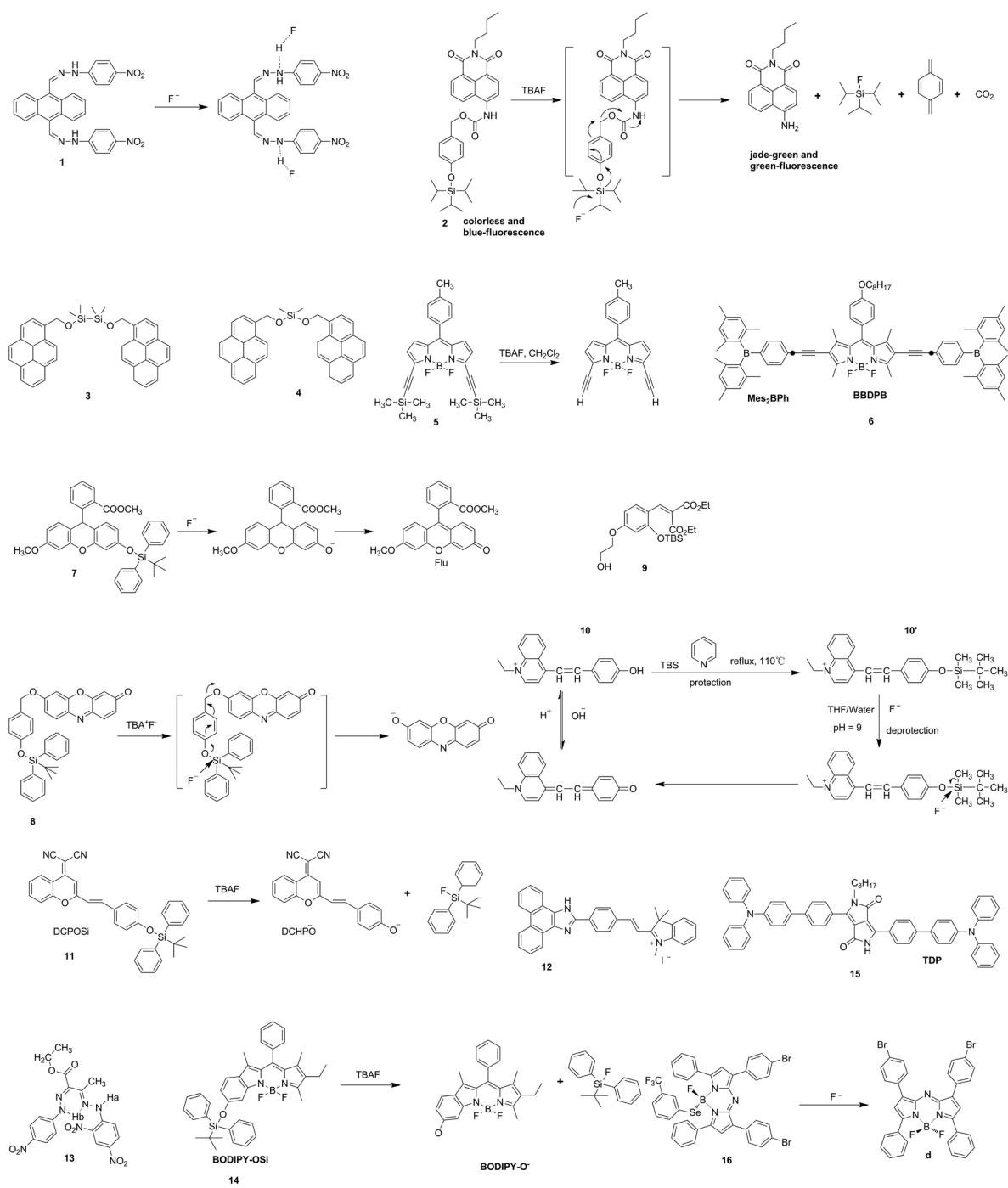
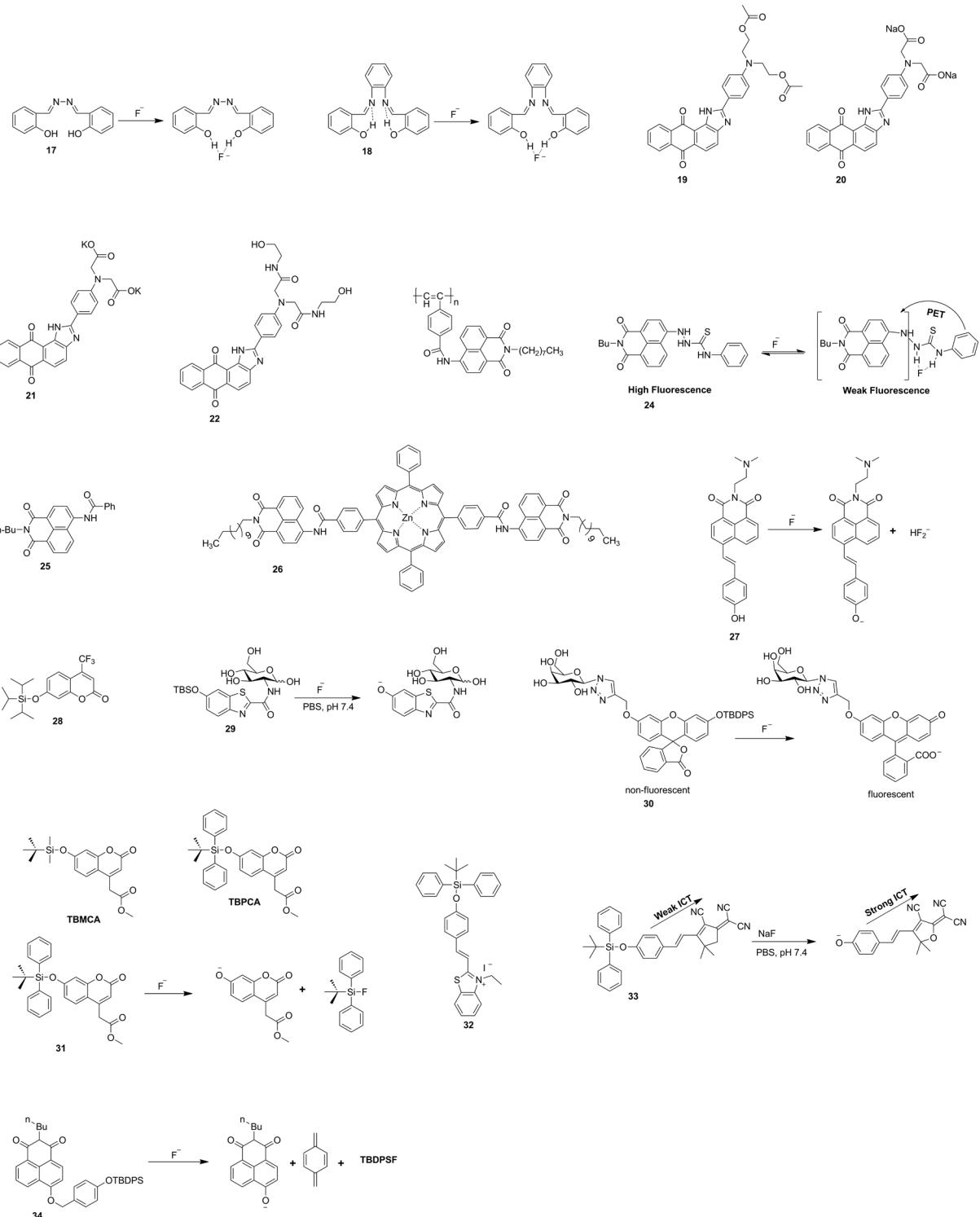


