

**The cytotoxic effects of dimethyl sulfoxide in mouse preimplantation embryos: a
mechanistic approach**

Min-Hee Kang¹, Joydeep Das¹, Sangiliyandi Gurunathan¹, Hwan Woo Park², Hyuk Song¹,
Chankyu Park¹, and Jin-Hoi Kim^{1,*}

¹Department of Stem Cell and Regenerative Biotechnology, Humanized Pig Research Center
(SRC), Konkuk University, Seoul, Korea.

²Department of Cell Biology, College of Medicine, Konyang University, Daejeon, Korea

Running title: Toxicity of DMSO in mouse preimplantation embryos

Number of Figures: 6

Supplementary Figures: 2

Supplementary Tables: 2

Supplementary Movie: 1

Corresponding authors*: Jin-Hoi Kim, PhD and Professor.

Tel.: +82-2-450-3687, Fax: +82-2-458-5414,

E-mail address: jhkim541@konkuk.ac.kr (J-H Kim).

Supplementary Table 1. List of primer sets used for RT-qPCR.

Genes	Forward	Reverse
<Oxidants and Anti-oxidants>		
<i>Gss</i>	AAAAGGACGACTATACTGCC	TGTAATCTGAGCGATTCCAGG
<i>Nox1</i>	GGCTTCTTCTGTAGCGTTTCG	ACCTGCTCATTTTGCAACCG
<i>Nox2</i>	TACCTTACTGGCTGGGATGA	TCACTTGCAATGGCTTTGAA
<i>Nox3</i>	CTGGGATGAAAGTCTGGATG	GGGTGATTGTAGGCAATCTG
<i>Nox4</i>	TCCAATCATTCCAGTGGTTT	CCCATCTGTTTGACTGAGGT
<i>Duox1</i>	GCTGAGAAGTTCGACCTCAG	CAGACTCCTGTTCAGCACCT
<i>Duox2</i>	CTCTGGAAGAGCCGCTACTG	GGTAGCCAAAGAAGACCCCC
<i>Ncf1</i>	GCCTTAGCCAGGACACCTAT	GATTGTCTCTGCCTCCAG
<i>Ncf2</i>	TCAGATGAGGATGTGGGACT	TTCTGTGTGACATGCAGCTT
<i>Gsr</i>	AGCTGTGAGGGTAAATTCAG	AGCTGTGAGGGTAAATTCAG
<i>Sod1</i>	GAAGAGAGGCATGTTGGAGA	CACGATCTTCAATGGACACA
<i>Sod2</i>	GAGTTGCTGGAGGCTATCAA	CGACCTTGCTCCTTATTGAA
<i>Gpx1</i>	CTCACCCGCTCTTACCTTC	AAGTTCAGGCAATGTCGTT
<i>Gpx2</i>	AATGTGGCGTCACTCTGAGG	GGGAAGCCGAGAACTACCAG
<i>Gpx3</i>	AGGCGAGAACTCGGAGATAC	GAGCTGGAATTAGGCACAA
<ER Stress>		
<i>Hspa5</i>	TCTGGTGATCAGGATACAGGTG	TTCAGCTGTCCTCGGAGAATA
<i>Hsp90b1</i>	AAGAATGAAGGAAAAACAGGACAAAA	CAAATGGAGAAGATTCGCC
<i>Atf4</i>	TCGATGCTCTGTTTCGAATG	AAGCAGCAGAGTCAGGCTTC
<i>Ddit</i>	GTCTTATGAGATGAGGAATG	AATGGCAGGATACTTCTT
<i>Edem1</i>	CTACCTGCGAAGAGGCCG	GTTCATGAGCTGCCACTGA
<i>Traf2</i>	GATGGGGTCTTCATCTGGAA	CCACTGCAACAGAGCATCAT
<i>Total Xbp1</i>	TGGCCGGTCTGCTGAGTCCG	GTCCATGGGAAGATGTTCTGG
<i>Spliced Xbp1</i>	CTGAGTCCGAATCAGGTGCAG	GTCCATGGGAAGATGTTCTGG
<i>Unspliced Xbp1</i>	GTCCATGGGAAGATGTTCTGG	CAGCACTCAGACTATGTGCA
<Autophagy>		
<i>Itr1</i>	TTGTTGATACTCGTGAAT	AGCATCACCATAGGATAA
<i>Atg3</i>	GCTTGATTGTGTCAGTT	ACAGACTAATATGGAGACT
<i>Atg4b</i>	CCACAAGGTAGCAAGAGA	GAAGTCAAGGGACAAGATATG
<i>Atg5</i>	GGAGAGAAGAGGAGCCAGGT	TGTTGCCTCCACTGAACTTG
<i>Atg6</i>	ACAGTTCTATAACAAGTGA	GGGTTCTTTGCTATACAT
<i>Atg7</i>	CAGTAGCCTGTAGAATAAC	GTAGGTGTGCTGTAATC
<i>Map1lc3b</i>	TTCTTCTCTGGTGAATGG	GTGGGTGCCTACGTTCTCAT
<Mitophagy>		
<i>PINK1</i>	GCCAACACTGAACTTTGCTT	GCTGGTTGCTGCTTACAAAT
<i>Parkin</i>	TCCGAAGATTCTACCTCC	AGGGGCTGCTCTGTAATCT
<Apoptosis>		
<i>Atf2</i>	ACATCCAATGGAGTCAGTTC	GCCATGACAATCTGTGAAAG

