

Supporting Information

Targeted Mesoporous Iron Oxide Nanoparticles-Encapsulated Perfluorohexane and a Hydrophobic Drug for Deep Tumor Penetration and Therapy

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Synthesis of Mesoporous Iron Oxide Nanoparticles (MIONs) without oleic acids.

In brief, 10 mmol of iron(III) chloride hexahydrate ($\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$, Acros) and 20 mmol of sodium acetate anhydrous (NaAc, Aldrich) were dissolved in 10 ml of ethylene glycol (EG, J. T. Baker) to form a clear solution. Then, the mixture was mixed at 60 °C for 24 hours to give a homogeneous solution. The mixture was replaced to a Teflon-lined stainless steel autoclave for the hydrothermal reactions at 220 °C for 8 h. After the reaction, the autoclave was cooled down to room temperature for another 3 h. The magnetic particles were collected by centrifugation at 7,000 rpm for 12 min and washed with excess ethanol 3 times afterwards. Before characterization and application, the particles were dried under vacuum overnight at room temperature. The morphologies of MIONs were characterized by a field emission scanning microscope (FE-SEM, JEOL-6700, Japan) as shown in **Figure S1**.

Magnetic Property and Stability of Lf-MIONs. The magnetization of dried particles was exhibited by magnetization (M)-field (H) curves, where the saturation at a field that is smaller than that observed in MION (**Fig. 2h**). The result shows a stronger inter-particle interaction for the MIONs. The lower magnetization of OA-MIONs and Lf-MIONs is due to the concentration dilution by oleic acid or lactoferrin. Based on the magnetic particle, OA-MIONs and Lf-MIONs have the identical magnetization property as that of MIONs. These properties indicate that the MION respond to the magnetic field in a similar way as OA-MIONs and Lf-MIONs but with a higher magnetic susceptibility. Furthermore, the stability of Lf-MIONs during a 4 °C storage was investigated in solution. As shown in the **Figure S2**, the magnetic property of Lf-MIONs after 3 weeks was compared with that of the original, exhibiting only slight difference of less than 4 %.

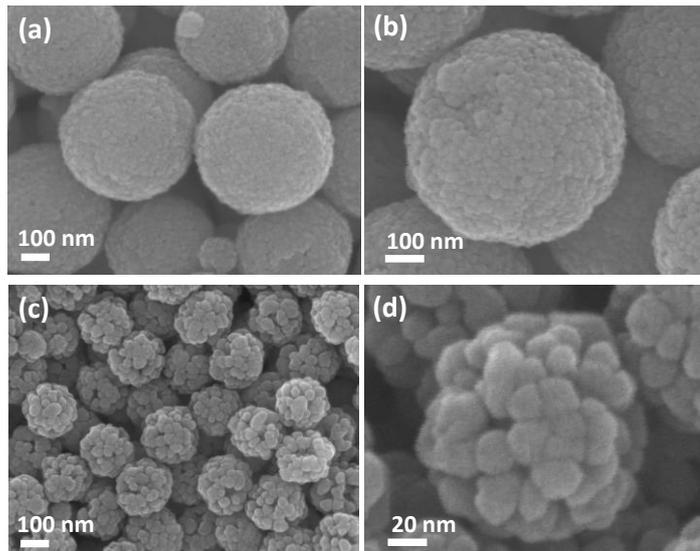


Figure S1 SEM images of resulting MIONs prepared by (a, b) without OA and (c, d) with OA in the synthesized processes.

