], Copyright 2018 Springer Nature.

3010

3011

3012

3013



3014

3015

3016 **Figure 9** Mechanism of ROS production by incident protons interacting with Au atoms
 3017 in a GNP, releasing primary electrons, X-rays, and Auger electrons, which may react
 3018 with surrounding water molecules to generate ROS. Adapted with permission from
 3019 [308], Copyright 2023 American Chemical Society.

3020

3021

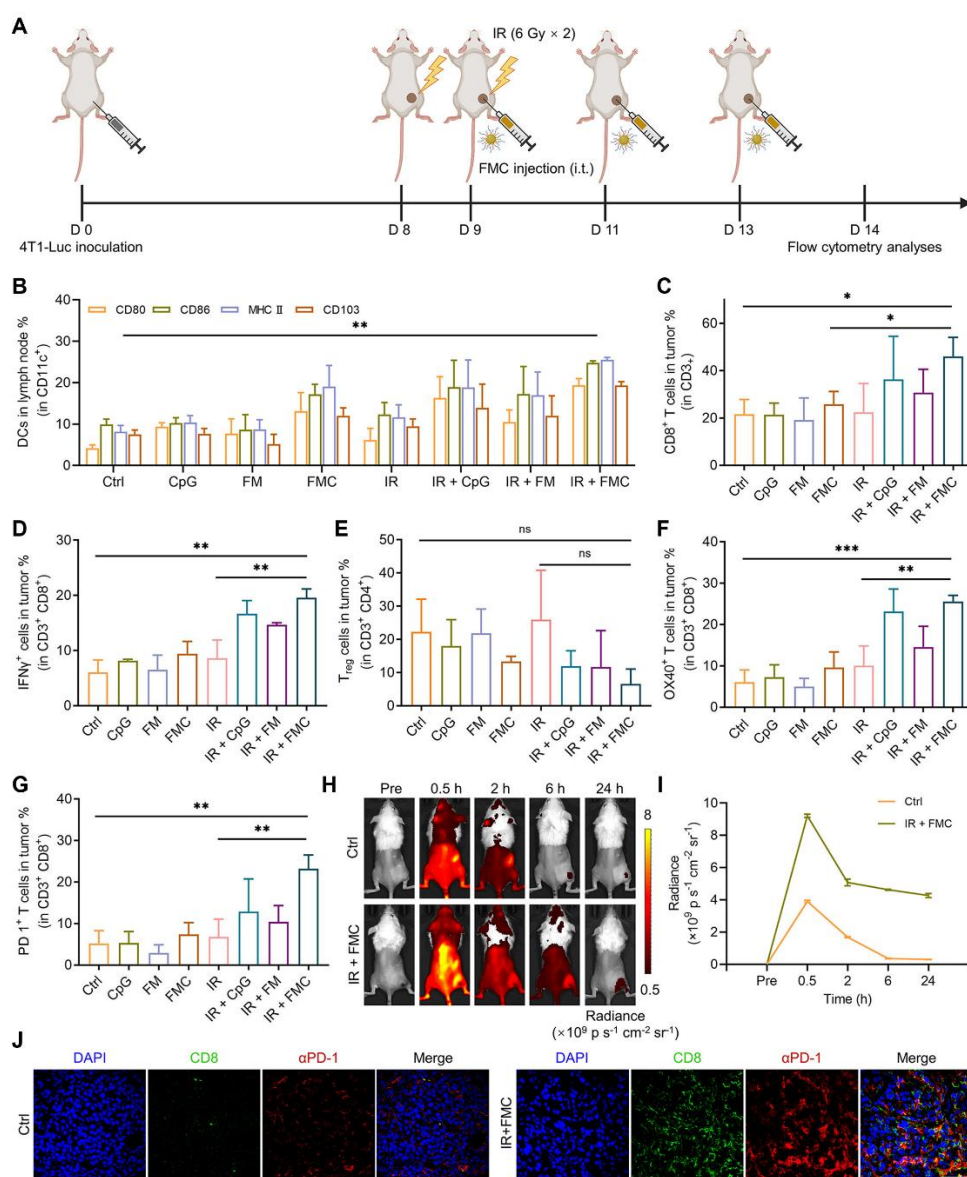
3022

3023

3024

3025

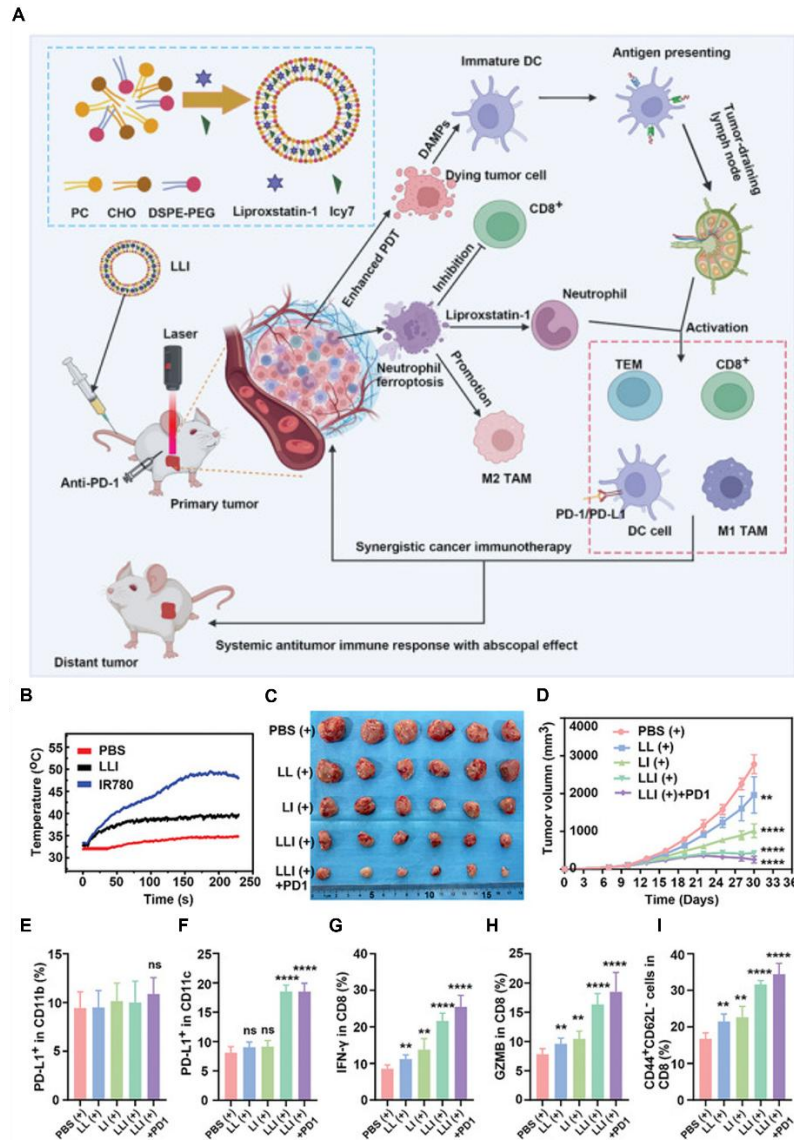
3026



3028

3029

3030 **Figure 10** Immune activation effects of FMC-based ISV in vivo. (A) Experimental
 3031 setup to assess immune responses triggered by FMC-based ISV, (B) percentage of
 3032 CD11c⁺ dendritic cells in tumor-draining lymph nodes after different treatments, (C–
 3033 G) flow cytometry analysis of tumor-infiltrating T cell subtypes after treatment: CD8⁺
 3034 T cells (C), IFN-γ⁺ CD8⁺ T cells (D), Tregs (E), OX40⁺ T cells (F), and PD-1⁺ T cells
 3035 (G), T cells were identified as CD3⁺CD8⁺, (H) in vivo FLI of Cy5-labeled αPD-1, (I)
 3036 changes in tumor-site fluorescence intensity over time, (J) fluorescence images of
 3037 tumor sections showing CD8⁺ cells (green, FITC), αPD-1 (red, Cy5), and nuclei (blue,
 3038 DAPI). Scale bar: 50 μm. Adapted with permission from [356], Copyright 2024
 3039 American Chemical Society.

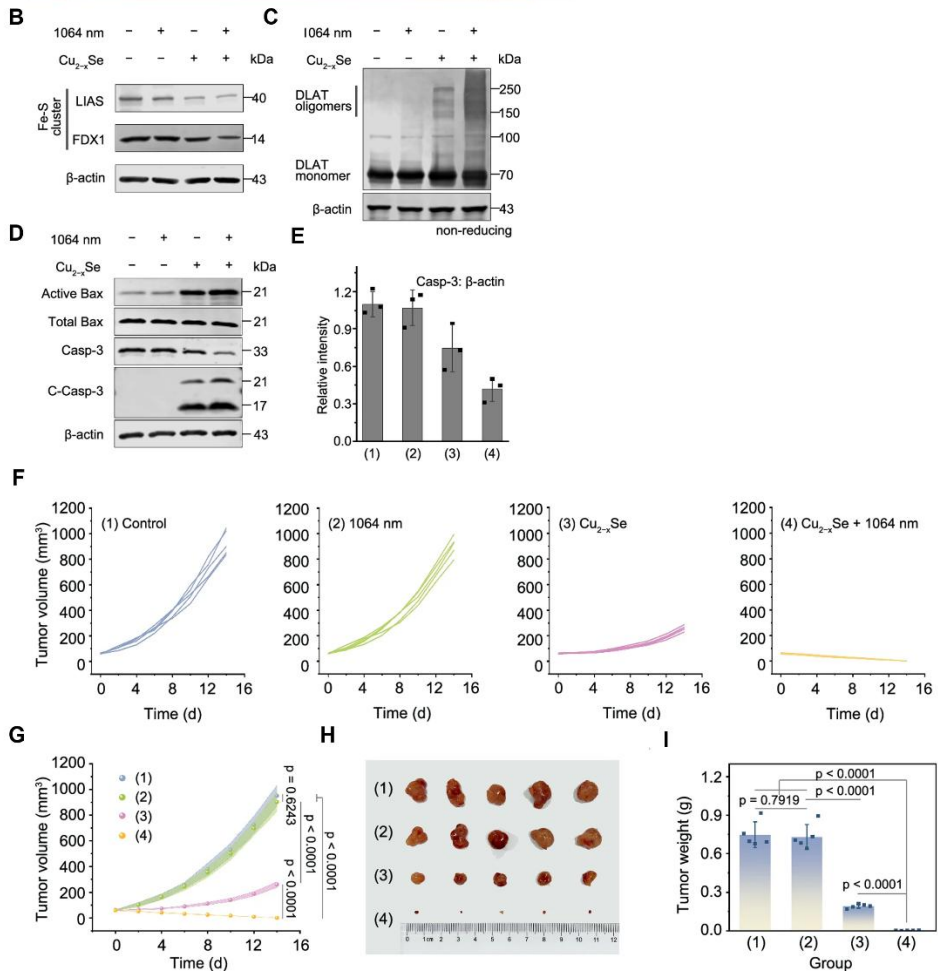
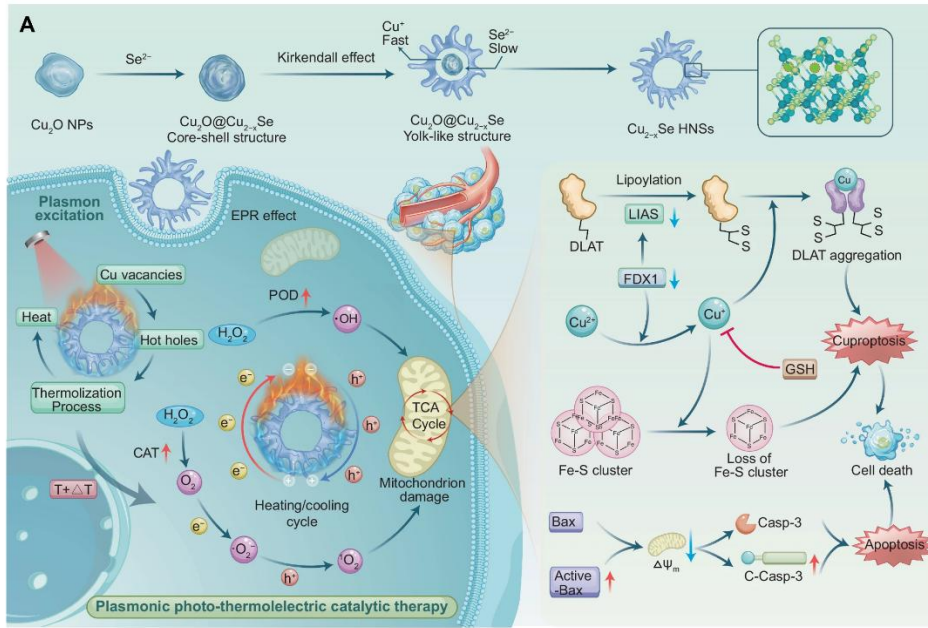


3040

3041

3042 **Figure 11 (A)** Schematic illustration of the preparation of multifunctional nanodrug
 3043 LLI, its immunogenic PDT, and systemic anti-tumor immunity with abscopal effects,
 3044 **(B)** tumor temperature changes at different time points after intravenous injection of
 3045 various solutions followed by laser irradiation (808 nm, 0.5 W·cm⁻²), **(C)** macroscopic
 3046 images of tumors from mice receiving different treatments, **(D)** tumor growth curves in
 3047 mice treated with various nanodrugs, **(E, F)** statistical analysis of PD-L1 expression on
 3048 CD11b⁺ and CD11c⁺ cells in tumor tissues after different treatments, **(G, H)** statistical
 3049 analysis of IFN- γ and GZMB expression in CD8⁺T cells after treatments, and **(I)**
 3050 statistical analysis of TEM percentages in spleens after different treatments. Adapted
 3051 with permission from [361], Copyright 2024 John Wiley and Sons.

3052



3053

3054

3055

3056 **Figure 12 (A)** Schematic illustration of the synthesis of Cu_{2-x}Se HNSs, and anti-tumor
3057 mechanism by synergistic therapies, **(B,C)** Western blotting analysis on the expression
3058 levels of cuproptosis-related proteins in CT26 cells treated with various
3059 conditions, **(D)** the expression levels of apoptosis-related proteins in CT26 cells
3060 treated with various conditions examined by western blotting, **(E)** Western blotting
3061 intensity ratio of Casp-3 and β -actin, tumor volume **(F, G)** individual and consolidated
3062 tumor growth curves of mouse undergoing various treatments, **(H)** photograph of the
3063 dissected tumor, and **(I)** tumor weight. Adapted with permission from [375], Copyright
3064 2024 Springer Nature.

3065

3066

3067

3068

3069

3070

3071

3072

3073

3074

3075

3076

3077

3078

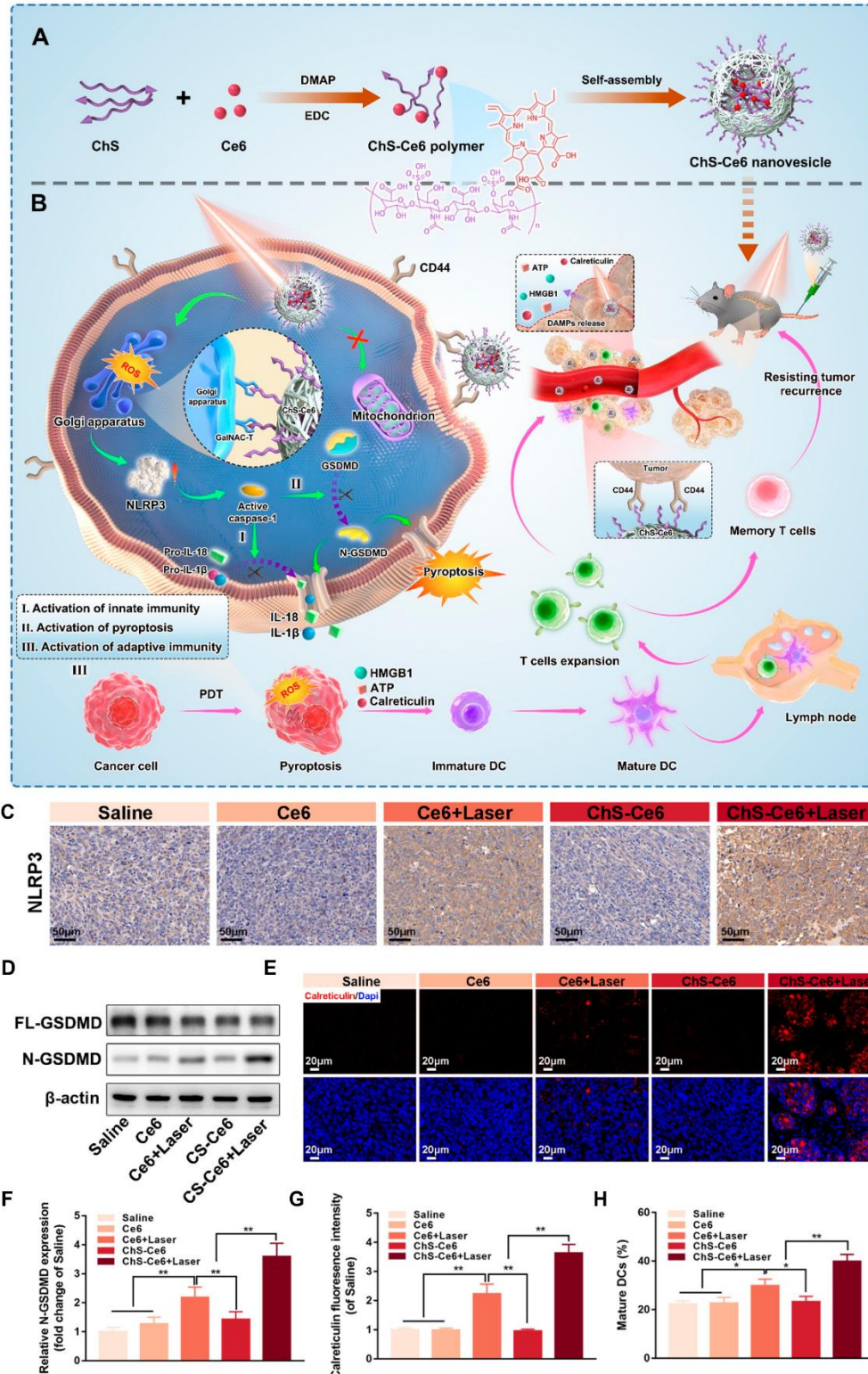
3079

3080

3081

3082

3083



3084

3085 **Figure 13 (A,B)** Schematic illustration of the synthesis and application of Golgi
 3086 apparatus-targeted ChS-Ce6 nanovesicles for NLRP3-dependent immunogenic
 3087 pyroptosis to remodel the tumor microenvironment, **(C)** representative
 3088 immunohistochemical staining of NLRP3 in tumor tissues, **(D, F)** Western blotting and

3089 quantification of N-GSDMD expression in tumor tissues **(E, G)** immunofluorescence
3090 staining and quantification of CRT expression in tumor tissues and **(H)** flow cytometry
3091 quantification of mature DC in tumor tissues. Adapted with permission from [381],
3092 Copyright 2023 American Chemical Society.

