Supplemental Information

Small-molecule activating SIRT6 elicits therapeutic effects and synergistically promotes anti-tumor activity of vitamin D₃ in colorectal cancer

Jialin Shang^{1,†}, Zhehui Zhu^{2,†}, Yingyi Chen^{1,†}, Jinglue Song², Yuji Huang², Kun Song¹, Jie Zhong¹, Xinyuan Xu¹, Jiacheng Wei¹, Chengxiang Wang¹, Long Cui², Chen-Ying Liu^{2,*}, and Jian Zhang^{1,3,4*}

¹State Key Laboratory of Oncogenes and Related Genes, Shanghai Jiao-Tong University School of Medicine, Shanghai 200025, China.

²Department of Colorectal and Anal Surgery, Xinhua Hospital, Shanghai Jiao-Tong University School of Medicine, Shanghai 200092, China.

³Medicinal Bioinformatics Center, Shanghai Jiao-Tong University School of Medicine, Shanghai 200025, China.

⁴School of Pharmaceutical Sciences, Zhengzhou University, Zhengzhou 450001, China.

[†]These authors contributed equally to this work.

*Corresponding author.

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Figure S1. Predicted binding modes of MDL-818, 813, 822, 821, 811 and 814 at the allosteric site on SIRT6. The compounds and pocket residues are displayed as sticks and colored in yellow and hotpink. Hydrogen bonds are shown as red dashed lines and hydrophobic interactions are shown as palegreen dashed lines.







MDL-800 SIRT6 deacetylation Figure S2. and MDL-811 activated on RHKK-Ac-AMC, determined FDL **HPLC** assays. as by and (A) Concentration-dependent effects of MDL-800/MDL-811 on the activation of SIRT6 deacetylation were assessed by FDL assays with the acetylated peptide RHKK-Ac-AMC (75 μ M). The data are shown as mean \pm s.d. from three independent experiments (MDL-811: $EC_{50} = 5.7 \pm 0.8 \mu M$; MDL-800: $EC_{50} = 12.3 \pm 0.7 \mu M$). (B) HPLC traces of SIRT6 deacetylation on RHKK-Ac-AMC in the absence or presence of MDL-800/MDL-811 at 25 µM. Data are the other two of three independent experiments.



Figure S3. Concentration-dependent effect of MDL-811 on SIRT6 demyristoylation. The effect of MDL-811 was assessed by FDL assay with a TNF α peptide containing a myristoyl group (EALPKK-Myr-AMC) at various concentrations of MDL-811. The data are shown as mean \pm s.d. from three independent experiments.



Figure S4. Dose-dependent deacetylation effects of MDL-811 in CRC cell lines. (A) Representative western blots indicating the thermal stabilization of the endogenous SIRT6 protein. The CETSA was performed in intact HCT116 cells in the absence or presence of 10 µM MDL-811 for 24 h. (B) Representative western blots of SIRT6, H3K9Ac, H3K18Ac, and H3K56Ac in HT29 and SW480 cells treated with the indicated concentrations of MDL-811 for 48 h, or treated with 10 µM MDL-811 at the indicated times. (C) Representative western blots of SIRT6, H3K9Ac, H3K18Ac, and H3K56Ac in various CRC cell lines treated with indicated concentrations of MDL-811 for 48 h. Histone H3 was used as the internal control, and β -actin was used as the loading control. (D) Pearson correlation analysis between the IC_{50} values of MDL-811 at 48 h and the levels of H3K9Ac or H3K56Ac deacetylation in various CRC cell lines after treatment with 10 µM MDL-811. Quantification of deacetylation levels was performed with ImageJ V4. Each deacetylation level was normalized to the corresponding histone H3 level. Each dot represents the mean \pm s.e.m. of two or three independent experiments in n = 26 CRC cell lines. The data were subjected to Pearson correlation analysis (10 μ M MDL-811: H3K9Ac, r = -0.4734, P = 0.0146; H3K56Ac, r = -0.4631, P = 0.0172).



Figure S5. Cell cytotoxicity assays of CRC cell lines treated with MDL-811. (A) Live-dead double staining assays of CRC cells (HCT116, HT29, and SW480) treated with DMSO for 48 h, with 5 or 10 μ M MDL-811 for 48 h or with 10 μ M doxorubicin for 12 h. Live cells were stained with the green fluorescent dye Calcein-AM, and dead cells were stained with the red dye propidium iodide. The images are representative fields from two independent experiments. Representative images (10× magnification) are shown. Scale bars, 100 μ m. (B) Cytotoxicity of MDL-811 to CRC cell lines (HCT116, HT29, SW480) and non-cancerous colon cell line FHC was confirmed by measuring the release of the cytosolic marker LDH. Cells were treated with the indicated concentrations of MDL-811 for 48 h or with 10 μ M doxorubicin for 12 h. The data are presented as the mean \pm s.d. of three independent experiments.



Figure S6. Cell proliferation of non-cancerous colon cell line treated with MDL-811. Dose response of the proliferation of non-cancerous colon cell line FHC and CRC cell line HCT116 exposed to MDL-811 for 48 h was normalized to the proliferation of the corresponding DMSO-treated controls. Cell proliferation was determined by CCK-8 assays. The data indicate the mean \pm s.e.m. of three independent experiments (FHC: IC₅₀ = 24.4 \pm 0.5 μ M; HCT116: IC₅₀ = 4.8 \pm 0.2 μ M).



Figure S7. Cell cycle distribution of CRC cell lines treated with MDL-811. (A) Cell cycle distribution of HCT116, HT29, and SW480 cells treated with DMSO or with 5 or 10 μ M MDL-811 for 48 h, as measured by PI staining. The data are presented as the mean \pm s.d. of three independent experiments (*, *P* < 0.05; **, *P* < 0.01; ***, *P* < 0.001; two-way ANOVA). (B) GSEA plots showing enrichment of gene expression changes by exposure to 10 μ M MDL-811 for 48 h in the DNA REPLICATION and CELL CYCLE signatures from the MSigDB v6.2 collection C2 curated gene sets (CP: KEGG gene sets). Positive normalized enrichment scores (NESs) indicate upregulation of gene sets, whereas negative NESs indicate downregulation of gene sets. FDR, false discovery rate.



Figure S8. Body weights of xenograft models treated with MDL-811. (A) Body weights of HCT116 CDX mice treated with either vehicle or MDL-811 are shown (n = 6 mice per group). (B) Body weights of PDX mice treated with either vehicle or MDL-811 are shown (n = 8 mice per group). The data is plotted as the mean \pm s.d..



Figure S9. Effect of SIRT6 overexpression on gene expressions in HCT116 cells. (A) Western blots showing effects on HCT116 cells transiently transfected with vector or pLVX-SIRT6-HA. (B) RT-qPCR analyses showing the mRNA levels of candidate genes in HCT116 cells transiently transfected with vector or pLVX-SIRT6-HA before being treated with 10 μ M MDL-811 for 48 h. The data are normalized to the β -actin levels and presented as mean \pm s.e.m. of two or three independent experiments (*, *P* < 0.05; **, *P* < 0.01; ***, *P* < 0.001, two-tailed student's unpaired t-test). (C) RT-qPCR analyses showing the mRNA levels of CYP24A1 and SIRT6 in HCT116 cells

transfected with vector or pLVX-SIRT6-HA and then treated with DMSO or 10 μ M MDL-811 for 48 h. The data are normalized to the β -actin levels and presented as mean \pm s.d. of three experiments. (D), (E) and (F) ChIP assays of HCT116 cells transfected with vector or pLVX-SIRT6-HA. Anti-H3K9Ac (D), anti-H3K18Ac (E), anti-SIRT6 or anti-HA (F) antibodies were used to detect the deacetylation levels of these residues in the indicated regions of CYP24A1. The data are presented as the mean values of the percentage of input \pm s.e.m. from one of two independent experiments with technical triplicates (*, P < 0.05; **, P < 0.01; ***, P < 0.001; two-tailed unpaired Student's t-test).



Figure S10. Effects of MDL-811 on the deacetylation of CYP24A1-binding histone marks in HCT116 cells. (A) ChIP assays using anti-H3K56Ac antibody to assess H3K56Ac occupancy in the indicated regions of CYP24A1 in HCT116 cells treated with DMSO or with 5 or 10 μ M MDL-811. The data are presented as the mean values of the percentage of input \pm s.e.m. from n = 3 independent experiments with technical triplicates. *P* values were determined by two-way ANOVA (*, *P* < 0.05). (B), (C) and (D) ChIP-qPCR primers were designed based on the CYP24A1 DNA sequence of peak regions of H3K9Ac (B), H3K18Ac (C) or H3K56Ac (D) using ChIP-seq data extracted from the Cistrome database.



Figure S11. Effect of CYP24A1 overexpression in HCT116 cells. Western blots (A) and RT-qPCR (B) analysis of HCT116 cells transiently transfected with vector or pcDNA3.1-CYP24A1-Flag for 48 h. The data are normalized to the β -actin levels and presented as mean \pm s.e.m. of two experiments.



Figure S12. Effect of MDL-811 and $1,25(OH)_2D_3$ cotreatment on CYP24A1 expression in CRC cell lines. RT-qPCR analysis showing the mRNA levels of CYP24A1 in HCT116 and HT29 cells treated with MDL-811 (5 μ M), $1,25(OH)_2D_3$ (5 μ M) or the combination for 48 h. The data are normalized to the β-actin levels and presented as mean ± s.e.m of three experiments (*, P < 0.05; **, P < 0.01; ***, P < 0.001, two-tailed student's unpaired t-test).



Figure S13. The superimposition of SIRT6 (violet) to SIRT1 (white), SIRT2 (skyblue), SIRT3 (paleyellow) and SIRT5 (limegreen) in cartoon mode. The allosteric site for MDL-811 is represented as red pentagram. The PDB codes of the SIRT1, SIRT2, SIRT3, SIRT5, and SIRT6 structures are 4KXQ, 3ZGV, 4BN4, 3RIY, and 5Y2F, respectively.

