| Table S1. Sequence of p | primers used in real-time P0 | CR |
|-------------------------|------------------------------|----|
|-------------------------|------------------------------|----|

| | Forward Primer | Reverse Primer | |
|---------------|-------------------------|-------------------------|--|
| Human IL1β | AGCTACGAATCTCCGACCAC | CGTTATCCCATGTGTCGAAGAA | |
| Human IL6 | ACTCACCTCTTCAGAACGAATTG | CCATCTTTGGAAGGTTCAGGTTG | |
| Human iNOS | TTCAGTATCACAACCTCAGCAAG | TGGACCTGCAAGTTAAAATCCC | |
| Human TNFα | CCTCTCTCTAATCAGCCCTCTG | GAGGACCTGGGAGTAGATGAG | |
| Human Arg-1 | GTGGAAACTTGCATGGACAAC | AATCCTGGCACATCGGGAATC | |
| Human CXCL9 | CCAGTAGTGAGAAAGGGTCGC | AGGGCTTGGGGCAAATTGTT | |
| Human CXCL10 | GTGGCATTCAAGGAGTACCTC | TGATGGCCTTCGATTCTGGATT | |
| Human IFNG | TCGGTAACTGACTTGAATGTCCA | TCGCTTCCCTGTTTTAGCTGC | |
| Human CCL2 | CAGCCAGATGCAATCAATGCC | TGGAATCCTGAACCCACTTCT | |
| Human β actin | CATGTACGTTGCTATCCAGGC | CTCCTTAATGTCACGCACGAT | |
| Mouse IL1β | GAAATGCCACCTTTTGACAGTG | TGGATGCTCTCATCAGGACAG | |
| Mouse IL6 | CTGCAAGAGACTTCCATCCAG | AGTGGTATAGACAGGTCTGTTGG | |
| Mouse iNOS | ACATCGACCCGTCCACAGTAT | CAGAGGGGTAGGCTTGTCTC | |
| Mouse TNFa | CAGGCGGTGCCTATGTCTC | CGATCACCCCGAAGTTCAGTAG | |
| Mouse Arg-1 | CTCCAAGCCAAAGTCCTTAGAG | GGAGCTGTCATTAGGGACATCA | |
| Mouse β actin | AAGGCCAACCGTGAAAAGAT | GTGGTACGACCAGAGGCATAC | |

Table S2. Array map of 40 human cytokines

| А | POS1 | POS2 | CXCL13 |
|---|---------|-----------|----------|
| В | Eotaxin | Eotaxin-2 | G-CSF |
| С | GM-CSF | I-309 | ICAM-1 |
| D | IFNg | IL-1a | IL-1b |
| E | IL-1ra | IL-2 | IL-4 |
| F | IL-5 | IL-6 | IL-6R |
| G | IL-7 | IL-8 | IL-10 |
| Н | IL-11 | IL-12p40 | IL-12p70 |
| I | IL-13 | IL-15 | IL-16 |
| J | IL-17 | MCP-1 | MCSF |
| К | MIG | MIP-1a | MIP-1b |
| L | MIP-1d | PDGF-BB | RANTES |
| М | TIMP-1 | TIMP-2 | TNFa |
| Ν | TNFb | TNF RI | TNF RII |

Supplemental Figures



Figure S1. CD68 and STING expression in GC patient samples. Upper panel, immunoblot analysis showing CD68 and STING expression in paired normal mucosa and tumor samples from GC patients. GAPDH was used as a loading control. Lower panel, the ratios of CD68/GAPDH and STING/GAPDH were quantified, and statistical significance was analyzed by comparing to the normal mucosa. Data are presented as the mean \pm SD (n=3). *, p < 0.05; **, p < 0.01; ***, p < 0.001.



Figure S2. Knocking-down and activating STING promotes both PBMC-DMs and BM-DMs differentiation into the pro-inflammatory subtype. (A) RT-PCR analysis of therapy-related cytokines in 8 pairs of adjacent normal mucosa and GC samples. ***, p < 0.001; **, p < 0.01; *, p < 0.05. (B) Immunoblot analysis showing STING expression in human peripheral blood-derived macrophages (PBMC-DMs) and mouse bone marrowderived macrophages (BM-DMs) that 1) had been knocked-down STING (shSTING); 2) were ectopically expressing STING (STING); and 3) were treated with the STING activator 2'3'-c-GAMP. α-Tubulin was used as a loading control. The control represents a sample of three undistinguishable controls including scrambled sequence (SC) for shSTING, empty vector (EV) for STING overexpression, and PBS treatment for 2'3'-c-GAMP. The expression of STING in the three control samples was similar, so only one was shown as the control for clarity. (C) The ratio of STING/Vinculin was quantified, and statistical significance was analyzed by comparing to the control (that is the average of three controls). *, p < 0.05; **, p < 0.01; ***, p < 0.001. (D) IFN- β expression measured by ELISA in the supernatants of human PBMC-DM (left) or mouse BM-DM (right) cultures treated as indicated. *, p < 0.05; ***, p < 0.001. (E, F) Flow cytometric analysis of surface marker expression for proinflammatory (CD11b⁺/CD80⁺) and anti-inflammatory macrophages (CD11b⁺/CD163⁺) in mouse BM-DMs with Sting knock-out based on the CRISPR-Cas9 system; quantification in (F). *, p < 0.05; **, p < 0.01. SC represents control with scrambled gRNA transfectioin. (G) Left panel, representative flow cytometric plots of human PBMC-DMs treated as indicated; right panel, quantification. ***, p < 0.001; **, p < 0.01; *, p < 0.05. Data in B-G are presented as the mean±SD (n=3).