3093

3094

3095

3096

3097

3098

3099

3100

3101

3102

3103

3104

3105

3106

3107

3108

3109

3110

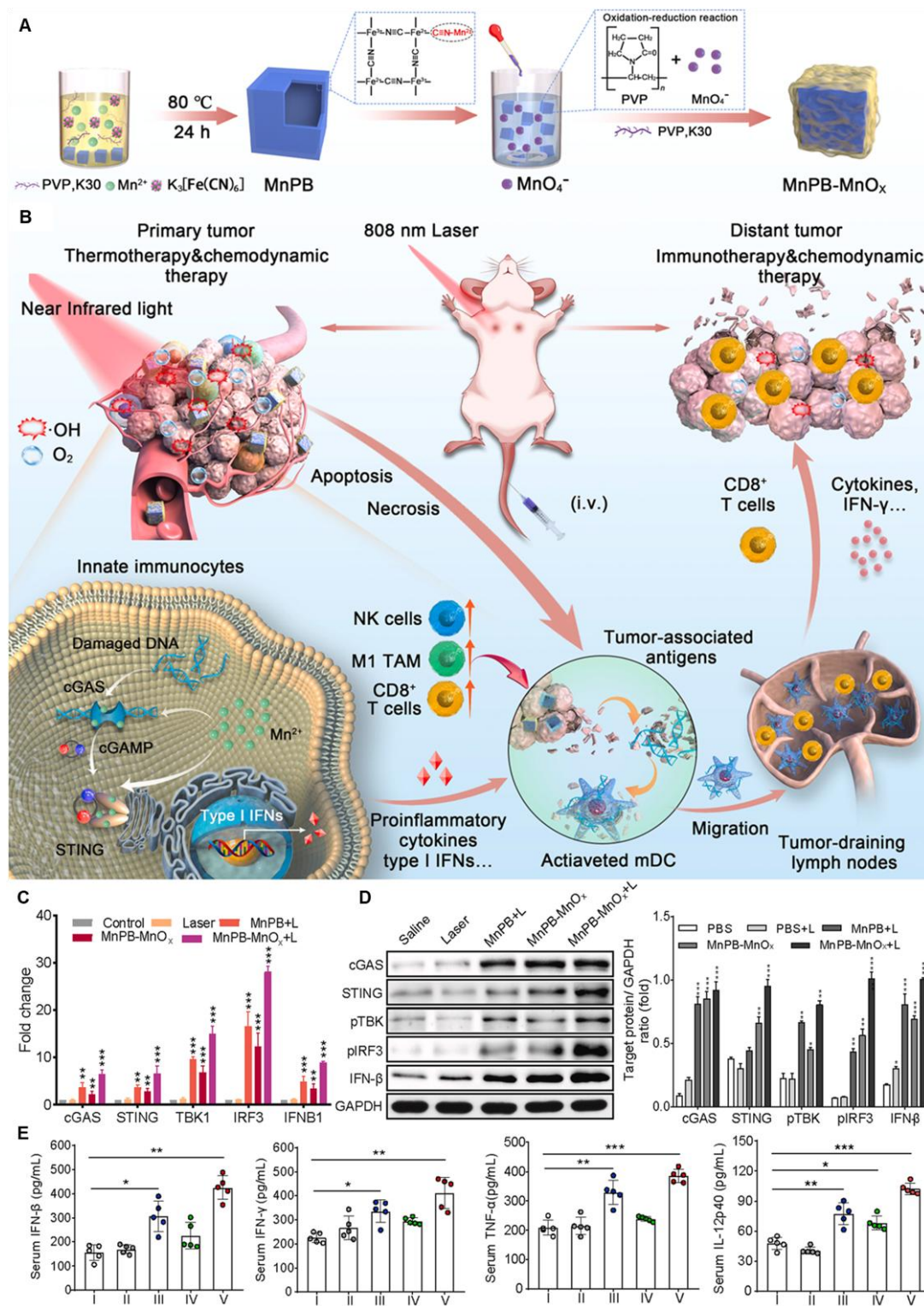
3111

3112

3113

3114

3115



3116

3117 **Figure 14 (A)** Schematic illustration of the preparation of MnPB-MnO_x nanosystems,
 3118 **(B)** mechanism of anti-tumor immune responses induced by combined noninvasive
 3119 photothermal ablation with Mn²⁺-augmented cGAS-STING pathway activation for
 3120 effective cancer immunotherapy, **(C)** QRT-PCR analysis of cGAS, STING, TBK1,
 3121 IRF3, and IFNB1 expression in primary tumors after the indicated treatments, **(D)**
 3122 expression levels of cGAS-STING-related proteins, and **(E)** cytokine levels in the

3123 serum of mice in each group on day 10 after various treatments. Adapted with
3124 permission from [396], Copyright 2023 Elsevier.

3125

3126

3127

3128

3129

3130

3131

3132

3133

3134

3135

3136

3137

3138

3139

3140

3141

3142

3143

3144

3145

3146

3147

3148

3149 **References**

- 3150 1. Siegel RL, Giaquinto AN, Jemal A. Cancer statistics, 2024. *CA Cancer J Clin.*
3151 2024; 74: 12-49.
- 3152 2. Juthani R, Punatar S, Mittra I. New light on chemotherapy toxicity and its
3153 prevention. *BJC Rep.* 2024; 2: 41.
- 3154 3. Nurgali K, Jagoe RT, Abalo R. Editorial: Adverse Effects of Cancer
3155 Chemotherapy: Anything New to Improve Tolerance and Reduce Sequelae? *Front*
3156 *Pharmacol.* 2018; 9: 245.
- 3157 4. Overchuk M, Weersink RA, Wilson BC, Zheng G. Photodynamic and
3158 Photothermal Therapies: Synergy Opportunities for Nanomedicine. *ACS Nano.* 2023;
3159 17: 7979-8003.
- 3160 5. Li X, Lovell JF, Yoon J, Chen X. Clinical development and potential of
3161 photothermal and photodynamic therapies for cancer. *Nat Rev Clin Oncol.* 2020; 17:
3162 657-74.
- 3163 6. Allamyradov Y, ben Yosef J, Annamuradov B, Ateyeh M, Street C, Whipple H,
3164 et al. Photodynamic Therapy Review: Past, Present, Future, Opportunities and
3165 Challenges. *Photochem.* 2024; 4: 434-61.
- 3166 7. Bian W, Wang Y, Pan Z, Chen N, Li X, Wong W-L, et al. Review of
3167 Functionalized Nanomaterials for Photothermal Therapy of Cancers. *ACS Appl Nano*
3168 *Mater.* 2021; 4: 11353-85.
- 3169 8. Zhao X, Liu J, Fan J, Chao H, Peng X. Recent progress in photosensitizers for
3170 overcoming the challenges of photodynamic therapy: from molecular design to
3171 application. *Chem Soc Rev.* 2021; 50: 4185-219.
- 3172 9. Zhao J, Zhong D, Zhou S. NIR-I-to-NIR-II fluorescent nanomaterials for
3173 biomedical imaging and cancer therapy. *J Mater Chem B.* 2018; 6: 349-65.
- 3174 10. Zhang Z, Du Y, Shi X, Wang K, Qu Q, Liang Q, et al. NIR-II light in clinical
3175 oncology: opportunities and challenges. *Nat Rev Clin Oncol.* 2024; 21: 449-67.
- 3176 11. Soleimany A, Aghmiouni D, Amirikhah M, Shokrgozar M, Khoee S, Sarmiento B.
3177 Two-Photon Mediated Cancer Therapy: A Comprehensive Review on Two-Photon
3178 Photodynamic Therapy and Two-Photon-Activated Therapeutic Delivery Systems.
3179 *Adv Funct Mater.* 2024; 34.
- 3180 12. Juvekar V, Joon Lee D, Gwan Park T, Samanta R, Kasar P, Kim C, et al. Two-
3181 photon excitation photosensitizers for photodynamic therapy: From small-molecules to
3182 nano-complex systems. *Coord Chem Rev.* 2024; 506: 215711.
- 3183 13. Chen H, Ding B, Ma Pa, Lin J. Recent progress in upconversion nanomaterials for
3184 emerging optical biological applications. *Adv Drug Deliv Rev.* 2022; 188: 114414.

- 3185 14. Zhang J, Du J, Jia S, Li Y, Ågren H, Prasad P, et al. Recent Advances in
3186 Upconversion Nanoparticles for Therapeutics: From Fundamentals to Cutting-Edge
3187 Applications. *Electron*. 2025; 3.
- 3188 15. Gao C, Zheng P, Liu Q, Han S, Li D, Luo S, et al. Recent Advances of
3189 Upconversion Nanomaterials in the Biological Field. *Nanomaterials (Basel)*. 2021; 11.
- 3190 16. Tsang CY, Zhang Y. Nanomaterials for light-mediated therapeutics in deep tissue.
3191 *Chem Soc Rev*. 2024; 53: 2898-931.
- 3192 17. Duan S, Hu Y, Zhao Y, Tang K, Zhang Z, Liu Z, et al. Nanomaterials for
3193 photothermal cancer therapy. *RSC Adv*. 2023; 13: 14443-60.
- 3194 18. Cui X, Ruan Q, Zhuo X, Xia X, Hu J, Fu R, et al. Photothermal Nanomaterials: A
3195 Powerful Light-to-Heat Converter. *Chem Rev*. 2023; 123: 6891-952.
- 3196 19. Omidian H, Dey Chowdhury S. Advances in Photothermal and Photodynamic
3197 Nanotheranostics for Precision Cancer Treatment. *J Nanotheranostics*. 2024; 5: 228-52.
- 3198 20. Gao D, Guo X, Zhang X, Chen S, Wang Y, Chen T, et al. Multifunctional
3199 phototheranostic nanomedicine for cancer imaging and treatment. *Mater Today Bio*.
3200 2020; 5: 100035.
- 3201 21. Bodei L, Herrmann K, Schöder H, Scott AM, Lewis JS. Radiotheranostics in
3202 oncology: current challenges and emerging opportunities. *Nat Rev Clin Oncol*. 2022;
3203 19: 534-50.
- 3204 22. Gupta D, Roy P, Sharma R, Kasana R, Rathore P, Gupta TK. Recent
3205 nanotheranostic approaches in cancer research. *Clin Exp Med*. 2024; 24: 8.
- 3206 23. Cui M, Zhu M, Tang D, Cao Z, Xiao H, Shang K. Nanotheranostics with
3207 Radionuclides for Cancer Diagnosis and Therapy. *Adv Funct Mater*. 2025; 35.
- 3208 24. Liu N, Su X, Sun X. Cerenkov radiation-activated probes for deep cancer
3209 theranostics: a review. *Theranostics*. 2022; 12: 7404-19.
- 3210 25. Wang Q, Liu N, Hou Z, Shi J, Su X, Sun X. Radioiodinated Persistent
3211 Luminescence NanoplatforM for Radiation-Induced Photodynamic Therapy and
3212 Radiotherapy. *Adv Healthc Mater*. 2021; 10: e2000802.
- 3213 26. Sun N, Wang T, Zhang S. Radionuclide-labelled nanoparticles for cancer
3214 combination therapy: a review. *J Nanobiotechnology*. 2024; 22: 728.
- 3215 27. Pijeira MSO, Viltres H, Kozempel J, Sakmár M, Vlk M, İlem-Özdemir D, et al.
3216 Radiolabeled nanomaterials for biomedical applications: radiopharmacy in the era of
3217 nanotechnology. *EJNMMI Radiopharm Chem*. 2022; 7: 8.
- 3218 28. Lemaître TA, Burgoyne AR, Ooms M, Parac-Vogt TN, Cardinaels T. Inorganic
3219 Radiolabeled Nanomaterials in Cancer Therapy: A Review. *ACS Appl Nano Mater*.
3220 2022; 5: 8680-709.
- 3221 29. Qi H, Li Y, Geng Y, Wan X, Cai X. Nanoparticle-mediated immunogenic cell
3222 death for cancer immunotherapy. *Int J Pharm*. 2024; 656: 124045.

- 3223 30. Ma C, Cheng Z, Tan H, Wang Y, Sun S, Zhang M, et al. Nanomaterials: leading
3224 immunogenic cell death-based cancer therapies. *Front Immunol.* 2024; 15: 1447817.
- 3225 31. Luobin L, Wanxin H, Yingxin G, Qinzhou Z, Zefeng L, Danyang W, et al.
3226 Nanomedicine-induced programmed cell death in cancer therapy: mechanisms and
3227 perspectives. *Cell Death Discov.* 2024; 10: 386.
- 3228 32. Li Y, Guo Y, Zhang K, Zhu R, Chen X, Zhang Z, et al. Cell Death Pathway
3229 Regulation by Functional Nanomedicines for Robust Antitumor Immunity. *Adv Sci*
3230 *(Weinh).* 2024; 11: e2306580.
- 3231 33. Ying X, Chen Q, Yang Y, Wu Z, Zeng W, Miao C, et al. Nanomedicines
3232 harnessing cGAS-STING pathway: sparking immune revitalization to transform 'cold'
3233 tumors into 'hot' tumors. *Mol Cancer.* 2024; 23: 277.
- 3234 34. Wang Y, Liu Y, Zhang J, Peng Q, Wang X, Xiao X, et al. Nanomaterial-mediated
3235 modulation of the cGAS-STING signaling pathway for enhanced cancer
3236 immunotherapy. *Acta Biomater.* 2024; 176: 51-76.
- 3237 35. Phipps J, Fan W, Kapoor M, Huynh D, Ma S. Photodynamic therapy and its
3238 evolving innovation. *Chem Commun (Camb).* 2026; 62: 3194-209.
- 3239 36. El-Sadek MZ, El-Aziz MKA, Shaaban AH, Mostafa SA, Wadan A-HS.
3240 Advancements and emerging trends in photodynamic therapy: innovations in cancer
3241 treatment and beyond. *Photochem Photobiol Sci.* 2025; 24: 1489-511.
- 3242 37. Tang Y, He C, Chen C, Zhuang J, Song W, Wang D, et al. NIR-II-Excited Type-I
3243 Conjugated Polymer Photosensitizer for Cancer Photodynamic Therapy. *Adv Mater.*
3244 2026; 38.
- 3245 38. Yang Y, Jiang S, Stanciu S, Peng H, Wu A, Yang F. Photodynamic therapy with
3246 NIR-II probes: Review on state-of-the-art tools and strategies. *Mater Horiz.* 2024; 11.
- 3247 39. Sangwan S, Kumar D, Thakur P, Thakur A. A comprehensive review on
3248 upconversion nanoparticles: From synthesis strategies to multifunctional applications.
3249 *Next Mater.* 2026; 11: 101697.
- 3250 40. Liu X, Jiang C, Chen G. Brightening Upconversion Nanoparticles. *Acc Chem Res.*
3251 2026.
- 3252 41. Lv K, Hou M, Kou Y, Yu H, Liu M, Zhao T, et al. Black Titania Janus Mesoporous
3253 Nanomotor for Enhanced Tumor Penetration and Near-Infrared Light-Triggered
3254 Photodynamic Therapy. *ACS Nano.* 2024; 18: 13910-23.
- 3255 42. Liu W, He S, Ma X, Lv C, Gu H, Cao J, et al. Near-Infrared Heptamethine Cyanine
3256 Photosensitizers with Efficient Singlet Oxygen Generation for Anticancer
3257 Photodynamic Therapy. *Angew Chem Int Ed Engl.* 2024; 63: e202411802.
- 3258 43. Wang W, Zhu C, Zhang B, Feng Y, Zhang Y, Li J. Correction to “Self-Assembled
3259 Nano-PROTAC Enables Near-Infrared Photodynamic Proteolysis for Cancer Therapy”.
3260 *J Am Chem Soc.* 2023; 145: 22855-6.

3261 44. Ding Q, Li M, Mei L, Hua S, Li Z, Mengji R, et al. Immune-Evasive NIR-II
3262 Nanoplatfoms for Repeatable Photodynamic Therapy. *J Am Chem Soc.* 2026; 148:
3263 11879-92.

3264 45. Zhao Y-Y, Xu Y, Wen J, Jeong H, Kim H, Li X, et al. An O₂-Independent
3265 Copper(II) Phototherapeutic Agent for Photoactivating H₂O₂ to Enhance Antitumor
3266 Immunotherapy. *J Am Chem Soc.* 2025; 147: 48072-86.

3267 46. Hu Q, Du J, Abedi SAA, Liu X, Long S, Sun W, et al. Unveiling the Power of
3268 Dark State Photocages: An Efficient Pathway to Triplet State Under Near-Infrared
3269 Light Irradiation. *Angew Chem Int Ed Engl.* 2025; 64: e202504670.

3270 47. Shen J, Xu B, Zheng Y, Zhao X, Qi H, Tang Y, et al. Near-Infrared Light-
3271 Responsive Immunomodulator Prodrugs Rejuvenating Immune Microenvironment for
3272 "Cold" Tumor Photoimmunotherapy. *Angew Chem Int Ed Engl.* 2025; 64: e202425309.

3273 48. Li CA, Nan J, Ye Q, Zheng B, Dai X, Li J, et al. Amplifying Anti-Tumor Immune
3274 Responses via Mitochondria-Targeting Near-Infrared Photodynamic Therapy. *Adv Sci*
3275 *(Weinh).* 2025; 12: e05525.

3276 49. Song CW, Park H, Griffin RJ. Improvement of tumor oxygenation by mild
3277 hyperthermia. *Radiat Res.* 2001; 155: 515-28.

3278 50. Feng L, Tao D, Dong Z, Chen Q, Chao Y, Liu Z, et al. Near-infrared light
3279 activation of quenched liposomal Ce6 for synergistic cancer phototherapy with
3280 effective skin protection. *Biomaterials.* 2017; 127: 13-24.

3281 51. Guo X, Zhang M, Qin J, Li Z, Rankl C, Jiang X, et al. Revealing the Effect of
3282 Photothermal Therapy on Human Breast Cancer Cells: A Combined Study from
3283 Mechanical Properties to Membrane HSP70. *ACS Appl Mater Interfaces.* 2023; 15:
3284 21965-73.