Chemical Characterization Synthesis of Compounds MDL analogs









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^aReagents and conditions: (a) pyridine, room temperature,8 h; (b) Fe, CH₃COOH, 50 °C 12 h; (c) pyridine, room temperature, 6 h; (d) LAH, THF, 50 °C, 12 h; (e) SOCl₂, 80 °C, 10 h; (f) secondary amine, THF, 50 °C, 6 h.

Methyl 2-(*N*-(5-bromo-4-fluoro-2-methylphenyl)sulfamoyl)-5-nitrobenzoate (3). To a solution of 5-bromo-4-fluoro-2-methylaniline (20 g, 98.0 mmol, 1) in 20 mL pyridine was added methyl 2-(chlorosulfonyl)-5-nitrobenzoate (32.9 g, 117.6 mmol, 2) under 0 °C and the reaction was stirred at the same temperature for about 1 hour. Then the reaction was moved to 25 °C and stirred for another 8 h. The reaction was cooled to 0 °C and adjusted the pH to 3-4 with 1 N hydrochloric acid solution. The precipitate formed was filtered, washed with water and dried to yield crude intermediate product 5, which was directly used to the next step without any purification. ¹H NMR (400 MHz, DMSO-d₆) δ 10.08 (s, 1H), 8.50-8.47 (m, 2H), 7.98-7.96 (dd, *J* = 2.0 Hz, *J* = 8.0 Hz, 1H), 7.36-7.34 (d, *J* = 8.0 Hz, 1H), 7.29-7.27 (d, *J* = 8.0 Hz, 1H), 3.78 (s, 3H), 1.96 (s, 3H).

Methyl 5-amino-2-(*N*-(5-bromo-4-fluoro-2-methylphenyl)sulfamoyl)benzoate (4). The crude intermediate product **3** (38.59 g, 86.53 mmol) dissolved in acetic acid was added iron powder (29.1 g, 519 mmol) at 50 °C. Then the solution was stirred at the same condition for 12 h. The system was filtered and the solvent was evaporated under reduced pressure. The residue was purified by column chromatography on silica gel (hexane: ethyl acetate = 2:1) to afford intermediate product **4** (29.66 g, two steps 73%) as a white powder. ¹H NMR (400 MHz, DMSO-d₆) δ 8.89 (s, 1H), 7.30-7.20 (m, 2H), 6.73 (s, 1H), 6.64-6.59 (m, 2H), 6.30 (s, 2H), 3.73 (s, 3H), 2.02 (s, 3H).

Methyl

2-(N-(5-bromo-4-fluoro-2-methylphenyl)sulfamoyl)-5-(3,5-dichlorophenylsulfona mido)benzoate solution (6). А of methvl 5-amino-2-(N-(5-bromo-4-fluoro-2-methylphenyl) sulfamoyl) benzoate (4, 13.9 g, 33.32 mmol) in 30 mL pyridine was added 3,5-dichlorobenzene-1-sulfonyl chloride (5, 16.45 g, 58.8 mmol) under 0 °C and the reaction was still stirred at the same temperature for about 1 hour. Then the reaction was moved to 25 °C and stirred for another 6 h. The reaction was cooled to 0 °C, and adjusted the pH to 3-4 with 2 N hydrochloric acid solution. The precipitate formed was filtered and subsequently purified by column chromatography on silica gel (hexane: ethyl acetate = 3:1) to afford 6 (17.6 g) as a white powder. Yield: 85%. ¹H NMR (400 MHz, DMSO- d_6) δ 11.36 (s, 1H), 9.46 (s, 1H), 8.01 (s, 1H), 7.81 (d, J = 2.0 Hz, 2H), 7.60-7.58 (d, J = 8.0Hz, 1H), 7.44-7.40 (dd, J = 2.0 Hz, J = 8 Hz, 1H), 7.27 (d, J = 2.0 Hz, 1H), 7.24-7.22 (d, J = 8.0 Hz, 1H), 7.18-7.16 (d, J = 8.0 Hz, 1H), 3.74 (s, 3H), 1.82 (s, 3H).

N-(5-bromo-4-fluoro-2-methylphenyl)-4-(3,5-dichlorophenylsulfonamido)-2

(hydroxymethyl)benzenesulfonamide (7). Within a flask was dissolved methyl 2-(*N*-(5-bromo-4-fluoro-2-methylphenyl)sulfamoyl)-5-(3,5-dichlorophenylsulfonamid o)benzoate (6, 1 g, 1.59 mmol) in 20 mL of tetrahydrofuran. Then was added Lithium aluminum hydride (242 mg, 6.38 mmol) slowly added at ice bath and the contents were heated at 60 °C for about 12 h. The reaction was allowed to cool to room temperature and poured into ice bath, and adjusted the pH to 2-3 with 3N hydrochloric acid solution. The precipitate formed was filtered and dried to yield crude 7 as a yellow solid without any purification. ¹H NMR (DMSO-d₆, 400 MHz) δ 11.09 (s, 1H), 9.74 (s, 1H), 8.04-8.03 (t, 1H), 7.79-7.78 (d, 2H), 7.63-7.62 (d, *J* = 4.0

Hz, 1H), 7.45-7.43 (d, J = 8.0 Hz, 1H), 7.16-7.06 (m, 3H), 5.58-5.55 (t, 1H), 4.75-4.74 (d, J = 4.0 Hz. 2H), 1.77 (s, 3H). LRMS (ESI-) 594.9 (M-H).

N-(5-bromo-4-fluoro-2-methylphenyl)-2-(chloromethyl)-4-(3,5-dichlorophenylsul fonamido)benzenesulfonamide (8).

N-(5-bromo-4-fluoro-2-methylphenyl)-4-(3,5-dichlorophenylsulfonamido)-2-(hydrox ymethyl)benzenesulfonamide (**7**, 580 mg, 0.969 mmol) was added 10 mL of thionyl chloride slowly at ice bath and the contents were heated at 60 °C for about 10 h. After the removal of the excess reagent, the crude product was recrystallized with ethyl acetate to afford 8 as a white solid (420 mg, two step 43%). ¹H NMR (DMSO-d₆, 400 MHz) δ 11.25 (s, 1H), 9.95 (s, 1H), 8.05-8.04 (t, 1H), 7.81-7.80 (d, 2H), 7.56-7.50 (m, 2H), 7.21-7.09 (m,3H), 4.99 (s, 2H), 1.78 (s, 3H) LRMS (ESI-) 612.8 (M-H).

(*R*)-*N*-(5-bromo-4-fluoro-2-methylphenyl)-4-(3,5-dichlorophenylsulfonamido)-2-((3-methylmorpholino)methyl)benzenesulfonamide (MDL-811).

N-(5-bromo-4-fluoro-2-methylphenyl)-2-(chloromethyl)-4-(3,5-dichlorophenylsulfon amido)benzenesulfonamide (8, 100 mg, 0.162 mmol)was dissolved in 5 mL of dried THF, and then (R)-3-methylmorpholine (33 mg, 0.324 mmol) and TEA (41 mg, 0.405 mmol) was added. The reaction was stirred at 50 °C for about 6 h. After concentration under reduced pressure, the residue was partitioned between ethyl acetate and water. The organic layer was further washed with saturated aq NaHCO₃, brine, dried over sodium sulfate, evaporated in vacuo, and subsequently purified by column chromatography on silica gel (ether: ethyl acetate = 3:1) to afford **MDL-811** as a white solid (80 mg, 73%). ¹H NMR (DMSO-d₆, 400 MHz) δ 11.07 (s, 1H), 9.86 (s, 1H), 8.04 (s, 1H), 7.77 (s, 2H), 7.71 (s, 1H), 7.59-7.57 (d, J = 8.0 Hz, 1H), 7.20-7.11 (m, 2H), 7.01-7.99 (d, J = 8.0 Hz, 1H), 3.98-3.94 (d, J = 16.0 Hz, 1H), 3.69-3.67 (m, 2H), 3.50-3.46 (m, 2H), 3.22-3.17 (m, 1H), 2.51-2.47 (m, 1H), 2.34-2.32 (d, J = 8.0 Hz, 1H), 2.08 (s, 2H), 1.82 (s, 3H), 0.85-0.84 (d, J = 4.0 Hz, 3H). ¹³C NMR (DMSO-d₆ 100 MHz) 156.7, 154.3, 140.9, 140.1, 139.0, 136.4 (*J* = 7), 134.1, 131.9 (*J* (J = 2), 131.7 (J = 5), 130.5 (J = 2), 129.6, 124.1, 118.5, 117.1 (J = 25), 116.4, 103.2 (J = 25), 106.2 (J = 25), 106.2 (J = 25), 106.= 22), 71.0, 65.3, 54.0, 52.6, 49.8, 15.8, 12.4 ppm. LRMS (ESI+) 680.0 (M+H)⁺ HRMS (ESI+) m/z calcd. for $C_{25}H_{25}BrCl_2FN_3O_5S_2$ (M+H)⁺ 679.9780, found 679.9850.

N-(5-bromo-4-fluoro-2-methylphenyl)-4-(3,5-dichlorophenylsulfonamido)-2-(pip erazin-1-ylmethyl)benzenesulfonamide (MDL-812).

