

Review

Combined-therapeutic strategies synergistically potentiate glioblastoma multiforme treatment via nanotechnology

Jun Yang^{1*}, Zhuyan Shi^{1,2*}, Ruiyuan Liu^{1,2}, Yanyue Wu^{1,2} and Xin Zhang¹

1. State Key Laboratory of Biochemical Engineering, Institute of Process Engineering, Chinese Academy of Sciences, Beijing 100190, PR China
2. University of Chinese Academy of Sciences, Beijing 100049, PR China

*These authors contributed equally to this work.

 Corresponding author: Xin Zhang, State Key Laboratory of Biochemical Engineering, Institute of Process Engineering, Chinese Academy of Sciences. Tel./fax: +86 10 82544853. Email: xzhang@ipe.ac.cn.

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Abstract

Glioblastoma multiforme (GBM) is a highly aggressive and devastating brain tumor characterized by poor prognosis and high rates of recurrence. Numerous therapeutic strategies and delivery systems are developed to prolong the survival time. They exhibit enhanced therapeutic effects in animal models, whereas few of them is applied in clinical trials. Taking into account the drug-resistance and high recurrence of GBM, combined-therapeutic strategies are exploited to maximize therapeutic efficacy. The combined therapies demonstrate superior results than those of single therapies against GBM. The co-therapeutic agents, the timing of therapeutic strategies and the delivery systems greatly affect the overall outcomes. Herein, the current advances in combined therapies for glioblastoma *via* systemic administration are exhibited in this review. And we will discuss the pros and cons of these combined-therapeutic strategies *via* nanotechnology, and provide the guidance for developing rational delivery systems to optimize treatments against GBM and other malignancies in central nervous system.

Key words: glioblastoma, combined therapy, nanotechnology, drug delivery

Introduction

Glioblastoma multiforme (GBM) is a primary malignant brain tumor of the central nervous system (CNS) with poor prognosis and high mortality [1, 2]. The median survival rate is around 12-18 months, and the long-term survival time is less than 5 years [3, 4]. The current clinical treatments for GBM consist of tumor resection, radiotherapy and chemotherapy. Maximal tumor resection improves the overall survival in GBM. But it is difficult to distinguish tumor from normal brain tissues. Several intraoperative technologies, such as 5-aminolevulinic acid (ALA) and fluorescein (FLCN) guided resection, facilitate the identification of tumor and simultaneous preservation of neurologic function [5, 6]. They

significantly improve the gross total resection with standard surgery from 36% to over 64% in high-grade gliomas (HGG). However, the infiltrative nature of GBM raises the challenge to realize a complete resection. Furthermore, the extent of resection must be balanced with the brain function [7]. The introduction of Stupp regimen and modified Stupp extends the median survival of the patients, whereas the overall survival is still poor [8-10].

Besides the standard therapy, the therapeutic strategies like gene therapy, immunotherapy, phototherapy and thermotherapy, have been applied for anti-glioma treatments [11-15]. They exhibit enhanced therapeutic effects in animal models, but

few of them is applied in clinical trials. Since the GBM is a complex disease with intricate mechanisms in their growth, progression and invasion processes, accumulating evidences indicate that single therapeutic strategy tends to result in drug resistances and tumor cell tolerance, which finally leads to tumor recurrence and metastasis [16, 17]. Therefore, combined-therapeutic strategies with various mechanisms of agents should be able to overcome these problems.

However, the current therapeutic agents have some critical problems, such as short half-life in circulation, hard transportation to the diseased areas and difficulty of controlled releasing the diverse drugs in the corresponding sites. These problems result in insufficient accumulation of therapeutic drugs in the tumor cells, and prevent adequate destruction of malignant tumors [18, 19]. Therefore, efficient drug delivery will be very critical and important for effective therapy [20]. For a typical cancer treatment, a five-step CAPIR cascade in drug delivery process is presented by Shen's group [21]. It includes circulation in the blood, accumulation in the tumor, penetration into tumor tissue, internalization into tumor cells and intracellular drug release. High efficiency at every step is very important to ensure the high therapeutic efficiency of the whole treatment. Furthermore, in view of the location of glioblastoma, the specific blood-brain barrier (BBB) also constructs a major and critical obstacle for the drug delivery [22, 23]. It is estimated that over 98% of the small molecular drugs and almost 100% of large molecular drugs cannot cross the BBB [24, 25]. Given this, the challenges of drug delivery in the GBM treatment should be composed of the following steps: long circulation in the blood, effective transportation across the BBB, efficient internalization into glioma cells and controlled drug release in the cells (Figure 1). All of these steps are the key factors to ensure sufficient therapeutic agents accumulating in GBM cells.

To overcome these physiological barriers in GBM treatments, nanotechnology has been explored to solve these problems and potentiate the therapeutic effects. These delivery systems can be classified as liposomes, polymer nanoparticles, lipopolymer nanoparticles, dendrimer nanoparticles, hybrid nanoparticles and so on [26-29]. Firstly, the diversity of these biomaterials can enable loading of various therapeutic agents simultaneously. Secondly, these nanodelivery systems can be modified with specific targeting, which shows the guidance to BBB and the GBM cells. The reported brain tumor targeting moieties include transferrin receptor (TfR), angiopep-2 peptide, TAT peptide, RGD peptide,

chlorotoxin and so on [30-32]. The modified nanodelivery systems can transport across the BBB by receptor-mediated endocytosis, adsorptive-mediated endocytosis and carrier-mediated transport [33]. These targeting modifications greatly improve the brain tumor targeting and reduce the side effects to normal tissues. Thirdly, stimuli-sensitive responses are introduced into delivery systems to ensure the maximal drug retention at the desired sites, such as pH, ROS, enzyme, light and thermal responses [34]. These strategies with nanotechnology prompt the drug accumulation in brain tumors. Intensive advances have exhibited the effective and promising outcomes for glioblastoma therapy these decades, but few of them is applied in clinical trials until now. Herein, in this review, we will focus on the combined-therapeutic strategies with diverse delivery systems, investigate the current combined strategies with improved therapeutic effects, and discuss the future guidance for glioblastoma treatments and other CNS diseases' therapies.