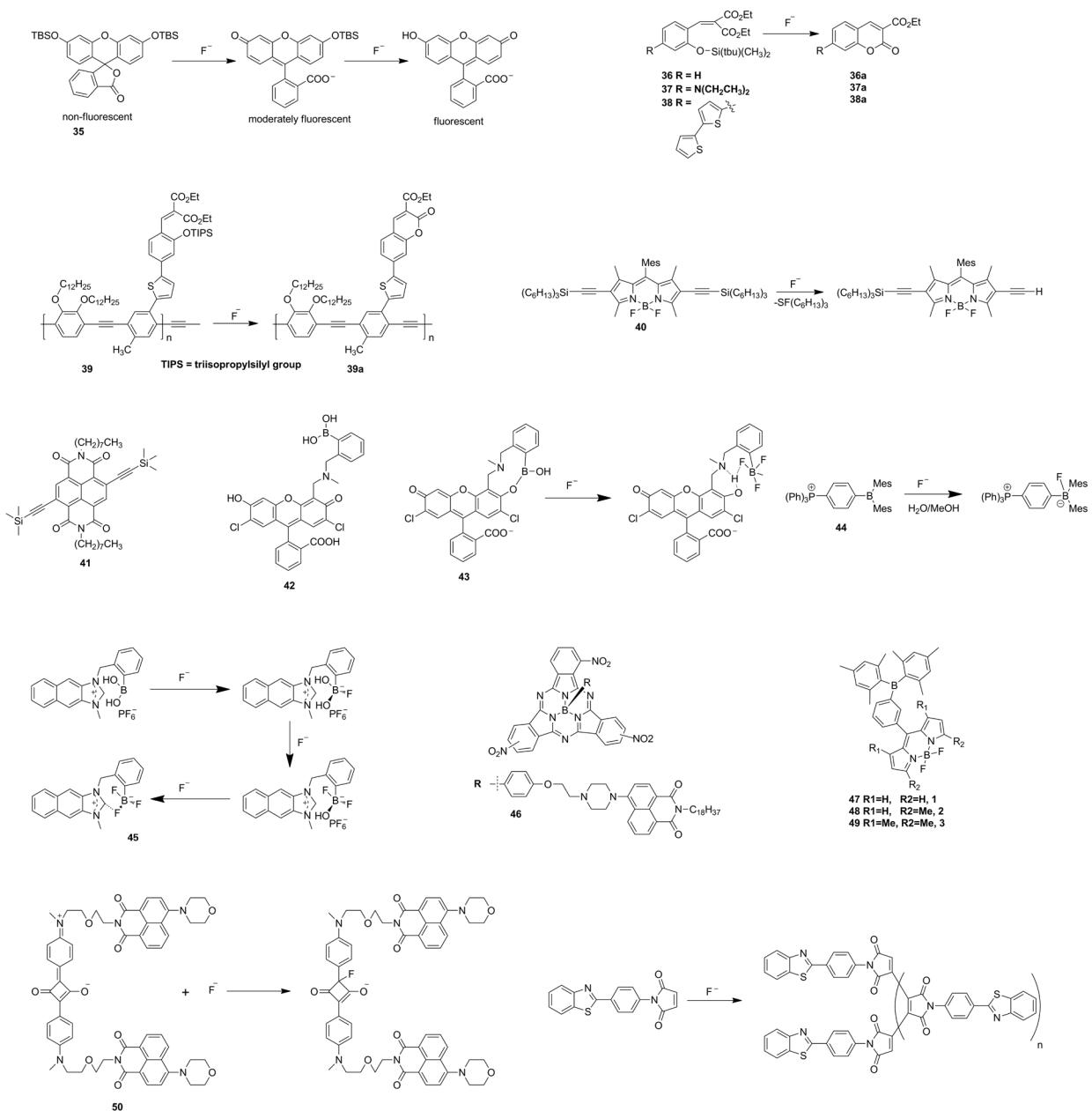
Supplementary materials



S Fig. 1

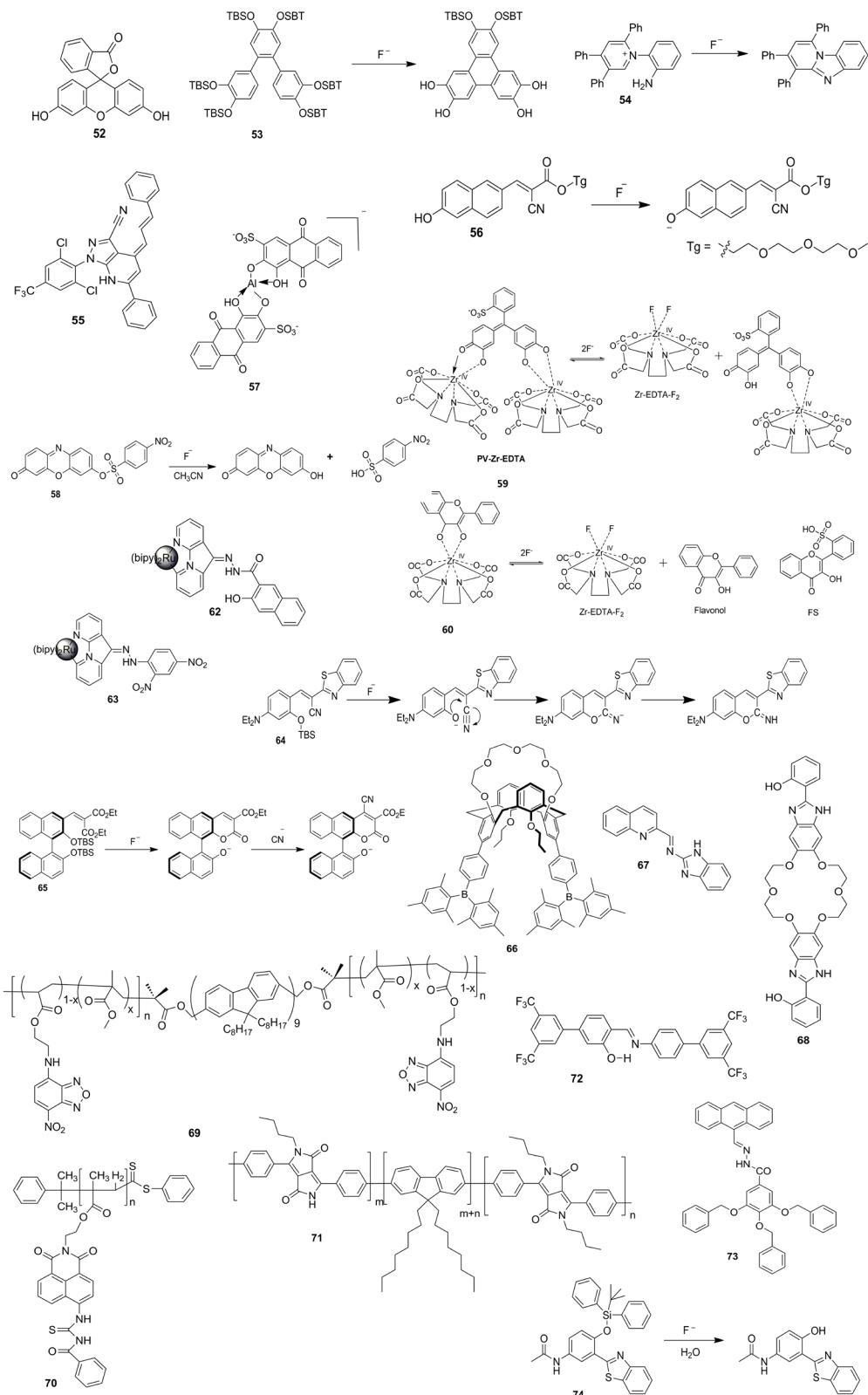


S Fig. 2



S Fig. 3

S Fig. 4



Supplementary Figure 1~4 Chemical formulas of the fluoride **Probe1~Probe 74** (except for **Probe 61**)

Supplementary Table 1 Comparison of three conventional analytical methods for F⁻

		Ion Chromatography	Ion Selective Electrode	Spectrophotometry
Principle	separating ions and polar molecules based on their charge differences	electrochemical reactions of the specific ions	measuring the absorbance or luminescence intensity of the light	
Detecting Range of Fluoride ($\mu\text{g/L}$)	$3.8 \sim 2.0 \times 10^3$	$0.041 \sim 1.9 \times 10^6$	$0 \sim 1.0 \times 10^6$	
Advantages	good sensitivity, wide linearity range, fast, simple, accurate, producible	most widely used, cheaper, fast, efficient, simple	stable, simple, fast, amenable to automation	
Disadvantages	weak binding between fluoride and ion exchangers	active surface of the electrode requires periodic regeneration, electrode drift and dissolution of the lanthanum fluoride membrane crystal	quantitative spectral overlap caused by the coexistence of spectral interference	
Detecting Targets	coffee, tea, spinach, fruits, alcoholic beverages, milk, mulberries, leaves, soil, toothpaste, and plaque biofilm	rain water, vegetables, fruit, tea, mulberries, medicinal materials, toothpaste, human blood, urine, pork, salt, air, cigarette smoke, and coal	mulberry leaves, toothpaste, minerals, ground water, and drinking water	
Ref.	Suppl [1-16]	Suppl [4, 16-26]	Suppl [4, 15, 27-32]	