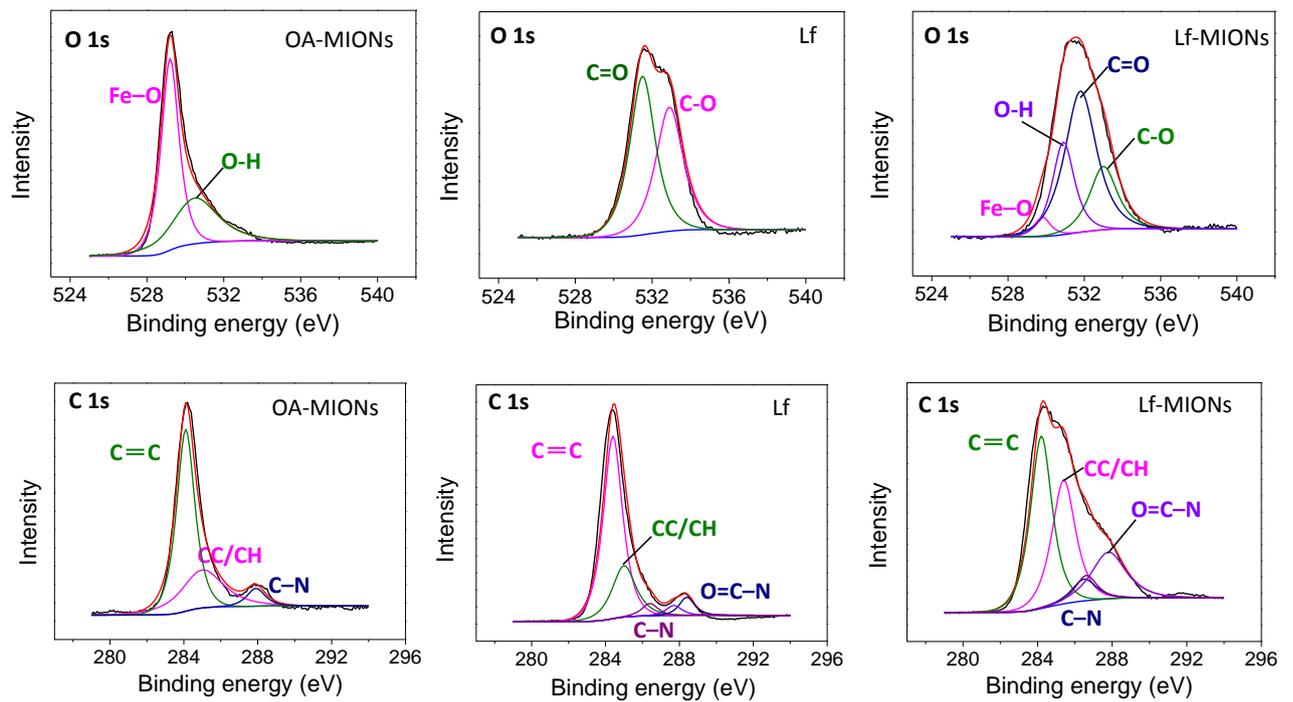


Figure S2 XPS survey scan spectra of OA-MIONs, lactoferrin (Lf), and Lf-MIONs.

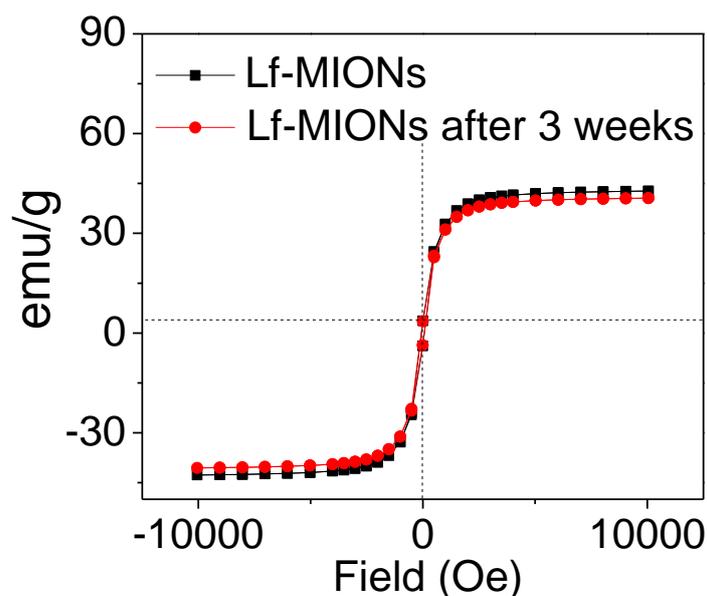


Figure S3 Field-dependent magnetization curve of Lf-MION after storage at 4 °C for 3 weeks. There was no significant change in the physical characteristic.

Table S1 The pore structure parameters before and after PTX loading to OA-MIONs.

Sample	Surface Area (m ² /g)	Pore Volume (cm ³ /g)	Pore Size (nm)
OA-MIONs	110.6	0.76	18.4
OA-MIONs/PTX	24.2	0.15	15.5