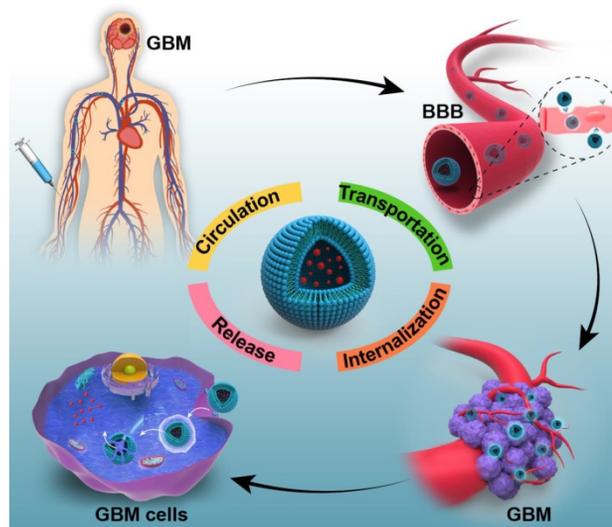


Figure 1. The schematic illustration exhibits the challenges of drug delivery via systemic administration for glioblastoma therapy. They generally include: (1) long circulation in the blood; (2) effective transportation across the BBB; (3) efficient internalization into GBM cells and (4) controlled drug release in the tumor cells.

Combined therapies against glioblastoma

The complexity of glioblastoma multiforme motivates researchers to develop various therapeutic strategies. Comparing with single therapy, combined therapies have emerged new issues to concern. The choice of the therapeutic strategy for combination is the first key factor. It contains the selection of combined methods and the therapeutic agents, and synergism of these strategies. The delivery systems based on the therapeutic agents are the second important issue to concern. These systems of combined therapies are required to load the multiple

therapeutic agents and deliver them to the corresponding targeted sites for synergistic treatments. Herein, various combined-therapeutic strategies for anti-glioma treatments are presented in the following sections. And the merits of the current delivery systems are discussed, which should inspire and guide to develop optimized delivery platforms for further clinical applications.

Combined chemotherapies

Chemotherapy was the most common treatment for glioblastoma. Single chemotherapy facilitated the drug resistance of the tumor after a certain period, which greatly hindered the successful treatment of GBM. Combination with diverse mechanisms of anti-cancer drugs should improve the GBM therapy, and some efforts had entered into clinical trials [35-38].

However, due to the physiological barriers of glioblastoma, some clinical trials on the combination therapies did not show survival advantages and failed to obtain the desirable outcomes [39-41]. Hence, numerous delivery systems were developed to overcome these challenges and obtained better therapeutic effects [42, 43]. For example, Guo et al. utilized a liposome (LP), which was composed of egg yolk phosphatidylcholine (EPC), cholesterol (Chol) and 1,2-distearoyl-*sn*-glycero-3-phosphoethanolamine-N-[methoxy(polyethylene glycol) 2000] (DSPE-PEG2000), to separately encapsulate the tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) and doxorubicin (DOX) [44]. The median survival time of the combination group (DOX-LP+TRAIL-LP) extended to 48 days, compared with the single groups TRAIL-LP (32 days) and DOX-LP (39 days). The combined chemotherapies showed improved therapeutic effects in the intracranial U87MG-bearing mice. The proportion of the drugs was easy to adjust by the detached drug load, but the synergism effects of these anticancer drugs *in vivo* might be hard to control.

To increase the drug accumulation in the brain tumors, active targeting was introduced into the nanosystems to enhance the drug concentration in tumor sites and reduce the systemic toxicity [45-47]. Erica Locatelli et al. developed a targeted delivery nanoparticle (Ag/Ali@PNP-Cltx-^{99m}Tc) based on the poly(lactic-co-glycolic acid)-block-polyethylene glycol (PLGA-*b*-PEG) copolymer [48]. The nanosystem conjugated with chlorotoxin (Cltx), which expressed the specific binding to matrix metalloproteinase-2 (MMP-2), a receptor overexpressed on the brain cancer cells. The *in vivo* biodistribution data exhibited that the targeted nanosystem Ag/Ali@PNP-Cltx-^{99m}Tc significantly increased drug concentration in tumors. The enhanced drug accumulation

obviously reduced the tumor size *in vivo*. The current approved anticancer drug temozolomide (TMZ) had been reported to associate with the introduction of O⁶-methylguanine-DNA methyltransferase (MGMT), which increased TMZ-resistance in chemotherapy [49-54]. Sang-Soo Kim et al. developed a cationic liposome (scL-p53) encapsulating the tumor suppressor gene p53 plasmid DNA to sensitize the cancer cells to TMZ [55]. This nanocomplex was modified with an antitransferrin receptor (TfR) single chain antibody fragment (scFv) to target the tumor cells. The combination of scL-p53 with TMZ obviously down-modulated the MGMT expression by 95% for 24 h *in vivo*. The mice treated with cycle administration of TMZ finished at day 45, whereas 63.6% of the mice were still alive when treated with this nanosystem. These studies demonstrated that the active brain targeting significantly enhanced the drug concentration in tumors with the same doses in comparison with the non-targeted systems.

Active brain targeting ensured the guidance of drug accumulation in tumor tissues, and the effective release of therapeutic drugs in tumor cells was also critical for efficient treatments against GBM [56]. Hence, plenty of stimuli-sensitive drug delivery systems were designed based on the microenvironment of GBM, including pH, ROS, enzyme, and so on [57-59]. For example, the anticancer drug doxorubicin (DOX) could potentiate the antitumor effect of tumor necrosis factor-related apoptosis-inducing ligand (TRAIL), which expressed by pORF-hTRAIL. Inspired by these, Jiang's group designed a targeting dendrigraft poly-L-lysine (DGL)-based nanosystem (DGDPT/pORF-hTRAIL) with pH-response to carry the chemotherapeutic drug DOX and the gene agent pORF-hTRAIL for anti-glioma therapy [60, 61]. This nanosystem was modified with peptide HAIYPRH (T7), a transferrin receptor-specific peptide, for brain tumor cells targeting. The target significantly increased the drug accumulation in tumors *in vivo*. The median survival of DGDPT/pORF-hTRAIL was extended to 57 days, in comparison with 34 days for the free DOX. More importantly, the dose of the anticancer drug DOX in DGDPT/pORF-hTRAIL was less than 0.35 mg/kg mice, whereas the dose of free DOX was 5 mg/kg mice. This nanosystem greatly increased the pharmaceutical effect of the DOX and reduced the side toxicity to normal tissues. Also, Shi's group designed a nanoparticle (3I-NM@siRNA) for combined siRNA delivery, which was stabilized by electrostatics, hydrogen bonds and hydrophobic interactions (Figure 2) [62]. The angiopep-2 targeting decorated 3I-NM@siRNA was prepared with poly(ethyleneglycol)-block-poly[(N-(3-methacrylamidopropyl)guanidinium-co-4-(4,4,5,5-tetramethyl-1,3,2-diox