<i>Mapk8</i>	CGTTGAGTAGTGTAAAGAA	CTTGTTCAGATAAATACCA
<i>Jun</i>	TTAACATTGACCAAGAACT	TTAAGGAGCACTACAGAA
<i>Casp3</i>	GATAATGTCTTAGAACTTGAATCC	CTTCATAAATCAGGTCCAA
<i>Casp9</i>	GACTTACAAGCAGATTCC	AGAATAATGAGGCAGAGA
<i>Trp53</i>	CAAACACGAACCTCAAAG	CTATCCTTACCATCATACA
<i>Bax</i>	CGAGCTGATCAGAACCATCA	GAAAAATGCCTTCCCTTC
<i>Bcl-2</i>	TAAGCTGTCACAGAGGGGCT	TGAAGAGTTCCTCCACCACC

<Wnt Signalling>

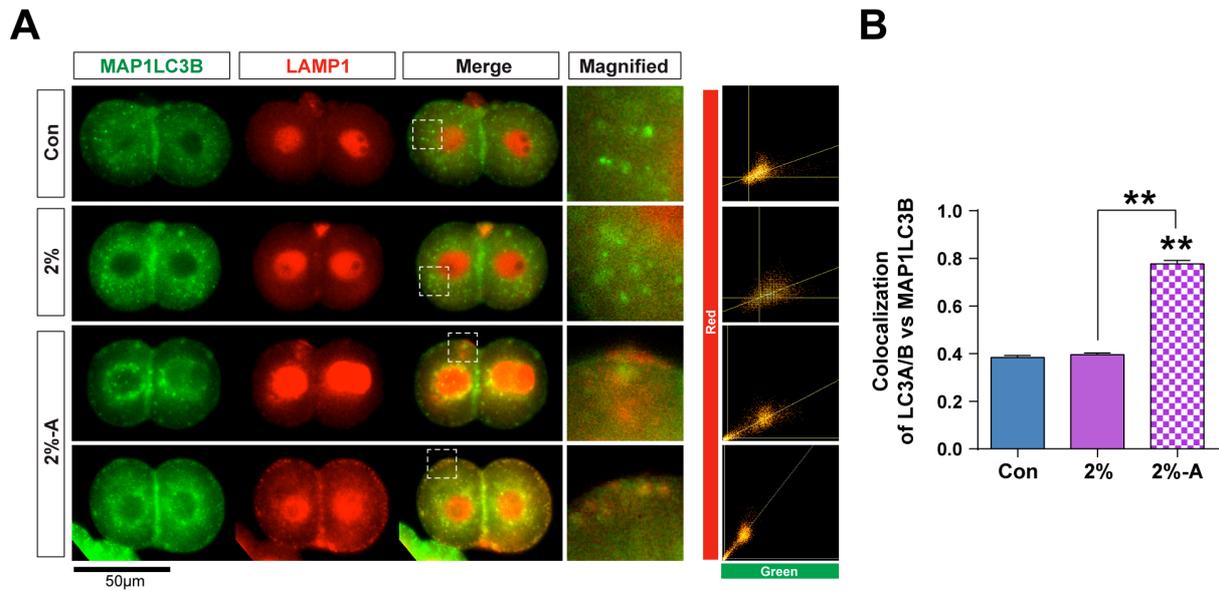
<i>B3gnt5</i>	ACGTGGGGCAATGAGAACTA	TTCAGTGGACCAGGAGTTCC
<i>Eomes</i>	GAGCTTCAACATAAACGGACTCAA	CGGCCAGAACCACTTCCA
<i>Lrp5</i>	GTGTGCAGTTGCAGGACAAT	CTCCAGGGGATCGTAGTCAA
<i>Lrp6</i>	GACAGACTGGGGAGAAGTGC	AACGTCAAGGCAAAAGGATG
<i>Wnt1</i>	CCGAGAAACAGCGTTCATCT	GGTTCATGAGGAAGCGTAGG
<i>Wnt3a</i>	GGTCTACTACGAGGCCTCAC	CATCTATGCCATGCGAGCTC