N-(5-bromo-4-fluoro-2-methylphenyl)-2-(chloromethyl)-4-(3,5-dichlorophenylsulfon amido)benzenesulfonamide (8, 100 mg, 0.162 mmol) was dissolved in 5 mL of dried THF, and then tert-butyl piperazine-1-carboxylate (60 mg, 0.324 mmol) and TEA (41 mg, 0.405 mmol) was added. The reaction was stirred at 50 °C for about 6 h. After concentration under reduced pressure, the residue was partitioned between ethyl acetate and water. The organic layer was further washed with saturated aq NaHCO₃, brine, dried over sodium sulfate, evaporated in vacuo, then the crude compound was dissolved in 3 mL of DCM and TFA (3 mL) was added at room temperature. The reaction was stirred at the same temperature for another 2 h. Then the solvent was almost moved subsequently out and purified by high performance liquid chromatography to afford MDL-812 as a white solid (51.6 mg, 48%). ¹H NMR (DMSO-d₆, 400 MHz) δ 7.68-7.65 (m, 3H), 7.40-7.38 (d, J = 8.0 Hz, 1H), 7.27 (s, 1H), 7.15-7.13 (d, J = 8.0 Hz, 1H), 6.97-6.95 (d, J = 8.0 Hz, 1H), 6.90-6.89 (d, J = 4.0 Hz, 1H), 3.72 (s, 2H), 3.03-3.00 (m, 4H), 2.56-2.55 (m, 4H), 2.03 (s, 3H). ¹³ C NMR (DMSO-d₆ 100 MHz) 172.1, 157.8, 155.8, 150.7, 148.6, 138.0 (J = 6), 137.2, 134.9, 133.6, 131.5, 130.7 (J = 23), 128.7, 122.5, 118.7, 118.4 (J = 18), 104.4 (J = 18), 59.0, 51.0, 44.3, 17.7 ppm. LRMS (ESI+) 665.0 (M+H)⁺ HRMS (ESI+) m/z calcd. for C₂₄H₂₄BrCl₂FN₄O₄S₂ (M+H)⁺ 664.9783, found 664.9858.

N-(5-bromo-4-fluoro-2-methylphenyl)-2-((butyl(ethyl)amino)methyl)-4-(3,5-dichl orophenylsulfonamido)benzenesulfonamide (MDL-813).

N-(5-bromo-4-fluoro-2-methylphenyl)-2-(chloromethyl)-4-(3,5-dichlorophenylsulfon amido)benzenesulfonamide (8, 100 mg, 0.162 mmol) was dissolved in 5 mL of dried THF, and then N-ethylbutan-1-amine (33 mg, 0.324 mmol) and TEA (41 mg, 0.405 mmol) was added. The reaction was stirred at 50 °C for about 6 h. After concentration under reduced pressure, the residue was partitioned between ethyl acetate and water. The organic layer was further washed with saturated aq NaHCO₃, brine, dried over sodium sulfate. evaporated in vacuo, and subsequently purified bv high performance liquid chromatography to afford MDL-813 as a white solid (56 mg, 51%). ¹H NMR (DMSO-d₆, 400 MHz) δ 7.98 (s, 1H), 7.75-7.74 (d, J = 4.0, 2H), 7.60-7.58 (m, 2H), 7.19-7.11 (m, 2H), 6.95-6.93(d, J = 8.0, 1H) 3.78(s, 2H), 2.47-2.42 (m,4H), 1.87 (s, 3H), 1.34-1.30 (m, 2H), 1.23-1.18 (m, 2H), 0.97-0.93 (m, 3H), 0.84-0.80 (t, 3H). ¹³C NMR (DMSO-d₆ 100 MHz) 157.9, 155.4, 137.6, 137.5, 135.7, 133.4 (J = 3), 133.2, 131.2, 131.0, 125.6, 125.6, 118.7, 118.4 (J = 9), 104.8, 104.6,99.9, 54.8, 52.9, 47.8, 20.4, 19.3, 17.6, 14.3, 11.4 ppm. LRMS (ESI+) 680.02 (M+H)⁺ HRMS (ESI+) m/z calcd. for $C_{26}H_{29}BrCl_2FN_3O_4S_2$ (M+H)⁺ 680.0144, found 680.0216.

2-([1,4'-bipiperidin]-1'-ylmethyl)-*N*-(5-bromo-4-fluoro-2-methylphenyl)-4-(3,5-di chlorophenylsulfonamido)benzenesulfonamide (MDL-814).

N-(5-bromo-4-fluoro-2-methylphenyl)-2-(chloromethyl)-4-(3,5-dichlorophenylsulfon amido)benzenesulfonamide (8, 100 mg, 0.162 mmol)was dissolved in 5 mL of dried THF, and then 1,4'-bipiperidine (55 mg, 0.324 mmol) and TEA (41 mg, 0.405 mmol) was added. The reaction was stirred at 50 °C for about 6 h. After concentration under reduced pressure, the residue was partitioned between ethyl acetate and water. The organic layer was further washed with saturated aq NaHCO₃, brine, dried over sodium sulfate. evaporated in vacuo, and subsequently purified by high performance liquid chromatography to afford MDL-814 as a white solid (55 mg, 46%). ¹H NMR (DMSO-d₆, 400 MHz) δ 7.72-7.71 (t, 1H), 7.64-7.63 (d, 2H), 7.23-7.22 (d, J = 4.0 Hz, 2H), 7.20 (s, 1H), 6.76-6.73 (m, 1H), 6.69-6.67 (d, J = 8.0Hz, 1H), 3.64 (s, 2H), 2.84-2.81 (m, 6H), 2.03 (s, 3H), 1.87-1.85 (d, J = 8.0 Hz, 2H), 1.62-1.58 (m, 4H), 1.44-1.38 (m, 4H). ¹³C NMR (DMSO-d₆ 100 MHz) 157.4, 154.9, 148.6, 137.9 (J = 8), 136.2, 134.1, 133.1, 130.6, 130.2, 129.6, 124.7, 122.1, 118.2, 128.1,117.9, 103.9, 103.7, 61.8, 58.9, 51.9, 49.4, 26.6, 24.1, 22.6, 17.3 ppm. LRMS (ESI+) 747.02 $(M+H)^+$ HRMS (ESI+) m/z calcd. for C₃₀H₃₄BrCl₂FN₄O₄S₂ $(M+H)^+$ 747.0637. N-(5-bromo-4-fluoro-2-methylphenyl)-4-(3,5-dichlorophenylsulfonamido)-2-((4-(3-methoxypropyl)piperazin-1-yl)methyl)benzenesulfonamide (MDL-815).

N-(5-bromo-4-fluoro-2-methylphenyl)-2-(chloromethyl)-4-(3,5-dichlorophenylsulfon amido)benzenesulfonamide (**8**, 100 mg, 0.162 mmol)was dissolved in 5 mL of dried THF, and then 1-(3-methoxypropyl)piperazine (51 mg, 0.324 mmol) and TEA (41 mg, 0.405 mmol) was added. The reaction was stirred at 50 °C for about 6 h. After concentration under reduced pressure, the residue was partitioned between ethyl acetate and water. The organic layer was further washed with saturated aq NaHCO₃, brine, dried over sodium sulfate, evaporated in vacuo, and subsequently purified by high performance liquid chromatography to afford **MDL-815** as a white solid (57 mg, 48%). ¹H NMR (DMSO-d₆, 400 MHz) δ 7.79 (s, 1H), 7.68 (s, 2H), 7.30-7.21 (m, 3H), 6.86-6.84 (d, *J* = 8.0, 1H), 6.68-6.66 (d, *J* = 8.0, 1H), 3.67 (s, 2H), 3.32-3.30 (4, 2H), 3.2 (s, 3H), 2.39-2.31(m, 8H), 2.01 (s, 3H), 1.65-1.62(m, 2H). ¹³C NMR (DMSO-d₆ 100 MHz) ¹³C NMR (100 MHz, DMSO) δ 157.6, 155.1, 146.9, 138.1 (*J* = 7), 136.5, 134.4, 132.7, 130.9, 130.5, 130.3, 127.6, 124.8, 121.9, 118.2, 117.9, 103.9 (*J* = 22), 69.9, 59.0, 57.8, 54.4, 52.4, 52.3, 26.2, 17.3 ppm. LRMS (ESI+) 737.04 (M+H)⁺ HRMS (ESI+) m/z calcd. for C₂₈H₃₂BrCl₂FN₄O₅S₂ (M+H)⁺ 737.0427.

N-(5-bromo-4-fluoro-2-methylphenyl)-2-((4-butylpiperazin-1-yl)methyl)-4-(3,5-di chlorophenylsulfonamido)benzenesulfonamide (MDL-816).

N-(5-bromo-4-fluoro-2-methylphenyl)-2-(chloromethyl)-4-(3,5-dichlorophenylsulfon amido) benzenesulfonamide (8, 100 mg, 0.162 mmol) was dissolved in 5 mL of dried THF, and then 1-butylpiperazine (46 mg, 0.324 mmol) and TEA (41 mg, 0.405 mmol) was added. The reaction was stirred at 50 °C for about 6 h. After concentration under reduced pressure, the residue was partitioned between ethyl acetate and water. The organic layer was further washed with saturated aq NaHCO₃, brine, dried over sodium sulfate. evaporated in vacuo, and subsequently purified by high performance liquid chromatography to afford MDL-816 as a white solid (50 mg. 43%). ¹H NMR (DMSO-d₆, 400 MHz) δ 7.87-7.86 (t, 1H), 7.73-7.72 (d, 2H), 7.40-7.36 (m, 2H), 7.19-7.17 (d, J = 8.0 Hz, 1H), 6.98-6.95 (m, 1H), 6.84-6.82 (d, J =8.0 Hz, 1H), 3.69 (s, 2H), 2.70-2.52 (m, 4H), 2.51-2.44 (m, 4H), 1.90 (s, 1H), 1.47-1.41 (m, 2H), 1.30-1.20 (m, 3H), 0.93-0.83 (m, 4H). ¹³C NMR (DMSO-d₆ 100 MHz) 164.4, 155.8, 145.5, 138.3 (J = 5), 137.8, 135.3, 132.9 (J = 3), 131.9 (J = 22), 131.1, 125.6, 121.5, 118.7, 118.5, 118.3, 104.6 (*J* = 23), 100.0, 58.7, 57.1, 52.6, 51.9, 29.5, 27.8, 20.3, 19.3, 17.7, 14.2 ppm. LRMS (ESI+) 721.1 (M+H)⁺ HRMS (ESI+) m/z calcd. for C₂₈H₃₂BrCl₂FN₄O₄S₂ (M+H)⁺ 721.0482.