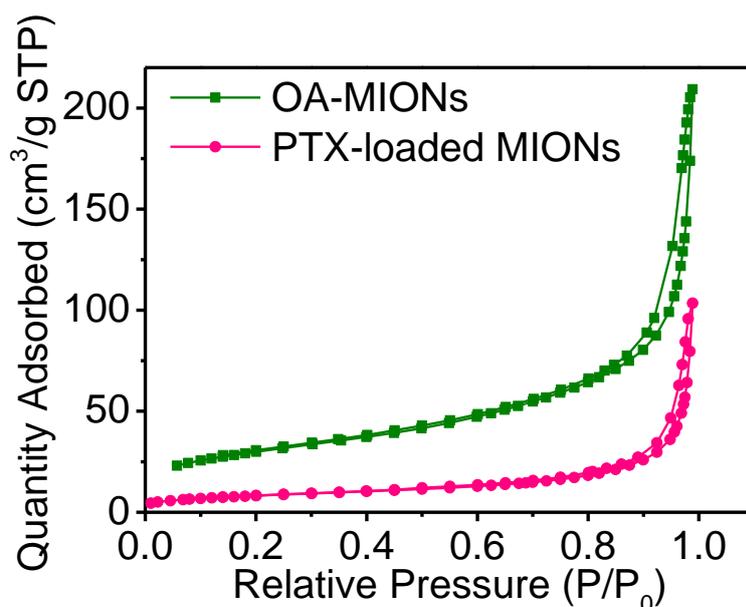


Figure S4 N₂ adsorption–desorption isotherms of OA-MIONs before and after PTX loading.

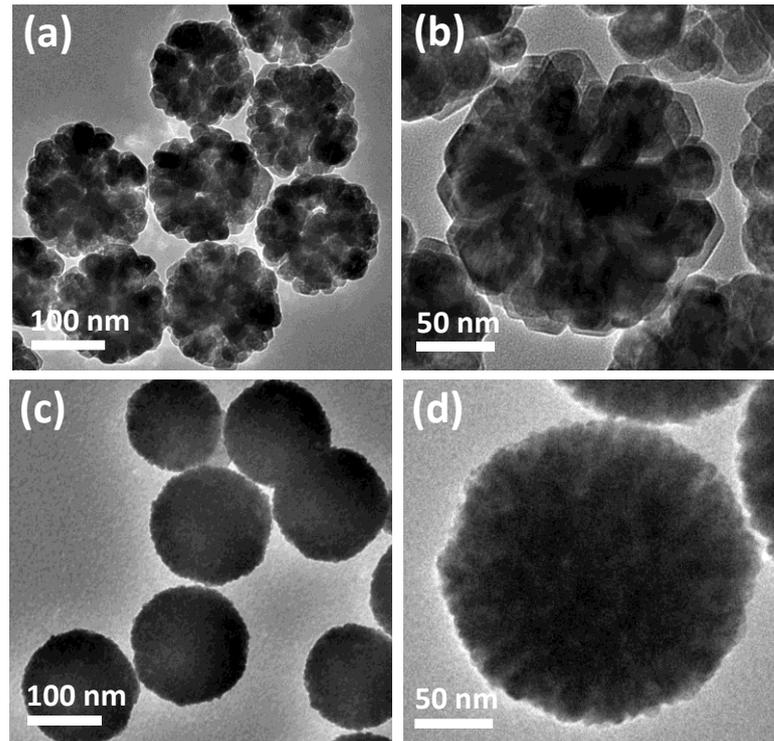


Figure S5 TEM images of OA-MIONs (a, b) before and (c, d) after PTX loading.

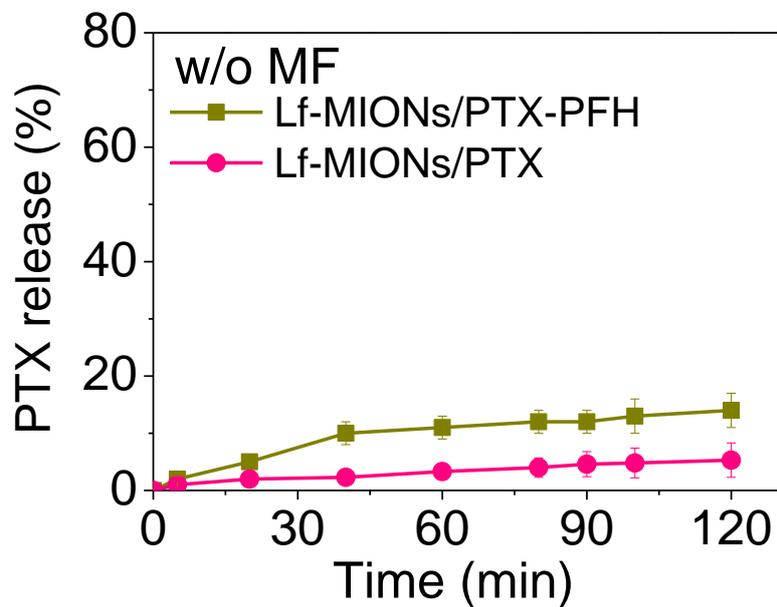


Figure S6 Cumulative PTX release from Lf-MIONs/PTX-PFH and Lf-MIONs/PTX without MF treatment.

