Supplementary Information

Supplementary Figures

Figure S1. ZFP91 overexpression suppresses HCC cell growth, colony formation, migration and invasion. (A) The ZFP91 and hnRNP A1 protein levels and PKM pre-mRNA splicing in both five HCC cell lines and three normal liver tissues were detected. (B-E) MHCC97H and MHCC-LM3 cells were transfected with ZFP91-Flag plasmids; ZFP91 protein level (B), cell growth (C), migration and invasion (D), and colony formation (E) was determined. Data are represented as mean ± SD.
**Figure S2.** ZFP91 decreased hnRNP A1 protein level, not mRNA level, and ECD attenuated the interaction of ZFP91 with hnRNP A1 and the ubiquitination of ZFP91 on hnRNP A1. (A) MHCC-LM3 HCC cells were transfected with ZFP91-Flag plasmid, and the hnRNP A1 mRNA level was determined. (B) SK-hep1 HCC cells were transfected with anti-ZFP91 siRNA, and the hnRNP A1 mRNA level was determined. (C) ZFP91 and hnRNP A1 protein levels in mouse xenograft tumor tissues in Figure 2E were detected. The correlations of ZFP91 with hnRNP A1 were analyzed in the right panel. (D) MHCC-LM3 HCC cells were transfected with the indicated plasmids, hnRNP A1-Flag complexes were co-immunoprecipitated with anti-Flag, and ZFP91-HA was detected using anti-HA antibody. (E) MHCC-LM3 HCC cells were transfected with the indicated plasmids for 36 h, followed by treatment with 10 μM MG132; the polyubiquitination level of hnRNP A1 was detected.
Figure S3. The ubiquitination modification lysine 8 site in the hnRNP A1 protein was identified by mass spectrometry. (A) Two representative MS/MS spectra of the identification the lysine 8 (K8) ubiquitination site in the hnRNP A1 protein. (B) Matching of the MS-identified peptides and their post-modifications with the acid amino sequence of hnRNP A1 protein.
**Figure S4.** HnRNP A1 promotes HCC cell growth, colony formation, migration and invasion. (A-D) SK-hep1 cells were transfected with hnRNP A1-HA plasmids; hnRNP A1 protein level (A), cell growth (B), migration and invasion (C), and colony formation (D) were determined. (E-H) MHCC-LM3 cells were transfected with anti-hnRNP A1 siRNAs; hnRNP A1 protein level (E), cell growth (F), migration and invasion (G), and colony formation (H) were determined (n = 3). Data are represented as mean ± SD.
Figure S5. HnRNP A1 promotes the alternative splicing of PKM pre-mRNA through hnRNP A1. (A, B) HCC SK-hep1 cells were transfected with hnRNP A1-HA plasmids; the protein levels of PKM1 and PKM2 (A) and PKM pre-mRNA splicing (B) were detected. (C, D) HCC MHCC-LM3 cells were transfected with anti-hnRNP A1 siRNAs; the protein levels of PKM1 and PKM2 (C) and PKM pre-mRNA splicing (D) were detected.
were detected. (E, F) Both ZFP91-HA and hnRNP A1-Flag plasmids were cotransfected into MHCC-LM3 cells; the protein levels of PKM1 and PKM2 (E) and PKM pre-mRNA splicing (F) were detected. (G, H) The protein levels of PKM1 and PKM2 (G) and PKM pre-mRNA splicing (H) were detected in mouse xenograft tumor tissues in Figure 2E. The correlations of ZFP91 protein levels with PKM1 and PKM2 protein levels were analyzed (right panel in G).
Figure S6. ZFP91 inhibits HCC cell proliferation, colony formation, migration and invasion through PKM2. HCC SK-hep1 cells were cotransfected with anti-ZFP91 siRNA together with anti-PKM2 siRNA; ZFP91 and PKM2 protein level (A), cell growth (B), migration and invasion (C), and colony formation (D) were determined (n = 3). Data are represented as mean ± SD.
Supplementary Tables

Table S1: Correlations between ZFP91 protein levels and clinicopathological features in 90 HCC cases.