<Mitochondrial Formation, Function and Copy Number>

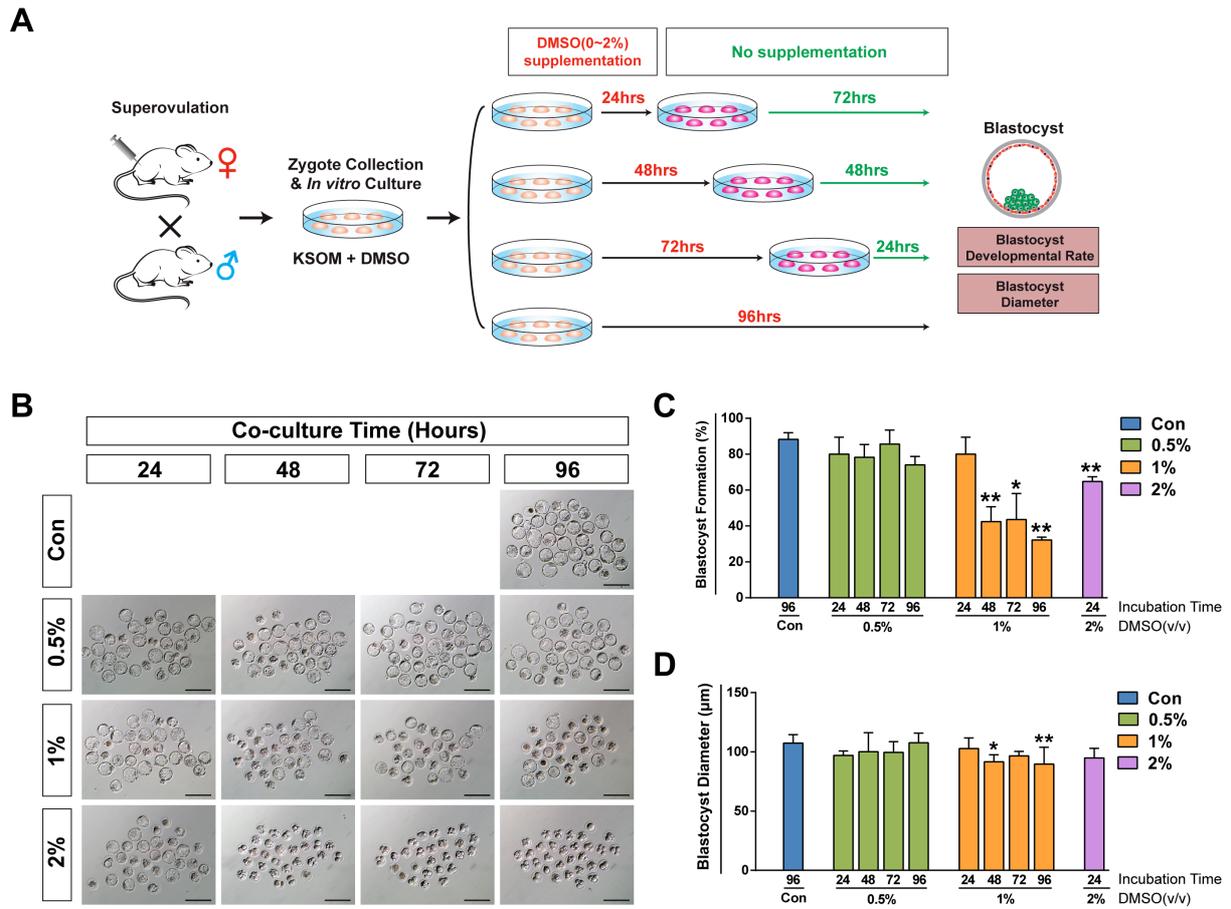
<i>Mpv17</i>	TCGGAGGCTGGTACAAAGTT	ATTGTCCTGGGCTGACATTC
<i>Cbr1</i>	AGTGGTGAATGTGTCCAGCA	CAGGACTGTCACCCCAATCT
<i>Imp11</i>	GGCATCCAAAGAGGTGACAT	ACATGACCTGTTGGCACGTA
<i>Mtfp1</i>	GCTGTGGTGTGGTTGAGCTA	ACACAGACGTTGATGGTGA
<i>Rhot2</i>	CAGTTACCCGCGAGAAGAAG	GGCTGTCAGCATTCACTCA
<i>Cy5</i>	TTCCACAACCCTCATGTGAA	TAAGGGTCCAAAACCAGTGC
<i>Atp5a1</i>	GCCCTCGGTAATGCTATTGA	CACAGAGATTGGGGGATAA
<i>Atp5e</i>	TCAGCTACATCCGGTTTCC	GGAGGTGAGGTTGATTCCA
<i>Atp5d</i>	CGGACAGATGCCTTCACCT	ACTTAGTCGTGGTGCCGTCT
<i>Atp5k</i>	CGGTTCAGGTCTCTCCACTC	TGACGCCTCACTGAGAATG
<i>Atp5f1</i>	TCAGAAGCGCCATTACCTCT	TTGGCAATGGTCTCCTTTTC
<i>Cytb</i>	ATTGACCTACCTGCCCATC	CTCGTCCGACATGAAGGAAT
<i>Acta2</i>	TCGCCATGGATGACGATA	CACGATGGAGGGGAATACAG

