Supplementary Information: ¹H-NMR characterization of MA-Chol, degradability assessment of MA-Chol in 10% FBS condition, siRNA encapsulation efficiency of SLNPs, serum stability of siRNA@SLNPs, time-course colloidal stability of siRNA@SLNPs in 10% FBS condition, comparison of siRNA encapsulation efficiency in SLNPs and DC-Chol/DOPE liposomes, intracellular uptake of TAMRA-siRNA@SLNPs by U87MG cells, tumor weights and changes in body weights in response to *in vivo* treatment with siRNA@SLNPs, and hemolysis assay.

Supplementary Information

Mono-arginine Cholesterol-based Small Lipid Nanoparticles as a Systemic siRNA Delivery Platform for Effective Cancer Therapy

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Figure S1. ¹H-NMR characterization of MA-Chol in DMSO-d⁶.



Figure S2. Evaluation of degradability of MA-Chol in 10% FBS. MA-Chol (10 mg/mL) was incubated in PBS (pH 7.4) containing 10% FBS at 37°C and aliquots were collected at each time points. Each aliquot (100 μ L) was mixed with 400 μ L of chloroform to extract organic compounds. The collecting chloroform layer was evaporated to concentrate the solvent. The remaining crude concentrate was developed in chloroform : methanol = 15 : 1 (v/v) by thin layer chromatography (TLC) and further stained with p-anisaldehyde solution for detection.



Figure S3. siRNA encapsulation efficiency of SLNPs. Encapsulation efficiency of non-modified cholesterol/DOPE liposomes and MA-Chol/DOPE (1:1 and 1:2) liposomes with siRNA were analyzed using OliGreen (Invitrogen) according to manufacturer's protocol. Total 4 μ mol lipid of Chol/DOPE or MA-Chol/DOPE liposomes were rehydrated with 3 nmol siRNA and the resulting liposome solutions were loaded onto Sepharose CL-4B column. (a) Any siRNA was encapsulated in Chol/DOPE liposomes. (b) MA-Chol/DOPE (1:1) and (c) MA-Chol/DOPE (1:2) liposomes revealed 100% of siRNA encapsulation efficiency.



Figure S4. Serum stability of siRNA@SLNPs. Equivalent amount of siRNA and siRNA@SLNPs were incubated with 30% fetal bovine serum (FBS)-conditioning buffer at 37°C for 0.5, 1, 3, 6, 12, and 24 h. After indicated incubation times, aliquots containing 1 μ g siRNA of each sample were visualized by 12% agarose gel electrophoresis. While naked siRNA showed rapid degradation within 3 h, siRNA encapsulated within SLNPs remained 75% intact for up to 6 h and 40% of siRNA was stable for up to 24 h.

Table S1. Colloidal stability of siRNA@SLNPs in 10% FBS condition. The integrity of siRNA@SLNPs upon incubation in PBS containing 10% FBS at 37°C was evaluated with respect to size and zeta potential at predetermined time points (0, 1, 3, 6, 12, and 24 h).

Time (h)	0	1	3	6	12	24
Size (nm)	49 ± 5.2	47 ± 8.0	49 ± 7.6	49 ± 5.8	59 ± 6.0	80 ± 7.3
Zeta potential (mV)	0.12 ± 1.8	0.15 ± 2.3	0.67 ± 2.3	1.3 ± 1.8	4.2 ± 1.3	8.9 ± 2.7



Figure S5. Comparison of siRNA encapsulation efficiency in SLNPs and DC-Chol/DOPE liposomes. SLNPs comprised of MA-Chol/DOPE (1:1) and DC-Chol/DOPE (1:1) liposomes were analyzed with respect to siRNA encapsulation efficiency as a function of lipid concentration. Both SLNPs and DC-Chol/DOPE liposomes were shown to have similar complexation ability and encapsulation efficiency for siRNA.



Figure S6. Intracellular uptake of TAMRA-siKSP@SLNPs by U87MG cells. Cells were treated with TAMRA-siKSP(100 nM)@SLNPs for 4 h and monitored for 24 h using a confocal microscope. At 24 h post transfection, many cancer cells started to become circularized and fragmented form of nucleus was observed.



Figure S7. Tumor weights and changes in body weight in response to *in vivo* treatment with siKSP@SLNPs in PC-3 tumor-bearing nude mice. * indicates p < 0.05.



Figure S8. Hemolysis study of SLNPs and DC-Chol/DOPE-based lipid nanoparticles. Hemolysis study was performed by plating SLNPs (only vehicle) or DC-Chol/DOPE-based lipid nanoparticles on horse blood agar plates under standard conditions. Both lipid nanoparticle systems did not induce hemolysis.