aborolan-2-yl) benzyl acrylate]] (PEG-*b*-P(Gu/Hb))/Ang-poly(ethylene glycol)-block-poly(N-(3-methacryl amidopropyl) guanidinium) (Ang-PEG-*b*-PGu). The nanoparticles could trigger drug release under ROS environment that enriched in the cancer cells. Treatment with Ang-3I-NM@(siPLK1+siVEGFR2) significantly increased the median survival time to 36 days, which was longer than the mice treated with 3I-NM@(siPLK1+siVEGFR2) (18 days), Ang-3I-NM@(siPLK1) (24 days) and Ang-3I-NM@(siVEGFR2) (26 days).

Numerous researches detected that the available drug therapies were dissatisfied because they failed to penetrate into depth of the tumor tissues and kill the brain cancer stem cells (BCSCs), which was the most responsible factor for tumor recurrence [63-65]. Intensive studies had been arisen to inhibit the growth of cancer stem cells for effective treatments to glioblastoma [66]. Lu et al. in our group designed a brain targeting and multifunctional nanoparticle

(CARD-B6) to synergistically aim at both the glioblastoma cells and glioblastoma stem cells (GSCs) (Figure 3) [67]. In this nanosystem, acid-sensitive poly(β -amino ester) (PAE) was utilized to load the antiangiogenic drug combretastain A4 (CA4), and triggered to release the cargo in the GBM microenvironment. The azobenzene (AZO) bond in the PAE, rapidly broken up in the hypoxic GSCs, was applied to conjugate with all-trans retinoic acid (ATRA), which could induce the GSCs differentiation into glioblastoma cells. The amide bond in the PAE was conjugated with anticancer drug doxorubicin (DOX) for controlled release in tumor cells. The designed CARD-B6 could spatiotemporally trigger to deliver these drugs to their corresponding target sites. And the survival time of the CARD-B6 treated mice significantly enhanced, with 60% survivals *vs.* 0% for all the other control groups.

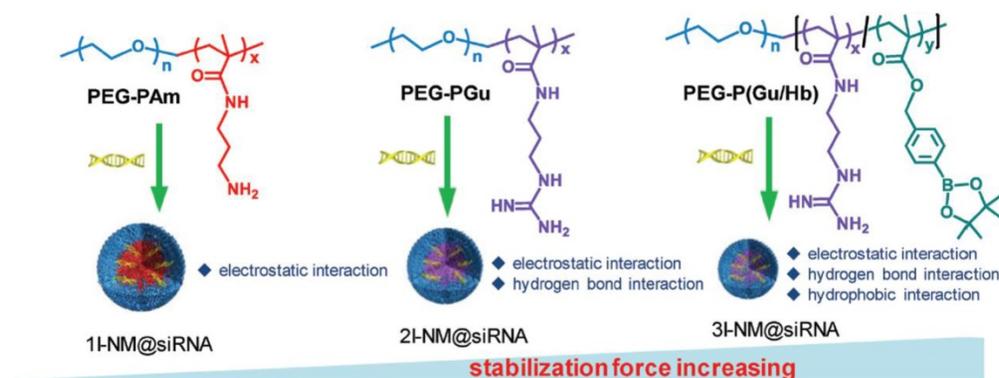


Figure 2. The schematic illustration of the formation of 1I-NM@siRNA, 2I-NM@siRNA and 3I-NM@siRNA utilizing different polymers to generate different numbers of stabilizing interactions. Adapted with permission from [62]. Copyright 2019 WILEY-VCH Verlag GmbH & Co.

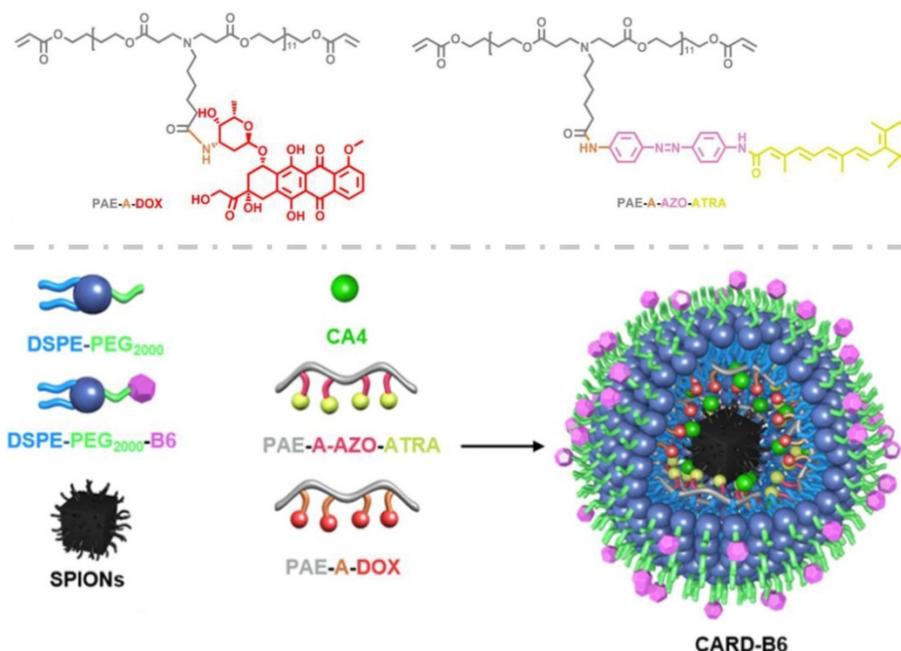


Figure 3. The structural composition and preparation route of the multifunctional nanoparticles CARD-B6. Adapted with permission from [67]. Copyright 2017 WILEY-VCH Verlag GmbH & Co.