Supplementary References

1. Heitkemper DT, Vela NP, Stewart KR, Westphal CS. Determination of total and speciated arsenic in rice by ion chromatography and inductively coupled plasma mass spectrometry. *Journal of Analytical Atomic Spectrometry*. 2001; 16: 299-306.
2. Hu ZY, Ye ML, Wu SC, Pan GW, Zhang TT, Liu L-Y. Determination of Impurity Anions in High Pure Fluoride Reagents by Ion Chromatography with Column-Switching. *Chinese Journal of Analytical Chemistry (Chinese Version)*. 2013; 40: 1703-8. doi:10.3724/sp.j.1096.2012.20213.
3. Jakubowski EM, McGuire JM, Evans RA, Edwards JL, Hulet SW, Benton BJ, et al. Quantitation of fluoride ion released sarin in red blood cell samples by gas chromatography-chemical ionization mass spectrometry using isotope dilution and large-volume injection. *Journal of Analytical Toxicology*. 2004; 28: 357-63.
4. Krishna MVB, Rao SV, Murthy VSN, Karunasagar D. A simple UV-photolysis digestion method for the determination of fluoride in fluorine-containing drugs by ion-selective electrode and spectrophotometry techniques. *Analytical Methods*. 2012; 4: 1565-72. doi:Doi 10.1039/C2ay05718b.
5. Lefler JE, Ivey MM. Ion chromatography detection of fluoride in calcium carbonate. *J Chromatogr Sci*. 2011; 49: 582-8.
6. Michalski R, Mathews B. Simultaneous determination of fluoride and monofluorophosphate in toothpastes by suppressed ion chromatography. *Central European Journal of Chemistry*. 2006; 4: 798-807. doi:10.2478/s11532-006-0042-0.
7. Müller ALH, Müller CC, Antes FG, Barin JS, Dressler VL, Flores EMM, et al. Determination of Bromide, Chloride, and Fluoride in Cigarette Tobacco by Ion Chromatography after Microwave-Induced Combustion. *Analytical Letters*. 2012; 45: 1004-15. doi:10.1080/00032719.2012.670800.
8. Neal M, Neal C, Wickham H, Harman S. Determination of bromide, chloride, fluoride, nitrate and sulphate by ion chromatography: comparisons of methodologies for rainfall, cloud water and river waters at the Plynlimon catchments of mid-Wales. *Hydrology and Earth System Sciences*. 2007; 11: 294-300.
9. Perring L, Bourqui B. An alternative method for fluoride determination by ion chromatography using post-column reaction. *American Laboratory*. 2002; 34: 28-+.
10. Pinkham DJ, Knapp MP. Determination of Soluble Fluoride and Monofluorophosphate in Dental Cream Using Gradient Elution Ion Chromatography. *Journal of Liquid Chromatography & Related Technologies*. 2000; 23: 1099-108. doi:10.1081/jlc-100101511.
11. Potter JJ, Hilliker AE, Breen GJ. Determination of fluoride and monofluorophosphate in toothpastes by ion chromatography. *J Chromatogr*. 1986; 367: 423-7. doi:10.1016/s0021-9673(00)94864-7.
12. Saha JK, Kundu S. Determination of Fluoride in Soil Water Extract Through Ion Chromatography. *Communications in Soil Science and Plant Analysis*. 2003; 34: 181-8. doi:10.1081/css-120017424.
13. Samanidou VF, Zacharis CK, Papadoyannis IN. Determination of Fluoride Ions by Single Column High Pressure Anion Chromatography in Dentifrice Preparations and Body Fluids: Saliva and Blood Serum. *Journal of Liquid Chromatography & Related Technologies*. 2002; 25: 803-18. doi:10.1081/jlc-120003037.
14. Sheen JF, Her GR. Analysis of neutral drugs in human plasma by fluoride attachment in liquid chromatography/negative ion electrospray tandem mass spectrometry. *Rapid Commun Mass Spectrom*. 2004; 18: 1911-8. doi:10.1002/rcm.1561.
15. Cassidy R, Elchuk S. Dynamically coated columns for the separation of metal ions and anions by ion chromatography. *Analytical Chemistry*. 1982; 54: 1558-63.
16. van den Hoop MAGT, Cleven RFMJ, van Staden JJ, Neele J. Analysis of fluoride in rain water comparison of capillary electrophoresis with ion chromatography and ion-selective electrode potentiometry. *Journal of Chromatography A*. 1996; 739: 241-8. doi:10.1016/0021-9673(96)00029-5.
17. Abudiak H, Robinson C, Duggal MS, Strafford S, Toumba KJ. Effect of fluoride sustained slow-releasing device on fluoride, phosphate and calcium levels in plaque biofilms over time measured using ion chromatography. *J Dent*. 2012; 40: 632-8. doi:10.1016/j.jdent.2012.04.008.
18. Giljanović J, Prkić A, Bralić M, Brkljača M. Determination of Fluoride Content in Tea Infusion by Using Fluoride Ion-Selective Electrode. *Int J Electrochem Sci*. 2012; 7: 2918-27.
19. Gotjamanos T, Orton V. Fluoride ion concentration in 40 per cent silver fluoride solutions determined by ion selective electrode and ion chromatography techniques. *Aust Dent J*. 1998; 43: 55-6. doi:10.1111/j.1834-7819.1998.tb00154.x.
20. Hara H, Kobayashi H, Maeda M, Ueno A, Kobayashi Y. Speciation of Aluminum in Rainwater Using a Fluoride Ion-Selective Electrode and Ion-Exchange Chromatography with Fluorometric Detection of the Aluminum-Lumogallion Complex. *Analytical Chemistry*. 2001; 73: 5590-5. doi:10.1021/ac010428w.
21. Rao MR, Mobin SM, Ravikanth M. Boron-dipyrromethene based specific chemodosimeter for fluoride ion. *Tetrahedron*. 2010; 66: 1728-34.
22. Srinivasan K, Rechnitz G. Reaction rate measurements with the fluoride ion selective membrane electrode. Mechanistic study of the iron (III)-iodide reaction in fluoride media. *Analytical Chemistry*. 1968; 40: 1955-9.
23. Srinivasan K, Rechnitz G. Reaction rate measurements with fluoride ion-selective membrane electrode. Formation kinetics of ferrous fluoride and aluminum fluoride complexes. *Analytical Chemistry*. 1968; 40: 1818-25.
24. Srinivasan K, Rechnitz GA. Activity measurements with a fluoride-selective membrane electrode. *Analytical Chemistry*. 1968; 40: 509-12.
25. Yuwono M. Determination of fluoride in black, green and herbal teas by ion-selective electrode using a standard-addition method. *Fluoride*. 2005; 52: 130-4.