3285 52. Tang Z, Zhao P, Ni D, Liu Y, Zhang M, Wang H, et al. Pyroelectric nanoplatfom
3286 for NIR-II-triggered photothermal therapy with simultaneous pyroelectric dynamic
3287 therapy. *Mater Horiz.* 2018; 5: 946-52.

3288 53. Younis MR, Wang C, An R, Wang S, Younis MA, Li Z-Q, et al. Low Power Single
3289 Laser Activated Synergistic Cancer Phototherapy Using Photosensitizer Functionalized
3290 Dual Plasmonic Photothermal Nanoagents. *ACS Nano.* 2019; 13: 2544-57.

3291 54. Zhang Y, Lei P, Zhu X, Zhang Y. Full shell coating or cation exchange enhances
3292 luminescence. *Nat Commun.* 2021; 12: 6178.

3293 55. Zhang Y, Zhu X, Zhang J, Wu Y, Liu J, Zhang Y. Synergistic upconversion
3294 photodynamic and photothermal therapy under cold near-infrared excitation. *J Colloid*
3295 *Interface Sci.* 2021; 600: 513-29.

3296 56. Wu C, Wu Y, Zhu X, Zhang J, Liu J, Zhang Y. Near-infrared-responsive
3297 functional nanomaterials: the first domino of combined tumor therapy. *Nano Today.*
3298 2021; 36: 100963.

3299 57. Hemmer E, Benayas A, L egar e F, Vetrone F. Exploiting the biological windows:
3300 current perspectives on fluorescent bioprobes emitting above 1000 nm. *Nanoscale*
3301 *Horiz.* 2016; 1: 168-84.

3302 58. Tian Q, Yao W, Wu W, Jiang C. NIR light-activated upconversion semiconductor
3303 photocatalysts. *Nanoscale Horiz.* 2019; 4: 10-25.

3304 59. Wang M, Hou Z, Al Kheraif AA, Xing B, Lin J. Mini Review of TiO(2) -Based
3305 Multifunctional Nanocomposites for Near-Infrared Light-Responsive Phototherapy.
3306 *Adv Healthc Mater.* 2018; 7: e1800351.

3307 60. Zheng S, Zhang H, Sheng T, Xiang Y, Wang J, Tang Y, et al. Photoswitchable
3308 upconversion nanoparticles with excitation-dependent emission for programmed
3309 stepwise NIR phototherapy. *iScience.* 2023; 26: 107859.

3310 61. Juengpanich S, Li S, Yang T, Xie T, Chen J, Shan Y, et al. Pre-activated
3311 nanoparticles with persistent luminescence for deep tumor photodynamic therapy in
3312 gallbladder cancer. *Nat Commun.* 2023; 14: 5699.

3313 62. Molkenova A, Choi HE, Lee G, Baek H, Kwon M, Lee SB, et al. Cold-Responsive
3314 Hyaluronated Upconversion Nanoplatfom for Transdermal Cryo-Photodynamic
3315 Cancer Therapy. *Adv Sci (Weinh).* 2024; 11: e2306684.

3316 63. Zhu T, Lin Z, Tang X, Liu J, Zhang Y, Zhong X. Programmable activation of
3317 berbamine and photosensitizers for enhanced photodynamic therapy using emission-
3318 switchable upconversion nanoparticles. *Int J Pharm.* 2024; 659: 124202.

3319 64. Chu Z, Tian T, Tao Z, Yang J, Chen B, Chen H, et al. Upconversion
3320 nanoparticles@AgBiS(2) core-shell nanoparticles with cancer-cell-specific
3321 cytotoxicity for combined photothermal and photodynamic therapy of cancers. *Bioact*
3322 *Mater.* 2022; 17: 71-80.

3323 65. Palanikumar L, Kalmouni M, Houhou T, Abdullah O, Ali L, Pasricha R, et al. pH-
3324 Responsive Upconversion Mesoporous Silica Nanospheres for Combined Multimodal
3325 Diagnostic Imaging and Targeted Photodynamic and Photothermal Cancer Therapy.
3326 *ACS Nano.* 2023; 17: 18979-99.

3327 66. Chu Z, Tian T, Tao Z, Yang J, Chen B, Chen H, et al. Upconversion
3328 nanoparticles@AgBiS2 core-shell nanoparticles with cancer-cell-specific cytotoxicity
3329 for combined photothermal and photodynamic therapy of cancers. *Bioact Mater.* 2022;
3330 17: 71-80.

3331 67. Mo C, Fu L, Zou H, Liu C, Zhu K, Cao J, et al. Near-infrared light-induced self-
3332 assembled lanthanide nanoparticles for boosted radiosensitization and persistent tumor
3333 therapy monitoring. *Nano Today.* 2025; 64: 102782.

3334 68. Wang F, Xu W, Liu Y, Zhu S, Liu W, Bo S, et al. Spatiotemporally Controlled
3335 Tumor Photodynamic/Immunotherapy Therapy Based on Upconversion Hybrid
3336 Nanosystem. *Adv Sci (Weinh).* 2026; 13: e15052.

- 3337 69. Yang M, Huang Y, Chen Z, Ye Q, Zeng Z, You X, et al. Synthetic carbon-based
3338 lanthanide upconversion nanoparticles for enhanced photothermal therapy. *Nat*
3339 *Commun.* 2025; 16: 6343.
- 3340 70. Juvekar V, Lee D, Park TG, Samanta R, Kasar P, Kim C, et al. Two-photon
3341 excitation photosensitizers for photodynamic therapy: From small-molecules to nano-
3342 complex systems. *Coord Chem Rev.* 2024; 506: 215711.
- 3343 71. Cheng SH, Hsieh CC, Chen NT, Chu CH, Huang CM, Chou PT, et al. Well-
3344 defined mesoporous nanostructure modulates three-dimensional interface energy
3345 transfer for two-photon activated photodynamic therapy. *Nano Today.* 2011; 6: 552-63.
- 3346 72. Xu Z, Jiang Y, Shen Y, Tang L, Hu Z, Lin G, et al. A biocompatible
3347 photosensitizer with a high intersystem crossing efficiency for precise two-photon
3348 photodynamic therapy. *Mater Horiz.* 2022; 9: 1283-92.
- 3349 73. Liu Y, Zhao J, Xu X, Xu Y, Cui W, Yang Y, et al. Emodin-Based
3350 Nanoarchitectonics with Giant Two-Photon Absorption for Enhanced Photodynamic
3351 Therapy. *Angew Chem Int Ed Engl.* 2023; 62: e202308019.
- 3352 74. Soleimany A, Khoee S, Dias S, Sarmiento B. Exploring Low-Power Single-Pulsed
3353 Laser-Triggered Two-Photon Photodynamic/Photothermal Combination Therapy
3354 Using a Gold Nanostar/Graphene Quantum Dot Nanohybrid. *ACS Appl Mater*
3355 *Interfaces.* 2023; 15: 20811-21.
- 3356 75. Fang F, Yuan Y, Wan Y, Li J, Song Y, Chen WC, et al. Near-Infrared Thermally
3357 Activated Delayed Fluorescence Nanoparticle: A Metal-Free Photosensitizer for Two-
3358 Photon-Activated Photodynamic Therapy at the Cell and Small Animal Levels. *Small.*
3359 2022; 18: e2106215.
- 3360 76. Kuang S, Wei F, Karges J, Ke L, Xiong K, Liao X, et al. Photodecaging of a
3361 Mitochondria-Localized Iridium(III) Endoperoxide Complex for Two-Photon
3362 Photoactivated Therapy under Hypoxia. *J Am Chem Soc.* 2022; 144: 4091-101.
- 3363 77. Wei F, Kuang S, Rees TW, Liao X, Liu J, Luo D, et al. Ruthenium(II) complexes
3364 coordinated to graphitic carbon nitride: Oxygen self-sufficient photosensitizers which
3365 produce multiple ROS for photodynamic therapy in hypoxia. *Biomaterials.* 2021; 276:
3366 121064.
- 3367 78. Xu Z, Li X, Yang Z, Zhang Z, Zhang Y, Fan M, et al. An NIR-II Two-Photon
3368 Excitable AIE Photosensitizer for Precise and Efficient Treatment of Orthotopic Small-
3369 Size Glioblastoma. *Adv Mater.* 2025; 37: e2413164.
- 3370 79. Wu YX, Zhang D, Hu X, Peng R, Li J, Zhang X, et al. Multicolor Two-Photon
3371 Nanosystem for Multiplexed Intracellular Imaging and Targeted Cancer Therapy.
3372 *Angew Chem Int Ed Engl.* 2021; 60: 12569-76.
- 3373 80. Ke L, Wei F, Xie L, Karges J, Chen Y, Ji L, et al. A Biodegradable Iridium(III)
3374 Coordination Polymer for Enhanced Two-Photon Photodynamic Therapy Using an

3375 Apoptosis-Ferroptosis Hybrid Pathway. *Angew Chem Int Ed Engl.* 2022; 61:
3376 e202205429.

3377 81. Li C, Liu J, Hong Y, Lin R, Liu Z, Chen M, et al. Click Synthesis Enabled Sulfur
3378 Atom Strategy for Polymerization-Enhanced and Two-Photon Photosensitization.
3379 *Angew Chem Int Ed Engl.* 2022; 61: e202202005.

3380 82. Mushtaq A, Iqbal MZ, Tang J, Sun W. The wonders of X-PDT: an advance route
3381 to cancer theranostics. *J Nanobiotechnology.* 2024; 22: 655.

3382 83. Souris JS, Leoni L, Zhang HJ, Pan A, Tanios E, Tsai H-M, et al. X-ray Activated
3383 Nanoplatfoms for Deep Tissue Photodynamic Therapy. *Nanomaterials.* 2023; 13: 673.

3384 84. He L, Yu X, Li W. Recent Progress and Trends in X-ray-Induced Photodynamic
3385 Therapy with Low Radiation Doses. *ACS Nano.* 2022; 16: 19691-721.

3386 85. Wang X, Sun W, Shi H, Ma H, Niu G, Li Y, et al. Organic phosphorescent
3387 nanoscintillator for low-dose X-ray-induced photodynamic therapy. *Nat Commun.*
3388 2022; 13: 5091.

3389 86. Li Y, Guo C, Yuan J, Yang X, Ji H, Wu M, et al. Recent advances and prospects
3390 of persistent luminescent materials in public health applications. *Chem Eng J.* 2024;
3391 487: 150424.

3392 87. Tan R, Wu J, Wang C, Zhao Z, Zhang X, Zhong C, et al. The develop of persistent
3393 luminescence nanoparticles with excellent performances in cancer targeted bioimaging
3394 and killing: a review. *J Nanobiotechnology.* 2025; 23: 299.

3395 88. Yao B, Xu F, Tian Z, Dai M, Song J, Li L, et al. Strong Persistent Luminescence
3396 NaYF₄-based Nanoparticles Combined with Manipulated Hyperfractionated
3397 Irradiation for X-ray-Excited Photodynamic Therapy Enhancement. *ACS Appl Mater*
3398 *Interfaces.* 2025; 17: 16561-75.

3399 89. Wang X, Sun W, Shi H, Ma H, Niu G, Li Y, et al. Organic phosphorescent
3400 nanoscintillator for low-dose X-ray-induced photodynamic therapy. *Nat Commun.*
3401 2022; 13: 5091.

3402 90. Cheng W, He S, Chen Q, Song X, Lu C, Yang H. X-ray Induced Persistent Type
3403 I Photodynamic Therapy with Enhanced Hypoxia Tolerance and Chemoradiotherapy.
3404 *Nano Lett.* 2025; 25: 4549-59.

3405 91. Zhang L, Gao F, Liu S, Ju M, Sun C, Sun G, et al. Synthesis of lanthanide-based
3406 scintillator@MOF nanocomposites for X-ray-induced photodynamic therapy. *Inorg*
3407 *Chem Front.* 2024; 11: 1607-15.

3408 92. Liu B-M, Lin Y, Liu Y, Zhang S, Wang J, Zhang H, et al. X-ray-activated ultra-
3409 long UVA persistent luminescence from a Bi-doped perovskite LaGaO₃ for
3410 photodynamic activation. *Inorg Chem Front.* 2024; 11: 2049-57.

- 3411 93. He L, Wang L, Yu X, Tang Y, Jiang Z, Yang G, et al. Full-course NIR-II imaging-
3412 navigated fractionated photodynamic therapy of bladder tumours with X-ray-activated
3413 nanotransducers. *Nat Commun.* 2024; 15: 8240.
- 3414 94. Yu Y, Xiang L, Zhang X, Zhang L, Ni Z, Zhu ZH, et al. Pure Organic AIE
3415 Nanoscintillator for X-ray Mediated Type I and Type II Photodynamic Therapy. *Adv*
3416 *Sci (Weinh).* 2023; 10: e2302395.
- 3417 95. Gu L, Wu H, Li X, Xu J, Wang M, Li C, et al. Hydrogen-Bonded Organic
3418 Framework Nanoscintillators for X-Ray-Induced Photodynamic Therapy in
3419 Hepatocellular Carcinoma. *Adv Mater.* 2025; 37: e2417001.
- 3420 96. Cairns RA, Harris IS, Mak TW. Regulation of cancer cell metabolism. *Nat Rev*
3421 *Cancer.* 2011; 11: 85-95.
- 3422 97. Pavlides S, Tsirogas A, Vera I, Flomenberg N, Frank PG, Casimiro MC, et al. Loss
3423 of stromal caveolin-1 leads to oxidative stress, mimics hypoxia and drives inflammation
3424 in the tumor microenvironment, conferring the "reverse Warburg effect": a
3425 transcriptional informatics analysis with validation. *Cell Cycle.* 2010; 9: 2201-19.
- 3426 98. Hsu PP, Sabatini DM. Cancer cell metabolism: Warburg and beyond. *Cell.* 2008;
3427 134: 703-7.
- 3428 99. Estrella V, Chen T, Lloyd M, Wojtkowiak J, Cornnell HH, Ibrahim-Hashim A, et
3429 al. Acidity Generated by the Tumor Microenvironment Drives Local Invasion. *Cancer*
3430 *Res.* 2013; 73: 1524-35.
- 3431 100. Park Y, Jeong EM. Glutathione Dynamics in the Tumor Microenvironment: A
3432 Potential Target of Cancer Stem Cells and T Cells. *Int J Stem Cells.* 2024; 17: 270-83.
- 3433 101. Kennedy L, Sandhu JK, Harper M-E, Cuperlovic-Culf M. Role of Glutathione in
3434 Cancer: From Mechanisms to Therapies. *Biomolecules.* 2020; 10: 1429.
- 3435 102. Jin M-Z, Jin W-L. The updated landscape of tumor microenvironment and drug
3436 repurposing. *Signal Transduct Target Ther.* 2020; 5: 166.
- 3437 103. Weinberg F, Ramnath N, Nagrath D. Reactive Oxygen Species in the Tumor
3438 Microenvironment: An Overview. *Cancers (Basel).* 2019; 11.
- 3439 104. Gonzalez-Avila G, Sommer B, García-Hernández AA, Ramos C. Matrix
3440 Metalloproteinases' Role in Tumor Microenvironment. *Adv Exp Med Biol.* 2020; 1245:
3441 97-131.
- 3442 105. Kessenbrock K, Plaks V, Werb Z. Matrix metalloproteinases: regulators of the
3443 tumor microenvironment. *Cell.* 2010; 141: 52-67.
- 3444 106. Niland S, Riscanevo AX, Eble JA. Matrix Metalloproteinases Shape the Tumor
3445 Microenvironment in Cancer Progression. *Int J Mol Sci.* 2021; 23.
- 3446 107. Singleton DC, Macann A, Wilson WR. Therapeutic targeting of the hypoxic
3447 tumour microenvironment. *Nat Rev Clin Oncol.* 2021; 18: 751-72.

- 3448 108. Chen Z, Han F, Du Y, Shi H, Zhou W. Hypoxic microenvironment in cancer:
3449 molecular mechanisms and therapeutic interventions. *Signal Transduct Target Ther.*
3450 2023; 8: 70.
- 3451 109. Ciepła J, Smolarczyk R. Tumor hypoxia unveiled: insights into microenvironment,
3452 detection tools and emerging therapies. *Clin Exp Med.* 2024; 24: 235.
- 3453 110. Li Z-H, Zhang X, Wu F-G. Hyperbaric Oxygen-Facilitated Cancer Treatment: A
3454 Minireview. *Adv Nanobiomed Res.* 2024; 4: 2300162.
- 3455 111. Moen I, Stuhr LE. Hyperbaric oxygen therapy and cancer--a review. *Target Oncol.*
3456 2012; 7: 233-42.
- 3457 112. P. N. N, Mehla S, Begum A, Chaturvedi HK, Ojha R, Hartinger C, et al. Smart
3458 Nanozymes for Cancer Therapy: The Next Frontier in Oncology. *Adv Healthc Mater.*
3459 2023; 12: 2300768.
- 3460 113. Shao L, Wang X, Du X, Yin S, Qian Y, Yao Y, et al. Application of
3461 Multifunctional Nanozymes in Tumor Therapy. *ACS Omega.* 2024; 9: 15753-67.
- 3462 114. Zhang X, Chen X, Zhao Y. Nanozymes: Versatile Platforms for Cancer Diagnosis
3463 and Therapy. *Nanomicro Lett.* 2022; 14: 95.
- 3464 115. Kim J-H, Jung E-A, Kim J-E. Perfluorocarbon-based artificial oxygen carriers for
3465 red blood cell substitutes: considerations and direction of technology. *J Pharm Investig.*
3466 2024; 54: 267-82.
- 3467 116. Charbe NB, Castillo F, Tambuwala MM, Prasher P, Chellappan DK, Carreño A,
3468 et al. A new era in oxygen therapeutics? From perfluorocarbon systems to
3469 haemoglobin-based oxygen carriers. *Blood Rev.* 2022; 54: 100927.
- 3470 117. Yang Y, Liu Y, Jiang Y. Recent Advances in Perfluorocarbon-Based Delivery
3471 Systems for Cancer Theranostics. *Mol Pharm.* 2023; 20: 3254-77.
- 3472 118. Liang X, Chen M, Bhattarai P, Hameed S, Dai Z. Perfluorocarbon@Porphyrin
3473 Nanoparticles for Tumor Hypoxia Relief to Enhance Photodynamic Therapy against
3474 Liver Metastasis of Colon Cancer. *ACS Nano.* 2020; 14: 13569-83.
- 3475 119. Liu L, Hou K, Lin S, Di Y, Zhuang Z, Zeng Z, et al. Hemoglobin based oxygen
3476 carrier and its application in biomedicine. *Coord Chem Rev.* 2025; 532: 216508.
- 3477 120. Grzegorzewski W, Czerniecka-Kubicka A, Gołda K, Niedźwiedzka A, Wollocko
3478 H, Majewski MS, et al. Hemoglobin-Based Oxygen Carriers: Selected Advances and
3479 Challenges in the Design of Safe Oxygen Therapeutics (A Focused Review). *Int J Mol*
3480 *Sci.* 2025; 26.
- 3481 121. Jose AD, Chong CH-N, Cheah E, Jaiswal J, Wu Z, Thakur SS. Formulation and
3482 evaluation of oxygen microbubbles stabilised in a hydrogel to potentiate radiotherapy.
3483 *Int J Pharm.* 2025; 674: 125443.
- 3484 122. Xiao Q, Jia Q, Inam M, Chen Q, Sun X. Nanobubbles: a bridge connecting
3485 nanomedicine and gas medicine. *Med Gas Res.* 2025; 15: 214-5.