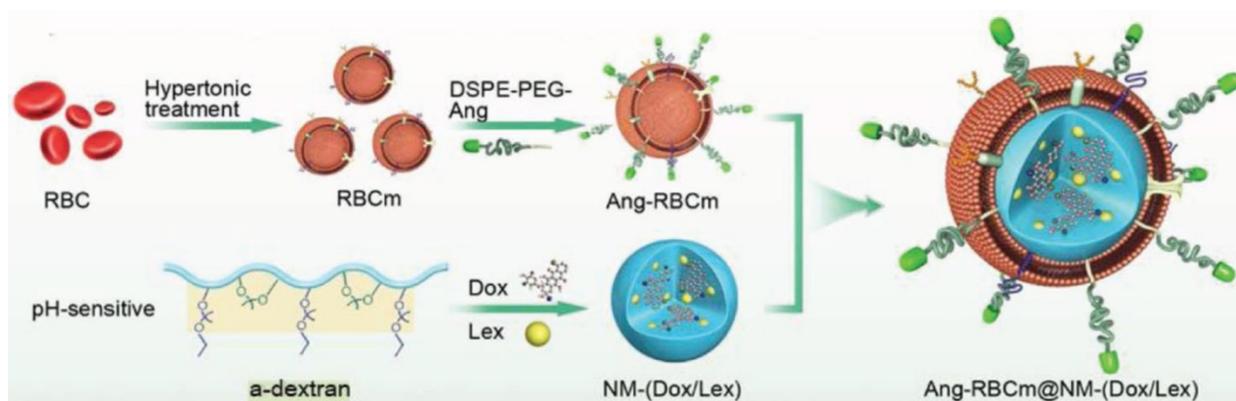


Figure 4. The schematic illustration of the formation of the biomimetic nanomedicine (Ang-RBCm@NM-(Dox/Lex)). Adapted with permission from [74]. Copyright 2018 WILEY-VCH Verlag GmbH & Co.

Recently, biomimetic nanosystems were exploited for drug delivery applications, including the utilization of red blood cell membranes, tumor cell membranes, platelet membranes and etc. [68-73]. They had good biocompatibility and avoided immunogenicity. For instance, Shi's group designed a biomimetic nanomedicine (Ang-RBCm@NM-(Dox/Lex)) by functionalizing with red blood cell membranes (RBCMs) (Figure 4) [74]. This nanosystem was equipped with angiopep-2 at the surface of red blood cell membranes. The target had high affinity for the low density lipoprotein receptor-related protein (LRP) receptor, which was overexpressed on both the endothelial cells of the BBB and glioblastoma cells. A pH-sensitive α -dextran that coloaded the anticancer drug doxorubicin (Dox) and lexican (Lex), and encapsulated in the RBCm, could transiently open the BBB to enhance the permeability of the nanomedicine Ang-RBCm@NM-(Dox/Lex). The Ang-RBCm@NM-(Dox/Lex) exhibited a longer blood circulation time, with the half lifetime of 9.3 h, whereas the circulation time of NM-(Dox/Lex) without RBCm camouflage was only 2.4 h. And the biodistribution of Ang-RBCm@NM-(Dox/Lex) in the orthotopic brain tumor was about 3.5-fold higher than that of NM-(Dox/Lex). The biomimetic nanomedicine enhanced the median survival time to 38 days, in comparison with RBCm@NM-(Dox/Lex) (28 d), Ang-RBCm@NM-Dox (25 d), NM-(Dox/Lex) (22 d), free Dox (22 d) or PBS (18 d) treated groups.

Chemotherapy was the most widely studied treatment for anti-glioma therapy. Mounting researches were reported from the non-targeting and non-sensitive delivery systems to the active targeting and stimuli-sensitive systems. Intelligent multifunctional delivery systems were designed in dependence on the specific characteristics of the GBM. They exhibited better drug accumulation and longer survival time for GBM treatments. However, GBM was a complex disease. The initiation, growth and

progression of the GBM were not completely understood until now. Hence the therapeutic drugs might not involve in every critical point of the whole treatments, and the designed drug delivery systems based on existing knowledge of GBM could not completely eliminate the GBM. Moreover, most of the biomaterials utilized in these systems were not approved by FDA, which aggravated difficulties for the clinical translations.

Biomimetic drug delivery systems offered new opportunities to mimic the biological vehicles. The naturally derived cell membranes decorated the drug delivery systems, and brought longer circulation and better biocompatibility for these nanosystems. However, they also generated new problems, like the preparation operations and the cost of the biological products, the strictness production conditions for the clinical translations and applications.

Combined chemotherapy with radiotherapy

Radiotherapy was one of the clinical treatments for GBM, and amount of efforts had been made to increase the efficacy [4]. However, the invasive tumor growth of the glioblastoma limited the radiotherapy's efficacy and induced tumor recurrence. Since the combination of radiotherapy with chemotherapy turned into the standard treatment strategy to inhibit GBM recurrence [75-78], the radiotherapy concomitant with free temozolomide or carmustine entered into clinical trials. But the regimens were still insufficient for the requirements [79].

Hence, delivery systems with nanotechnology were exploited for the combined therapies to maximize the therapeutic effects. Shi et al. developed an integrin targeting ^{177}Lu radiolabeled 3PRGD₂ (a dimeric RGD peptide with 3 PEG₄ linkers) for radiotherapy [80]. The cellular uptake of ^{177}Lu -3PRGD₂ in U87MG tumors was about 6%ID/g for 1 h. The combination with the anti-angiogenic agent Endostar exhibited significant tumor inhibition

in comparison with the control group. Later, Matteo Tamborini et al. reported that combining the chlorotoxin-nanovectors and radiation could exhibit a synergistic effect for *in vivo* GBM growth (Figure 5) [81]. Chlorotoxin, a selective target and anticancer drug, was conjugated on poly(lactic-co-glycolic acid) (PLGA) nanoparticles (PNP). Low dose of radiation (2 Gy) rendered the brain extracellular matrix permeable and facilitated the accumulation of the nanovectors (Ag-PNP-CTX). The combination therapy significantly inhibited approximately 50% tumor growth in U87MG-bearing mice.

As the clinical standard treatment, radiotherapy often resulted in the modest improvement for glioblastoma treatments without reducing the quality of life or cognition. From the available better treatments, radiotherapy with concomitant chemotherapy was the current standard of care. However, the reported clinical trials regarding the combination of radiotherapy and chemotherapy exhibited diverse results. For instance, the addition of bevacizumab to radiotherapy-temozolomide did not improve the survival against GBM in phase 3 study [79]. And sorafenib combined with radiotherapy-temozolomide exhibited severe side effects in phase 1 study [82]. These might be due to the mechanisms of these drugs or the insufficient drug concentrations in the tumors, which counteracted the therapeutic effects. Hence, the definition of tumor margins, the choice of the chemotherapeutic drugs and drug delivery systems were very important for the regimen.