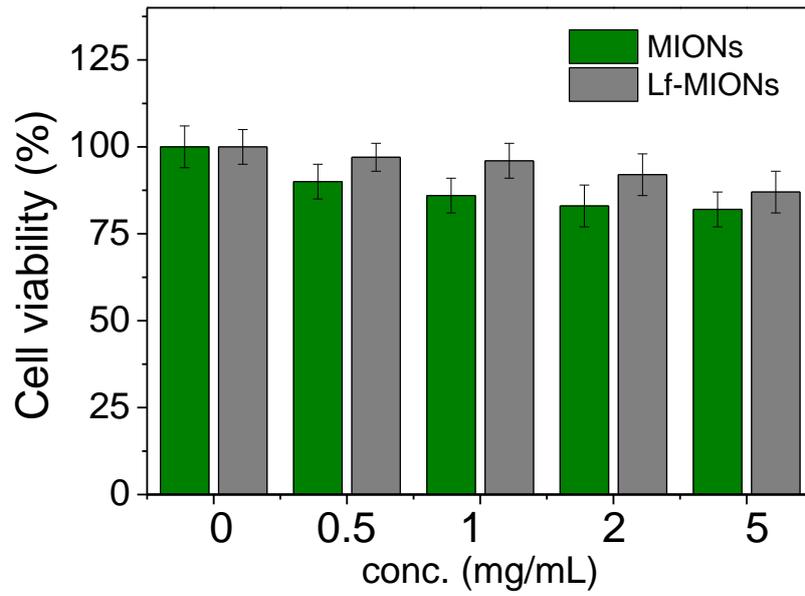


Figure S7 Cell viability of RG2 cells after 24 h incubation with various concentrations of MIONs and Lf-MIONs.

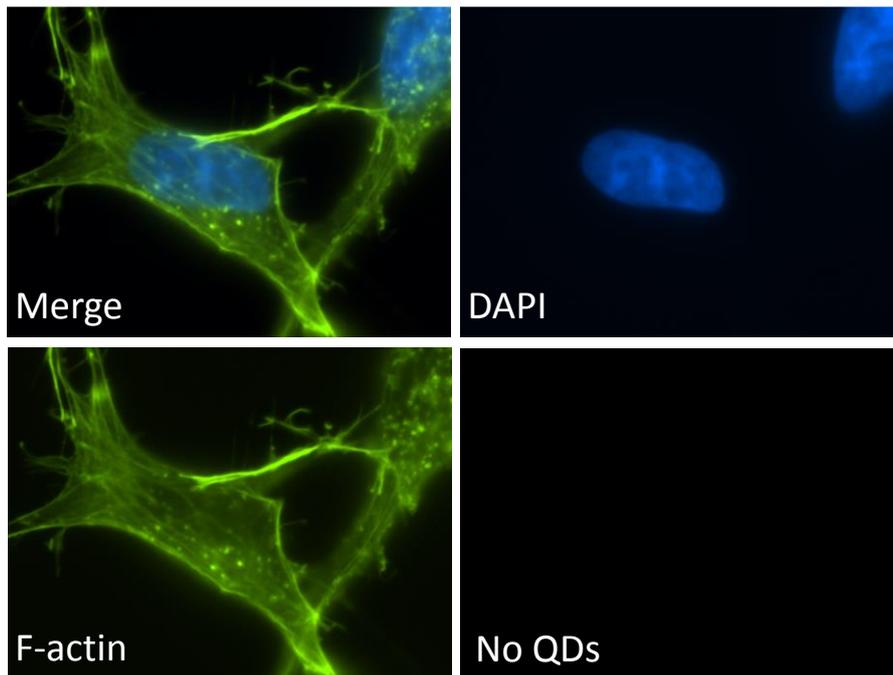


Figure S8 Control experiment on RG2 cells without any particles.

