## Supplementary information

Table S1. Physiochemical characterization of PBAE/pDNA NPs suspended in deionized

Entry	Medium	Z-average(nm)	PDI	ζpotential(mV)
NPs/pCXCR4	DI H <sub>2</sub> O	179.7±0.32	0.142±0.04	43.22±2.33
	ADSCs medium	207.7±0.75	0.149±0.01	-7.55±1.36
NPs/pEGFP	DI H <sub>2</sub> O	183.5±0.34	0.136±0.03	42.84±1.24
	ADSCs medium	206.1±2.31	0.281±0.01	-8.77 ±0.41

water or full-component culture medium (n = 3).



Figure S1. Map of pcDNA3.1(+)-CXCR4



**Figure.S2.** Flow cytometry analyses indicates hADSCs at passage 3 were positive for CD44 (A), CD29 (B), CD105 (C), and CD73 (D), and negative for CD34 (E) and CD31 (F). 2×106 events were acquired and the frequency of positive cells was determined using FlowJo.



**Figure S3. (A) Transfection efficiency visualized using fluorescence microsco**py. In vitro transfection efficiency with PBAE NPs or a leading commercially available transfection agent (Lipofectamine 2000; Lipo 2k). EGFP was used as a reporter to optimize the transfection formulation, and pcDNA was used as a vector control. Optimal PBAE/pEGFP NPs led to higher transfection efficiency than achieved with Lipo 2k. No fluorescence was detected in pcDNA-treated hADSCs. Scale bars = 200 μm. (B) Flow cytometry of hADSCs 48 h after transfection with pEGFP mediated by PBAE NPs or Lipo 2k. hADSCs treated with 20:1 PBAE:pEGFP consistently demonstrated ~5.4-fold higher transfection efficiency than cells transfected with Lipo 2k. \*\*p<0.01. (C) Cell viability was analyzed via the MTS assay 48 h after transfection with NPs/pEGFP or Lipo 2k/pEGFP. hADSCs transfected with NPs/pEGFP exhibited no significant change in viability compared with naïve controls. The viability of untransfected, unmanipulated parallel cultures of hADSCs was defined as100% viability.



**Figure S4.** CXCR4<sup>+</sup> hADSCs (passae 3) determined using Flow Cytometry (without permeabilization 48h after NPs-mediated transfection. (A) Non-treated hADSCs (no transfection and no immunostaining); (B) untransfected hADSCs without immunostaining; (C) Transfected hADSCs with immunostaining.



Figure S5. More hADSCs migrate toward conditioned medium in which the malignant GBM cell line U87MG was cultured under normoxic conditions (O2, 20%) than toward fresh Dulbecco's Modified Eagle Medium (control). CXCR4 overexpression further enhanced this migration. Pre-incubation with the CXCR4 antagonist AMD3100 did not affect the migration of naïve hADSCs toward conditioned medium. Cell membranes were stained red with the florescent dye PKH26; nuclei were labeled blue with Hoechst 33342. Scale bar =  $200 \mu m$ .



**Figure S6. hADSCs did not migrate towards tumor tissues 5 days after injection into the tail veins of mice with brain glioma**. (A) Schematic of experimental design. i.c., intracranial; i.v., intravenous; SQ, subcutaneous. (B) Representative sections of the tropism of CXCR4-overexpressing hADSCs and naïve hADSCs toward malignant cells in the brain tissue of a xenograft mouse model 5 days after intravenous injection.