Combined chemotherapy with phototherapy

Phototherapy was composed of the photothermal therapy (PTT) and the photodynamic therapy (PDT), which was a promising non-invasive strategy for cancer treatments. The former one utilized the photothermal agents to generate heat and kill tumor cells under light absorption. And the latter one produced cytotoxic reactive oxygen species, such as singlet oxygen (1O_2), free radicals or peroxides, to

induce cell death [83-85]. Nanotechnology assisted the phototherapy into further applications since most of the photosensitizers and photothermal agents were hydrophobic and had low tumor selectivity [15, 86-89].

However, phototherapy alone could not kill the tumor cells entirely due to the uneven light distribution and hypoxic condition in the tumor tissues, and it was easy to induce local recurrence and distant metastasis, especially for glioblastoma [90, 91]. Hence, combination of the phototherapy with chemotherapy could highlight the potential approaches for malignant glioblastoma therapies [92, 93]. For instance, Yu et al. designed an organoplatinum (II) metallacage (M) coordinated by photosensitizer 5, 10, 15, 20-tetra(4-pyridyl)porphyrin (TPP), therapeutic drug cis-(PEt₃)₂Pt(OTf)₂ (cPt), and disodium terephthalate (DSTP) [94]. The metallacage self-assembled with mPEG-*b*-PEBP and RGD-PEG-*b*-PEBP to form metallacage-loaded nanoparticles (MNPs) for long blood circulation and active targeting (Figure 6). The MNPs exhibited superior anticancer results against U87MG with no recurrence after single injection, which attributed to the synergistic photochemotherapy.

The inherent proteins were applied as drug carriers due to their superior biocompatibility. Liu's group utilized the human serum albumin (HSA) as the carrier to conjugate with the photosensitizer chlorin e6 (Ce6) and self-assemble with the anticancer drug paclitaxel (PTX) [95]. An acyclic Arg-Gly-Asp (cRGDyK) peptide, targeting to $\alpha v\beta 3$ -integrin overexpressed on tumor angiogenic endothelia, was chosen as the active targeting (HSA-Ce6-PTX-RGD). The accumulation of the nanoparticles was about 2.4 times higher than the non-targeting one *in vivo*. The survival time after the combination therapy prolonged to 40 days without a single death, while the mice treated with control groups exhibited lifetimes about 15-30 days.

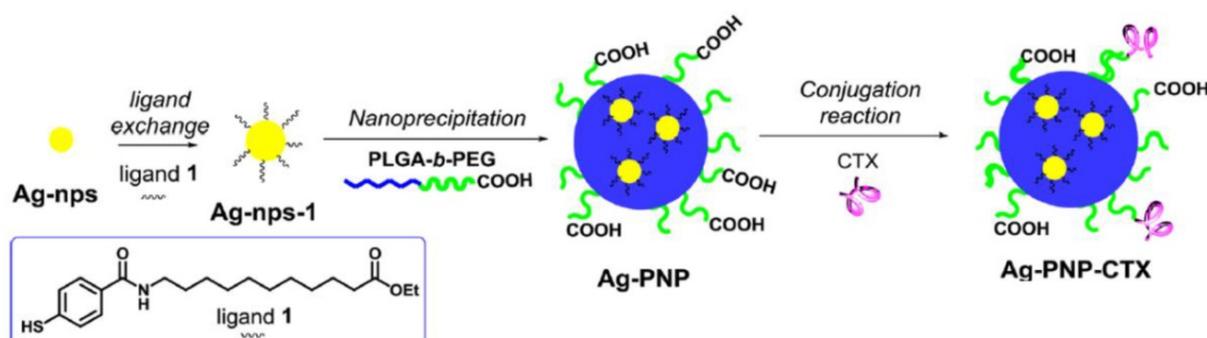


Figure 5. Schematic procedure for the synthesis of chlorotoxin-nanovectors (Ag-PNP-CTX). Adapted with permission from [81]. Copyright 2016 American Chemical Society.

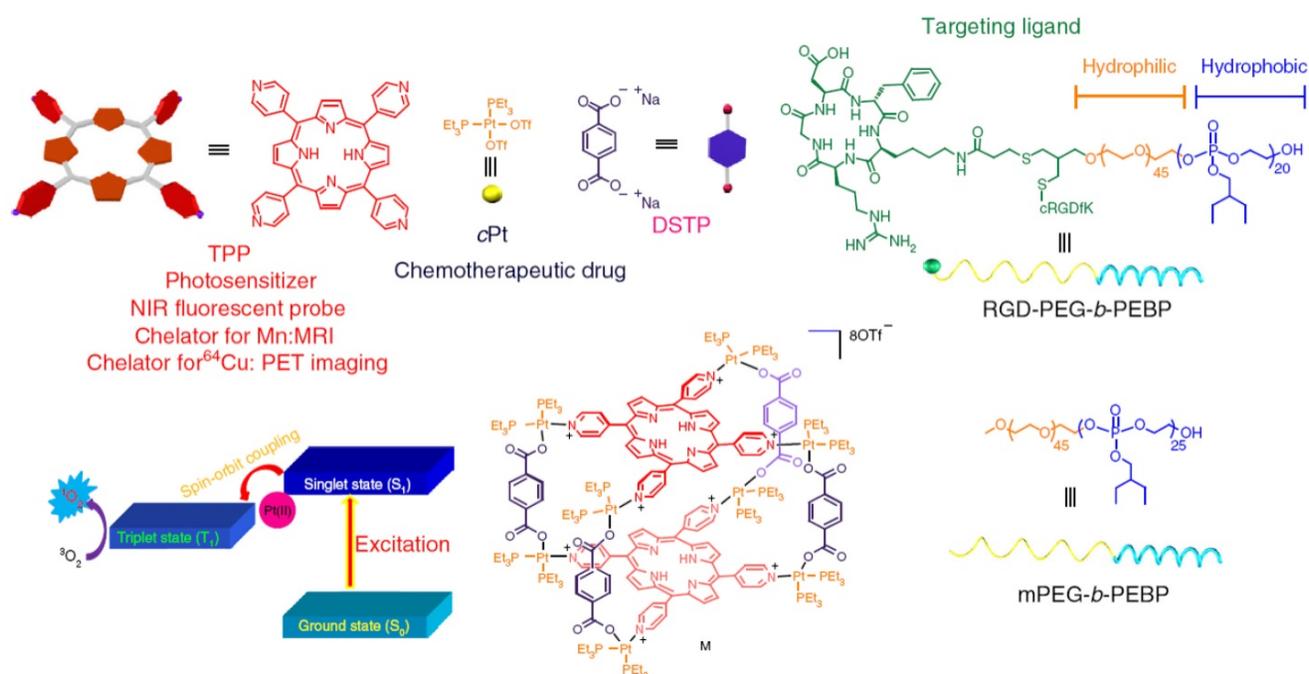


Figure 6. Schematic diagrams of the MNPs served as a multifunctional theranostic platform. The structures of TPP, cPt, DSTP, M, mPEG-*b*-PEBP and RGD-PEG-*b*-PEBP were illustrated. Adapted with permission from [94]. Copyright 2018 Springer Nature.