Supplementary Table 2. Antibodies used for immunostaining and blotting.

Antibody	Cat#	Company
PDI	ab31811	Abcam
p-p53	9286s	Cell Signalling
CYCS/Cytochrome C	ADI-AAP-170	Enzo Life Sciences
Bcl-2	ab59348	Abcam
BAX	ab10813	Abcam
PINK1	ab23707	Abcam
Parkin	ab15954	Abcam
MAP1LC3B	4108s	Cell Signalling
LAMP1	ab24170	Abcam
GRP78/BiP	ab21685	Abcam
p38	ab27986	Abcam
p-p38	sc-7973	Santa Cruz
TOM20	sc-17764	Santa Cruz
ATG5/12	NB110-53818	Novus Biologicals
Beclin1	ab16998	Abcam
SQSTM1/p62	ab56416	Abcam
OCT3/4	sc-9081	Santa Cruz
CDX2	MU392A-UC	BioGenex
ACTB	ab8227	Abcam



Supplementary Figure 1. (A) MAP1LC3B and LAMP1 were co-immunostained in 2-cell embryos treated with DMSO for 24 and 36 h (2%-Arrested). **(B)** Fluorescence intensities of both of MAP1LC2B and LAMP1 were analyzed for co-localization. **: $p < 0.01$.



Supplementary Figure 2. (A) Schematic experimental diagram for rescuing embryos from DMSO toxicity. Zygotes are cultured with different concentration (0%, 0.5%, 1%, and 2% v/v) of DMSO during 24, 48, and 72 h and then moved to fresh KSOM medium without DMSO for further culture, respectively. **(B)** A representative embryo picture cultured according to (A)'s schedule. **(C, D)** Developmental rate and diameter of blastocysts developed after exposure to 0%, 0.5%, 1%, and 2% DMSO for 24, 48, 72, and 96 h. Scale bar = 200 µm. *: $p < 0.05$, **: $p < 0.01$. Even though any blastocyst stage of embryos could not be obtained from 2% of DMSO supplementation, approximately 70% of zygotes, which are exposed 2% of DMSO for 24h and then move to fresh KSOM medium, can develop to fully hatched blastocyst stage of embryos.

Supplementary Movie 1. Time lapse observation of preimplantation embryo development in KSOM supplemented with 0%, 0.5%, 1%, and 2% DMSO.