<table>
<thead>
<tr>
<th>Clinical character</th>
<th>Clinical groups</th>
<th>All cases</th>
<th>ZFP91</th>
<th>( x^2 )</th>
<th>( P ) value*</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Low</td>
<td>High</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Age (Years)</td>
<td>≤ 50</td>
<td>42</td>
<td>29</td>
<td>13</td>
<td>2.686</td>
</tr>
<tr>
<td></td>
<td>&gt; 50</td>
<td>48</td>
<td>25</td>
<td>23</td>
<td></td>
</tr>
<tr>
<td>Gender</td>
<td>Female</td>
<td>16</td>
<td>8</td>
<td>8</td>
<td>0.811</td>
</tr>
<tr>
<td></td>
<td>Male</td>
<td>74</td>
<td>46</td>
<td>28</td>
<td></td>
</tr>
<tr>
<td>Cirrhosis</td>
<td>YES</td>
<td>78</td>
<td>47</td>
<td>31</td>
<td>0.016</td>
</tr>
<tr>
<td></td>
<td>NO</td>
<td>12</td>
<td>7</td>
<td>5</td>
<td></td>
</tr>
<tr>
<td>Histological Grade</td>
<td>G1-2, G2</td>
<td>58</td>
<td>33</td>
<td>25</td>
<td>0.655</td>
</tr>
<tr>
<td></td>
<td>G2-3, G3</td>
<td>32</td>
<td>21</td>
<td>11</td>
<td></td>
</tr>
<tr>
<td>Tumor size (cm)</td>
<td>≤ 3</td>
<td>29</td>
<td>17</td>
<td>12</td>
<td>0.034</td>
</tr>
<tr>
<td></td>
<td>&gt; 3</td>
<td>61</td>
<td>37</td>
<td>24</td>
<td></td>
</tr>
<tr>
<td>Tumor number (n)</td>
<td>≤ 3</td>
<td>74</td>
<td>43</td>
<td>31</td>
<td>0.621</td>
</tr>
<tr>
<td></td>
<td>&gt; 3</td>
<td>16</td>
<td>11</td>
<td>5</td>
<td></td>
</tr>
<tr>
<td>AFP (μg/L)</td>
<td>&lt; 400</td>
<td>57</td>
<td>32</td>
<td>25</td>
<td>0.965</td>
</tr>
<tr>
<td></td>
<td>≥ 400</td>
<td>33</td>
<td>22</td>
<td>11</td>
<td></td>
</tr>
<tr>
<td>Cirrhosis Nodule size (mm)</td>
<td>≤ 3</td>
<td>50</td>
<td>29</td>
<td>21</td>
<td>0.188</td>
</tr>
<tr>
<td></td>
<td>&gt; 3</td>
<td>40</td>
<td>25</td>
<td>15</td>
<td></td>
</tr>
<tr>
<td>Clinical stage#</td>
<td>I</td>
<td>61</td>
<td>31</td>
<td>30</td>
<td>6.648</td>
</tr>
<tr>
<td></td>
<td>II</td>
<td>29</td>
<td>23</td>
<td>6</td>
<td></td>
</tr>
<tr>
<td>TNM stage</td>
<td>I</td>
<td>54</td>
<td>27</td>
<td>27</td>
<td>5.625</td>
</tr>
<tr>
<td></td>
<td>II</td>
<td>36</td>
<td>27</td>
<td>9</td>
<td></td>
</tr>
<tr>
<td>Recurrence of cancer</td>
<td>YES</td>
<td>37</td>
<td>17</td>
<td>20</td>
<td>5.171</td>
</tr>
<tr>
<td></td>
<td>NO</td>
<td>53</td>
<td>37</td>
<td>16</td>
<td></td>
</tr>
</tbody>
</table>

ZFP91 low score: 0-3; ZFP91 high score: 4-7;
*Pearson Chi-square test;
#Clinical stage: American Joint Committee on Cancer classification (AJCC).
siRNA sequences

ZFP91 si#1:
Sense: 5’- CAUUGCUGCAUCUAGACCUUTT-3’
Anti-sense: 5’- AGGUCUAGAUGCAGCAUGTT-3’

ZFP91 si#2:
Sense: 5’- GCAGCUCAUUUGCAAGUCATT-3’
Anti-sense: 5’- UGACUUGCAAAUGAGCUGCTT-3’

hnRNP A1 si#1:
Sense: 5’-GCUGUGUAAAGUUAGUCUATT-3’
Anti-sense: 5’-UAGACUAACUUUACACAGCTT-3’

hnRNP A2 si#2:
Sense: 5’-GUGUAGUUGAACUGAUAGUTT-3’
Anti-sense: 5’-ACUAUCAGUUCAACUACACCTT-3’

NC siRNA:
Sense: 5’-GCACAAGCUGGAGUACAACATT -3’
Anti-sense: 5’- UGUAGUUGUACUCGCUUGUGCTT-3’

RT-PCR primers

ZFP91
Forward: TCCTTGCCCATCCTCGCTATT
Reverse: TGTTTGGCATGTGCAGAAGT

β-actin
Forward: GAGAAAATCTGGCACACACC
Reverse: GGATAGCACAAGCTGGATAGCAA

PKM
Forward: CTGAAGGCAGTGATGTGGCC
Reverse: ACCCGGAGGTCCACGTCCTC

Vector construction primers

ZFP91-Flag
Forward: CGGGGTACCGACGGACAAGCCCCGATGC
Reverse:
CGGAATTCTTACTTGTCACTCGTCTCCTTGTAGTCAGGTCCGGCAGAGTCTGAATC
ZFP91-HA
Forward: CGGGGTACCGACGGACAAGCCCCCGATGC
Reverse:
CGGAATTCTTAAGCGTAATCTGGAACATCGTATGGGTAAGGTCCGGCAGA
GTCTGAATC
hnRNP A1-Flag
Forward: CGGGGTACCACGTTCGTCAGCTTGCTCCTT
Reverse:
CGGAATTCTTACTTGTCACTCGTCGTCCTTGTAGTCAAATCTTCTGCCACTGC
CATAGC
hnRNP A1-HA
Forward: CGGGGTACCACGTTCGTCAGCTTGCTCCTT
Reverse:
CGGAATTCTTAAAGCGTAATCTGGAACATCGTATGGGTAAAATCTTCTGCCATAGC
hnRNP A1-Flag K8R mutation
Forward: TGCCGTCAGATCTAAGTCAGAGTCTAAAGTGCAGGCCACTG
Reverse: ACTTAGATCTGACGGCAGGGTGGAAGAGAGACT