N-(5-bromo-4-fluoro-2-methylphenyl)-4-(3,5-dichlorophenylsulfonamido)-2-((3-(dimethylamino) pyrrolidin-1-yl) methyl) benzenesulfonamide (MDL-817). *N*-(5-bromo-4-fluoro-2-methylphenyl)-2-(chloromethyl)-4-(3,5-dichlorophenylsulfon amido) benzenesulfonamide (8, 100 mg, 0.162 mmol) was dissolved in 5 mL of dried THF, and then *N*,*N*-dimethylpyrrolidin-3-amine (37 mg, 0.324 mmol) and TEA (41 mg, 0.405 mmol) was added. The reaction was stirred at 50 °C for about 6 h. After concentration under reduced pressure, the residue was partitioned between ethyl acetate and water. The organic layer was further washed with saturated aq NaHCO₃, brine, dried over sodium sulfate, evaporated in vacuo, and subsequently purified by high performance liquid chromatography to afford **MDL-817** as a white solid (53 mg, 48%). ¹H NMR (DMSO-d₆, 400 MHz) δ 7.81-7.80 (t, 1H), 7.69-7.68 (d, 2H), 7.36-7.34 (d, J = 8.0, 1H), 7.24-7.17 (m, 2H), 6.92-6.87 (m, 2H), 3.82-3.71 (m, 2H), 3.29-3.19 (m, 1H), 2.69-2.59 (m, 2H), 2.55-2.52 (m, 1H), 2.42-2.36 (m, 7H), 2.01-1.93 (m, 4H), 1.79-1.74 (m, 1H). ¹³C NMR (DMSO-d₆ 100 MHz) 157.4, 154.9, 146.7, 137.7, 137.4 (J = 8), 134.4, 132.7, 131.0, 130.7, 130.3, 127.9, 124.8, 120.9, 117.9 (J = 17), 117.7, 104.9 (J = 22), 64.3, 56.0, 55.9, 52.2, 41.7, 26.9, 17.1 ppm. LRMS (ESI+) 693.06 (M+H)⁺ HRMS (ESI+) m/z calcd. for C₂₆H₂₈BrCl₂FN₄O₄S₂ (M+H)⁺ 693.0167.

N-(5-bromo-4-fluoro-2-methylphenyl)-4-(3,5-dichlorophenylsulfonamido)-2-((di methylamino)methyl)benzenesulfonamide (MDL-818).

N-(5-bromo-4-fluoro-2-methylphenyl)-2-(chloromethyl)-4-(3,5-dichlorophenylsulfon amido) benzenesulfonamide (**8**, 100 mg, 0.162 mmol) was dissolved in 5 mL dried THF, and then dimethylamine absolute in tetrahydrofuran solution (2 M, 0.245 mL) was added. The reaction was stirred at 50 °C for about 6 h. After concentration under reduced pressure, the residue was partitioned between ethyl acetate and water. The organic layer was further washed with saturated aq NaHCO₃, brine, dried over sodium sulfate, evaporated in vacuo, and subsequently purified by column chromatography on silica gel (petroleum ether: ethyl acetate = 2:1) to afford **MDL-818** as a white solid (66 mg, 65%).¹H NMR (DMSO-*d*₆, 500 MHz) δ 7.92 (s, 1H), 7.76-7.75 (d, 2H), 7.49-7.48 (d, *J* = 5.0, 1H), 7.33 (s, 1H), 7.20-7.18 (d, *J* = 10.0, 1H), 7.10-7.07 (m, 1H), 6.91-6.89 (d, *J* = 10.0, 1H) 3.84 (s, 2H), 2.31 (s, 6H), 1.90 (s, 3H). ¹³C NMR (DMSO-*d*₆, 125 MHz) 156.1 (*J* = 242.5), 136.96 (*J* = 9), 134.91, 132.96, 132.08, 130.57, 130.42, 129.55, 125.08, 122.09, 118.55, 118.03 (*J* = 22.5), 104.02 (*J* = 22.5), 59.62, 43.95, 17.09 ppm. LRMS (ESI+) 624.0 (M+H)⁺, HRMS (ESI+) m/z calcd. for C₂₂H₂₂BrCl₂FN₃O₄S₂⁺(M+H)⁺623.9591, found 623.9585.

N-(5-bromo-4-fluoro-2-methylphenyl)-4-(3,5-dichlorophenylsulfonamido)-2-((eth yl(methyl)amino)methyl)benzenesulfonamide (MDL-819).

N-(5-bromo-4-fluoro-2-methylphenyl)-2-(chloromethyl)-4-(3,5-dichlorophenylsulfon amido) benzenesulfonamide (**8**, 100 mg, 0.162 mmol) was dissolved in 5 mL of dried THF, and then N-methylethanamine (29 mg, 0.486 mmol) was added. The reaction was stirred at 50 ° C for about 6 h. After concentration under reduced pressure, the residue was partitioned between ethyl acetate and water. The organic layer was further washed with saturated aq NaHCO₃, brine, dried over sodium sulfate, evaporated in vacuo, and subsequently purified by column chromatography on silica gel (petroleum ether: ethyl acetate = 2:1) to afford **MDL-819** as a white solid (78 mg, 76%). ¹H NMR (DMSO-*d*₆, 500 MHz) δ 7.65-7.62 (m, 3H), 7.23-7.21 (m, 2H), 7.00-6.99 (d, *J* = 5.0, 1H), 6.74-6.72 (m, 1H), 6.67-6.65 (d, *J* = 10.0, 1H), 3.70 (s, 2H), 2.47-2.43 (m, 2H), 2.11-2.10 (d, 6H), 0.98-0.95 (t, 3H). ¹³C NMR (DMSO-*d*₆ 125 MHz) 156.16(*J* = 241.3), 150.45, 137.53, 136.14, 134.45, 139.94 (*J* = 87.5), 129.41, 125.17, 124.16, 118.64, 118.44, 104.22 (*J* = 22.5), 60.06, 51.00, 41.04, 18.09, 11.87 ppm. LRMS (ESI+) 638.0 (M+H)⁺, HRMS (ESI+) m/z calcd. for C₂₃H₂₄BrCl₂FN₃O₄S₂⁺ (M+H)⁺ 637.9747, found 637.9742.

N-(5-bromo-4-fluoro-2-methylphenyl)-4-(3,5-dichlorophenylsulfonamido)-2-((methyl(propyl)amino)methyl)benzenesulfonamide(MDL-820).

N-(5-bromo-4-fluoro-2-methylphenyl)-2-(chloromethyl)-4-(3,5-dichlorophenylsulfon

amido) benzenesulfonamide (**8**, 100 mg, 0.162 mmol) was dissolved in 5 mL of dried THF, and then N-methylpropan-1-amine (36 mg, 0.486 mmol) was added. The reaction was stirred at 50 °C for about 6 h. After concentration under reduced pressure, the residue was partitioned between dichloromethane and water. The organic layer was further washed with saturated aq NaHCO₃, brine, dried over sodium sulfate, evaporated in vacuo, and subsequently purified by column chromatography on silica gel(petroleum ether: ethyl acetate = 2:1) to afford **MDL-820** as a white solid (76 mg, 72%). ¹H NMR (DMSO-*d*₆, 500 MHz) δ 7.96 (s, 1H), 7.76-7.75 (d, 2H), 7.55-7.53 (d, *J* = 10.0, 1H), 7.46 (s, 1H), 7.20-7.18 (d, *J* = 10.0, 1H), 7.13-7.11 (m, 1H), 6.90-6.89 (d, *J* = 5.0, 1H), 3.78 (s, 2H), 2.42-2.41 (m, 2H), 2.15 (s, 3H), 1.89 (s, 3H), 1.48-1.43 (m, 2H), 0.85-0.82 (t, 3H). ¹³C NMR (DMSO-*d*₆ 125 MHz) 156.74 (*J* = 242.5), 143.64, 137.81, 137.75, 135.58, 133.12 (*J* = 40), 131.25, 131.09, 125.66, 121.83, 118.64 (*J* = 22.5), 104.64 (*J* = 21.3), 59.30, 58.42, 41.60, 19.70, 17.62, 11.94 ppm. LRMS (ESI+) 651.9 (M+H)⁺, HRMS (ESI+) m/z calcd. for C₂₄H₂₆BrCl₂FN₃O₄S₂⁺ (M+H)⁺ 651.9904, found 651.9904.

N-(5-bromo-4-fluoro-2-methylphenyl)-4-(3,5-dichlorophenylsulfonamido)-2-(pip eridin-1-ylmethyl)benzenesulfonamide (MDL-821).

