## Overcoming EZH2 inhibitor resistance by taxane in PTEN-mutated prostate cancer

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## Figure S1. EZH2 represses expression of FOXO1, but not FOXO3 and FOXO4.

(A, B) RT-qPCR analysis of *DAB2IP* and *BRACHYURY* mRNA expression in C4-2 and 22Rv1 cell lines transfected with non-specific siRNA (siNS) or a pool of EZH2-specific siRNA for 48 h. (C) RT-qPCR analysis of *FOXO1* mRNA expression in LNCaP prostate cancer cell line and BPH1 benign prostatic epithelial cell line transfected with siNS or a pool of EZH2-specific siRNA for 48 h. (D) Western blot analysis of FOXO1 and EZH2 proteins in indicated cell lines transfected with siNS or a pool of EZH2-specific siRNA for 48 h. (D) Western blot analysis of FOXO1 and EZH2 proteins in indicated cell lines transfected with siNS or a pool of EZH2-specific siRNA for 48 h. ERK2 was used as a loading control. (E, F) RT-qPCR analysis of expression of *FOXO3* (E) and *FOXO4* (F) mRNA in C4-2 and 22Rv1 cell lines transfected with siNS or a pool of EZH2-specific siRNA for 48 h. *GAPDH* gene was utilized as an internal control for RT-qPCR. Data are shown as means  $\pm$  SEM. The *P* value was performed by the unpaired two-tailed Student's t-test. \* *P*<0.05; \*\* *P*<0.01; \*\*\* *P*<0.001; n.s., no significance.



Figure S2. The inhibition of EZH2 and HDAC promote FOXO1 expression. (A) C4-2 and 22Rv1 cells were treated with different concentrations of EZH2 inhibitor GSK126 for 72 h and harvested for RT-qPCR analysis of mRNA expression of DAB2IP. (B) C4-2 and 22Rv1 cells were treated with different concentrations of EZH2 inhibitor GSK343 for 72 h and harvested for RT-qPCR analysis of mRNA expression of DAB2IP. (C) C4-2 and 22Rv1 cells were treated with different concentrations of EZH2 ASO for 48 h and harvested for RT-qPCR analysis of mRNA expression of DAB2IP. (D) C4-2 and 22Rv1 cells were treated with different concentrations of HDAC inhibitor SAHA for 48 h and harvested for RT-qPCR analysis of mRNA expression of DAB2IP. In the above experiments, GAPDH was utilized as an internal control for RT-qPCR. (E, F) C4-2 and 22Rv1 cells were treated with 2 µM of GSK126 for 48 h and harvested for ChIP-qPCR analysis with H3K27ac antibody (E) and H3K27me3 antibody (F). (G, H) HCT116 (colorectal cancer), MCF7 (breast cancer) and PANC1 (pancreatic cancer) cell lines were treated with 2 µM of GSK126 for 48 h and harvested for ChIP-qPCR analysis with H3K27ac antibody (G) and H3K27me3 antibody (H). (I) HCT116, MCF7 and PANC1 were treated with the indicated concentrations of GSK126 for 24 h and harvested for WB with the indicated antibodies. Data are shown as means  $\pm$ SEM. The P value was performed by the unpaired two-tailed Student's t-test. \* P<0.05; \*\* *P*<0.01; \*\*\* *P*<0.001; n.s., no significance.



Figure S3. AKT signaling pathway causes the resistance to EZH2 inhibition. (A - G)22Rv1 and DU145 cells were infected with lentivirus expressing shNS or shFOXO1 and selected with puromycin for stable cell lines. Cells were harvested to detect the transfection efficiency by western blot analysis with the indicated antibodies in (A) and MTS with the indicated concentrations of GSK126 in (B) and (C). Furthermore, the infected 22Rv1 cells were plated into Matrigel for 3D culture assay with the representative images taken at day 7 (D) and quantification results are shown in (E). Similarly, the infected DU145 cells were plated into Matrigel for 3D culture assay with the representative images taken at day 7 (F) and quantification results are shown in (G). Data are shown as means ± SEM. The *P* value was performed by the unpaired two-tailed Student's t-test. \* *P*<0.05; \*\* *P*<0.01; \*\*\* *P*<0.001; n.s., no significance.



