NIR-II fluorescence microscopic imaging of cortical

vasculature in non-human primates

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Supplementary Figure 1. Hydrodynamic mean size distributions of rhesus macaque serum (pink) and ICG in rhesus macaque serum (yellow) by dynamic light scattering. These distributions are significantly different (RMS mean: 5.4 nm, ICG in RMS mean: 5.76 nm; ($X^2 = 131.6$, p < 0.001).



Supplementary Figure 2. NIR-II fluorescence wide-field microscopic image of cerebral vessels showing the blood flow directions and determination of arteries and veins. Depth = $180 \mu m$. Scale bar: $100 \mu m$.



Supplementary Figure 3. NIR-II fluorescence wide-field microscopic images of cerebral blood vessels of the rhesus macaque at various depths (40 μ m, 100 μ m, 160 μ m, 240 μ m, 320 μ m, and 400 μ m). Scale bars: 100 μ m.



Supplementary Figure 4. A and B NIR-II fluorescence wide-field microscopic images of cerebral blood vessels of the rhesus macaque at two typical depths (170 μ m and 300 μ m). C-D The cross-sectional fluorescence intensity profiles (black) and the related Gaussian fitting (red) along the capillaries indicated by the white-dashed lines in **A** and **B**. Scale bars: 100 μ m.



Supplementary Figure 5. Measurement of cardiac impulse period of the rhesus macaque based on NIR-II fluorescence wide-field microscopic brain vascular imaging. A Two typical images with the fluorescence bright/dark border in the highest (top image) and lowest (bottom image) position in one cerebral blood vessel. Depth = 130 μ m. Scale bars: 100 μ m. B A plot of the position of the bright/dark boundary in the vessel as a function of time. C A plot of the peak timepoints of each impulse shown in B. The linear fit reveals an average cardiac impulse period of 502.9 ms/pulse, matching the 120 pulses/minute recorded on the heart rate monitor.

NIR-II fluorescence confocal microscopy in mice.

Prior to using the NIR-II fluorescence confocal microscope in primates, we first evaluated its performance in mice. As shown in Supplementary Fig. 6 and Supplementary Fig. 7, we imaged cerebral vasculature through a cranial window on a mouse which had received an intravenous injection of ICG (1 mg/mL, 200 μ L). Images at various depths were obtained and a 3D image was reconstructed, revealing a clear vascular network (Supplementary Fig. 6). In addition, the spatial resolution and SBR were analyzed (Supplementary Fig. 7). This verified the superior imaging performance of our confocal setup.



Supplementary Figure 6. NIR-II fluorescence confocal microscopic images of cerebral blood vessels of mouse at various depths and a 3D reconstructed image over depths of 0-400 μ m. Scale bars: 100 μ m.



Supplementary Figure 7. A and B NIR-II fluorescence confocal microscopic images of cerebral blood vessels of the mouse at two typical depths (100 μ m and 200 μ m). C-D The cross-sectional fluorescence intensity profiles (black) and the related Gaussian fitting (red) along the capillary vessels indicated by the white-dashed lines in A and B. Scale bars: 100 μ m.

MOV S1. A movie showing the NIR-II fluorescence wide-field microscopic *in vivo* imaging the flow of cerebral blood vessels in the rhesus macaque, with the objective magnification of $25 \times$.

MOV S2. NIR-II fluorescence wide-field microscopic *in vivo* imaging showing the blood flow directions in cerebral vessels, as well as the discrimination of artery and vein.

MOV S3. NIR-II fluorescence wide-field microscopic *in vivo* imaging showing the moving of fluorescence bright/dark border in one cerebral blood vessel acompanied with the cardiac impulse.

MOV S4. 3D reconstructed NIR-II fluorescence confocal microscopic *in vivo* images of cerebral blood vessels of the rhesus macaque.