26. Zhang J, Zhao SL, Zhang WC. [Evaluating the uncertainty in urinary fluoride measurement by ion selective electrode method]. Zhonghua Lao Dong Wei Sheng Zhi Ye Bing Za Zhi. 2011; 29: 867-8.
27. Yamaguchi H, Itoh S, Hasegawa R, Kobayashi T. Determination of trace silicon in high purity aluminium by molybdosilicic acid blue spectrophotometry after fluoride separation. Tetsu to Hagane-Journal of the Iron and Steel Institute of Japan. 2001; 87: 129-31.
28. Rocklin RD, Johnson EL. Determination of cyanide, sulfide, iodide, and bromide by ion chromatography with electrochemical detection. Analytical Chemistry. 1983; 55: 4-7.
29. Briska M, Hoffmeister W. Determination of copper in ammonium fluoride solutions by extraction and atomic-absorption spectrophotometry. Talanta. 1973; 20: 895-7.
30. Chen JT, Shen ST, Chung CS, Chang H, Wang SM, Li NC. Achatina fulica hemocyanin and its interactions with imidazole, potassium cyanide, and fluoride as studied by spectrophotometry and nuclear magnetic resonance and resonance raman spectrometry. Biochemistry. 1979; 18: 3097-101. doi:10.1021/bi00581a029.
31. Capitan-Vallvey LF, Valencia MC, Bosque Sendra JM. Microdetermination of fluoride in water by ion-exchange spectrophotometry. Analyst. 1988; 113: 419-22. doi:10.1039/an9881300419.
32. Yamaguchi H, Kiyokawa M, Hasegawa R. Determination of Trace Silicon in High Purity Titanium and Chromium by Molybdosilicic Acid Blue Spectrophotometry after Fluoride Separation. Bunseki Kagaku. 1995; 44: 647-50.

Supplementary Table 2 Spectroscopic and analytical parameters of fluorescent fluoride probes

Fluorescent probe	Quantum yield	Reaction medium	$\lambda_{\text{ex}}(\text{nm}) / \lambda_{\text{em}}(\text{nm})$	Liner detection range / (LOD) (μM)	Selectivity	Mechanism	Analytical applications	Ref.
1	0.76	DMSO	520 / 623	2.0-1.2×10 ² / 2.0	AcO ⁻ , H ₂ PO ₄ ⁻ , Cl ⁻ , Br ⁻ , I ⁻	ICT PET	toothpaste	[20]
2	0.41	CH ₃ CN	390 / 449	NM	Cl ⁻ , Br ⁻ , I ⁻ , HSO ₄ ⁻ , NO ₃ ⁻ , PF ₆ ⁻ , ClO ₄ ⁻ , AcO ⁻ , H ₂ PO ₄ ⁻	ICT	NM	[25]
3	NM	THF/H ₂ O (50/50, v/v)	335 / 470-480	NM	Cl ⁻ , Br ⁻ , I ⁻ , CO ₃ ²⁻ , NO ₂ ⁻ , NO ₃ ⁻ , SCN ⁻ , ClO ₄ ⁻ , SO ₄ ²⁻ , HPO ₄ ²⁻ , H ₂ PO ₄ ⁻	NM	NM	[27]
4	NM	THF/ H ₂ O (50/50, v/v)	335 / 470-480	NM	Cl ⁻ , Br ⁻ , I ⁻ , CO ₃ ²⁻ , NO ₃ ⁻ , NO ₂ ⁻ , SCN ⁻ , ClO ₄ ⁻ , SO ₄ ²⁻ , HPO ₄ ²⁻ , H ₂ PO ₄ ⁻	NM	HeLa cells	[27]
5	0.29	CH ₂ Cl ₂	425 / 584,564	NM	Br ⁻ , Cl ⁻ , I ⁻ , HPO ₄ ²⁻ , ClO ₄ ⁻	NM	NM	[30]
6	0.65	CH ₂ Cl ₂	430 / 602	NM / 2.0×10 ⁻¹	Br ⁻ , Cl ⁻ , I ⁻ , H ₂ PO ₄ ⁻ , CH ₃ COO ⁻ , NO ₃ ⁻ , ClO ₄ ⁻	PET	NM	[31]
7	0.55	THF/H ₂ O (1/9,v/v)	NM / 520	NM / 18.0	AcO ⁻ , NO ₂ ⁻ , NO ₃ ⁻ , SO ₃ ²⁻ , CO ₃ ²⁻ , SO ₄ ²⁻ , PO ₄ ³⁻ , Cl ⁻ , IO ₃ ⁻ , ClO ₃ ⁻ , Br ⁻ , HPO ₄ ²⁻ , HSO ₃ ⁻ , S ²⁻ , S ₂ O ₃ ²⁻ , I ⁻ , SCN ⁻ , CN ⁻	NM	HeLa cells	[34]
8	NM	CH ₃ CN/H ₂ O (50/50, v/v)	550 / 585	NM	Cl ⁻ , Br ⁻ , I ⁻ , HSO ₄ ⁻ , NO ⁻ , N ⁻	NM	NM	[37]
9	0.86	THF	365 / 402	NM / 0-15.0 ppm (unimers), 0-8.0 ppm(micelles)	Cl ⁻ , Br ⁻ , I ⁻ , CO ₃ ²⁻ , NO ₃ ⁻ , HSO ₄ ⁻ , HPO ₄ ²⁻ , H ₂ PO ₄ ⁻ , AcO ⁻ ,	NM	aqueous media	[38]
10	0.98	THF/water (7/3, v/v)	NM	NM / 1.0×10 ⁻¹	Cl ⁻ , Br ⁻ , I ⁻ , SO ²⁻ , SCN ⁻ , PO ₃ ⁻	NM	water samples	[40]
11	0.43	DMSO/H ₂ O (95/5, v/v)	645 / 718	NM / 8.5×10 ⁻²	Cl ⁻ , Br ⁻ , I ⁻ , SO ²⁻ , NO ⁻ , N ⁻ , AcO ⁻ , HPO ₄ ²⁻	NM	NM	[42]
12	0.12	DCM	555 / 738	NM	Cl ⁻ , Br ⁻ , I ⁻	NM	NM	[43]
13	NM	DMSO	NM / 675	2.0-4.0 / 2.59×10 ⁻¹	CH ₃ COO ⁻ , C ₆ H ₅ COO ⁻ , H ₂ PO ₄ ⁻ , Cl ⁻ , Br ⁻ , I ⁻ , HSO ₄ ⁻ , ClO ₄ ⁻ , BF ₄ ⁻ , PF ₆ ⁻	NM	NM	[44]
14	NM	CH ₂ Cl ₂	644 / 676	NM / 1.18×10 ⁻¹	Cl ⁻ , Br ⁻ , I ⁻ , HSO ₄ ⁻ , HPO ₄ ²⁻ , H ₂ PO ₄ ⁻ , N ₃ ⁻ , NO ₃ ⁻	NM	NM	[45]
15	0.27	THF	800 / 534, 594,645	NM	Cl ⁻ , Br ⁻ , I ⁻	NM	NM	[46]
16	NM	PBS buffer solution	660 / 690	0.10-1.0 / 7.4×10 ⁻²	C ₂ O ₄ ²⁻ , NO ₂ ⁻ , NO ₃ ⁻ , I ⁻ , HCO ₃ ⁻ , Br ⁻ , CO ₃ ²⁻ , Cl ⁻ , CN ⁻ , SO ₄ ²⁻	PET	HepG2 cells	[47]
17	0.85	DMSO	396 / 520	NM	Cl ⁻ , Br ⁻ , I ⁻ , AcO ⁻ , ClO ₄ ⁻ , HSO ₄ ⁻ , H ₂ PO ₄ ⁻	NM	NM	[49]
18	0.82	DMSO	390 /	NM	NM	NM	NM	[49]