AKT

ERK2

0.0

ò

24

48 Time (h)

96

72

0.0

ò

24 48 72 Time (h)

96



Figure S4. Taxane sensitizes prostate cancer cells to EZH2 inhibitor. (A) 22Rv1 cells were treated with GSK126 (10  $\mu$ M) for 72 h and/or DTX (2 nM) for 30 min prior to IFC. The cell membrane was co-stained with anti-E-cadherin while the nucleus was counterstained with DAPI. FNS stands for FOXO1 nuclear staining. Scale bar: 25  $\mu$ m. (B, C) AR negative DU145 (PTEN+/-) and PC-3 (PTEN-/-) cell lines were treated with the indicated concentrations of GSK126 followed by MTS assay at different time points. (D) DU145 and PC-3 cells were treated with different concentrations of GSK126 for 72 h and harvested for western blot analysis with the indicated antibodies. (E) 22Rv1 cells were treated with GSK126 (5  $\mu$ M), DTX (2 nM) and the combination of GSK126 (5  $\mu$ M) and DTX (2 nM) for MTS assay at the indicated time points. Data are shown as means ± SEM. The *P* value was performed by the unpaired two-tailed Student's t-test. \* *P*<0.05; \*\* *P*<0.01; \*\*\* *P*<0.001; n.s., no significance.



## **Figure S5. FOXO1 plays a critical role in the sensitivity of EZH2 inhibition. (A, B)** C4-2 and 22Rv1 cells were treated with 5 μM of non-specific control ASO and subjected for MTS at the different time points. **(C, D)** Mice with C4-2 xenograft tumors were treated with vehicle (20% Captisol), GSK126 (50 mg/kg), DTX (5 mg/kg) or the combination of GSK126 (50 mg/kg) and DTX (5 mg/kg) five days a week for three consecutive weeks. The tumors isolated at day 24 are shown in **(C)** with quantification results shown in **(D)**.

Gene	Usage	Sequences (5'-3')		
EZH2	siRNA	GGAAAGAACUGAAACCUUA		
		CAGAAGAGCUGAUGAAGUA		
FOXO1	shRNA-#1	CCGGATCTACGAGTGGATGGTCAACTCGAGTTGACCATCCACTCGTAGATCTTTTTG		
	shRNA-#2	CCGGGCCGGAGTTTAGCCAGTCCAACTCGAGTTGGACTGGCTAAACTCCGGCTTTTTG		
EED	shRNA-#1	CCGGCCAGAGACATACATAGGAATTCTCGAGAATTCCTATGTATG		
	shRNA-#2	CCGGCCAGTGAATCTAATGTGACTACTCGAGTAGTCACATTAGATTCACTGGTTTTT		
SUZ12	shRNA-#1	CCGGCGGAATCTCATAGCACCAATACTCGAGTATTGGTGCTATGAGATTCCGTTTTTG		
	shRNA-#2	CCGGGCTGACAATCAAATGAATCATCTCGAGATGATTCATTTGATTGTCAGCTTTTTG		

Table S1. Sequences of siRNAs and shRNAs used in the manuscript.

## Table S2. Sequences of primers used in the manuscript.

Gene	Usage	Sequences (5' – 3')	Sequences (5' – 3')
FOX01	ChIP-qPCR	GGGGTAGTGGGGTGTTTTTC	AGTACTCGGCTCTGCTGCTC
F0X01	RT-qPCR	TTATGACCGAACAGGATGATCTTG	TGTTGGTGATGAGAGAAGGTTGAG
F0X03	RT-qPCR	GTTTGAACGTGGGGAACTTCACTG	AACGGTATCACTGTCCACTTGCTG
F0X04	RT-qPCR	ACGAGTGGATGGTCCGTACT	GGTTCAGCATCCACCAAGAG
DAB2IP	RT-qPCR	TGGACGATGTGCTCTATGCC	GGATGGTGATGGTTTGGTAG
BRACHYURY	RT-qPCR	GGAGCTCACCAATGAGATGA	CGAGTCGGGGTGGATGTAGA
GAPDH	RT-qPCR	ACCCACTCCTCCACCTTTGAC	TGTTGCTGTAGCCAAATTCGTT
EZH1	sgRNA	CACCGTCGACAACTTAAACGGCTTC	AAACGAAGCCGTTTAAGTTGTCGAC