N-(5-bromo-4-fluoro-2-methylphenyl)-2-(chloromethyl)-4-(3,5-dichlorophenylsulfon amido) benzenesulfonamide (**8**, 100 mg, 0.162 mmol) was dissolved in 5 mL of dried THF, and then piperidine (42 mg, 0.486 mmol) was added. The reaction was stirred at 50 °C for about 6 h. After concentration under reduced pressure, the residue was partitioned between ethyl acetate and water. The organic layer was further washed with saturated aq NaHCO₃, brine, dried over sodium sulfate, evaporated in vacuo, and subsequently purified by column chromatography on silica gel (petroleum ether: ethyl acetate = 2:1) to afford **MDL-821** as a white solid (69 mg, 64%). ¹H NMR (DMSO-*d*₆, 500 MHz) δ 7.99 (s, 1H), 7.78-7.77 (d, 2H), 7.49-7.45 (m, 2H), 7.22-7.20 (d, *J* = 10.0, 1H), 7.11-7.09 (m, 1H), 6.71 (s, 1H), 3.79 (s, 2H), 2.44(s, 4H), 1.93 (s, 3H), 1.50-1.42 (m, 6H). ¹³C NMR (DMSO-*d*₆ 125 MHz) 156.98 (*J* = 243.8) 138.61 (*J* = 9.0), 135.63, 133.04, 131.51, 131.22, 125.70, 122.01, 118.75 (*J* = 22.5), 104.54 (*J* = 21.3), 59.63, 54.07, 25.40, 23.57, 17.71 ppm. LRMS (ESI+) 664.0 (M+H)⁺, HRMS (ESI+) m/z calcd. for C₂₅H₂₆BrCl₂FN₃O₄S⁺ (M+H)⁺ 663.9904, found 663.9900.

N-(5-bromo-4-fluoro-2-methylphenyl)-4-(3,5-dichlorophenylsulfonamido)-2-((2-(hydroxymethyl)pyrrolidin-1-yl)methyl)benzenesulfonamide (MDL-822). *N*-(5-bromo-4-fluoro-2-methylphenyl)-2-(chloromethyl)-4-(3,5-dichlorophenylsulfon amido) benzenesulfonamide (**8**, 100 mg, 0.162 mmol) was dissolved in 5 mL of dried THF, and then piperidine (49 mg, 0.486 mmol) was added. The reaction was stirred at 50 °C for about 6 h. After concentration under reduced pressure, the residue was partitioned between ethyl acetate and water. The organic layer was further washed with saturated aq NaHCO₃, brine, dried over sodium sulfate, evaporated in vacuo, and subsequently purified by column chromatography on silica gel (petroleum ether: ethyl acetate = 2:1) to afford **MDL-822** as a white solid (66 mg, 60%). ¹H NMR (DMSO-*d*₆, 500 MHz) δ 7.98 (s, 1H), 7.77-7.76 (d, 2H), 7.50-7.48 (d, *J* = 10.0, 1H), 7.44 (s, 1H), 7.20-7.18 (d, *J* = 10.0, 1H), 7.08-7.06 (m, 1H), 6.94-6.93 (d, *J* = 5.0, 1H), 4.49-4.46 (d, *J* = 15.0, 1H), 3.62-3.60 (m, 2H), 3.49-3.43 (m, 3H), 2.75-2.60 (m, 2H), 2.18-2.13 (m, 1H), 1.90-1.88 (m, 4H), 1.72-1.57 (m, 2H). ¹³C NMR (DMSO- d_6 125 MHz) 156.27 (J = 242.5), 137.79 (J = 9), 135.07, 132.51, 131.13, 130.72, 125.12, 121.41, 118.12 (J = 22.5), 104.96 (J = 22.5), 65.55, 62.20, 55.47, 53.76, 33.19, 27.13, 22.43, 17.12ppm. LRMS (ESI+) 680.0 (M+H)⁺, HRMS (ESI+) m/z calcd. for C₂₅H₂₆BrCl₂FN₃O₅S₂⁺ (M+H)⁺ 679.9853, found 679.9841.

NMR spectra of compounds MDL analogs N-(5-bromo-4-fluoro-2-methylphenyl)-4-(3,5-dichlorophenylsulfonamido)-2-(hyd roxymethyl)benzenesulfonamide (7)





N-(5-bromo-4-fluoro-2-methylphenyl)-2-(chloromethyl)-4-(3,5-dichlorophenylsul fonamido)benzenesulfonamide (8)



10.5









MDL-814













MDL-818



11

MDL-819








Relate to "Synthesis of compounds MDL analogs". NMR spectra of compounds MDL analogs were shown.

HPLC analysis data of compounds MDL analogs

HPLC analysis data of compounds MDL analogs. The purities of identified compounds that were essential to the conclusions drawn in the text and determined by one standard instrumentation with one system given in the following table. The peak purity was checked with UV spectra.

		Method		
Equ	iipment	Agilent 1260 with quaternary pump, photodiode array detector (DAD)		
Co	olumn	Agilent Zorbax Exlipse Plus C18 (100×4.6 mm, 3.5 µm		
		particle size)		
System	n condition	a.	b.	
		CH ₃ CN(0.1% TFA)/H ₂ O	CH ₃ CN(0.1% T	FA)/H ₂ O
		(0.1% TFA), 10% (v/v) of	(0.1% TFA), 10	% (v/v) of
		$CH_3CN(0.1\% \text{ TFA})$ at the	CH ₃ CN(0.1% T	FA) at the
		first time. Next, percentage of	first time. Next,	percentage of
		CH ₃ CN(0.1% TFA) was	CH ₃ CN(0.1% T	TFA) was
		slowly increased to 100% in	slowly increase	d to 100% in
		15 min and the condition was	15 min and the	condition was
		maintained at 5 min, flow	maintained at 5	min, flow
		rate: 1.0 mL/min, calculated	rate: 1.0 mL/mi	n, calculated
		the relative purity of each	the relative puri	ty of each
		compound at 254 nM.	compound at 28	30 nM.
		С.	d.	
		CH ₃ CN(0.1% TFA)/H ₂ O	CH ₃ CN(0.1% T	TFA)/H ₂ O
		(0.1% TFA), 10% (v/v) of	(0.1% TFA), 10	% (v/v) of
		$CH_3CN(0.1\% \text{ TFA})$ at the	CH ₃ CN(0.1% T	FA) at the
		first time. Next, percentage of	first time. Next,	percentage of
		CH ₃ CN(0.1% TFA) was	CH ₃ CN(0.1% T	TFA) was
		slowly increased to 100% in	slowly increase	d to 100% in
		15 min and the condition was	15 min and the	condition was
		maintained at 5 min, flow	maintained at 5	min, flow
		rate: 1.2 mL/min, calculated	rate: 1.2 mL/mi	n, calculated
		the relative purity of each	the relative puri	ty of each
		compound at 254 nM.	compound at 28	30 nM.
Result	Compound	Retention time (min)	Relactive	purity (%)
			254 nM	280 nM
	811	15.132	^a 98.28	^b 98.48
	812	13.736	^a 99.51	^b 99.82
	813	16.439	^a 98.53	^b 98.42
	814	13.361	^a 96.00	^b 95.92
	815	15.186	^a 99.55	^b 99.47
	816	15.963	^a 98.00	^b 98.35

817	13.224	^a 95.09	^b 95.31
818	11.772	°98.86	^d 99.41
819	12.009	°99.38	^d 99.43
820	12.483	°98.76	^d 99.18
821	12.474	°99.58	^d 99.57
822	10.483	°97.78	^d 99.22

HPLC UV spectra data of purity of compounds MDL analogs.

MDL-811

Data File C:\CHEM32\1\DATA\FCOMPOUND\TEST 2019-05-20 16-17-05\811.D Sample Name: 811

Acq. Operator : SYSTEM	Seq. Line : 2
Acq. Instrument : 12601c	Location : Vial 25
Injection Date : 5/20/2019 4:40:57 FM	Inj Volume : 2.000 µl
Different Inj Volume from Sample Entry! Actu	al Inj Volume : 5.000 µl
Method : C:\CHEM32\1\DATA\FCOMPOUND\	TEST 2019-05-20 16-17-05\TEST12.M (Sequence
Method) Last changed • 5/20/2019 4.17.05 PM by SVS	TEM
Additional Info : Peak(s) manually integrated	114
DAD1 A, Sig=254,4 Ref=550,100 (FCOMPOUND\TEST 2019-05	-20 16-17-05\811.D)
mAU	-132
	τρ T
500 -	
400	
400 -	
300 -	
-	
200 -	
-	
100 -	
	4.250
0	
2.5 5 7.5	10 12.5 15 17.5 11
Area Percent Report	
Sorted By : Signal	
Multiplier : 1.0000	
Dilution : 1.0000	
Use Multiplier & Dilution factor with isibs	
Signal 1: DAD1 A, Sig=254,4 Ref=550,100	
Dook DotTimo Timo Width Aroo Hoight	A rco.
# [min] [min] [mAU*s] [mAU]	%
1 14.250 BB 0.0551 44.22514 12.393	42 1.7187
2 15.132 VB 0.0705 2528.98633 550.993	41 98.2813
Totals: 2573.21146 563.386	83
*** End of Report *	

12601c 11/21/2019 4:39:34 PM SYSTEM

Data File C:\CHEM32\1\DATA\FCOMPOUND\TEST 2019-05-20 16-17-05\811.D Sample Name: 811



12601c 11/21/2019 4:39:56 PM SYSTEM

Data File C:\CHEM32\1\DATA\FCOMPOUND\TEST 2019-05-20 16-17-05\812.D Sample Name: 812

Acq. Operator : SYSTEM Seq. Line : 3 Acq. Instrument : 12601c Location : Vial 26 Injection Date : 5/20/2019 5:02:27 PM Inj : 1 Inj Volume : 2.000 µl Different Inj Volume from Sample Entry! Actual Inj Volume : 10.000 µl : C:\CHEM32\1\DATA\FCOMPOUND\TEST 2019-05-20 16-17-05\TEST12.M (Sequence Method Method) : 5/20/2019 4:17:05 PM by SYSTEM Last changed Additional Info : Peak(s) manually integrated DAD1 A, Sig=254,4 Ref=550,100 (FCOMPOUND\TEST 2019-05-20 16-17-05\812.D) mAU 13.736 1000 800 600 400 200 14.782 16.734 13.174 0 2.5 5 7.5 10 12.5 15 17.5 min _____ Area Percent Report Signal Sorted By : Multiplier : 1.0000 Dilution 1.0000 : Use Multiplier & Dilution Factor with ISTDs Signal 1: DAD1 A, Sig=254,4 Ref=550,100 Peak RetTime Type Width Area Height Area # [min] [min] [mAU*s] [mAU] S ----|-----|-----|-----| 1 13.174 BB 0.0547 6.77659 1.87163 0.1058
 2
 13.736 VB
 0.0822 6374.10400 1186.53284 99.5194