			420					
19	0.8	CH ₃ CN/DMSO (5/95, v/v)	360 / 430	NM	Cl ⁻ , Br ⁻ , I ⁻ , AcO ⁻ , H ₂ PO ₄ ⁻ , NO ₃ ⁻ , ClO ₄ ⁻ , HSO ₄ ⁻	ICT	aqueous medium	[51]
20	0.9	CH ₃ CN/DMSO (5/95, v/v)	360 / 430	NM	Cl ⁻ , Br ⁻ , I ⁻ , AcO ⁻ , H ₂ PO ₄ ⁻ , NO ₃ ⁻ , CN ⁻ , ClO ₄ ⁻ , HSO ₄ ⁻	ICT	aqueous medium	[50]
21	0.8	CH ₃ CN/DMSO (5/95, v/v)	360 / 430	NM	Cl ⁻ , Br ⁻ , I ⁻ , AcO ⁻ , H ₂ PO ₄ ⁻ , NO ₃ ⁻ , CN ⁻ , ClO ₄ ⁻ , HSO ₄ ⁻	ICT	aqueous medium	[50]
22	0.8	CH ₃ CN/DMSO (5/95, v/v)	360 / 430	NM	Cl ⁻ , Br ⁻ , I ⁻ , AcO ⁻ , H ₂ PO ₄ ⁻ , NO ₃ ⁻ , ClO ₄ ⁻ , HSO ₄ ⁻	ICT	aqueous medium	[50]
23	0.89	CH ₃ CN	360 / 460	NM / 10.0	Cl ⁻ , Br ⁻ , I ⁻	ICT	NM	[51]
24		CH ₃ CN	390 / 450,650	NM	Cl ⁻ , Br ⁻ , I ⁻	ICT, PET	NM	[52]
25	0.75	CH ₃ CN	340 / 468,583	2.0×10 ⁴ -1.0×10 ⁵ / NM	Cl ⁻ , Br ⁻ , I ⁻	ICT	NM	[53]
26	0.037→0.026	THF	365 / 608 (655)	NM	Cl ⁻ , Br ⁻ , I ⁻	ICT, PET, FRET	NM	[54]
27	NM	DMSO	490 / 582	NM	Cl ⁻ , Br ⁻ , I ⁻ , NO ₃ ⁻ , H ₂ PO ₄ ⁻ , AcO ⁻ , SO ₄ ²⁻ , BF ₄ ⁻ , CN ⁻ , OH ⁻	ICT	RAW 264.7 cells, 4T1 cells, HeLa cells	[56]
28	0.67	CH ₃ CN	410 / 500	NM	Cl ⁻ , Br ⁻ , I ⁻ , CN ⁻ , AcO ⁻ , NO ₃ ⁻ , PhCO ₂ ⁻ , SCN ⁻ , H ₂ PO ₄ ⁻ , HSO ₄ ⁻	NM	aqueous medium	[57]
29	NM	DMSO/PBS (1/199, v/v)	380 / 508	1.0×10 ² -1.0×10 ³ / NM	Cl ⁻ , Br ⁻ , I ⁻ , H ₂ PO ₄ ⁻ , N ₃ ⁻ , NO ₂ ⁻ , NO ₃ ⁻ , AcO ⁻ , SO ₄ ²⁻	NM	KB cells	[58]
30	0.86	CTAB/PBS (1/10, v/v)	470 / 520	NM / 10.5	Cl ⁻ , H ₂ PO ₄ ⁻ , I ⁻ , SO ₄ ²⁻ , NO ₃ ⁻ , CO ₃ ²⁻ , N ₃ ⁻ , HPO ₄ ²⁻ , CN ⁻ , HCO ₃ ⁻ , AcO ⁻ , Br ⁻	NM	water samples, HepG2 cells	[59]
31	NM	HEPES	365 / 461	NM / 1.86×10 ⁻⁵	Cl ⁻ , Br ⁻ , I ⁻ , AcO ⁻ , NO ₃ ⁻ , N ₃ ⁻ , H ₂ PO ₄ ⁻	ICT	A549 cells	[60]
32	NM	C ₂ H ₅ OH/H ₂ O (7/3, v/v)	540-580 / 470-510	5.0×10 ² -2.8×10 ⁴ / 80.0	CO ₃ ²⁻ , SO ₄ ²⁻ , SCN ⁻ , NO ₃ ⁻ , N ₃ ⁻ , Cl ⁻ , Br ⁻ , I ⁻ , Cys, GSH, BSA, HSA	ICT	RAW 264.7 cells	[61]
33	0.58	C ₂ H ₅ OH/H ₂ O (7/3, v/v)	560 / 612	0-6.0×10 ³ / 70.0	Cl ⁻ , Br ⁻ , I ⁻ , SCN ⁻ , N ₃ ⁻ , NO ₃ ⁻ , HCO ₃ ⁻	ICT	HeLa cells	[62]
34	NM	CH ₃ CN/H ₂ O (50/50, v/v)	475 / 560	0-7.2×10 ² / 0.35 mg L ⁻¹	Cl ⁻ , Br ⁻ , I ⁻ , CN ⁻ , AcO ⁻ , H ₂ PO ₄ ⁻ , NO ₃ ⁻ , SO ₄ ²⁻	NM	A549 cells, toothpaste	[63]
35	0.676	DMF/H ₂ O (7/3, v/v)	480 / 532	0.1-2.0 / 4.1×10 ⁻²	Cl ⁻ , Br ⁻ , I ⁻	NM	toothpaste	[64]
36	0.0001	NM	322 / NM	NM	NM	FRET	NM	[65]
37	0.002	THF	386 / 450	NM	NM	FRET	NM	[65]
38	0.008	NM	384 / 489	NM	NM	FRET	NM	[65]
39	0.12	THF	378 / 482	NM	NM	FRET	NM	[65]
40	0.71	C ₃ H ₆ O	400 / 555	NM / 6.74×10 ⁻²	Cl ⁻ , Br ⁻ , I ⁻ , ClO ₄ ⁻ , H ₂ PO ₄ ⁻ , CH ₃ COO ⁻ , SCN ⁻ , CO ₃ ²⁻ , NO ₃ ⁻	NM	NM	[66]