3486 123. Liang J, Lai X, Mei Y, Liu X, Wen S, Zhou Y, et al. Oxygenation:
3487 Nanotechnological Strategies for Conquering Tumor Hypoxia in Photodynamic
3488 Therapy. *Int J Nanomed.* 2026; 21: 569340.

3489 124. Mancebo JG, Sack K, Hartford J, Dominguez S, Balcarcel-Monzon M, Chartier E,
3490 et al. Systemically injected oxygen within rapidly dissolving microbubbles improves
3491 the outcomes of severe hypoxaemia in swine. *Nat Biomed Eng.* 2024; 8: 1396-411.

3492 125. Michalczyk M, Humeniuk E, Adamczuk G, Korga-Plewko A. Hyaluronic Acid as
3493 a Modern Approach in Anticancer Therapy-Review. *Int J Mol Sci.* 2022; 24.

3494 126. Bhattacharyya M, Jariyal H, Srivastava A. Hyaluronic acid: More than a carrier,
3495 having an overpowering extracellular and intracellular impact on cancer. *Carbohydr*
3496 *Polym.* 2023; 317: 121081.

3497 127. Zhao J, Chen J, Li C, Xiang H, Miao X. Hyaluronidase overcomes the extracellular
3498 matrix barrier to enhance local drug delivery. *Eur J Pharm Biopharm.* 2024; 203:
3499 114474.

3500 128. Li X, Xue H, Liu Q, Ren L, Sun Y. Triple-remodeling of tumor microenvironment
3501 through hyaluronidase-assisted folate-targeted lipid nanoparticle-mediated
3502 siVEGF/siPD-L1 for enhanced tumor immunotherapy. *J Transl Med.* 2026; 24: 51.

3503 129. Gong H, Chao Y, Xiang J, Han X, Song G, Feng L, et al. Hyaluronidase To
3504 Enhance Nanoparticle-Based Photodynamic Tumor Therapy. *Nano Lett.* 2016; 16:
3505 2512-21.

3506 130. Xu F, Wang M, Dotse E, Chow KT, Lo PC. Inducing Immunogenic Cancer Cell
3507 Death through Oxygen-Economized Photodynamic Therapy with Nitric Oxide-
3508 Releasing Photosensitizers. *Angew Chem Int Ed Engl.* 2024; 63: e202404561.

3509 131. Fan Z, Wu S, Deng H, Li G, Huang L, Liu H. Light-Triggered Nanozymes
3510 Remodel the Tumor Hypoxic and Immunosuppressive Microenvironment for
3511 Ferroptosis-Enhanced Antitumor Immunity. *ACS Nano.* 2024; 18: 12261-75.

3512 132. Luo T, Yang H, Wang R, Pu Y, Cai Z, Zhao Y, et al. Bifunctional Cascading
3513 Nanozymes Based on Carbon Dots Promotes Photodynamic Therapy by Regulating
3514 Hypoxia and Glycolysis. *ACS Nano.* 2023; 17: 16715-30.

3515 133. Qu C, Yuan H, Tian M, Zhang X, Xia P, Shi G, et al. Precise Photodynamic
3516 Therapy by Midkine Nanobody-Engineered Nanoparticles Remodels the
3517 Microenvironment of Pancreatic Ductal Adenocarcinoma and Potentiates the
3518 Immunotherapy. *ACS Nano.* 2024; 18: 4019-37.

3519 134. Sun Q, Yang J, Shen W, Lu H, Hou X, Liu Y, et al. Engineering mitochondrial
3520 uncoupler synergistic photodynamic nanoplatform to harness immunostimulatory pro-
3521 death autophagy/mitophagy. *Biomaterials.* 2022; 289: 121796.

3522 135. Dai W, Zhou X, Zhao J, Lei L, Huang Y, Jia F, et al. Tumor microenvironment-
3523 modulated nanoparticles with cascade energy transfer as internal light sources for
3524 photodynamic therapy of deep-seated tumors. *Biomaterials*. 2025; 312: 122743.

3525 136. Gao Z, Miao Z, Jia S, Zhang D, Zhang H, Tian J, et al. An Activatable and
3526 Covalent Tumor-Associated Antigen Capturer Enabling Systemic Injection In Vivo for
3527 Promoted Antitumor Immunity. *J Am Chem Soc*. 2025; 147: 34659-71.

3528 137. Attar GS, Bhalla V, Kumar M. Tumor Microenvironment-Modulated Self-
3529 illuminating Photodynamic Therapy: A Promising Tool for Deep-Seated and Metastatic
3530 Cancer. *ACS Appl Bio Mater*. 2026; 9: 5-26.

3531 138. Zahra K, Deng F, Deng W, Sang R. Advances in photodynamic therapy and its
3532 combination strategies for breast cancer. *Acta Biomater*. 2025; 205.

3533 139. Zhao R, Hou Y, Li B, Pan Z, Qiu J, Wang Q, et al. Bioengineered hybrid dual-
3534 targeting nanoparticles reprogram the tumour microenvironment for deep glioblastoma
3535 photodynamic therapy. *Nat Commun*. 2025; 16: 7672.

3536 140. Xiaohui Z, Xuelin T, Bai L, Zhao R, Chen Y, Tian X. Iridium complex-loaded
3537 biomimetic vesicles enable enhanced photodynamic therapy and immune modulation.
3538 *Microsyst Nanoeng*. 2026; 12: 33.

3539 141. Pei Y, Pan Y, Zhang Z, Zhu J, Sun Y, Zhang Q, et al. Leveraging Tumor
3540 Microenvironment to Boost Synergistic Photodynamic Therapy, Ferroptosis Anti-
3541 Tumor Efficiency Based on a Functional Iridium(III) Complex. *Adv Sci (Weinh)*. 2025;
3542 12: e2413879.

3543 142. Zhao Y-Y, Xu Y, Zhang X, Chen Z, Kim H, Li X, et al. A Hypoxia-Triggered
3544 Bioreduction of Hydrophilic Type I Photosensitizer for Switchable In Vivo
3545 Photoacoustic Imaging and High-Specificity Cancer Phototherapy. *Angew Chem Int
3546 Ed Engl*. 2025; 64: e202506412.

3547 143. Liu J, Liu Y, Zhi S, Yang Y, Kim H, Wu D, et al. A Nanotherapeutic Agent for
3548 Synergistic Tumor Therapy: Co-Activation of Photochemical-Biological Effects.
3549 *Angew Chem Int Ed Engl*. 2025; 64: e202425631.

3550 144. Zhang Y, Wu X, Wang K, Tang Y, Lu X, Sun F, et al. Simultaneous Reversal of
3551 T Lymphocytes and Cancer Cells Metabolism Via a Biomimetic Heavy-Atom-Free
3552 Photosensitizers-Based Combination Therapies to Boost Cancer Photoimmunotherapy.
3553 *Adv Sci (Weinh)*. 2025; 12: 2416143.

3554 145. Zhang Y, Feng G, He T, Yang M, Lin J, Huang P. Traceable Lactate-Fueled Self-
3555 Acting Photodynamic Therapy against Triple-Negative Breast Cancer. *Research (Wash
3556 D C)*. 2024; 7: 0277.

3557 146. Li Y, Qu F, Wan F, Zhong C, Rao J, Liu Y, et al. Aggregation control of anionic
3558 pentamethine cyanine enabling excitation wavelength selective NIR-II fluorescence
3559 imaging-guided photodynamic therapy. *Nat Commun*. 2025; 16: 762.

3560 147. Feng N, Peng Z, Zhang X, Lin Y, Hu L, Zheng L, et al. Strategically engineered
3561 Au(I) complexes for orchestrated tumor eradication via chemo-phototherapy and
3562 induced immunogenic cell death. *Nat Commun.* 2024; 15: 8187.

3563 148. Tang Y, Yu X, He L, Tang M, Yue W, Chen R, et al. A high-valence bismuth(V)
3564 nanoplatfrom triggers cancer cell death and anti-tumor immune responses with
3565 exogenous excitation-free endogenous H₂O₂- and O₂-independent ROS
3566 generation. *Nat Commun.* 2025; 16: 860.

3567 149. Nuthalapati K, Raviraj V, Shanmugam M, Thangudu S, Chiang C-S, Hwang K.
3568 Engineering Tumor-Specific Nanotheranostic Agent with MR Image-Guided NIR-II &
3569 -III Photodynamic Therapy to Combat Against Deeply Seated Orthotopic Glioblastoma.
3570 *Small Sci.* 2024; 4.

3571 150. Li Z, Zhao R, Pei Q, Xie Z, Zheng M. Near-Infrared Afterglow Imaging-Guided
3572 Surgical Resection and Synergistic Photodynamic-Chemo Therapy of Breast Cancer.
3573 *Adv Sci (Weinh).* 2025; 12: e03883.

3574 151. Diao S, Zhang Z, Zhao S, Li Q, Zhang X, Yang X, et al. Dual-Activatable Nano-
3575 Immunomodulator for NIR-II Fluorescence Imaging-Guided Precision Cancer
3576 Photodynamic Immunotherapy: *Adv Sci (Weinh).* 2024; 11: 2409833.

3577 152. Yu D, Ding Q, Xiang C, Wang D, Hu L, Wang J, et al. NIR-II Engineered
3578 Exosome Nanotheranostic Probes for "Oriented Blasting" in Orthotopic Glioblastoma.
3579 *ACS Nano.* 2025; 19: 22900-13.

3580 153. Yang Z, Qin R, Ruan D, Hu C, Li W, Zhou J, et al. Ce6-DNAzyme-Loaded Metal-
3581 Organic Framework Theranostic Agents for Boosting miRNA Imaging-Guided
3582 Photodynamic Therapy in Breast Cancer. *ACS Nano.* 2025; 19: 27873-89.

3583 154. Zhang C, Shi Y, Zhu Z, Yang T, Wang Y, Hu S, et al. Electrostatically Stabilized
3584 Light-Activated Membrane Delivery System: Overcoming Membrane Flexibility and
3585 Self-Repair to Enhance Tumor Therapy. *ACS Nano.* 2025; 19: 12119-37.

3586 155. Zhu Z, Wang Q, Chen X, Wang Q, Yan C, Zhao X, et al. An Enzyme-Activatable
3587 Aggregation-Induced-Emission Probe: Intraoperative Pathological Fluorescent
3588 Diagnosis of Pancreatic Cancer via Specific Cathepsin E. *Adv Mater.* 2022; 34:
3589 e2107444.

3590 156. Wang Y, Liao J, Lyu Y, Guo Q, Zhu Z, Wu X, et al. An AIE Photosensitizer with
3591 Simultaneous Type I and Type II ROS Generation: Efficient Bacterial Elimination and
3592 Hypoxic Tumor Ablation. *Adv Funct Mater.* 2023; 33.

3593 157. Badir A, Refki S, Sekkat Z. Utilizing gold nanoparticles in plasmonic
3594 photothermal therapy for cancer treatment. *Heliyon.* 2025; 11: e42738.

3595 158. Chuang Y-C, Lin C-J, Lo S-F, Wang J-L, Tzou S-C, Yuan S-S, et al. Dual
3596 functional AuNRs@MnMEIOs nanoclusters for magnetic resonance imaging and
3597 photothermal therapy. *Biomaterials.* 2014; 35: 4678-87.

3598 159. Zu H, Wu Y, Meng H, Cheng X, Wang Y, Zhang LW, et al. Tumor Metabolism
3599 Aiming Cu₂-xS Nanoagents Mediate Photothermal-Derived Cuproptosis and Immune
3600 Activation. *ACS Nano*. 2024; 18: 23941-57.

3601 160. Deng X, Zhao R, Tang Y, Yi M, Wang D, Lin W, et al. FeS(2)@COF based
3602 nanocarrier for photothermal-enhanced chemodynamic/thermodynamic tumor therapy
3603 and immunotherapy via reprogramming tumor-associated macrophages. *J*
3604 *Nanobiotechnology*. 2024; 22: 711.

3605 161. Chen S, Zhang P, Bai H, Yi W. Recent advances in nano-molybdenum oxide for
3606 photothermal cancer therapy. *Nanomedicine (Lond)*. 2025; 20: 883-901.

3607 162. Bahadur R, Singh B, Rai D, Srivastava R. Influence of PEGylation on WS₂
3608 Nanosheets and Its Application in Photothermal Therapy. *ACS Appl Bio Mater*. 2023;
3609 6: 4740-8.

3610 163. Butt IA, Yin P, Chen N, Zhang W. α -Fe Nanoparticles and Multiwalled Carbon
3611 Nanotubes Composite with Improved Photothermal Conversion Efficiency for Tumor
3612 Therapy. *ACS Appl Bio Mater*. 2025; 8: 6291-304.

3613 164. Khan S, Batool H, Tariq H, Noor A. Graphene Oxide-Based Photothermal and
3614 Photodynamic Therapy-A Systematic Review. *J Biomed Mater Res B Appl Biomater*.
3615 2025; 113: e35656.

3616 165. Xie C, Wang L, Liu Y, Chen M, Du P, Wang Y, et al. Fullerene Covalent
3617 Passivation of Black Phosphorus Nanosheets toward Enhanced Near-Infrared-II
3618 Photothermal Therapy. *ACS Appl Mater Interfaces*. 2023; 15: 20686-96.

3619 166. Lee S, Min S, Kim G, Lee S. Recent advances in the design of organic
3620 photothermal agents for cancer treatment: A review. *Coord Chem Rev*. 2024; 506:
3621 215719.

3622 167. Li Y, Qi H, Geng Y, Li L, Cai X. Research progress of organic photothermal
3623 agents delivery and synergistic therapy systems. *Colloids Surf B Biointerfaces*. 2024;
3624 234: 113743.

3625 168. Zhou R, Zhang M, Xi J, Li J, Ma R, Ren L, et al. Gold Nanorods-Based
3626 Photothermal Therapy: Interactions Between Biostructure, Nanomaterial, and Near-
3627 Infrared Irradiation. *Nanoscale Res Lett*. 2022; 17: 68.

3628 169. Nejabat M, Samie A, Ramezani M, Alibolandi M, Abnous K, Taghdisi SM. An
3629 Overview on Gold Nanorods as Versatile Nanoparticles in Cancer Therapy. *J Control*
3630 *Release*. 2023; 354: 221-42.

3631 170. Taylor ML, Wilson RE, Jr., Amrhein KD, Huang X. Gold Nanorod-Assisted
3632 Photothermal Therapy and Improvement Strategies. *Bioengineering (Basel)*. 2022; 9.

3633 171. Wang X, Zhu L, Gu Z, Dai L. Carbon nanomaterials for phototherapy.
3634 *Nanophotonics*. 2022; 11: 4955-76.

3635 172. Balou S, Shandilya P, Priye A. Carbon dots for photothermal applications. *Front*
3636 *Chem.* 2022; 10: 1023602.

3637 173. Naief MF, Mohammed SN, Mayouf HJ, Mohammed AM. A review of the role of
3638 carbon nanotubes for cancer treatment based on photothermal and photodynamic
3639 therapy techniques. *J Organomet Chem.* 2023; 999: 122819.

3640 174. Báez DF. Graphene-Based Nanomaterials for Photothermal Therapy in Cancer
3641 Treatment. *Pharmaceutics.* 2023; 15: 2286.

3642 175. Liu Y, Li J, Chen M, Chen X, Zheng N. Palladium-based nanomaterials for cancer
3643 imaging and therapy. *Theranostics.* 2020; 10: 10057-74.

3644 176. Phan TTV, Huynh T-C, Manivasagan P, Mondal S, Oh J. An Up-To-Date Review
3645 on Biomedical Applications of Palladium Nanoparticles. *Nanomaterials (Basel).* 2020;
3646 10: 66.

3647 177. Beniwal N, Verma A, Putta CL, Rengan AK. Recent Trends in Bio-nanomaterials
3648 and Non-invasive Combinatorial Approaches of Photothermal Therapy against Cancer.
3649 *Nanotheranostics.* 2024; 8: 219-38.

3650 178. Chen S, Zhu L, Du Z, Ma R, Yan T, Alimu G, et al. Polymer encapsulated clinical
3651 ICG nanoparticles for enhanced photothermal therapy and NIR fluorescence imaging
3652 in cervical cancer. *RSC Adv.* 2021; 11: 20850-8.

3653 179. Teng CW, Huang V, Arguelles GR, Zhou C, Cho SS, Harmsen S, et al.
3654 Applications of indocyanine green in brain tumor surgery: review of clinical evidence
3655 and emerging technologies. *Neurosurg Focus.* 2021; 50: E4.

3656 180. Du W, Chong Y, Hu X, Wang Y, Zhu Y, Chen J, et al. Increasing Photothermal
3657 Efficacy by Simultaneous Intra- and Intermolecular Fluorescence Quenching. *Adv*
3658 *Funct Mater.* 2020; 30: 1908073.

3659 181. Zhang X-S, Cheng K, Wei J-S, Huang Z-Y, Hou X-L, Hu Y-G, et al. Nanoplaform
3660 based on ultra-small Au regulating phototoxicity and fluorescence off-on function of
3661 Ag₂S for multi-modal diagnosis and treatment of tumor. *Chem Eng J.* 2022; 431:
3662 133212.

3663 182. Huang Y, Liu Q, Wang Y, He N, Zhao R, Choo J, et al. Gold nanorods
3664 functionalized by a glutathione response near-infrared fluorescent probe as a promising
3665 nanoplaform for fluorescence imaging guided precision therapy. *Nanoscale.* 2019; 11:
3666 12220-9.

3667 183. Qu R, Zhen X, Jiang X. Emerging Designs of Aggregation-Induced Emission
3668 Agents for Enhanced Phototherapy Applications. *CCS Chem.* 2022; 4: 401-19.

3669 184. Tian S, He J, Lyu D, Li S, Xu Q-H. Aggregation enhanced photoactivity of
3670 photosensitizer conjugated metal nanoparticles for multimodal imaging and synergistic
3671 phototherapy below skin tolerance threshold. *Nano Today.* 2022; 45: 101534.