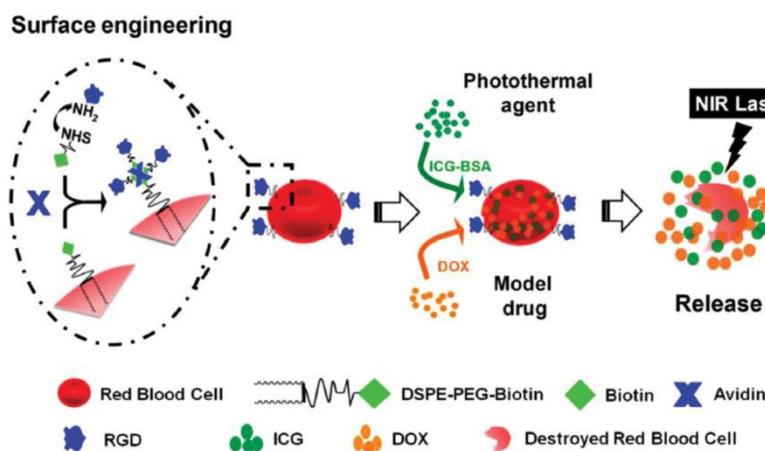


Figure 7. Schematic illustration for the preparation of the RBC-based system (IB&D@RBC-RGD) and the remotely controlled drug release under the NIR laser. Adapted with permission from [103]. Copyright 2015 WILEY-VCH Verlag GmbH & Co.

Gold nanoparticles were the attractive photothermal agents for cancer therapies due to their strong absorbance of NIR light and higher heat conversion efficiency [96-99]. Yongwei Hao et al. developed a tumor-targeting hybrid nanosystem by modifying Au on the surface of docetaxel (DTX)-loaded poly(lactide-co-glycolide) (PLGA) nanoparticles (ANG/GS/PLGA/DTX NPs) [100]. The U87MG cells treated with nanosystem showed a striking temperature increase for 12.3°C when exposed to 808 nm irradiation. And the tumor inhibition rate of the ANG/GS/PLGA/DTX NPs under irradiation was about 70%, which was the highest among all of the control groups for *in vivo* studies.

The biomimetic drug delivery systems coated with native cell membranes also attracted researchers'

attentions to fight against cancers with phototherapy [101, 102]. Liu's group exploited a RBC-based system (IB&D@RBC-RGD) with a tumor angiogenesis targeting (Figure 7) [103]. Anticancer drug DOX and photothermal agent indocyanine green-bovine serum albumin nanocomplexes were co-loaded into the modified RBCs. With deep penetrability of near-infrared (NIR) light, the IB&D@RBC-RGD significantly released the cargos and enhanced cytotoxicity against U87MG and 4T1 cells.

Phototherapy for GBM was restricted due to the limited tissue penetration and photosensitizers' delivery. Nanosystems were applied in photosensitizers' delivery for targeting transportation and reducing the non-specific accumulation in normal tissues. However, due to the highly invasive growth

of GBM, there usually had tumor recurrence and metastasis after phototherapy. Combined therapies by nanotechnology maximized the anti-glioma efficacy, which were attributed to the significant increase of chemotherapeutic drugs and photosensitizers/photothermal agents in the tumor cells. And these were confirmed by a growing amount of studies. But the challenges for this combined-therapeutic strategy against GBM still remained, including the ratio of the therapeutic drugs and photosensitizers/photothermal agents, the drug-to-light interval and the synergistic interaction of the therapeutic methods. All of these should greatly affect the anti-glioma effects and need comprehensive studies in future.

Combined chemotherapy with immunotherapy

Immunotherapy had received tremendous attentions in the treatment of cancers in the decades [104]. It could activate the body's own immune systems and induce the specific immune responses with tumor antigens to eliminate the tumor cells [105, 106]. Most importantly, it had the potential for long-term reduction of cancer metastasis and recurrence [107]. The immunotherapeutic strategies included the cancer vaccines, monoclonal antibodies, oncolytic virus, engineered T-cells and immunomodulation [11, 107, 108]. However, the major challenges of immunotherapy for glioblastoma were the lack of specific tumor antigens, limited immunogenicity of the cancer cells and the immunosuppressive environment of the tumors [109-111]. Intensive studies had been reported to promote the specific immune responses and exhibit effective immunotherapy for glioblastoma [112, 113].

The epidermal growth factor receptor variant III (EGFRvIII), an oncogenic variant of the EGFR, expressed in 20-30% of all glioblastoma [114]. It was exploited to cancer vaccine (Rindopepimut) and showed an encouraging progression-free survival in clinical trials [115, 116]. However, the combination of Rindopepimut and temozolomide in phase 3 trial did not show significant difference in overall survival for EGFRvIII-expressing glioblastoma [117]. This might be due to inappropriate combination of the drugs, which counteracted the immunological and chemical effects. And the unspecific tumor antigens, which were not sufficient to cover the heterogeneous brain tumors, also influenced the therapeutic effects of GBM.

The immune checkpoint was a costimulatory factor for regulating the antigen recognition in the process of immune responses. The inhibition of the checkpoint could enhance the anticancer immune activation [118-120]. Combination of immune check-

point with chemotherapy could stimulate the tumor immunity and sensitize the tumor to chemical agents [121-123]. For instance, Jing Kuang et al. designed an iRGD-modified silica nanoparticles (DOX@MSN-SS-iRGD&1MT) to simultaneously deliver the chemical agent doxorubicin (DOX) and the immune checkpoint inhibitor 1-methyltryptophan (1MT) for anti-glioma treatment (Figure 8) [119]. The active targeting iRGD guided the nanoparticles to penetrate across BBB and improved drug accumulation in orthotopic brain tumors. The nanoparticles induced antitumor immune responses and regulated the immunosuppressive microenvironment. The chemo-immunotherapeutic therapy significantly extended the medium survival with a 50% durable cure rate.