					SO_4^{2-}			
41	0.98	CH_2Cl_2	289 / 455	NM	Cl^- , Br^- , HSO_4^- , AcO^- , H_2PO_4^-	NM	NM	[67]
42	0.63	$\text{C}_2\text{H}_3\text{N}/\text{CH}_3\text{OH}$ (9/1, v/v)	483 / 522	NM	Cl^- , Br^- , I^-	PET	NM	[33]
43	0.6	THF	324 / 531	NM	NM	NM	NM	[68]
44	0.63	$\text{H}_2\text{O}/\text{CH}_3\text{OH}$ (9/1, v/v)	NM	NM	Cl^- , Br^- , I^- , NO_3^- , HPO_4^{2-} , HSO_4^-	NM	NM	[69]
45	NM	CH_3CN	323 / 440	NM	Cl^- , Br^- , CH_3COO^- , HSO_4^- , H_2PO_4^-	NM	aqueous medium	[70]
46	NM	THF	390 / 495, 500 / 578/	NM	Cl^- , Br^- , I^- , ClO_4^-	NM	NM	[71]
47	0.48	DCM	350 / 408, 519	NM / 1.2 ppm	Cl^- , Br^- , I^- , ClO_4^- , H_2PO_4^- , HSO_4^- , NO_3^- , PF_6^- , CN^-	FRET	NM	[72]
48	0.55	DCM	350 / 405, 450, 491, 526 408, 515	NM / 2.3 ppm	Cl^- , Br^- , I^- , ClO_4^- , H_2PO_4^- , HSO_4^- , NO_3^- , PF_6^- , CN^-	FRET	NM	[72]
49	0.72	DCM	350 / 408, 519	NM / 0.2 ppm	Cl^- , Br^- , I^- , ClO_4^- , H_2PO_4^- , HSO_4^- , NO_3^- , PF_6^- , CN^-	FRET	NM	[72]
50	0.059	CH_2Cl_2	402 / 659	NM / 2.0×10^2	Br^- , I^- , OH^- , PO_4^{3-} , SCN^- , CH_3COO^- , ClO_4^- , NO_3^- , $\text{C}_2\text{O}_4^{2-}$, SO_4^{2-} , S^{2-} , CN^-	FRET, PET	NM	[73]
51	0.89	DMSO	365 / 412	NM / 5.0×10^4	CN^- , Cl^- , OH^- , Br^- , I^- , PF_6^- , BF_4^- , HSO_4^- , OAc^-	NM	NM	[74]
52	0.61	CH_3CN	480 / 532	NM	Cl^- , Br^- , I^- , HSO_4^-	NM	NM	[75]
53	0.86	THF	294 / 393	NM / 2.0	Cl^- , Br^- , I^- , CN^- , HSO_4^- , H_2PO_4^- , NO_3^- , OAc^-	NM	NM	[76]
54	0.46	CH_3CN	366 / 469	NM / 2.72	Cl^- , Br^- , I^- , NO_3^- , H_2PO_4^- , AcO^-	NM	NM	[77]
55	0.15	CH_2Cl_2	324 / 400	NM / 8.54×10^{-3}	Cl^- , Br^- , I^- , NO_3^- , HSO_4^-	NM	NM	[78]
56	0.94	PBS buffer	388 / 490	NM	NO_3^- , Br^- , Cl^- , HSO_4^- , CO_3^{2-} , S^{2-} , I^- , N_3^- , PO_4^{3-} , SCN^- , CH_3COO^-	NM	PC3 cells	[79]
57	NM	$\text{C}_2\text{H}_5\text{OH}/\text{H}_2\text{O}$ (1/5, v/v)	247 / 532	6.0×10^5 - 8.0×10^7 / 6.0×10^5	Cl^- , SO_4^{2-} , NO_3^- , PO_4^{3-} , I^- , Br^-	NM	NM	[80]
58	0.58	CH_3CN	485 / 550, 591	NM / 1.9	Cl^- , Br^- , I^- , AcO^- , NO_3^- , N_3^- , ClO_4^- , HSO_4^-	NM	NM	[81]
59	NM	aqueous solution		15.0 - 1.5×10^2 / 4.5234×10^2	SO_4^{2-} , SO_3^{2-} , CO_3^{2-} , Cl^- , NO_3^- , NO_2^- , PO_4^{3-}	NM	NM	[83]
60	NM	aqueous	377 /	0.5 - 1.0×10^3 /	Cl^- , Br^- , NO_3^- ,	NM	toothpaste,	[84]