3672 185. Yang M, Deng J, Su H, Gu S, Zhang J, Zhong A, et al. Small organic molecule-
3673 based nanoparticles with red/near-infrared aggregation-induced emission for
3674 bioimaging and PDT/PTT synergistic therapy. *Mater Chem Front.* 2021; 5: 406-17.
3675 186. Farajollahi A, Baharvand M. Advancements in photoacoustic imaging for cancer
3676 diagnosis and treatment. *Int J Pharm.* 2024; 665: 124736.
3677 187. Jain PK, Lee KS, El-Sayed IH, El-Sayed MA. Calculated Absorption and
3678 Scattering Properties of Gold Nanoparticles of Different Size, Shape, and
3679 Composition: Applications in Biological Imaging and Biomedicine. *J Phys Chem B.*
3680 2006; 110: 7238-48.
3681 188. Cao Y, Chen Z, Ran H. In vivo photoacoustic image-guided tumor photothermal
3682 therapy and real-time temperature monitoring using a core-shell polypyrrole@CuS
3683 nanohybrid. *Nanoscale.* 2022; 14: 12069-76.
3684 189. Liu X, Duan Y, Liu B. Nanoparticles as contrast agents for photoacoustic brain
3685 imaging. *Aggregate (Hoboken).* 2021; 2: 4-19.
3686 190. Wang R, Huang Z, Xiao Y, Huang T, Ming J. Photothermal therapy of copper
3687 incorporated nanomaterials for biomedicine. *Biomater Res.* 2023; 27: 121.
3688 191. Xu M, Zhao R, Liu B, Geng F, Wu X, Zhang F, et al. Ultrasmall copper-based
3689 nanoplatfoms for NIR-II light-triggered photothermal/ photodynamic and amplified
3690 nanozyme catalytic therapy of hypoxic tumor. *Chem Eng J.* 2024; 491: 151776.
3691 192. Wen S, Shi Y, Zhang Y, Chang Q, Hu H, Deng X, et al. pH-Activated Ce-Doped
3692 Molybdenum Oxide Nanoclusters for Tumor Microenvironment Responsive
3693 Photothermal and Chemodynamic Therapy. *Langmuir.* 2023; 39: 10145-53.
3694 193. Zhu L, Yang Y, Li X, Zheng Y, Li Z, Chen H, et al. Facile preparation of
3695 indocyanine green and tiny gold nanoclusters co-loaded nanocapsules for targeted
3696 synergistic sono-/photo-therapy. *J Colloid Interface Sci.* 2022; 627: 596-609.
3697 194. Zhang C, Wu J, Liu W, Zhang W, Lee CS, Wang P. New Xanthene Dyes with
3698 NIR-II Emission Beyond 1200 nm for Efficient Tumor Angiography and Photothermal
3699 Therapy. *Small.* 2022; 18: e2202078.
3700 195. Egloff-Juras C, Bezdetnaya L, Dolivet G, Lassalle HP. NIR fluorescence-guided
3701 tumor surgery: new strategies for the use of indocyanine green. *Int J Nanomedicine.*
3702 2019; 14: 7823-38.
3703 196. Tong S, Li C, Wang K, Wu F. Indocyanine green-loaded porphyrin covalent
3704 organic frameworks for photothermal cancer therapy. *Mater Chem Front.* 2024; 8:
3705 1390-9.
3706 197. Wang Y, Yang Y, Yang L, Lin Y, Tian Y, Ni Q, et al. Gold Nanostar@Polyaniline
3707 Theranostic Agent with High Photothermal Conversion Efficiency for Photoacoustic
3708 Imaging-Guided Anticancer Phototherapy at a Low Dosage. *ACS Appl Mater*
3709 *Interfaces.* 2022; 14: 28570-80.

- 3710 198. Quiñones ED, Lu T-Y, Liu K-T, Fan Y-J, Chuang E-Y, Yu J. Glycol chitosan/iron
3711 oxide/polypyrrole nanoclusters for precise chemodynamic/photothermal synergistic
3712 therapy. *Int J Biol Macromol.* 2022; 203: 268-79.
- 3713 199. Chen H, Cai G, Guo A, Zhao Z, Kuang J, Zheng L, et al. Low Band Gap Donor–
3714 Acceptor Conjugated Polymers with Indanone-Condensed Thiadiazolo[3,4-
3715 g]quinoxaline Acceptors. *Macromolecules.* 2019; 52: 6149-59.
- 3716 200. Kang D, Kim HS, Han S, Lee Y, Kim Y-P, Lee DY, et al. A local water molecular-
3717 heating strategy for near-infrared long-lifetime imaging-guided photothermal therapy
3718 of glioblastoma. *Nat Commun.* 2023; 14: 2755.
- 3719 201. Lee KK, Park KW, Lee SC, Lee CS. Perfluorocarbon-polyepinephrine core-shell
3720 nanoparticles as a near-infrared light activatable theranostic platform for bimodal
3721 imaging-guided photothermal/chemodynamic synergistic cancer therapy. *Theranostics.*
3722 2025; 15: 1077-93.
- 3723 202. Chuang Y-C, Hsia Y, Chu C-H, Maharajan S, Hsu F-C, Lee H-L, et al.
3724 Photothermal Temperature-Modulated Cancer Metastasis Harnessed Using Proteinase-
3725 Triggered Assembly of Near-Infrared II Photoacoustic/Photothermal Nanotheranostics.
3726 *ACS Appl Mater Interfaces.* 2024; 16: 40611-27.
- 3727 203. Xi S, Xiao H, Duan Z, Li L, Chen J, Hu T, et al. Effective One-for-All
3728 Phototheranostic Agent for Hypoxia-Tolerant NIR-II Fluorescent/PA Image-Guided
3729 Phototherapy. *Small.* 2025; 21: 2406226.
- 3730 204. Kang X, Zhang Y, Song J, Wang L, Li W, Qi J, et al. A photo-triggered self-
3731 accelerated nanoplatform for multifunctional image-guided combination cancer
3732 immunotherapy. *Nat Commun.* 2023; 14: 5216.
- 3733 205. Zeng S, Chen J, Ren Y, Gu J, Zhang J, Liu L, et al. Multiscale imaging on proton
3734 pump-driven acidity for assessing tumor progression and metastasis. *Nat Commun.*
3735 2026; 17: 1785.
- 3736 206. Shi J, Hu L, Zhang Y, Chen X, Hu T, Wang C, et al. Halogenation-Engineered
3737 Semiconducting Polymer for Near-Infrared Fluorescence Imaging-Guided
3738 Photothermal Cancer Therapy. *ACS Appl Polym Mater.* 2025; 7: 9433-44.
- 3739 207. Zhao M, Li J, He X, Yang R, Zhang Y, Wang J, et al. NIR Image-Guided
3740 Multimodal Nanotherapy Synergizing Photothermal, NO Gas, and Ferroptosis
3741 Enhances Immunogenic Cell Death in Triple-Negative Breast Cancer. *Mater Des.* 2025;
3742 260: 115071.
- 3743 208. Zou M, Wang Y, Jiao J, Lv M, Mo T. Advances in the near-infrared II for in vivo
3744 fluorescence imaging applications: A review. *Talanta.* 2026; 297: 128630.
- 3745 209. Yan H, Ma D, Ma H, Zhang J, Zhang X-D, Wang H. Visualized diagnosis in the
3746 second near-infrared window. *Coord Chem Rev.* 2026; 559: 217788.

3747 210. Hong G, Robinson JT, Zhang Y, Diao S, Antaris AL, Wang Q, et al. In vivo
3748 fluorescence imaging with Ag₂S quantum dots in the second near-infrared region.
3749 *Angew Chem Int Ed Engl.* 2012; 51: 9818-21.

3750 211. Ding C, Huang Y, Shen Z, Chen X. Synthesis and Bioapplications of Ag(2) S
3751 Quantum Dots with Near-Infrared Fluorescence. *Adv Mater.* 2021; 33: e2007768.

3752 212. Li H, Liu Y, Huang B, Zhang C, Wang Z, She W, et al. Highly Efficient GSH-
3753 Responsive “Off–On” NIR-II Fluorescent Fenton Nanocatalyst for Multimodal
3754 Imaging-Guided Photothermal/Chemodynamic Synergistic Cancer Therapy. *Anal*
3755 *Chem.* 2022; 94: 10470-8.

3756 213. Liu T, Zhang X, Liu D, Chen B, Ge X, Gao S, et al. Self-Assembled Ag₂S-QD
3757 Vesicles for In Situ Responsive NIR-II Fluorescence Imaging-Guided Photothermal
3758 Cancer Therapy. *Adv Opt Mater.* 2021; 9: 2100233.

3759 214. Wang W, Zhang X, Ni X, Zhou W, Xie C, Huang W, et al. Semiconducting
3760 polymer nanoparticles for NIR-II fluorescence imaging-guided
3761 photothermal/thermodynamic combination therapy. *Biomater Sci.* 2022; 10: 846-53.

3762 215. Zhang Q, O'Brien S, Grimm J. Biomedical Applications of Lanthanide
3763 Nanomaterials, for Imaging, Sensing and Therapy. *Nanotheranostics.* 2022; 6: 184-94.

3764 216. Sharma N, Mohammad W, Le Guével X, Shanavas A. Gold Nanoclusters as High
3765 Resolution NIR-II Theranostic Agents. *Chem Biomed Imaging.* 2024; 2: 462-80.

3766 217. Zhao R, Zhang R, Feng L, Dong Y, Zhou J, Qu S, et al. Constructing virus-like
3767 SiO_x/CeO₂/VO_x nanozymes for 1064 nm light-triggered mild-temperature
3768 photothermal therapy and nanozyme catalytic therapy. *Nanoscale.* 2022; 14: 361-72.

3769 218. Chen Y, Yu Z, Zheng K, Ren Y, Wang M, Wu Q, et al. Degradable mesoporous
3770 semimetal antimony nanospheres for near-infrared II multimodal theranostics. *Nat*
3771 *Commun.* 2022; 13: 539.

3772 219. Zhu Y, Wang Z, Zhao R, Zhou Y, Feng L, Gai S, et al. Pt Decorated Ti₃C₂T_x
3773 MXene with NIR-II Light Amplified Nanozyme Catalytic Activity for Efficient
3774 Phototheranostics. *ACS Nano.* 2022; 16: 3105-18.

3775 220. Song W, He Y, Feng Y, Wang Y, Li X, Wu Y, et al. Image-Guided Photothermal
3776 and Immune Therapy of Tumors via Melanin-Producing Genetically Engineered
3777 Bacteria. *Small.* 2024; 20: 2305764.

3778 221. Liu L, Wu Y, Ye J, Fu Q, Su L, Wu Z, et al. Synthesis of magnesium nanoparticle
3779 for NIR-II-photoacoustic-imaging-guided synergistic burst-like and H₂ cancer therapy.
3780 *Chem.* 2022; 8: 2990-3007.

3781 222. Wu X, Suo Y, Shi H, Liu R, Wu F, Wang T, et al. Deep-Tissue Photothermal
3782 Therapy Using Laser Illumination at NIR-IIa Window. *Nanomicro Lett.* 2020; 12: 38.

3783 223. Zhang W, Sun X, Huang T, Pan X, Sun P, Li J, et al. 1300 nm absorption two-
3784 acceptor semiconducting polymer nanoparticles for NIR-II photoacoustic imaging

3785 system guided NIR-II photothermal therapy. *Chem Commun (Camb)*. 2019; 55: 9487-
3786 90.

3787 224. Liu Y, Li Y, Koo S, Sun Y, Liu Y, Liu X, et al. Versatile Types of
3788 Inorganic/Organic NIR-IIa/IIb Fluorophores: From Strategic Design toward Molecular
3789 Imaging and Theranostics. *Chem Rev*. 2022; 122: 209-68.

3790 225. Xu W, Wang D, Tang BZ. NIR-II AIEgens: A Win–Win Integration towards
3791 Bioapplications. *Angew Chem Int Ed Engl*. 2021; 60: 7476-87.

3792 226. Huang B, Hu J, Li H, Luo M-Y, Chen S, Zhang M, et al. Near-Infrared IIb Emitting
3793 Nanoprobe for High-Resolution Real-Time Imaging-Guided Photothermal Therapy
3794 Triggering Enhanced Anti-tumor Immunity. *ACS Appl Bio Mater*. 2020; 3: 1636-45.

3795 227. Lou H, Ji A, Qu C, Liu H, Jiang L, Chen H, et al. A Small-Molecule Based Organic
3796 Nanoparticle for Photothermal Therapy and Near-Infrared-IIb Imaging. *ACS Appl*
3797 *Mater Interfaces*. 2022; 14: 35454-65.

3798 228. Wang P, Li J, Wei M, Yang R, Lou K, Dang Y, et al. Tumor-microenvironment
3799 triggered signal-to-noise boosting nanoprobes for NIR-IIb fluorescence imaging guided
3800 tumor surgery and NIR-II photothermal therapy. *Biomaterials*. 2022; 287: 121636.

3801 229. Ma Y, Xiao J, Chen GJ, Dang H, Zhang Y, He X, et al. Ultrafine fiber-mediated
3802 transvascular interventional photothermal therapy using indocyanine green for
3803 precision embolization treatment. *Biomater Sci*. 2025; 13: 1091-100.

3804 230. Huang X, Yang F, Gao B, Ge W, Gao L, Wu J, et al. Free carrier-enhanced
3805 Bi/Bi₂S₃ nanoparticles enable precise OCT catheter-guided interventional
3806 photothermal therapy for colorectal cancer. *Acta Biomater*. 2025; 198: 401-12.

3807 231. Singh N, Kulkarni PP, Tripathi P, Agarwal V, Dash D. Nanogold-coated stent
3808 facilitated non-invasive photothermal ablation of stent thrombosis and restoration of
3809 blood flow. *Nanoscale Adv*. 2024; 6: 1497-506.

3810 232. Cho YC, Kang JM, Park W, Kim D-H, Shin JH, Kim DH, et al. Photothermal
3811 therapy via a gold nanoparticle-coated stent for treating stent-induced granulation tissue
3812 formation in the rat esophagus. *Sci Rep*. 2021; 11: 10558.

3813 233. Hu H-T, Park J-H, Wang Z, Bakheet N, Xu S-J, Lee EJ, et al. Localized
3814 Photothermal Ablation Therapy of Obstructive Rectal Cancer Using a
3815 Nanofunctionalized Stent in a Mouse Model. *ACS Biomater Sci Eng*. 2021; 7: 5890-8.

3816 234. Zhang M, Zhang Y, Hang L, Zhang T, Luo C, Li W, et al. Bionic nanotheranostic
3817 for multimodal imaging-guided NIR-II-photothermal cancer therapy. *Nanoscale*. 2024;
3818 16: 6095-108.

3819 235. Ma G, Ding Q, Wang Y, Zhang Z, Zhang Y, Shi H, et al. Precision Photothermal
3820 Therapy at Mild Temperature: NIR-II Imaging-Guided, H₂O₂-Responsive Stealth
3821 Nanobomb (*Adv. Healthcare Mater.* 26/2025). *Adv Healthc Mater*. 2025; 14: 70239.

3822 236. Wang Z, Liu Y, He C, Zhang X, Li X, Li Y, et al. Small-Molecule
3823 Phototheranostic Agent with Extended π -Conjugation for Efficient NIR-II
3824 Photoacoustic-Imaging-Guided Photothermal Therapy. *Small*. 2024; 20: e2307829.

3825 237. Liu Z, Hong C, Pan C, Sun Y, Lv Y, Wei Y, et al. Synergistically Enhanced
3826 Chemodynamic Therapy Induced by NIR-II Photothermal Therapy with MRI/CT
3827 Imaging Guidance for Colorectal Cancer. *ACS Mater Lett*. 2024; 6: 1593-605.

3828 238. Santhamoorthy M, Vy Phan TT, Ramkumar V, Raorane CJ, Thirupathi K, Kim
3829 SC. Thermo-Sensitive Poly (N-isopropylacrylamide-co-polyacrylamide) Hydrogel for
3830 pH-Responsive Therapeutic Delivery. *Polymers (Basel)*. 2022; 14.

3831 239. Biscari G, Re AL, Fiorica C, Darcos V, Dejean S, Palumbo FS, et al. Nanofibers
3832 of polyester–polydopamine copolymers enable combined photothermal effect and
3833 chemotherapy for local cancer treatment. *Int J Pharm*. 2026; 693: 126689.

3834 240. Srikhao N, Theerakulpisut S, Chindaprasirt P, Okhawilai M, Narain R, Kasemsiri
3835 P. Green synthesis of nano silver-embedded carboxymethyl starch waste/poly vinyl
3836 alcohol hydrogel with photothermal sterilization and pH-responsive behavior. *Int J Biol*
3837 *Macromol*. 2023; 242: 125118.

3838 241. Zhou G, Chen Y, Chen W, Wu H, Yu Y, Sun C, et al. Renal Clearable Catalytic
3839 2D Au-Porphyrin Coordination Polymer Augmented Photothermal-Gas Synergistic
3840 Cancer Therapy. *Small*. 2023; 19: e2206749.

3841 242. Zhao X, Zheng Y, Liu Y, Li Y, Lin Z, Li H, et al. Potent Amphiphilic Poly(Amino
3842 Acid) Nanoadjuvant Delivers Biomineralized Ovalbumin for Photothermal-
3843 Augmented Immunotherapy. *ACS Nano*. 2024; 18: 32088-102.

3844 243. Zhang S, Yu S, Sun J, Huang T, Lin H, Li Z, et al. Au@CuS Nanoshells for
3845 Surface-Enhanced Raman Scattering Image-Guided Tumor Photothermal Therapy with
3846 Accelerated Hepatobiliary Excretion. *Pharmaceutics*. 2024; 16.

3847 244. Yuan G, Luo X, He K, Tan Y, Luo C, Liu B, et al. Intratumoral self-assembly of
3848 renal-clearable gold nanoparticles as precise photothermal nanomedicine for liver
3849 tumor therapy. *Sci Adv*. 2025; 11: eadw7032.

3850 245. Zhang Y, Li Z, Huang Y, Zou B, Xu Y. Amplifying cancer treatment: advances in
3851 tumor immunotherapy and nanoparticle-based hyperthermia. *Front Immunol*. 2023;
3852 Volume 14 - 2023.

3853 246. Marangon I, Silva AA, Guilbert T, Kolosnjaj-Tabi J, Marchiol C, Natkhunarajah
3854 S, et al. Tumor Stiffening, a Key Determinant of Tumor Progression, is Reversed by
3855 Nanomaterial-Induced Photothermal Therapy. *Theranostics*. 2017; 7: 329-43.