 3
 14.782 BB
 0.0595 10.87952 2.78801 0.1699

 4
 16.734 BB
 0.1150 13.12411 1.73899 0.2049
Totals : 6404.88423 1192.93146

12601c 11/21/2019 4:40:19 PM SYSTEM

Data File C:\CHEM32\1\DATA\FCOMPOUND\TEST 2019-05-20 16-17-05\812.D Sample Name: 812



12601c 11/21/2019 4:41:02 PM SYSTEM

Data File C:\CHEM32\1\DATA\FCOMPOUND\TEST 2019-05-20 16-17-05\813.D Sample Name: 813

Acq. Operator : SYSTEM Seq. Line : 4 Acq. Instrument : 12601c Location : Vial 27 Injection Date : 5/20/2019 5:24:16 PM Inj : 1 Inj Volume : 2.000 µl Different Inj Volume from Sample Entry! Actual Inj Volume : 8.000 µl : C:\CHEM32\1\DATA\FCOMPOUND\TEST 2019-05-20 16-17-05\TEST12.M (Sequence Method Method) : 5/20/2019 4:17:05 PM by SYSTEM Last changed Additional Info : Peak(s) manually integrated DAD1 A, Sig=254,4 Ref=550,100 (FCOMPOUND\TEST 2019-05-20 16-17-05\813.D) 16.439 mAU 600 500 400 300 200 100 15.620 5.829 0 4 6 10 12 14 16 18 2 8 min _____ Area Percent Report Signal Sorted By : Multiplier : 1.0000 Dilution 1.0000 : Use Multiplier & Dilution Factor with ISTDs Signal 1: DAD1 A, Sig=254,4 Ref=550,100 Peak RetTime Type Width Area Height Area # [min] [min] [mAU*s] [mAU] S ----|-----|-----|-----| 1 5.829 BB 0.1166 11.75607 1.54736 0.3027 2 15.620 VB 0.0544 45.33770 12.93765 1.1674 3 16.439 BB 0.0856 3826.52124 691.46991 98.5299 Totals : 3883.61501 705.95492 _____

12601c 11/21/2019 4:41:26 PM SYSTEM

Data File C:\CHEM32\1\DATA\FCOMPOUND\TEST 2019-05-20 16-17-05\813.D Sample Name: 813



12601c 11/21/2019 4:41:40 PM SYSTEM

Data File C:\CHEM32\1\DATA\FCOMPOUND\TEST 2019-05-20 16-17-05\814.D Sample Name: 814

Acq. Operator : SYSTEM Seq. Line : 5 Acq. Instrument : 12601c Location : Vial 28 Injection Date : 5/20/2019 5:46:06 PM Inj : 1 Inj Volume : 2.000 µl Different Inj Volume from Sample Entry! Actual Inj Volume : 8.000 µl : C:\CHEM32\1\DATA\FCOMPOUND\TEST 2019-05-20 16-17-05\TEST12.M (Sequence Method Method) : 5/20/2019 4:17:05 PM by SYSTEM Last changed Additional Info : Peak(s) manually integrated DAD1 A, Sig=254,4 Ref=550,100 (FCOMPOUND\TEST 2019-05-20 16-17-05\814.D) mAU 13.361 700 600 500 400 300 200 12.550 100 13.258 0 2.5 7.5 10 12.5 15 17.5 5 mir _____ Area Percent Report Signal Sorted By : Multiplier : 1.0000 Dilution 1.0000 : Use Multiplier & Dilution Factor with ISTDs Signal 1: DAD1 A, Sig=254,4 Ref=550,100 Peak RetTime Type Width Area Height Area # [min] [min] [mAU*s] [mAU] S ----|-----|-----|-----| 1 12.550 BB 0.0480 148.06168 47.30540 3.8326 2 13.258 BV 0.0507 6.27082 1.86637 0.1623 3 13.361 VB 0.0749 3708.91333 760.61719 96.0051 Totals : 3863.24583 809.78896 _____ _____

12601c 11/21/2019 4:42:03 PM SYSTEM

Data File C:\CHEM32\1\DATA\FCOMPOUND\TEST 2019-05-20 16-17-05\814.D Sample Name: 814



12601c 11/21/2019 4:42:22 PM SYSTEM

Data File C:\CHEM32\1\DATA\FCOMPOUND\TEST 2019-05-20 16-17-05\815.D Sample Name: 815

Acq. Operator : SYSTEM Seq. Line : 6 Acq. Instrument : 12601c Location : Vial 29 Injection Date : 5/20/2019 6:07:55 PM Inj : 1 Inj Volume : 2.000 µl Different Inj Volume from Sample Entry! Actual Inj Volume : 10.000 µl : C:\CHEM32\1\DATA\FCOMPOUND\TEST 2019-05-20 16-17-05\TEST12.M (Sequence Method Method) Last changed : 5/20/2019 4:17:05 PM by SYSTEM Additional Info : Peak(s) manually integrated DAD1 A, Sig=254,4 Ref=550,100 (FCOMPOUND\TEST 2019-05-20 16-17-05\815.D) mAU 186 цĠ 400 300 200 100 5.816 0 2.5 7.5 10 12.5 15 17.5 5 min _____ Area Percent Report _____ Signal 1.0000 Sorted By : Multiplier : Dilution 1.0000 : Use Multiplier & Dilution Factor with ISTDs Signal 1: DAD1 A, Sig=254,4 Ref=550,100 Height Area Peak RetTime Type Width Area # [min] [min] [mAU*s] [mAU] % 1 5.816 BB 0.1004 9.23906 1.18561 0.4516 2 15.186 BB 0.0677 2036.49268 464.31644 99.5484 Totals : 2045.73174 465.50205 _____ _____

*** End of Report ***

12601c 11/21/2019 4:42:46 PM SYSTEM

Data File C:\CHEM32\1\DATA\FCOMPOUND\TEST 2019-05-20 16-17-05\815.D Sample Name: 815



12601c 11/21/2019 4:43:08 PM SYSTEM

Data File C:\CHEM32\1\DATA\FCOMPOUND\TEST 2019-05-20 16-17-05\816.D Sample Name: 816

Acq. Operator : SYSTEM Seq. Line : 7 Acq. Instrument : 12601c Location : Vial 30 Injection Date : 5/20/2019 6:29:46 PM Inj : 1 Inj Volume : 2.000 µl Different Inj Volume from Sample Entry! Actual Inj Volume : 6.000 µl : C:\CHEM32\1\DATA\FCOMPOUND\TEST 2019-05-20 16-17-05\TEST12.M (Sequence Method Method) : 5/20/2019 4:17:05 PM by SYSTEM Last changed Additional Info : Peak(s) manually integrated DAD1 A, Sig=254,4 Ref=550,100 (FCOMPOUND\TEST 2019-05-20 16-17-05\816.D) mAU 15.963 350 300 250 200 150 100 50 14.995 16.558 0 2.5 5 7.5 10 12.5 15 17.5 min _____ Area Percent Report _____ Signal Sorted By : Multiplier : 1.0000 Dilution 1.0000 : Use Multiplier & Dilution Factor with ISTDs Signal 1: DAD1 A, Sig=254,4 Ref=550,100 Peak RetTime Type Width Area Height Area # [min] [min] [mAU*s] [mAU] S ----|-----|-----|-----| 1 14.995 BB 0.0542 28.12886 8.06285 1.6854 2 15.963 BB 0.0668 1635.56042 375.72974 97.9955 3 16.558 BB 0.0608 5.32693 1.30085 0.3192 Totals : 1669.01622 385.09344 _____ _____

12601c 11/21/2019 4:43:28 PM SYSTEM

Data File C:\CHEM32\1\DATA\FCOMPOUND\TEST 2019-05-20 16-17-05\816.D Sample Name: 816



12601c 11/21/2019 4:43:41 PM SYSTEM

Data File C:\CHEM32\1\DATA\FCOMPOUND\TEST 2019-05-20 16-17-05\817.D Sample Name: 817

Acq. Operator : SYSTEM Seq. Line : 8 Acq. Instrument : 12601c Location : Vial 31 Injection Date : 5/20/2019 6:51:36 PM Inj : 1 Inj Volume : 2.000 µl Different Inj Volume from Sample Entry! Actual Inj Volume : 6.000 µl : C:\CHEM32\1\DATA\FCOMPOUND\TEST 2019-05-20 16-17-05\TEST12.M (Sequence Method Method) Last changed : 5/20/2019 4:17:05 PM by SYSTEM Additional Info : Peak(s) manually integrated DAD1 A, Sig=254,4 Ref=550,100 (FCOMPOUND\TEST 2019-05-20 16-17-05\817.D) mAU 13.224 350 300 250 200 150 100 50 0 2.5 7.5 10 12.5 15 17.5 5 mir _____ Area Percent Report Signal Sorted By : Multiplier : 1.0000 Dilution 1.0000 : Use Multiplier & Dilution Factor with ISTDs Signal 1: DAD1 A, Sig=254,4 Ref=550,100 Peak RetTime Type Width Area Height Area 90 # [min] [min] [mAU*s] [mAU] ----|-----|-----|-----| 1 12.311 MM 0.0636 56.24012 14.74519 4.0391 2 13.097 VV 0.0638 12.09847 2.77612 0.8689 3 13.224 VB 0.0552 1324.04688 370.23776 95.0920 Totals : 1392.38546 387.75907 _____