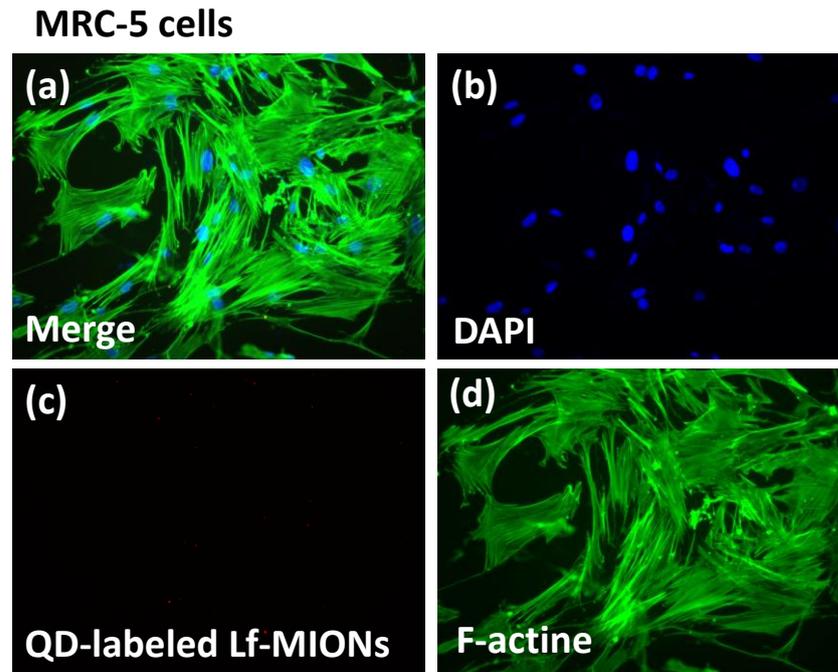


Figure S9 Cellular uptake of quantum dot (QD)-loaded Lf-MIONs in MRC-5 (a human lung fibroblast, normal cell) cells after 2 h of incubation.

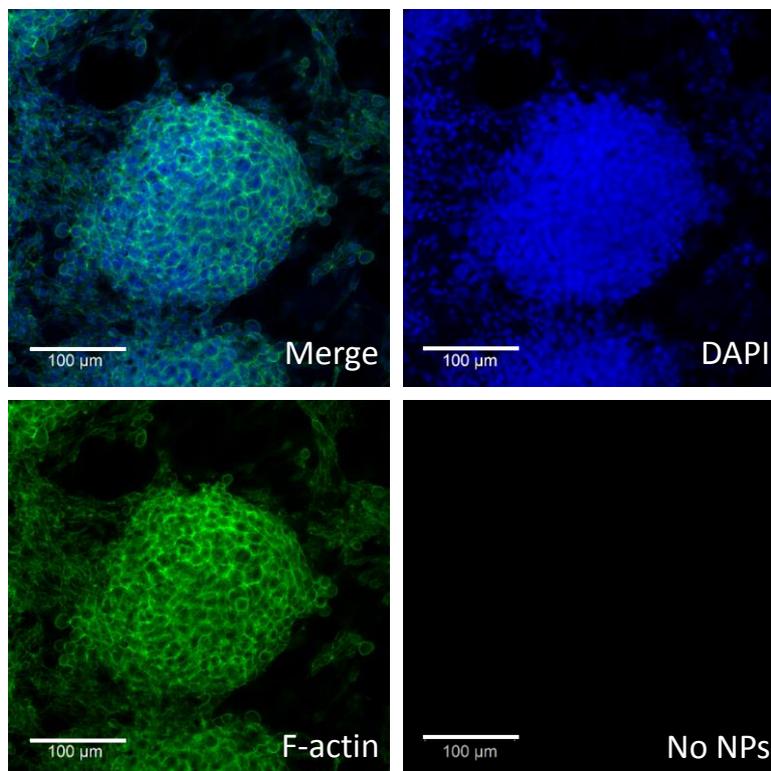


Figure S10 Control experiment on RG2 cells without any particles.

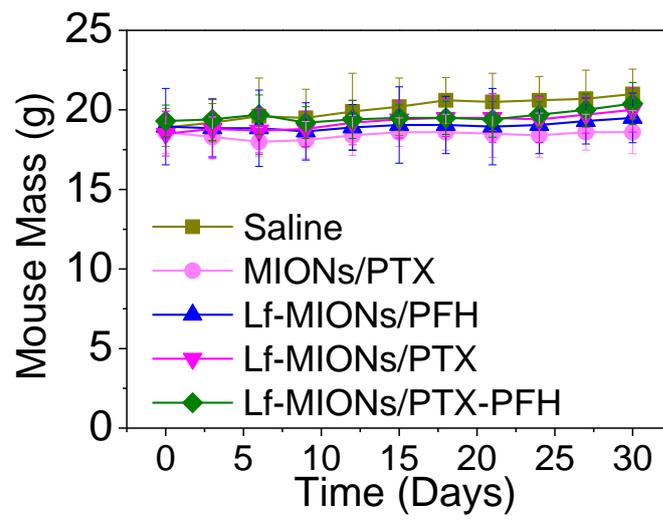


Figure S11 Mouse mass after treating by saline, MIONs/PTX, Lf-MIONs/PTX, and Lf-MIONs/PTX-PFH for 30 days.