3856 247. Lo JH, von Maltzahn G, Douglass J, Park J-H, Sailor MJ, Ruoslahti E, et al.
3857 Nanoparticle amplification via photothermal unveiling of cryptic collagen binding sites.
3858 *J Mater Chem B*. 2013; 1: 5235-40.

3859 248. Gavilán H, Avugadda SK, Fernández-Cabada T, Soni N, Cassani M, Mai BT, et
3860 al. Magnetic nanoparticles and clusters for magnetic hyperthermia: optimizing their
3861 heat performance and developing combinatorial therapies to tackle cancer. *Chem Soc*
3862 *Rev.* 2021; 50: 11614-67.

3863 249. Jiang J, Hu J, Li M, Luo M, Dong B, Sitti M, et al. NIR-II Fluorescent
3864 Thermophoretic Nanomotors for Superficial Tumor Photothermal Therapy. *Adv Mater.*
3865 2025; 37: e2417440.

3866 250. Shi N, Zhang R, Yao S, Sun Q, Wu Y, Yang X, et al. Multiple rotor-based
3867 photothermal agents for NIR-I/NIR-II fluorescence imaging-guided tumor
3868 phototherapy. *J Mater Chem B.* 2025; 13: 9883-92.

3869 251. Patnaik C, Reddy BPK, Gupta A, Rath S, Neekhara S, Chakrabarty S, et al. Non-
3870 invasive imaging-guided preclinical assessments reveal high performance
3871 photothermal effect of an Au-solid lipid nanomaterial. *Nanoscale.* 2025; 17: 26658-77.

3872 252. Sun L, Wu H, Zhang Z, Wu K, Sun J, Dong X, et al. A Smart Visualized
3873 Phototherapy Switch: From NIR-I Imaging-Guided Photodynamic Therapy to NIR-II-
3874 Guided Photothermal Therapy for Enhanced Cascade Tumor Photoablation. *Aggregate*
3875 (Hoboken). 2025; 6.

3876 253. Chen P, Rong J, Chen K, Huang T, Shen Q, Sun P, et al. Photo-Amplified Plasma
3877 Membrane Rupture by Membrane-Anchoring NIR-II Small Molecule Design for
3878 Improved Cancer Photoimmunotherapy. *Angew Chem Int Ed Engl.* 2025; 64:
3879 e202418081.

3880 254. Chen S, Zhan R, Zhou W, Wang L, Zhang W, Tian J. TME-Triggered Degradable
3881 Phototheranostic NanoplatforM for NIR-II Fluorescence Bioimaging-Guided
3882 Phototherapies and Immune Activation. *ACS Macro Lett.* 2024; 13: 768-74.

3883 255. Badir A, Refki S, Sekkat Z. Utilizing gold nanoparticles in plasmonic
3884 photothermal therapy for cancer treatment. *Heliyon.* 2025; 11.

3885 256. Han X, Feng G, Li X, Mo S, Xu C, Yan J, et al. Functional nanomaterials for
3886 enhanced tumor photothermal therapy - the mechanisms and applications. *Front*
3887 *Pharmacol.* 2025; 16 - 2025.

3888 257. Kim M, Suh Y, Nam J-M. Plasmonic Photothermal Nanoparticles for Biomedical
3889 Applications. *Adv Sci (Weinh).* 2019; 6: 1900471.

3890 258. Kratochwil C, Giesel FL, Bruchertseifer F, Mier W, Apostolidis C, Boll R, et al.
3891 ²¹³Bi-DOTATOC receptor-targeted alpha-radionuclide therapy induces remission in
3892 neuroendocrine tumours refractory to beta radiation: a first-in-human experience. *Eur*
3893 *J Nucl Med Mol Imaging.* 2014; 41: 2106-19.

3894 259. Chan HS, Konijnenberg MW, Daniels T, Nysus M, Makvandi M, de Blois E, et al.
3895 Improved safety and efficacy of (213)Bi-DOTATATE-targeted alpha therapy of

3896 somatostatin receptor-expressing neuroendocrine tumors in mice pre-treated with L-
3897 lysine. *EJNMMI Res.* 2016; 6: 83.

3898 260. Cieslik PA, Roth D, Nisli E, Genz J, Berton C, Grundler PV, et al. Light-Induced
3899 Synthesis and Radiotheranostic Treatment of Gastric Cancer with ¹⁶¹Tb-Labeled
3900 Monoclonal Antibodies. *JACS Au.* 2025; 5: 2606-18.

3901 261. Sakmár M, Kozempel J, Kučka J, Janská T, Štíbr M, Vlk M, et al. Biodistribution
3902 study of (²¹¹Pb) progeny released from intravenously applied (²²³Ra) labelled TiO₂
3903 nanoparticles in a mouse model. *Nucl Med Biol.* 2024; 130-131: 108890.

3904 262. Wang R, Liu H, Antal B, Wolterbeek HT, Denkova AG. Ultrasmall Gold
3905 Nanoparticles Radiolabeled with Iodine-125 as Potential New Radiopharmaceutical.
3906 *ACS Appl Bio Mater.* 2024; 7: 1240-9.

3907 263. Westrøm S, Malenge M, Jorstad IS, Napoli E, Bruland Ø S, Bønsdorff TB, et al.
3908 Ra-224 labeling of calcium carbonate microparticles for internal α -therapy: Preparation,
3909 stability, and biodistribution in mice. *J Labelled Comp Radiopharm.* 2018; 61: 472-86.

3910 264. Suchánková P, Kukleva E, Nykl E, Nykl P, Sakmár M, Vlk M, et al.
3911 Hydroxyapatite and Titanium Dioxide Nanoparticles: Radiolabelling and In Vitro
3912 Stability of Prospective Theranostic Nanocarriers for (²²³Ra) and (^{99m}Tc).
3913 *Nanomaterials (Basel).* 2020; 10.

3914 265. Cui Z, Wang L, Liu W, Xu D, Zhang T, Ma B, et al. Imageable Brachytherapy
3915 with Chelator-Free Radiolabeling Hydrogel. *Adv Healthc Mater.* 2024; 13: e2401438.

3916 266. Li Y, Shan S, Zhang R, Sun C, Hu X, Fan J, et al. Imaging and Downstaging
3917 Bladder Cancer with the ¹⁷⁷Lu-Labeled Bioorthogonal Nanoprobe. *ACS Nano.* 2024;
3918 18: 17209-17.

3919 267. Hu K, Xie L, Zhang Y, Hanyu M, Yang Z, Nagatsu K, et al. Marriage of black
3920 phosphorus and Cu²⁺ as effective photothermal agents for PET-guided combination
3921 cancer therapy. *Nat Commun.* 2020; 11: 2778.

3922 268. Wauters AC, Scheerstra JF, van Leent MMT, Teunissen AJP, Priem B, Beldman
3923 TJ, et al. Polymersomes with splenic avidity target red pulp myeloid cells for cancer
3924 immunotherapy. *Nat Nanotechnol.* 2024; 19: 1735-44.

3925 269. Pei P, Shen W, Zhou H, Sun Y, Zhong J, Liu T, et al. Radionuclide labeled gold
3926 nanoclusters boost effective anti-tumor immunity for augmented radio-immunotherapy
3927 of cancer. *Nano Today.* 2021; 38: 101144.

3928 270. Simó C, Serra-Casablancas M, Hortelao AC, Di Carlo V, Guallar-Garrido S,
3929 Plaza-García S, et al. Urease-powered nanobots for radionuclide bladder cancer therapy.
3930 *Nat Nanotechnol.* 2024; 19: 554-64.

3931 271. Wang T, Wang C, Wang Y, Zhang X, Cai X, Guo Z, et al. Radioiodine-Mediated
3932 Transition Metal Valence Conversion for Enhanced Chemodynamic Therapy. *Adv
3933 Funct Mater.* 2024; 34.

3934 272. Jiang QR, Pu XQ, Deng CF, Wang W, Liu Z, Xie R, et al. Microfluidic
3935 Controllable Preparation of Iodine-131-Labeled Microspheres for Radioembolization
3936 Therapy of Liver Tumors. *Adv Healthc Mater.* 2023; 12: e2300873.

3937 273. Jia G, Wang T, Li R, Li X, Sun G, Chen W, et al. Radioiodine-131-Labeled
3938 Theranostic Nanoparticles for Transarterial Radioembolization and
3939 Chemoembolization Combination Therapy of VX2 Liver Tumor. *Adv Healthc Mater.*
3940 2023; 12.

3941 274. Shao S, Xu X, Lin G, Liu G. Radio-granular hydrogels for image-guided tumor
3942 brachytherapy. *Iliver.* 2023; 2: 177-9.

3943 275. Xiao L, Li Y, Geng R, Chen L, Yang P, Li M, et al. Polymer composite
3944 microspheres loading (177)Lu radionuclide for interventional radioembolization
3945 therapy and real-time SPECT imaging of hepatic cancer. *Biomater Res.* 2023; 27: 110.

3946 276. Li X, Qian Y, Lu X, Xu M, He S, Zhang J, et al. Iodine-131 radioembolization
3947 boosts the immune activation enhanced by icaritin/resiquimod in hepatocellular
3948 carcinoma. *J Control Release.* 2025; 378: 849-63.

3949 277. Zhao L, Pang Y, Zhou Y, Chen J, Fu H, Guo W, et al. Antitumor efficacy and
3950 potential mechanism of FAP-targeted radioligand therapy combined with immune
3951 checkpoint blockade. *Signal Transduct Target Ther.* 2024; 9: 142.

3952 278. Zhang J, Zhang S, Cheng C, Zhu C, Wang T, Tang L, et al. Targeting senescence
3953 with radioactive 223Ra/Ba SAzymes enables senolytics-unlocked One-Two punch
3954 strategy to boost anti-tumor immunotherapy. *Biomaterials.* 2025; 315: 122915.

3955 279. Pei P, Zhang Y, Jiang Y, Shen W, Chen H, Yang S, et al. Pleiotropic
3956 Immunomodulatory Functions of Radioactive Inactivated Bacterial Vectors for
3957 Enhanced Cancer Radio-immunotherapy. *ACS Nano.* 2022; 16: 11325-37.

3958 280. Shen J, Feng K, Yu J, Zhao Y, Chen R, Xiong H, et al. Responsive and traceless
3959 assembly of iron nanoparticles and 131I labeled radiopharmaceuticals for ferroptosis
3960 enhanced radio-immunotherapy. *Biomaterials.* 2025; 313: 122795.

3961 281. Shaffer TM, Pratt EC, Grimm J. Utilizing the power of Cerenkov light with
3962 nanotechnology. *Nat Nanotechnol.* 2017; 12: 106-17.

3963 282. Teng M, Liang X, Liu H, Li Z, Gao X, Zhang C, et al. Cerenkov radiation shining
3964 a light for cancer theranostics. *Nano Today.* 2024; 55: 102174.

3965 283. Xu P, Lin S, Wang Y, Abdukayum A, Wang Y. Radionuclide-based Cerenkov
3966 luminescence in biomedicine: Current research progress and future perspectives.
3967 *Trends Analyt Chem.* 2024; 170: 117452.

3968 284. Thorek DL, Ogirala A, Beattie BJ, Grimm J. Quantitative imaging of disease
3969 signatures through radioactive decay signal conversion. *Nat Med.* 2013; 19: 1345-50.

3970 285. Joaqui-Joaqui MA, Sands GG, Śmiłowicz D, Gork MJ, Aluicio-Sarduy E,
3971 Barnhart TE, et al. A Trifunctional, Rare-Earth Theranostic Chelator Platform to Enable

3972 Diagnostic Nuclear Imaging, Surgical Resection, and Radiotherapy. *J Am Chem Soc.*
3973 2025; 147: 45303-14.

3974 286. Guo R, Cao T, Yang S, Liu H, Guo Q, Lan X, et al. Improved X-ray absorption
3975 capability of Core-shell nano-transducer in situ enhances γ -ray-excited
3976 radioluminescence imaging in vivo. *J Nanobiotechnology.* 2026; 24: 141.

3977 287. Zhang GQ, Feng G, Xu C, Gao Z, Zhang J, Li L, et al. Radioisotope-mimetic
3978 molecular afterglow probe for downregulated cancer biomarker imaging. *Nat Mater.*
3979 2026.

3980 288. Qian R, Wang K, Guo Y, Li H, Zhu Z, Huang X, et al. Minimizing adverse effects
3981 of Cerenkov radiation induced photodynamic therapy with transformable
3982 photosensitizer-loaded nanovesicles. *J Nanobiotechnology.* 2022; 20: 203.

3983 289. Li J, Dai S, Qin R, Shi C, Ming J, Zeng X, et al. Ligand Engineering of Titanium-
3984 Oxo Nanoclusters for Cerenkov Radiation-Reinforced Photo/Chemodynamic Tumor
3985 Therapy. *ACS Appl Mater Interfaces.* 2021; 13: 54727-38.

3986 290. Guo R, Jiang D, Gai Y, Qian R, Zhu Z, Gao Y, et al. Chlorin e6-loaded goat milk-
3987 derived extracellular vesicles for Cerenkov luminescence-induced photodynamic
3988 therapy. *Eur J Nucl Med Mol Imaging.* 2023; 50: 508-24.

3989 291. Kotagiri N, Cooper ML, Rettig M, Egbulefu C, Prior J, Cui G, et al. Radionuclides
3990 transform chemotherapeutics into phototherapeutics for precise treatment of
3991 disseminated cancer. *Nat Commun.* 2018; 9: 275.

3992 292. Goel S, Ferreira CA, Chen F, Ellison PA, Siamof CM, Barnhart TE, et al.
3993 Activatable Hybrid Nanotheranostics for Tetramodal Imaging and Synergistic
3994 Photothermal/Photodynamic Therapy. *Adv Mater.* 2018; 30.

3995 293. Zhang Q, Pratt EC, Tamura R, Ogirala A, Hsu H-T, Farahmand N, et al. Ultrasmall
3996 Downconverting Nanoparticle for Enhanced Cerenkov Imaging. *Nano Lett.* 2021; 21:
3997 4217-24.

3998 294. Su W, Wang H, Zhao S, Wang T, Hsu JC, Ji X, et al. Cerenkov radiation modulates
3999 the extracellular matrix for improved pancreatic cancer chemotherapy. *Cell Biomater.*
4000 2025: 100221.

4001 295. Zhu Z, Liu Q, Zhu K, Wang K, Lin L, Chen Y, et al. Aggregation-induced
4002 emission photosensitizer/bacteria biohybrids enhance Cerenkov radiation-induced
4003 photodynamic therapy by activating anti-tumor immunity for synergistic tumor
4004 treatment. *Acta Biomater.* 2023; 167: 519-33.

4005 296. Liu H, Wang Q, Guo J, Feng K, Ruan Y, Zhang Z, et al. Prodrug-based strategy
4006 with a two-in-one liposome for Cerenkov-induced photodynamic therapy and
4007 chemotherapy. *J Control Release.* 2023; 364: 206-15.

4008 297. Jo C, Ahn H, Kim JH, Lee YJ, Kim JY, Lee KC, et al. Cancer therapy by antibody-
4009 targeted Cerenkov light and metabolism-selective photosensitization. *J Control Release*.
4010 2022; 352: 25-34.

4011 298. Rosenkrans ZT, Hsu JC, Aluicio-Sarduy E, Barnhart TE, Engle JW, Cai W.
4012 Amplification of Cerenkov luminescence using semiconducting polymers for cancer
4013 theranostics. *Adv Funct Mater*. 2023; 33.

4014 299. Cao C, Treechairusame T, Safavi AH, Wu Y, Zhang Z, Shamseddine A, et al.
4015 Intensity-modulated proton therapy vs intensity-modulated radiotherapy in
4016 nasopharyngeal carcinoma: a case–control study. *Lancet Reg Health Am*.
4017 2026; 54.

4018 300. Vasudevan SS, Deeb H, Katta A, Olinde L, Pang J, Asarkar AA, et al. Efficacy
4019 and safety of proton therapy versus intensity-modulated radiation therapy in the
4020 treatment of head and neck tumors: A systematic review and meta-analysis. *Head Neck*.
4021 2024; 46: 2616-31.

4022 301. Mohan R. A Review of Proton Therapy - Current Status and Future Directions.
4023 *Precis Radiat Oncol*. 2022; 6: 164-76.

4024 302. Masuda T, Kataoka J, Arimoto M, Takabe M, Nishio T, Matsushita K, et al.
4025 Measurement of nuclear reaction cross sections by using Cherenkov radiation toward
4026 high-precision proton therapy. *Sci Rep*. 2018; 8: 2570.

4027 303. Carbone GG, Mariano S, Gabriele A, Cennamo S, Primiceri V, Aziz MR, et al.
4028 Exploring the Potential of Gold Nanoparticles in Proton Therapy: Mechanisms,
4029 Advances, and Clinical Horizons. *Pharmaceutics*. 2025; 17.

4030 304. Laughlin BS, Golafshar M, Prince M, Liu W, Kuttyreff CJ, Ahmed SK, et al.
4031 Dosimetric comparison between proton beam therapy, intensity modulated radiation
4032 therapy, and 3D conformal therapy for soft tissue extremity sarcoma. *Acta Oncol*. 2023;
4033 62: 473-9.

4034 305. Zavestovskaya IN, Filimonova MV, Popov AL, Zelepukin IV, Shemyakov AE,
4035 Tikhonowski GV, et al. Bismuth nanoparticles-enhanced proton therapy: Concept and
4036 biological assessment. *Mater Today Nano*. 2024; 27: 100508.

4037 306. Zwiehoff S, Johny J, Behrends C, Landmann A, Mentzel F, Bäumer C, et al.
4038 Enhancement of Proton Therapy Efficiency by Noble Metal Nanoparticles Is Driven by
4039 the Number and Chemical Activity of Surface Atoms. *Small*. 2022; 18: e2106383.

4040 307. Peukert D, Kempson I, Douglass M, Bezak E. Gold Nanoparticle Enhanced Proton
4041 Therapy: Monte Carlo Modeling of Reactive Species' Distributions Around a Gold
4042 Nanoparticle and the Effects of Nanoparticle Proximity and Clustering. *Int J Mol Sci*.
4043 2019; 20.

4044 308. Lo C-Y, Tsai S-W, Niu H, Chen F-H, Hwang H-C, Chao T-C, et al. Gold-
4045 Nanoparticles-Enhanced Production of Reactive Oxygen Species in Cells at Spread-
4046 Out Bragg Peak under Proton Beam Radiation. *ACS Omega*. 2023; 8: 17922-31.

4047 309. Lee DW, Kim JE, Lee G-H, Son A, Park HC, Oh D, et al. High-Throughput 3D
4048 Tumor Spheroid Array Platform for Evaluating Sensitivity of Proton-Drug
4049 Combinations. *Int J Mol Sci*. 2022; 23: 587.