12601c 11/22/2019 4:36:09 PM SYSTEM

Data File C:\CHEM32\1\DATA\FCOMPOUND\TEST 2019-05-20 16-17-05\817.D Sample Name: 817



12601c 11/22/2019 4:36:32 PM SYSTEM

Data File C:\CHEM32\1\DATA\FCOMPOUND\TEST 2020-03-03 11-11-11\ME-ME.D Sample Name: Me-Me

Acq. Operator : SYSTEM Seq. Line : 5 Acq. Instrument : 12601c Location : Vial 65 Injection Date : 3/3/2020 12:40:06 PM Inj : 1 Inj Volume : 2.000 µl Different Inj Volume from Sample Entry! Actual Inj Volume : 6.000 µl : C:\CHEM32\1\DATA\FCOMPOUND\TEST 2020-03-03 11-11-11\TEST12.M (Sequence Method Method) Last changed : 3/3/2020 11:11:11 AM by SYSTEM Additional Info : Peak(s) manually integrated DAD1 A, Sig=254,4 Ref=550,100 (FCOMPOUND\TEST 2020-03-03 11-11-11\ME-ME.D) mAU 44.772 200 175 150 125 100 75 50 25 11.068 16.620 0 12.5 15 17.5 2.5 5 7.5 10 min _____ Area Percent Report Signal Sorted Bv : Multiplier : 1.0000 : 1.0000 Dilution Use Multiplier & Dilution Factor with ISTDs Signal 1: DAD1 A, Sig=254,4 Ref=550,100 Peak RetTime Type Width Area Height Area # [min] [min] [mAU*s] [mAU] % 1 11.068 BB 0.0647 11.79943 2.71694 0.6496 2 11.772 BB 0.1294 1795.72339 224.37698 98.8628 3 16.620 VB 0.0917 8.85562 1.42138 0.4875 1816.37844 228.51530 Totals :

12601c 3/26/2020 9:46:02 AM SYSTEM

Data File C:\CHEM32\1\DATA\FCOMPOUND\TEST 2020-03-03 11-11-11\ME-ME.D Sample Name: Me-Me



12601c 3/26/2020 9:46:25 AM SYSTEM

Data File C:\CHEM32\1\DATA\FCOMPOUND\TEST 2020-03-03 11-11-11\ME-ET.D Sample Name: Me-Et

_____ Acq. Operator : SYSTEM Seq. Line : 6 Location : Vial 66 Acq. Instrument : 12601c Injection Date : 3/3/2020 1:01:53 PM Inj : 1 _ . _ Inj Volume : 2.000 µl Different Inj Volume from Sample Entry! Actual Inj Volume : 6.000 µl Method : C:\CHEM32\1\DATA\FCOMPOUND\TEST 2020-03-03 11-11-11\TEST12.M (Sequence Method) : 3/3/2020 11:11:11 AM by SYSTEM Last changed Additional Info : Peak(s) manually integrated DAD1 A, Sig=254,4 Ref=550,100 (FCOMPOUND\TEST 2020-03-03 11-11-11\ME-ET.D) mAU 12.009 200 150 100 50 11.346 0 2.5 5 7.5 10 12.5 15 17.5 min ------Area Percent Report _____ _____ Signal : Sorted Bv : 1.0000 : 1.0000 Multiplier Dilution Use Multiplier & Dilution Factor with ISTDs Signal 1: DAD1 A, Sig=254,4 Ref=550,100 Peak RetTime Type Width Area Height Area # [min] [min] [mAU*s] [mAU] 응 1 11.346 BB 0.0637 12.47002 2.95845 0.6193 2 12.009 BB 0.1371 2001.16431 234.39781 99.3807 2013.63433 237.35626 Totals : *** End of Report ***

12601c 3/26/2020 9:46:36 AM SYSTEM

Data File C:\CHEM32\1\DATA\FCOMPOUND\TEST 2020-03-03 11-11-11\ME-ET.D Sample Name: Me-Et



12601c 3/26/2020 9:46:48 AM SYSTEM

Data File C:\CHEM32\1\DATA\FCOMPOUND\TEST 2020-03-03 11-11-11\ME-PR.D Sample Name: Me-Pr

_____ Acq. Operator : SYSTEM Seq. Line : 7 Location : Vial 67 Acq. Instrument : 12601c Injection Date : 3/3/2020 1:23:40 PM Inj : 1 Inj Volume : 2.000 µl Different Inj Volume from Sample Entry! Actual Inj Volume : 6.000 µl : C:\CHEM32\1\DATA\FCOMPOUND\TEST 2020-03-03 11-11-11\TEST12.M (Sequence Method Method) : 3/3/2020 11:11:11 AM by SYSTEM Last changed Additional Info : Peak(s) manually integrated DAD1 A, Sig=254,4 Ref=550,100 (FCOMPOUND\TEST 2020-03-03 11-11-11\ME-PR.D) mAU -12.483 175 150 125 100 75 -50 25 11.852 15.647 0 2.5 7.5 10 12.5 15 17.5 min 5 ------Area Percent Report _____ _____ Signal Sorted Bv : Multiplier : 1.0000 : 1.0000 Dilution Use Multiplier & Dilution Factor with ISTDs Signal 1: DAD1 A, Sig=254,4 Ref=550,100 Peak RetTime Type Width Area Height Area # [min] [min] [mAU*s] [mAU] 응
 1
 11.852 BB
 0.0667
 14.50987
 3.27323
 0.9185

 2
 12.483 BB
 0.1312
 1560.13611
 192.28294
 98.7560

 3
 15.647 BV
 0.0710
 5.14208
 1.15293
 0.3255
1579.78806 196.70911 Totals : _____

12601c 3/26/2020 9:46:59 AM SYSTEM

Data File C:\CHEM32\1\DATA\FCOMPOUND\TEST 2020-03-03 11-11-11\ME-PR.D Sample Name: Me-Pr



12601c 3/26/2020 9:48:07 AM SYSTEM

Data File C:\CHEM32\1\DATA\FCOMPOUND\TEST 2020-03-03 11-11-11\PEP-1.D Sample Name: PEP-1

_____ Acq. Operator : SYSTEM Seq. Line : 8 Location : Vial 68 Acq. Instrument : 12601c Injection Date : 3/3/2020 1:45:27 PM Inj : 1 _ . _ Inj Volume : 2.000 µl Different Inj Volume from Sample Entry! Actual Inj Volume : 6.000 µl Method : C:\CHEM32\1\DATA\FCOMPOUND\TEST 2020-03-03 11-11-11\TEST12.M (Sequence Method) : 3/3/2020 11:11:11 AM by SYSTEM Last changed Additional Info : Peak(s) manually integrated DAD1 A, Sig=254,4 Ref=550,100 (FCOMPOUND\TEST 2020-03-03 11-11-11\PEP-1.D) mAU _ 12:474 200 150 100 50 11.848 0 2.5 5 7.5 10 12.5 15 17.5 min ------Area Percent Report _____ _____ : Signal : 1.0000 : 1.0000 Sorted Bv Multiplier Dilution Use Multiplier & Dilution Factor with ISTDs Signal 1: DAD1 A, Sig=254,4 Ref=550,100 Peak RetTime Type Width Area Height Area # [min] [min] [mAU*s] [mAU] 9
 1
 11.848 BB
 0.0644
 9.53240
 2.18448
 0.4227

 2
 12.474 BB
 0.1475
 2245.37402
 245.92307
 99.5773
2254.90643 248.10755 Totals : *** End of Report ***

12601c 3/26/2020 9:47:33 AM SYSTEM

Data File C:\CHEM32\1\DATA\FCOMPOUND\TEST 2020-03-03 11-11-11\PEP-1.D Sample Name: PEP-1



12601c 3/26/2020 9:47:53 AM SYSTEM

Data File C:\CHEM32\1\DATA\FCOMPOUND\TEST 2020-03-10 15-04-31\822.D Sample Name: 822

_____ Acq. Operator : SYSTEM Seq. Line : 8 Location : Vial 8 Acq. Instrument : 12601c Injection Date : 3/10/2020 5:38:57 PM Inj : 1 Inj Volume : 2.000 µl Different Inj Volume from Sample Entry! Actual Inj Volume : 8.000 µl : C:\CHEM32\1\DATA\FCOMPOUND\TEST 2020-03-10 15-04-31\TEST12.M (Sequence Method Method) : 3/10/2020 3:04:58 PM by SYSTEM Last changed Additional Info : Peak(s) manually integrated DAD1 A, Sig=254,4 Ref=550,100 (FCOMPOUND\TEST 2020-03-10 15-04-31\822.D) mAU 10.483 300 250 200 150 100 50 9.603 9.882 10.195 14.358 11.267 0 4 6 8 10 12 14 16 18 min _____ Area Percent Report _____ _____ Sorted By : Signal Multiplier : 1.0000 : 1.0000 Dilution Use Multiplier & Dilution Factor with ISTDs Signal 1: DAD1 A, Sig=254,4 Ref=550,100 Peak RetTime Type Width Area Height Area # [min] [min] [mAU*s] [mAU] 옹 1 9.603 BB 0.0885 7.52870 1.13340 0.3884 16.56597 2 9.882 BB 0.0581 3 10.195 BB 0.0912 4.28472 0.8546 1.02264 0.3400 6.59114 4 10.483 BB 0.1027 1895.50354 299.96573 97.7795
 5
 11.267
 VB
 0.0595
 5.00499
 1.19285
 0.2582

 6
 14.358
 BB
 0.0687
 7.35488
 1.64479
 0.3794
Totals : 1938.54922 309.24412 12601c 3/26/2020 9:49:18 AM SYSTEM Page 1 of 2





12601c 3/26/2020 9:49:29 AM SYSTEM

Supplementary Tables

Table S1. Plasma pharmacokinetic parameters of MDL-800 and MDL-811 in

Compound	MDL-800	MDL-811
Administration	IP 30 mg/mL	IP 30 mg/mL
t _{max} (h)	0.50	0.50
C _{max} (ng/mL)	6610.46	5330.67
AUC_{0-t} (h × ng/mL)	17831.51	19580.01
$AUC_{0-\infty}$ (h × ng/mL)	17883.09	19669.85
t _{1/2} (h)	2.07	1.38
MRT (h)	2.47	2.63
F (%)	71.33	92.96

C57BL/6J m	ice. '	۲
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 * The data represent the mean value from five C57BL/6J mice per group.