Figure S3. STING knock-out in THP1-derived macrophages promotes pro-inflammatory subtype differentiation. Representative flow cytometric plots of surface marker expression (CD11b⁺CD80⁺) in human THP1-derived macrophages treated as indicated. Control stands for the average of undistinguishable controls of scrambled sequence (SC; for shSTING), and PBS treatment (for 2'3'-c-GAMP treatment), empty vector (EV) was used as a control of STING overexpression.



Figure S4. Macrophages with STING alteration had no effect on cell cycle arrest of GC cells or phenotype of T cells. (A-C) Left panels, immunoblot analysis of cleaved-PARP and total PARP expression in human PBMC-DMs (A), mouse BM-DMs (B) and THP1-DMs (C) treated as indicated. GAPDH was used as a loading control. Right panels, the ratios of Cleaved PARP/Total PARP were quantified; *, p < 0.05; **, p < 0.01; ***, p < 0.001. (D) Cell cycle analysis of human PBMC-DMs (left) and mouse BM-DMs (right) treated as indicated. (E) Isolated CD3⁺ total T cells from human PBMCs were stimulated with ImmunoCultTM Human CD3/CD28 T Cell Activator and cultured in ImmunoCultTM-XF T Cell Expansion Medium. The activation of viable CD3⁺ T cells was assessed by CD25 expression using flow cytometry. (F) Representative flow cytometric plots of CD4/CD8 T cells in cocultures of human T cells with human PBMC-DMs treated as indicated; quantification of the CD4/CD8 ratio in (G). (A, B, D, F, G) Control stands for a representative sample of undistinguishable controls of scrambled sequence (for shSTING), empty vector (for STING overexpression), and PBS treatment (for 2'3'-c-GAMP treatment). (C) Control stands for the average of undistinguishable controls of scrambled sequence (SC; for shSTING), and PBS

treatment (for 2'3'-c-GAMP treatment), empty vector (EV) was used as a control of STING overexpression. Data in A, B, C, D, G are presented as the mean±SD (n=3).



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Figure S5. The effects of STING knockdown and activation on macrophage differentiation are mediated through the JAK-STAT-IL24 pathway. (A) Upper panel, whole pictures of cytokine arrays in Figure 5A. Lower panel, enriched pathways of altered cytokines. Control stands for a representative sample of undistinguishable controls of scrambled sequence (for shSTING), empty vector (for STING overexpression), and PBS treatment (for 2'3'-c-GAMP treatment). (B) Representative flow cytometric plots of human PBMC-DMs in Figure 5C. (C, D) Left panels, immunoblot analysis of cleaved-PARP and total PARP expression in human PBMC-DMs treated as indicated. GAPDH was used as a loading control. Right panels, the ratios of Cleaved PARP/Total PARP were quantified; **, p < 0.01; ***, p < 0.001. Data in C and D are presented as the mean±SD (n=3) (B-D) Control stands for the average of undistinguishable controls of scrambled sequence (SC; for shSTING), and PBS treatment (for 2'3'-c-GAMP treatment), empty vector (EV) was used as a control of STING overexpression.



Figure S6. Macrophages with Sting knocking-down or activation have killing effects on cancer cells of spontaneous gastric tumors in mice. (A) Representative pictures of CD68 IHC staining in the spleens of mice treated as indicated(n=8). Scale bars represent 200 μm. **(B)** Body weight of mice with indicated treatments(n=8). **(C)** Immunofluorescence images indicating the engulfment of pHrodoTM red bioparticles by treated mice. Data are presented as the mean±SD (n=10). **(D)** Flow cytometric analysis of surface marker expression for total T cells (CD3⁺) in the blood and spleens of mice treated as indicated. **(E)** Representative flow cytometric plots of mouse CD8⁺ and CD4⁺ T cells in monocytes (upper) and splenocytes (lower) in Figure 6E and F. (C-E) Control stands for a representative sample of undistinguishable controls of scrambled sequence (for shSting), empty vector (for Sting overexpression), and PBS treatment (for 2'3'-c-GAMP treatment).