4050 310. Tudor M, Popescu RC, Irimescu IN, Rzyanina A, Tarba N, Dinischiotu A, et al.
4051 Enhancing Proton Radiosensitivity of Chondrosarcoma Using Nanoparticle-Based
4052 Drug Delivery Approaches: A Comparative Study of High- and Low-Energy Protons.
4053 *Int J Mol Sci*. 2024; 25: 11481.

4054 311. Silva CR, Berionni G, Ribeiro MS, Heuskin A-C. Porphyrin-coated gold
4055 nanoparticles associated with X-rays and proton therapy in the treatment of triple-
4056 negative breast cancer. *Nanoscale Adv*. 2026; 8: 2031-42.

4057 312. Fagundes MA, Zerey MM, Gal O, Wohl J, Rodrigues MAM, Kaiser A, et al. Dose-
4058 Escalated Intensity Modulated Proton Therapy (IMPT) for Node-Positive Prostate
4059 Cancer – First Report of Clinical Outcomes. *Int J Radiat Oncol Biol Phys*.
4060 2025; 123: e548.

4061 313. Parwaie W, Molazadeh M, Mortezaadeh T. Evaluation of the impact of energy,
4062 radiation type, and concentration on dose enhancement by Gold Nanoparticles. *Cancer*
4063 *Treat Res Commun*. 2025; 43: 100933.

4064 314. Bonvalot S, Le Pechoux C, De Baere T, Kantor G, Buy X, Stoeckle E, et al. First-
4065 in-Human Study Testing a New Radioenhancer Using Nanoparticles (NBTXR3)
4066 Activated by Radiation Therapy in Patients with Locally Advanced Soft Tissue
4067 Sarcomas. *Clin Cancer Res*. 2017; 23: 908-17.

4068 315. Huang CW, Hu HM, Hsu WH, Chen CY, Huang MY, Chen CP, et al. A phase
4069 Ib/II trial of PEP503 (NBTXR3, radioenhancer) with radiotherapy and chemotherapy
4070 in patients with rectal cancer. *Nanomedicine (Lond)*. 2023; 18: 511-24.

4071 316. Bonvalot S, Rutkowski PL, Thariat J, Carrère S, Ducassou A, Sunyach MP, et al.
4072 Final Safety and Health-Related Quality of Life Results of the Phase 2/3 Act.In.Sarc
4073 Study With Preoperative NBTXR3 Plus Radiation Therapy Versus Radiation Therapy
4074 in Locally Advanced Soft-Tissue Sarcoma. *Int J Radiat Oncol Biol Phys*. 2022; 114:
4075 422-32.

4076 317. Bagley AF, Ludmir EB, Maitra A, Minsky BD, Li Smith G, Das P, et al. NBTXR3,
4077 a first-in-class radioenhancer for pancreatic ductal adenocarcinoma: Report of first
4078 patient experience. *Clin Transl Radiat Oncol*. 2022; 33: 66-9.

4079 318. Hoffmann C, Calugaru V, Borcoman E, Moreno V, Calvo E, Liem X, et al. Phase
4080 I dose-escalation study of NBTXR3 activated by intensity-modulated radiation therapy

4081 in elderly patients with locally advanced squamous cell carcinoma of the oral cavity or
4082 oropharynx. *Eur J Cancer*. 2021; 146: 135-44.

4083 319. Chajon E, Pracht M, Rolland Y, Nguyen F, Bronowicki JP, Durand-Labrunie J, et
4084 al. Radioenhancing Hafnium Oxide Nanoparticles (NBTXR3) Followed by Stereotactic
4085 Body Radiation Therapy in Patients With Hepatocellular Carcinoma and Liver
4086 Metastases (NBTXR3-103): Phase 1 Dose-Escalation Trial. *Adv Radiat Oncol*. 2026;
4087 11: 101937.

4088 320. Selbo PK, Korbelik M. Enhancing antitumour immunity with photodynamic
4089 therapy. *Photochem Photobiol Sci*. 2025; 24: 227-34.

4090 321. Yun WS, Park JH, Lim DK, Ahn CH, Sun IC, Kim K. How Did Conventional
4091 Nanoparticle-Mediated Photothermal Therapy Become "Hot" in Combination with
4092 Cancer Immunotherapy? *Cancers (Basel)*. 2022; 14.

4093 322. Syed Altaf RR, Mohan A, Palani N, Mendonce KC, Monisha P, Rajadesingu S. A
4094 review of innovative design strategies: Artificial antigen presenting cells in cancer
4095 immunotherapy. *Int J Pharm*. 2025; 669: 125053.

4096 323. Hato L, Vizcay A, Eguren I, Pérez-Gracia JL, Rodríguez J, Gállego Pérez-Larraya
4097 J, et al. Dendritic Cells in Cancer Immunology and Immunotherapy. *Cancers (Basel)*.
4098 2024; 16: 981.

4099 324. Qian J, Liu Y. Recent advances in adoptive cell therapy for cancer immunotherapy.
4100 *Front Immunol*. 2025; 16: 2025.

4101 325. Sharma P, Goswami S, Raychaudhuri D, Siddiqui BA, Singh P, Nagarajan A, et
4102 al. Immune checkpoint therapy—current perspectives and future directions. *Cell*. 2023;
4103 186: 1652-69.

4104 326. Babamohamadi M, Mohammadi N, Faryadi E, Haddadi M, Merati A,
4105 Ghobadinezhad F, et al. Anti-CTLA-4 nanobody as a promising approach in cancer
4106 immunotherapy. *Cell Death Dis*. 2024; 15: 17.

4107 327. Parvez A, Choudhary F, Mudgal P, Khan R, Qureshi KA, Farooqi H, et al. PD-1
4108 and PD-L1: architects of immune symphony and immunotherapy breakthroughs in
4109 cancer treatment. *Front Immunol*. 2023; 14 - 2023.

4110 328. Wolchok J. Putting the Immunologic Brakes on Cancer. *Cell*. 2018; 175: 1452-4.

4111 329. Zhao Z, He Y, Shi C, Li J, Liu Q, Zhong Y, et al. Immunotherapy augmentation
4112 through photothermal nanomaterial-mediated tumor microenvironment modulation.
4113 *Coord Chem Rev*. 2026; 547: 217118.

4114 330. Yu S, Wang Y, He P, Shao B, Liu F, Xiang Z, et al. Effective Combinations of
4115 Immunotherapy and Radiotherapy for Cancer Treatment. *Front Oncol*. 2022; 12:
4116 809304.

4117 331. Cui M, He J, Zhang F, Zhang Y, Gao A, Liu J. Abscopal effect induced by
4118 conventional fractionated radiotherapy following anti-PD-1 immunotherapy in

4119 pulmonary metastatic thymic squamous cell carcinoma: a case report and literature
4120 review. *Front Immunol.* 2026; 17: 1733066.

4121 332. He M, Chen S, Yu H, Fan X, Wu H, Wang Y, et al. Advances in nanoparticle-
4122 based radiotherapy for cancer treatment. *iScience.* 2025; 28: 111602.

4123 333. Su JX, Li SJ, Zhou XF, Zhang ZJ, Yan Y, Liu SL, et al. Chemotherapy-induced
4124 metastasis: molecular mechanisms and clinical therapies. *Acta Pharmacol Sin.* 2023;
4125 44: 1725-36.

4126 334. Li Q, Liu Y, Huang Z, Guo Y, Li Q. Triggering Immune System With
4127 Nanomaterials for Cancer Immunotherapy. *Front Bioeng Biotechnol.* 2022; 10: 878524.

4128 335. Zhu W, Pan S, Zhang J, Xu J, Zhang R, Zhang Y, et al. The role of hyperthermia
4129 in the treatment of tumor. *Crit Rev Oncol Hematol.* 2024; 204: 104541.

4130 336. Bouwstra R, van Meerten T, Bremer E. CD47-SIRP α blocking-based
4131 immunotherapy: Current and prospective therapeutic strategies. *Clin Transl Med.* 2022;
4132 12: e943.

4133 337. Lax BM, Palmeri JR, Lutz EA, Sheen A, Stinson JA, Duhamel L, et al. Both
4134 intratumoral regulatory T cell depletion and CTLA-4 antagonism are required for
4135 maximum efficacy of anti-CTLA-4 antibodies. *Proc Natl Acad Sci USA.* 2023; 120:
4136 e2300895120.

4137 338. Liu R, Li HF, Li S. PD-1-mediated inhibition of T cell activation: Mechanisms
4138 and strategies for cancer combination immunotherapy. *Cell Insight.* 2024; 3: 100146.

4139 339. Liu Y, Maccarini P, Palmer GM, Etienne W, Zhao Y, Lee C-T, et al. Synergistic
4140 Immuno Photothermal Nanotherapy (SYMPHONY) for the Treatment of Unresectable
4141 and Metastatic Cancers. *Sci Rep.* 2017; 7: 8606.

4142 340. Wilky BA, Julian KA, Maleddu A, Mailhot AC, Cartwright CR, Gao D, et al. A
4143 Single-Arm Phase 2 Trial of Doxorubicin Plus Zalifrelimab (Anti-CTLA-4 Antibody)
4144 and Balstilimab (Anti-PD-1 Antibody) in Advanced/Metastatic Soft Tissue Sarcomas.
4145 *Clin Cancer Res.* 2025; 31: 2945-56.

4146 341. Wang JW, Feng YF, Liu JH. CTLA-4 and PD-1 combined blockade therapy for
4147 malignant melanoma brain metastases: mechanisms, challenges, and prospects. *Front*
4148 *Immunol.* 2025; 16: 1629879.

4149 342. Cai X, Leung AWN, Lv L, Xu C. A mini-review: photodynamic therapy-induced
4150 immune activation. *Front Pharmacol.* 2026; 17 - 2026.

4151 343. Bayless S, Travers JB, Sahu RP, Rohan CA. Inhibition of photodynamic therapy
4152 induced-immunosuppression with aminolevulinic acid leads to enhanced outcomes of
4153 tumors and pre-cancerous lesions. *Oncol Lett.* 2021; 22: 664.

4154 344. Zhang Y, Sriramaneni RN, Clark PA, Jagodinsky JC, Ye M, Jin W, et al.
4155 Multifunctional nanoparticle potentiates the in situ vaccination effect of radiation

4156 therapy and enhances response to immune checkpoint blockade. *Nat Commun.* 2022;
4157 13: 4948.

4158 345. Lin G, Tillman L, Luo T, Jiang X, Fan Y, Liu G, et al. Nanoscale Metal-Organic
4159 Layer Reprograms Cellular Metabolism to Enhance Photodynamic Therapy and
4160 Antitumor Immunity. *Angew Chem Int Ed Engl.* 2024; 63: e202410241.

4161 346. Zhu W, Guo Y, Huang J, Zhang Y, Ni Z, Wei M, et al. NIR-II photothermal
4162 therapy mediated by polymer-based nanoparticles to enhance checkpoint inhibitor
4163 immunotherapy for inhibiting lymph node metastasis in oral squamous cell carcinoma.
4164 *Nano Today.* 2024; 57: 102351.

4165 347. Galluzzi L, Kepp O, Hett E, Kroemer G, Marincola FM. Immunogenic cell death
4166 in cancer: concept and therapeutic implications. *J Transl Med.* 2023; 21: 162.

4167 348. Wang K, Li Y, Wang X, Zhang Z, Cao L, Fan X, et al. Gas therapy potentiates
4168 aggregation-induced emission luminogen-based photoimmunotherapy of poorly
4169 immunogenic tumors through cGAS-STING pathway activation. *Nat Commun.* 2023;
4170 14: 2950.

4171 349. Turubanova VD, Mishchenko TA, Balalaeva IV, Efimova I, Peskova NN,
4172 Klapshina LG, et al. Novel porphyrazine-based photodynamic anti-cancer therapy
4173 induces immunogenic cell death. *Sci Rep.* 2021; 11: 7205.

4174 350. Li W, Yang J, Luo L, Jiang M, Qin B, Yin H, et al. Targeting photodynamic and
4175 photothermal therapy to the endoplasmic reticulum enhances immunogenic cancer cell
4176 death. *Nat Commun.* 2019; 10: 3349.

4177 351. Yang W, Zhang F, Deng H, Lin L, Wang S, Kang F, et al. Smart Nanovesicle-
4178 Mediated Immunogenic Cell Death through Tumor Microenvironment Modulation for
4179 Effective Photodynamic Immunotherapy. *ACS Nano.* 2020; 14: 620-31.

4180 352. Deng Z, Xi M, Zhang C, Wu X, Li Q, Wang C, et al. Biomineralized MnO₂
4181 Nanoplatfoms Mediated Delivery of Immune Checkpoint Inhibitors with STING
4182 Pathway Activation to Potentiate Cancer Radio-Immunotherapy. *ACS Nano.* 2023; 17:
4183 4495-506.

4184 353. Chen Q, Chen J, Zhang Q, Yang P, Gu R, Ren H, et al. Combining High-Z
4185 Sensitized Radiotherapy with CD73 Blockade to Boost Tumor Immunotherapy. *ACS*
4186 *Nano.* 2023; 17: 12087-100.

4187 354. Huang Z, Wang Y, Yao D, Wu J, Hu Y, Yuan A. Nanoscale coordination polymers
4188 induce immunogenic cell death by amplifying radiation therapy mediated oxidative
4189 stress. *Nat Commun.* 2021; 12: 145.

4190 355. Li J, Tan C, Yang J, Xiang Z, Wang Y, Shen M, et al. Radiotherapy-
4191 immunomodulated nanoplatfom triggers both hypoxic and normoxic tumor associated
4192 antigens generation for robust abscopal effect and sustained immune memory.
4193 *Biomaterials.* 2025; 316: 123005.

4194 356. Xu P, Ma J, Zhou Y, Gu Y, Cheng X, Wang Y, et al. Radiotherapy-Triggered In
4195 Situ Tumor Vaccination Boosts Checkpoint Blockaded Immune Response via Antigen-
4196 Capturing Nanoadjuvants. *ACS Nano*. 2024; 18: 1022-40.

4197 357. Tang D, Chen X, Kang R, Kroemer G. Ferroptosis: molecular mechanisms and
4198 health implications. *Cell Res*. 2021; 31: 107-25.

4199 358. Tang D, Kroemer G. Ferroptosis. *Curr Biol*. 2020; 30: R1292-r7.

4200 359. Tang Y, Zhou Q, Fan Y, Liu T-M, Zhu L, Liu M. Nanomaterial-mediated
4201 ferroptosis as a promising strategy for cancer therapy. *Nanoscale*. 2025; 17: 27066-84.

4202 360. Wang Z, Xu H, Li M, Lei S, Cai Y, Wu Y, et al. Nanotechnology for tumor
4203 ferroptosis. *Cell Biomater*. 2026; 2.

4204 361. Zhu X, Zheng W, Wang X, Li Z, Shen X, Chen Q, et al. Enhanced Photodynamic
4205 Therapy Synergizing with Inhibition of Tumor Neutrophil Ferroptosis Boosts Anti-PD-
4206 1 Therapy of Gastric Cancer. *Adv Sci (Weinh)*. 2024; 11: e2307870.

4207 362. Zhuang J, Wang B, Chen H, Zhang K, Li N, Zhao N, et al. Efficient NIR-II Type-
4208 I AIE Photosensitizer for Mitochondria-Targeted Photodynamic Therapy through
4209 Synergistic Apoptosis–Ferroptosis. *ACS Nano*. 2023; 17: 9110-25.

4210 363. Chen W, Xie W, Gao Z, Lin C, Tan M, Zhang Y, et al. Mild-Photothermal Effect
4211 Induced High Efficiency Ferroptosis-Boosted-Cuproptosis Based on
4212 Cu₂O@Mn₃Cu₃O₈ Nanozyme. *Adv Sci (Weinh)*. 2023; 10: 2303694.

4213 364. Chen C, Wang Z, Jia S, Zhang Y, Ji S, Zhao Z, et al. Evoking Highly Immunogenic
4214 Ferroptosis Aided by Intramolecular Motion-Induced Photo-Hyperthermia for Cancer
4215 Therapy. *Adv Sci (Weinh)*. 2022; 9: 2104885.

4216 365. Aishajiang R, Liu Z, Liang Y, Du P, Wei Y, Zhuo X, et al. Concurrent
4217 Amplification of Ferroptosis and Immune System Activation Via Nanomedicine-
4218 Mediated Radiosensitization for Triple-Negative Breast Cancer Therapy. *Adv Sci*
4219 *(Weinh)*. 2024: e2407833.

4220 366. Wang D, Tang L, Chen M, Gong Z, Fan C, Qu H, et al. Nanocarriers Targeting
4221 Circular RNA ADARB1 Boost Radiosensitivity of Nasopharyngeal Carcinoma through
4222 Synergically Promoting Ferroptosis. *ACS Nano*. 2024; 18: 31055-75.

4223 367. Bae C, Hernández Millares R, Ryu S, Moon H, Kim D, Lee G, et al. Synergistic
4224 Effect of Ferroptosis-Inducing Nanoparticles and X-Ray Irradiation Combination
4225 Therapy. *Small*. 2024; 20: 2310873.

4226 368. Tang D, Kroemer G, Kang R. Targeting cuproplasia and cuproptosis in cancer.
4227 *Nat Rev Clin Oncol*. 2024; 21: 370-88.

4228 369. Tang D, Chen X, Kroemer G. Cuproptosis: a copper-triggered modality of
4229 mitochondrial cell death. *Cell Res*. 2022; 32: 417-8.

4230 370. Wang Y, Zhang L, Zhou F. Cuproptosis: a new form of programmed cell death.
4231 *Cell Mol Immunol*. 2022; 19: 867-8.

4232 371. Li W, Xiao Y, Guo G, Peng J, Zhu N, Chen Z, et al. Cuprous oxide
4233 nanocomposites with photothermal (PTT) and chemical dynamics (CDT) effects induce
4234 cuproptosis in breast cancer using the strategy of increasing inflow and reducing
4235 outflow. *Nano Today*. 2024; 56: 102223.

4236 372. Zhou J, Yu Q, Song J, Li S, Li XL, Kang BK, et al. Photothermally Triggered
4237 Copper Payload Release for Cuproptosis-Promoted Cancer Synergistic Therapy.
4238 *Angew Chem Int Ed Engl*. 2023; 62: e202213922.

4239 373. Wu L, Lin H, Cao X, Tong Q, Yang F, Miao Y, et al. Bioorthogonal Cu Single-
4240 Atom Nanozyme for Synergistic Nanocatalytic Therapy, Photothermal Therapy,
4241 Cuproptosis and Immunotherapy. *Angew Chem Int Ed Engl* . 2024; 63: e202405937.