Enzyme	Effect [*]
HDAC1	ND
HDAC2	ND
HDAC3	ND
HDAC4	ND
HDAC5	ND
HDAC6	ND
HDAC7	ND
HDAC8	ND
HDAC9	ND
HDAC10	ND
HDAC11	ND
SIRT1	ND
SIRT2	ND
SIRT3	ND
SIRT5	ND
SIRT6	Activation
SIRT7	ND

Table S2. Effect of MDL-811 among histone deacetylase enzymes.

*ND represents no detectable activation or inhibition effect on the targets in the presence of MDL-811 at 100 μ M. The data are from three independent experiments.

Table S3. Melting temperatures of SIRT6 treated with or without MDL-811 inthe CETSA.

	Melting Temperature (T _m) (°C) *
DMSO	45.8 ± 0.3
10 µM MDL-811	47.6 ± 0.1

*The data represent the mean \pm s.e.m. from four independent experiments.

CRC cell line	$IC_{50} \pm s.d. (\mu M)^*$
NCI-H716	61.0 ± 2.3
Colo205	38.1 ± 4.4
Colo320DM	33.2 ± 0.9
LS1034	31.9 ± 0.7
Caco2	28.7 ± 1.7
LS513	28.6 ± 0.6
LS174T	23.8 ± 2.0
HCT15	20.5 ± 1.3
Colo201	20.3 ± 1.4
HCT8	20.1 ± 2.0
LS123	19.8 ± 0.5
LS180	19.1 ± 0.4
DLD-1	18.0 ± 0.1
SW620	16.7 ± 1.4
SW1417	15.1 ± 1.9
SW1463	13.0 ± 1.5
T84	13.0 ± 1.4
RKO	12.3 ± 1.8
SKCO1	12.1 ± 0.1

Table S4. IC₅₀ values of MDL-811 in CRC cell lines.

SW48	10.8 ± 1.5
NCI-H508	9.7 ± 1.2
SW480	9.3 ± 1.7
LOVO	7.8 ± 0.2
НТ29	7.0 ± 1.1
SW1116	5.7 ± 0.3
HCT116	4.7 ± 0.2

^{*}The IC_{50 ±} s.d. values of MDL-811 to the indicated CRC cell lines were determined by CCK8 assays. The value of each MDL-811 treatment group was calculated as a percentage change of the DMSO controls, which represents the proliferation of the CRC cells. The data represent the mean \pm s.d. from two or three independent experiments.

Cell name	Phase	0 μΜ	5μΜ	10 µM
	G ₀ /G ₁	49.5 ± 3.5	$63.2 \pm 2.1^{**}$	$66.9 \pm 1.4^{***}$
HCT116	S	30.1 ± 4.4	$19.0 \pm 2.3^{*}$	$19.0 \pm 4.3^{*}$
	G_2/M	20.4 ± 4.5	17.9 ± 4.0	14.1 ± 3.8
	G_0/G_1	40.5 ± 2.6	54.5 ± 4.1*	$59.9 \pm 2.7^{***}$
HT29	S	36.9 ± 4.6	25.3 ± 4.5	$19.6 \pm 2.3^{**}$
	G_2/M	22.6 ± 6.7	20.2 ± 4.2	20.5 ± 3.7
	G_0/G_1	47.7 ± 1.3	$62.9 \pm 6.0^{*}$	$70.0 \pm 3.8^{***}$
SW480	S	38.3 ± 3.6	26.3 ± 5.1	$18.1 \pm 4.9^{***}$
	G_2/M	13.9 ± 1.8	10.8 ± 4.4	11.9 ± 6.4

Table S5. Statistical data of cell cycle phase distribution in CRC cell lines treated with MDL-811 for 48 h.^a

^aThe data are shown as mean \pm s.d. from three independent experiments (*, *P* < 0.05; **, *P* < 0.01; ***, *P* < 0.001, two-way ANOVA analysis).

Antibody name	Company name	Catalog number	Dilution
SIRT6	Cell Signaling Technology	12486	1:2000
H3K9Ac	Abcam	ab32129	1:2000
H3K18Ac	Abcam	ab1191	1:2000
НЗК56Ас	Active Motif	39281	1:2000
Histone H3	Abcam	ab10799	1:2000
β-actin	Proteintech	HRP-60008	1:5000
His-Tag	Proteintech	HRP-66005	1:10000
Flag-Tag	Sigma-Aldrich	F1804	1:1000

Table S6. Primary antibodies used for western blots.
Table S7. Pri	imer sequences	used for	RT-qPCR.
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Gene nan	ne Forward primer (5' to 3')	Reverse primer (5' to 3')
АСТВ	TGACTGACTACCTCATGAAGATCC	CCATCTCTTGCTCGAAGTCCAG
LDHA	AGGAGAAACACGCCTTGATTTAG	ACGAGCAGAGTCCAGATTACAA
GLUT1	TGGACCCATGTCTGGTTGTA	ATGGAGCCCAGCAGCAA
PDK1	GGAGGTCTCAACACGAGGTC	GTTCATGTCACGCTGGGTAA
PKM2	ATGTCGAAGCCCCATAGTGAA	TGGGTGGTGAATCAATGTCCA
PCNA	AGGGCTCCATCCTCAAGAAGG	TGGTGCTTCAAATACTAGCGC
CDC2	CAGTCTTCAGGATGTGCTTATGC	GAGGTTTTAAGTCTCTGTGAAGAACTC
CCNA2	GAAGACGAGACGGGTTGCA	AGGAGGAACGGTGACATGCT
CDC25C	GAACAGGCCAAGACTGAAGC	GCCCCTGGTTAGAATCTTCC
c-MYC	GGCTCCTGGCAAAAGGTCAGAGT	CTGCGTAGTTGTGCTGATGTGT
AKT1	CACAAACGAGGGGGGGGGAGTACATC	GCCATCATTCTTGAGGAGGAAGT
AKT2	TCCAGAACACCAGGCACCC	ATTGTCCTCCAGCACCTCA
MTOR	AGTGGACCAGTGGAAACAGG	TTCAGCGATGTCTTGTGAGG
CYP24A1	CCGTAATCCCCAAGTGCAAC	CCCAGAACTGTTGCCTTGTC
SIRT6	TACGCGGACAAGGGCAAG	ACTTGGGGGGCCAGACCTCGC

Antibody name	Company name	Catalog number	Dilution
SIRT6	Abcam	ab62739	1:200
Н3К9Ас	Abcam	ab32129	1:100
H3K18Ac	Abcam	ab1191	1:100
H3K56Ac	Active Motif	39281	1:100
HA-Tag	Cell Signaling Technology	#3724	1:50

Table S8. Primary antibodies used for ChIP assays.

Primer name	Forward primer (5' to 3')	Reverse primer (5' to 3')
H3K9Ac_1	CGCTGTCCCACCTGATCACA	AGGACATGACCGCTTTCTTCAA
H3K9Ac_2	СТСССТТТСТСТТТТССТТТАСТСС	ACGCATAACCCCTGTACCCT
H3K9Ac_3	CAGCTATCCTGAGGTGTGCC	AGGAGCTGTGTCCAGAATTGG
H3K9Ac_4	TGGGTTCAGGGATTTTGAGGT	TTGCTGGTGATGGGGGTGTTC
H3K9Ac_5	CTAGCGGTAAAAGGGGGGCAT	TGAAGCCCACACCAATGAGT
H3K9Ac_6	TCTCCATGTTCCTATGCCCAG	TCGCTCACCTCGCTGACT
H3K9Ac_7	CGTAAAGCGGCAACAACGAA	GTAGAAGAACAGAGGCGGGC
H3K9Ac_8	CCGGATTGCAGAGGAAAGCA	TCCTGGAAGCGGGATCAAAA
H3K18Ac_1	CGCTGTCCCACCTGATCACA	AGGACATGACCGCTTTCTTCAA
H3K18Ac_2	TTCAGGTCTTGCTACATCGC	CAGGACGAGGTCATTAGGGG
H3K18Ac_3	GTCATGTGAGCCCTGGAAGC	GCACGGAGTCAAAGGGAGTT
H3K18Ac_4	CATACTTCTTGTGGTACTCCACCT	TGGTGCGCTTCGGCGT
H3K18Ac_5	AATGCACGTAAAGCGGCAAC	TCAAGCATCGTTGGTGCAAG
H3K18Ac_6	TCTAGGCTGGGCCCTAAATAGT	CTGCCCCACACATACTGACAT
H3K18Ac_7	CAAGGGACCACCCATGACAA	GCAGAGAAACCAGCCCTTGA
H3K56Ac_1	CAGCTATCCTGAGGTGTGCC	AGGAGCTGTGTCCAGAATTGG
H3K56Ac_2	TTTGACTCCGTGCTGGCTAA	GGCATGAAATGACGTGGTGT
H3K56Ac_3	CATACTTCTTGTGGTACTCCACCT	TGGTGCGCTTCGGCGT
H3K56Ac_4	CTCTCCATGTTCCTATGCCCAG	CTCGCTGACTCCATCCTCCTTC
H3K56Ac_5	CGTAAAGCGGCAACAACGAA	GTAGAAGAACAGAGGCGGGC

Table S9. Primer sequences used for ChIP-qPCR.