The immunomodulation of the microenvironment of the glioblastoma was another promising approach for tumor regression [124]. For example, Padma Kadiyala et al. reported a nanodiscs (DTX-sHDL-CpG) based on high-density lipoprotein (HDL), the synthetic apolipoprotein-I (ApoA-1) [125], which encompassed the anticancer drug docetaxel (DTX) and the immune activator CpG (Figure 9). The nanosystem could elicited antitumor CD8⁺ T cell responses and developed the anti-GBM immunological memory. The DTX-sHDL-CpG showed moderate effect on tumor growth, and the combination of DTX-sHDL-CpG with radiation significantly increased the long-term survival in 80% of GBM-bearing mice.

Current advances demonstrated that the chemotherapy with temozolomide (TMZ) could induce and aggravate the immunosuppressive tumor microenvironment [126, 127]. Regulating the tumor microenvironment by immunotherapy enabled the GBM more sensitive to chemotherapy. Qiao et al. in our team exploited a dual targeting and ROS responsive nanosystem (ALBTA) for immunochemotherapy (Figure 10) [128]. SiRNA against tumor growth factor β (siTGF- β) was employed to modify the immune microenvironment of glioblastoma and improve the efficacy of TMZ. To deliver these two drugs in a controlled manner, firstly, ROS-responsive poly[(2-acryloyloxyethyl)(p-boronic acid benzyl) diethylammonium bromide] (BAP) was chosen to assemble with siTGF- β and triggered to release in cytoplasm of tumor cells. Secondly, zwitterionic lipid distearoyl phosphoethanol-aminepolycarboxybetaine (DSPE-PCB) based envelope (ZLE) was selected to enhance the delivery of TMZ and BAP/siTGF- β (AN@siTGF- β) into cytoplasm. The core-shell structural nanoparticles (ALBTA) significantly improved the immunosuppressive microenvironment *in vivo*, and increased the medium survival time from 19 d (for untreated control group) to 36 d without obvious systemic toxicity in the intracranial glioblastoma mice.

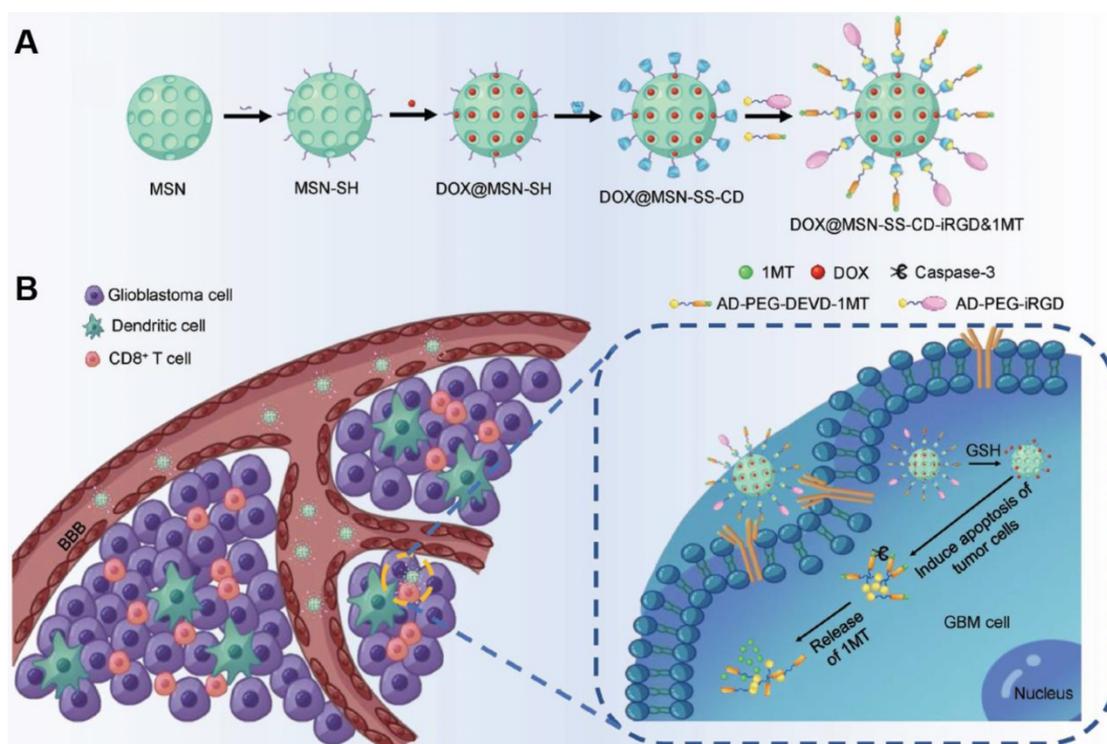


Figure 8. A) The synthetic route of the iRGD-modified silica nanoparticles DOX@MSN-SS-iRGD&1MT. B) The illustration of DOX@MSN-SS-iRGD&1MT showed active targeting and drug release. Adapted with permission from [119]. Copyright 2018 WILEY-VCH Verlag GmbH & Co.

Preparation of DTX-sHDL-CpG nanodiscs

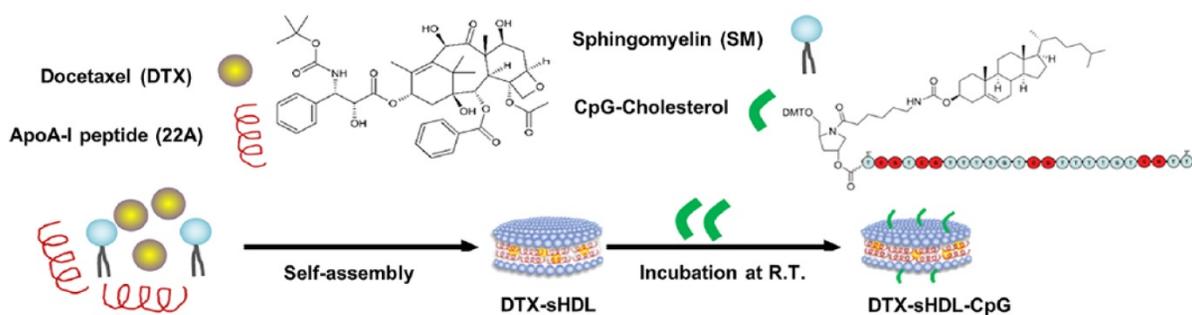


Figure 9. The formulation route of the nanodiscs DTX-sHDL-CpG. Adapted with permission from [125]. Copyright 2019 American Chemical Society.