4242 374. Liao Y, Wang D, Gu C, Wang X, Zhu S, Zheng Z, et al. A cuproptosis nanocapsule
4243 for cancer radiotherapy. *Nat Nanotechnol*. 2024; 19: 1892-902.

4244 375. Yang L, Zhao Z, Tian B, Yang M, Dong Y, Zhou B, et al. A singular plasmonic-
4245 thermoelectric hollow nanostructure inducing apoptosis and cuproptosis for catalytic
4246 cancer therapy. *Nat Commun*. 2024; 15: 7499.

4247 376. Wang X, Wang D, Liao Y, Guo X, Song Q, Liu W, et al. Hafnium oxide-based
4248 sensitizer with radiation-triggered cuproptosis for radiotherapy. *Nano Today*. 2025; 61:
4249 102626.

4250 377. Wei X, Xie F, Zhou X, Wu Y, Yan H, Liu T, et al. Role of pyroptosis in
4251 inflammation and cancer. *Cell Mol Immunol*. 2022; 19: 971-92.

4252 378. Kong Q, Zhang Z. Cancer-associated pyroptosis: A new license to kill tumor.
4253 *Front Immunol*. 2023; 14.

4254 379. Liu D, Liang M, Tao Y, Liu H, Liu Q, Bing W, et al. Hypoxia-accelerating
4255 pyroptosis nanoinducers for promoting image-guided cancer immunotherapy.
4256 *Biomaterials*. 2024; 309: 122610.

4257 380. Xiao Y, Zhang T, Ma X, Yang QC, Yang LL, Yang SC, et al. Microenvironment-
4258 Responsive Prodrug-Induced Pyroptosis Boosts Cancer Immunotherapy. *Adv Sci*
4259 (Weinh). 2021; 8: e2101840.

4260 381. Hu Z-C, Wang B, Zhou X-G, Liang H-F, Liang B, Lu H-W, et al. Golgi Apparatus-
4261 Targeted Photodynamic Therapy for Enhancing Tumor Immunogenicity by Eliciting
4262 NLRP3 Protein-Dependent Pyroptosis. *ACS Nano*. 2023; 17: 21153-69.

4263 382. Wang B, Zhou H, Chen L, Ding Y, Zhang X, Chen H, et al. A Mitochondria-
4264 Targeted Photosensitizer for Combined Pyroptosis and Apoptosis with NIR-II
4265 Imaging/Photoacoustic Imaging-Guided Phototherapy. *Angew Chem Int Ed Engl*. 2024;
4266 63: e202408874.

4267 383. Xing G, Yu X, Zhang Y, Sheng S, Jin L, Zhu D, et al. Macrophages-Based
4268 Biohybrid Microrobots for Breast Cancer Photothermal Immunotherapy by Inducing
4269 Pyroptosis. *Small*. 2024; 20: 2305526.

4270 384. Liu R, Wang R, Zhao M, Liu Y, Zhu X, Wu X, et al. Ultra-small radiosensitizers
4271 deliver epigenetic drugs to induce pyroptosis and boost triple-negative breast cancer
4272 radiotherapy. *Nano Today*. 2023; 52: 101997.

4273 385. Samson N, Ablasser A. The cGAS–STING pathway and cancer. *Nat Cancer*. 2022;
4274 3: 1452-63.

4275 386. Guo Z, Liu T, Gao Q, Wang C, Wang Q, Du R, et al. Targeting the cGAS-STING
4276 Pathway for Cancer Immunotherapy: From Small-Molecule Agonists to Advanced
4277 Nanomaterials. *Mol Pharm*. 2025; 22: 7262-84.

4278 387. Li Y, Li X, Yi J, Cao Y, Qin Z, Zhong Z, et al. Nanoparticle-Mediated STING
4279 Activation for Cancer Immunotherapy. *Adv Healthc Mater*. 2023; 12: e2300260.

4280 388. Qiao Y, Wei L, Su Y, Tan Q, Yang X, Li S. Nanoparticle-Based Strategies to
4281 Enhance the Efficacy of STING Activators in Cancer Immunotherapy. *Int J*
4282 *Nanomedicine*. 2025; 20: 5429-56.

4283 389. Sun X, Zhang S, Li Q, Yang M, Qiu X, Yu B, et al. Bimetallic infinite coordination
4284 nanopolymers via phototherapy and STING activation for eliciting robust antitumor
4285 immunity. *J Colloid Interface Sci*. 2023; 642: 691-704.

4286 390. Liu X, Shen M, Bing T, Zhang X, Li Y, Cai Q, et al. A Bioactive Injectable
4287 Hydrogel Regulates Tumor Metastasis and Wound Healing for Melanoma via NIR-
4288 Light Triggered Hyperthermia. *Adv Sci (Weinh)*. 2024; 11: 2402208.

4289 391. Tao H, Tan J, Zhang H, Ren H, Cai Z, Liu H, et al. cGAS-STING Pathway
4290 Activation and Systemic Anti-Tumor Immunity Induction via Photodynamic
4291 Nanoparticles with Potent Toxic Platinum DNA Intercalator Against Uveal Melanoma.
4292 *Adv Sci (Weinh)*. 2023; 10: 2302895.

4293 392. Luo T, Nash GT, Jiang X, Feng X, Mao J, Liu J, et al. A 2D Nanoradiosensitizer
4294 Enhances Radiotherapy and Delivers STING Agonists to Potentiate Cancer
4295 Immunotherapy. *Adv Mater*. 2022; 34: 2110588.

4296 393. Liu Y, Crowe WN, Wang L, Lu Y, Petty WJ, Habib AA, et al. An inhalable
4297 nanoparticulate STING agonist synergizes with radiotherapy to confer long-term
4298 control of lung metastases. *Nat Commun*. 2019; 10: 5108.

4299 394. Zhan M, Yu X, Zhao W, Peng Y, Peng S, Li J, et al. Extracellular matrix-degrading
4300 STING nanoagonists for mild NIR-II photothermal-augmented chemodynamic-
4301 immunotherapy. *J Nanobiotechnology*. 2022; 20: 23.

4302 395. Xia J, Wang L, Shen T, Li P, Zhu P, Xie S, et al. Integrated manganese (III)-doped
4303 nanosystem for optimizing photothermal ablation: Amplifying hyperthermia-induced
4304 STING pathway and enhancing antitumor immunity. *Acta Biomater*. 2023; 155: 601-
4305 17.

4306 396. Zheng Y, Chen J, Song X-R, Chang M-Q, Feng W, Huang H, et al. Manganese-
4307 enriched photonic/catalytic nanomedicine augments synergistic anti-TNBC

4308 photothermal/nanocatalytic/immuno-therapy via activating cGAS-STING pathway.
4309 Biomaterials. 2023; 293: 121988.

4310 397. Wu C, Feng D, Xu H, He Z, Hou J. Optimized Bionic Drug-Delivery-Inducing
4311 Immunogenic Cell Death and cGAS-STING Pathway Activation for Enhanced
4312 Photodynamic-Chemotherapy-Driven Immunotherapy in Prostate Cancer. ACS Appl
4313 Mater Interfaces. 2024; 16: 43257-71.

4314 398. Zhou Q, Dutta D, Cao Y, Ge Z. Oxidation-Responsive PolyMOF Nanoparticles
4315 for Combination Photodynamic-Immunotherapy with Enhanced STING Activation.
4316 ACS Nano. 2023; 17: 9374-87.

4317 399. Peng Y, Liang S, Liu D, Ma K, Yun K, Zhou M, et al. Multi-Metallic Nanosheets
4318 Reshaping Immunosuppressive Tumor Microenvironment through Augmenting cGAS-
4319 STING Innate Activation and Adaptive Immune Responses for Cancer Immunotherapy.
4320 Adv Sci (Weinh). 2024; 11: 2403347.

4321 400. Wang L, Zhou H, Chen Q, Lin Z, Jiang C, Chen X, et al. STING Agonist-Loaded
4322 Nanoparticles Promotes Positive Regulation of Type I Interferon-Dependent
4323 Radioimmunotherapy in Rectal Cancer. Adv Sci (Weinh). 2024; 11: e2307858.

4324 401. Ding F, Liu J, Ai K, Xu C, Mao X, Liu Z, et al. Simultaneous Activation of
4325 Pyroptosis and cGAS-STING Pathway with Epigenetic/ Photodynamic
4326 Nanotheranostic for Enhanced Tumor Photoimmunotherapy. Adv Mater. 2024; 36:
4327 e2306419.

4328 402. Ma W, Sun R, Tang L, Li Z, Lin L, Mai Z, et al. Bioactivable STING Nanoagonists
4329 to Synergize NIR-II Mild Photothermal Therapy Primed Robust and Long-Term
4330 Anticancer Immunity. Adv Mater. 2023; 35: e2303149.

4331 403. Guo X, Tu P, Wang X, Du C, Jiang W, Qiu X, et al. Decomposable Nanoagonists
4332 Enable NIR-Elicited cGAS-STING Activation for Tandem-Amplified Photodynamic-
4333 Metalloimmunotherapy. Adv Mater. 2024; 36: e2313029.

4334 404. Mou X, Leeman SM, Roye Y, Miller C, Musah S. Fenestrated Endothelial Cells
4335 across Organs: Insights into Kidney Function and Disease. Int J Mol Sci. 2024; 25.

4336 405. Satchell SC, Braet F. Glomerular endothelial cell fenestrations: an integral
4337 component of the glomerular filtration barrier. Am J Physiol Renal Physiol. 2009; 296:
4338 F947-56.

4339 406. Liu J, Yu M, Zhou C, Zheng J. Renal clearable inorganic nanoparticles: A new
4340 frontier of bionanotechnology. Mater Today. 2013; 16: 477-86.

4341 407. Adhipandito CF, Cheung SH, Lin YH, Wu SH. Atypical Renal Clearance of
4342 Nanoparticles Larger Than the Kidney Filtration Threshold. Int J Mol Sci. 2021; 22:
4343 11182.

4344 408. Cabral H, Li J, Miyata K, Kataoka K. Controlling the biodistribution and clearance
4345 of nanomedicines. Nat Rev Bioeng. 2024; 2: 214-32.

4346 409. Tang S, Huang Y, Zheng J. Salivary Excretion of Renal-Clearable Silver
4347 Nanoparticles. *Angew Chem Int Ed Engl.* 2020; 59: 19894-8.

4348 410. Yang Q, Chen D, Liu X, Li W, Zheng H, Cai X, et al. Identification of nanoparticle
4349 infiltration in human breast milk: Chemical profiles and trajectory pathways. *Proc Natl*
4350 *Acad Sci U S A.* 2025; 122: e2500552122.

4351 411. Teow Y, Asharani PV, Hande MP, Valiyaveetil S. Health impact and safety of
4352 engineered nanomaterials. *Chem Commun.* 2011; 47: 7025-38.

4353 412. Souris JS, Lee CH, Cheng SH, Chen CT, Yang CS, Ho JA, et al. Surface charge-
4354 mediated rapid hepatobiliary excretion of mesoporous silica nanoparticles.
4355 *Biomaterials.* 2010; 31: 5564-74.

4356 413. Zhang Y-N, Poon W, Tavares AJ, McGilvray ID, Chan WCW. Nanoparticle–liver
4357 interactions: Cellular uptake and hepatobiliary elimination. *J Control Release.* 2016;
4358 240: 332-48.

4359 414. Uzhytchak M, Smolková B, Lunova M, Frtús A, Jirsa M, Dejneka A, et al.
4360 Lysosomal nanotoxicity: Impact of nanomedicines on lysosomal function. *Adv Drug*
4361 *Deliv Rev.* 2023; 197: 114828.

4362 415. Wu Y, Cao X, Du H, Guo X, Han Y, McClements DJ, et al. Adverse effects of
4363 titanium dioxide nanoparticles on beneficial gut bacteria and host health based on
4364 untargeted metabolomics analysis. *Environ Res.* 2023; 228: 115921.

4365 416. Liu Q, Xu C, Ji G, Liu H, Mo Y, Tollerud DJ, et al. Sublethal effects of zinc oxide
4366 nanoparticles on male reproductive cells. *Toxicol In Vitro.* 2016; 35: 131-8.

4367 417. Nabeshi H, Yoshikawa T, Matsuyama K, Nakazato Y, Arimori A, Isobe M, et al.
4368 Amorphous nanosilicas induce consumptive coagulopathy after systemic exposure.
4369 *Nanotechnology.* 2012; 23: 045101.

4370 418. Huang C, Shao N, Huang Y, Chen J, Wang D, Hu G, et al. Overcoming challenges
4371 in the delivery of STING agonists for cancer immunotherapy: A comprehensive review
4372 of strategies and future perspectives. *Mater Today Bio.* 2023; 23: 100839.

4373 419. Wang Z, Wang Y, He Z, Liu C. Emerging cGAS-STING Agonist-Based
4374 Nanotherapeutics: Mechanistic Insights and Applications in Cancer Combination
4375 Therapy. *Adv Sci (Weinh).* 2025; 12: e09890.

4376 420. Zhang M, Ji Y, Liu M, Liu Y, Tong S, Cai Y, et al. Nanodelivery Strategies for
4377 STING Agonists: Toward Efficient Cancer Immunotherapy. *Int J Nanomedicine.* 2025;
4378 20: 12805-29.

4379 421. Dosta P, Cryer AM, Dion MZ, Shiraishi T, Langston SP, Lok D, et al.
4380 Investigation of the enhanced antitumour potency of STING agonist after conjugation
4381 to polymer nanoparticles. *Nat Nanotechnol.* 2023; 18: 1351-63.

4382 422. Huang A, Zhou W. Mn-based cGAS-STING activation for tumor therapy. *Chin J*
4383 *Cancer Res.* 2023; 35: 19-43.

4384 423. Zhao Z, Ma Z, Wang B, Guan Y, Su X-D, Jiang Z. Mn²⁺ Directly Activates cGAS
4385 and Structural Analysis Suggests Mn²⁺ Induces a Noncanonical Catalytic Synthesis of
4386 2'3'-cGAMP. *Cell Rep.* 2020; 32: 108053.

4387 424. Hooy RM, Massaccesi G, Rousseau KE, Chattergoon MA, Sohn J. Allosteric
4388 coupling between Mn²⁺ and dsDNA controls the catalytic efficiency and fidelity of
4389 cGAS. *Nucleic Acids Res.* 2020; 48: 4435-47.

4390 425. Ling K, Zheng J, Jiang X, Huang W, Mai Y, Liao C, et al. Mn²⁺/CpG
4391 Oligodeoxynucleotides Codecorated Black Phosphorus Nanosheet Platform for
4392 Enhanced Antitumor Potency in Multimodal Therapy. *ACS Nano.* 2024; 18: 2841-60.

4393 426. Wang Y, Li S, Hu M, Yang Y, McCabe E, Zhang L, et al. Universal STING mimic
4394 boosts antitumour immunity via preferential activation of tumour control signalling
4395 pathways. *Nat Nanotechnol.* 2024; 19: 856-66.

4396 427. Yan S, He Y, Zhu Y, Ye W, Chen Y, Zhu C, et al. Human patient derived
4397 organoids: an emerging precision medicine model for gastrointestinal cancer research.
4398 *Front Cell Dev Biol.* 2024; 12: 1384450.

4399 428. Natesh NR, Varghese S. Advances and challenges in human 3D solid tumor
4400 models. *Adv Funct Mater.* 2025; 35.

4401 429. Xia M, Wu G, Wu D, Hu W, Deng H, Wang S. Organoids, organ-on-a-chip, and
4402 microtumors: Biomimetic 3D tumor models advancing drug development and precision
4403 medicine. *Acta Pharm Sin B.* 2026.

4404 430. Domínguez-Oliva A, Hernández-Ávalos I, Martínez-Burnes J, Olmos-Hernández
4405 A, Verduzco-Mendoza A, Mota-Rojas D. The Importance of Animal Models in
4406 Biomedical Research: Current Insights and Applications. *Animals.* 2023; 13: 1223.

4407 431. Liu Y, Wu W, Cai C, Zhang H, Shen H, Han Y. Patient-derived xenograft models
4408 in cancer therapy: technologies and applications. *Signal Transduct Target Ther.* 2023;
4409 8: 160.

4410 432. Lange S, Inal JM. Animal Models of Human Disease. *Int J Mol Sci.* 2023; 24.

4411 433. Xuan L, Ju Z, Skonieczna M, Zhou PK, Huang R. Nanoparticles-induced potential
4412 toxicity on human health: Applications, toxicity mechanisms, and evaluation models.
4413 *MedComm (2020).* 2023; 4: e327.

4414 434. Egbuna C, Parmar VK, Jeevanandam J, Ezzat SM, Patrick-Iwuanyanwu KC,
4415 Adetunji CO, et al. Toxicity of Nanoparticles in Biomedical Application:
4416 Nanotoxicology. *J Toxicol.* 2021; 2021: 9954443.

4417 435. Makharadze D, del Valle LJ, Katsarava R, Puiggali J. The Art of PEGylation:
4418 From Simple Polymer to Sophisticated Drug Delivery System. *Int J Mol Sci.* 2025; 26:
4419 3102.

4420 436. Shen J, Zhang S, Wang X, Huang H, Xie L, Zhang Y, et al. Precise PEGylation
4421 Modulates the in Vivo Fate of Peptide Radiopharmaceuticals. *Small*. 2025; 21:
4422 2410410.

4423 437. Lipsa D, Ruiz Moreno A, Desmet C, Bianchi I, Geiss O, Colpo P, et al. Inter-
4424 Individual Variations: A Challenge for the Standardisation of Complement Activation
4425 Assays. *Int J Nanomedicine*. 2023; 18: 711-20.

4426 438. Bhattacharya K, Kiliç G, Costa PM, Fadeel B. Cytotoxicity screening and cytokine
4427 profiling of nineteen nanomaterials enables hazard ranking and grouping based on
4428 inflammogenic potential. *Nanotoxicology*. 2017; 11: 809-26.

4429 439. Szebeni J, Bedocs P, Dézsi L, Urbanics R. A porcine model of complement
4430 activation-related pseudoallergy to nano-pharmaceuticals: Pros and cons of translation
4431 to a preclinical safety test. *Precis Nanomed*. 2018; 1: 63-73.

4432 440. Hachhach M, Bayou S, El Kasmi A, Saidi MZ, Akram H, Hanafi M, et al. Towards
4433 Sustainable Scaling-Up of Nanomaterials Fabrication: Current Situation, Challenges,
4434 and Future Perspectives. *Eng*. 2025; 6: 149.

4435 441. Stavis SM, Fagan JA, Stopa M, Liddle JA. Nanoparticle Manufacturing –
4436 Heterogeneity through Processes to Products. *ACS Appl Nano Mater*. 2018; 1: 4358-
4437 85.

4438

4439

4440

4441