		solution	460	NM	CH_3COO^- , SO_4^{2-} , H_2PO_4^-		water samples	
61	NM	HAc/NaAc buffer solution	NM / 562.4, 519.9	0.10-10.0 / 0.031	Cl^- , I^- , Br^- , NO_3^- , H_2PO_4^- , HCO_3^- , HPO_4^{2-}	NM	toothpaste, tap water samples	[85]
62	NM	CH_3CN	355 / 530	NM	Cl^- , I^- , Br^- , HSO_4^- , NO_3^- , H_2PO_4^-	PET, ESIPT	NM	[86]
63	0.6	CH_3CN	465 / 610	NM	HSO_4^- , NO_3^- , Cl^- , Br^- , I^-	PET	aqueous medium	[87]
64	0.0008 →0.0709	$\text{C}_2\text{H}_5\text{OH}/\text{H}_2\text{O}$ (1/1, v/v)	460 / 523	NM / 1.177×10^{-1}	Cl^- , Br^- , I^- , AcO^- , ClO_4^- , CF_3SO_3^- , NO_3^- , HSO_4^- , H_2PO_4^- , BF_4^- , N_3^- , SCN^- , OH^-	ACRR	NM	[88]
65	0.009	THF	330 / 480	5.0-45.0 / 1.86	Cl^- , Br^- , I^- , CN^- , AcO^- , ClO_4^- , CF_3SO_3^- , NO_3^- , HSO_4^- , H_2PO_4^- , BF_4^- , N_3^- , SCN^- , HSCH_2COOH	ARR	NM	[89]
66	0.70	CH_2Cl_2	342 / 430	NM	Cl^- , Br^- , I^- , H_2PO_4^- , AcO^-	NM	NM	[90]
67	0.43	$\text{CH}_3\text{CN}/\text{H}_2\text{O}$ (1/4, v/v)	405 / 472	NM	Cl^- , Br^- , I^- , CN^- , H_2PO_4^- , NO_3^- , NO_2^- , SO_4^{2-} , HSO_3^- , S^{2-}	NM	HeLa cells	[92]
68	0.62→0.57	CH_3CN	330 / 455	NM / 4.0×10^{-2}	H_2PO_4^- , AcO^- , NO_3^- , Cl^- , Br^- , I^-	ESIPT	HeLa cells	[93]
69	0.83	$\text{C}_3\text{H}_6\text{O}$	390 / 515	NM / 4.78	Cl^- , Br^- , I^- , CH_3COO^- , ClO_4^- , HSO_4^- , H_2PO_4^-	FRET	NM	[95]
70	0.93	$\text{CH}_2\text{Cl}_2/\text{DMSO}$ (9/1, v/v)	340 / 468/, 490 / 580	NM	Cl^- , Br^- , I^-	ICT	NM	[99]
71	0.98	THF	532 / 570	1.5 $\mu\text{g}/\text{mL}$ -15.0 $\mu\text{g}/\text{mL}$ / NM	AcO^- , H_2PO_4^-	FRET, ICT	KB cells	[102]
72	0.82	THF	405 / 588	NM	Cl^- , Br^- , I^- , NO_3^- , H_2PO_4^- , AcO^-	ESIPT	NM	[107]
73	NM	THF	470 / 490→ 505	NM	Cl^- , Br^- , I^- , ClO_4^- , CH_3COO^- , H_2PO_4^- , HSO_4^-	NM	NM	[108]
74	0.52	aqueous solution	360 / 468	NM	anions (sodium salts) and cations (nitrate salts)	ESIPT	water samples	[109]

ACRR=anion to cation relay recognition

ARR=anion relay recognition

A549 cells=A549 human lung carcinoma cell lines

BSA=bovine serum albumin

CTAB=cetyl trimethyl ammonium bromide

Cys=cysteine

DCM=Dichloromethane

DMF=N,N-Dimethylformamide

DMSO=(dimethyl sulfoxide

FRET=fluorescence resonance energy transfer

GSH=glutathione
HeLa cells =human cervical cancer cell line cells
HepG2 cells=Human hepatocellular liver carcinoma cells
HAS=human serum albumin
ICT=internal charge transfer
LOD=limit of detection
NM=not mentioned
PBS=phosphate buffered saline
PET=photoinduced electron transfer
RAW 264.7 cells =mouse monocyte macrophage cell line cells
TBA=tetrabutylammonium
THF=Tetrahydrofuran
4T1 cells =mouse breast cancer cell line cells