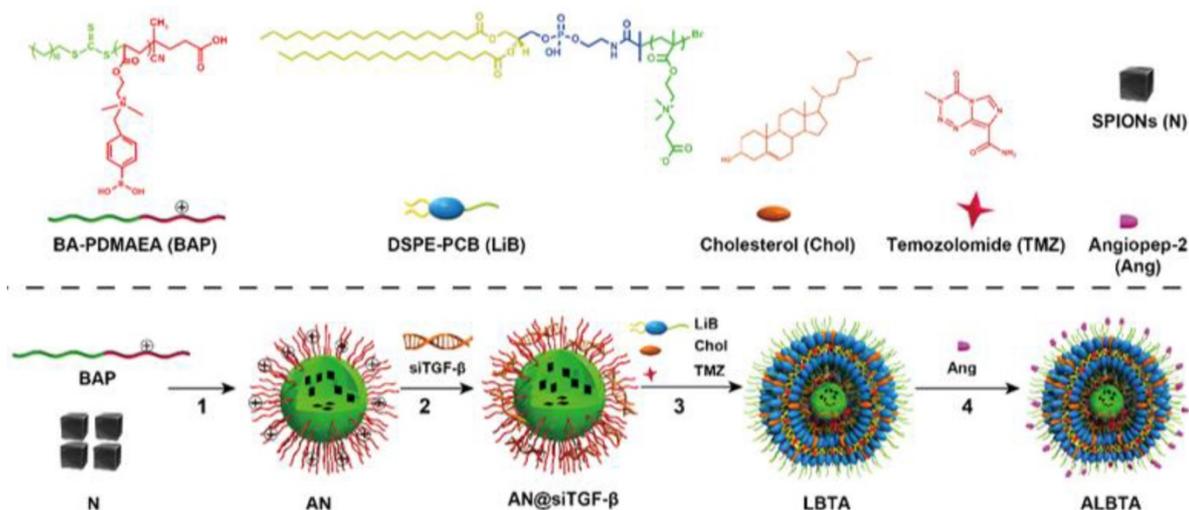


Figure 10. The chemical structural formula of the components and the preparation route of the nanosystem ALBTA. Adapted with permission from [128]. Copyright 2018 WILEY-VCH Verlag GmbH & Co.

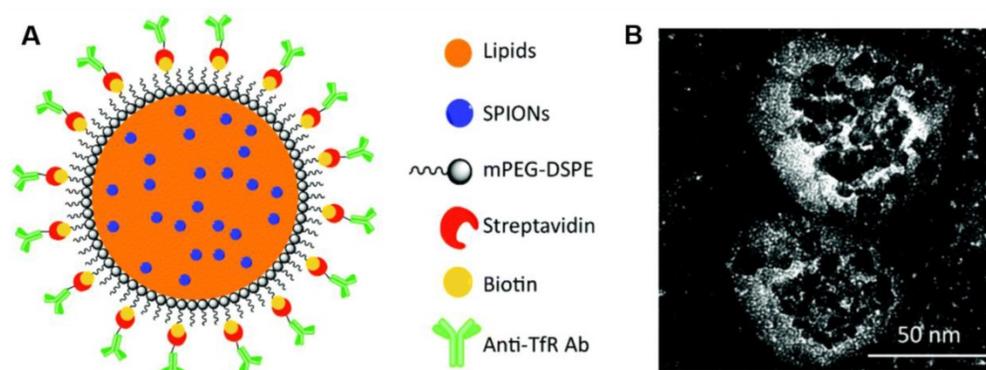


Figure 11. A) The illustration of the lipid magnetic nanovectors (LMNVs) loaded with SPIONs and functionalized with an anti-transferrin receptor antibody (anti-TfR Ab). B) The high magnification transmission electron microscopy (TEM) images of functionalized nanovectors (AbLMNVs). Adopted with permission from [143]. Copyright 2019 The Royal Society of Chemistry.

Despite the impressive advances in GBM treatments, the efficacy of immunotherapy still needed further improvements for desirable overall survival. For example, the identification of the specific glioblastoma antigens was required to reinforce the specific immunity responses. Combination of immunotherapy with chemotherapy *via* nanotechnology obviously enhanced the sensitivity of the GBM cells to chemotherapeutic drugs, and regulated the microenvironment of tumors. However, the timing of the immunotherapy for combination [127, 129] and the delivery strategies had critical influences on the therapeutic effects, which needed more efforts to investigate.

Combined chemotherapy with magnetothermal therapy

Hyperthermia therapy had been developed as a safe and effective complementary therapy for cancer treatments [130]. Several magnetic nanomaterials had been used in hyperthermia applications, including iron oxide nanoparticles, cobalt ferrite, iron platinum nanoparticles and so on [131-133]. Iron oxide nanoparticles were usually applied as the thermo-agents in cancer treatments and entered into clinical trials [134-136]. Since the most thermo-agents were the metal nanomaterials, they were preferable for phagocytosis by macrophages rather than the glioblastoma cells. Moreover, the challenges for magnetothermal therapy were the focused heating on the tumor sites with optimum temperature, and not harmed the surrounding normal tissues [137, 138]. Hence, nanotechnology was introduced to this therapy for active targeting and inducing more thermo-agents into diseased areas, which promoted the anti-tumor effects [139-141].

Plenty of evidences showed that the magnetothermal therapy against GBM was more effective by combining with chemotherapy [142]. They could enhance the sensitivity of the tumor cells to therapeutic

drugs. For example, Gianni Ciofani's group developed a lipid magnetic nanovectors (LMNVs) with 1,2-distearoyl-*sn*-glycero-3-phosphoethanolamine-conjugated poly(ethylene glycol) (DSPE-PEG) (Figure 11) [143, 144]. It was functionalized with the antibody against the transferrin receptor (TfR) for dual targeting to endothelial cells of BBB and GBM cells. The superparamagnetic iron oxide nanoparticles (SPIONs) and temozolomide (TMZ) were co-loaded in the lipid nanovectors (LMNVs). The anti-TfR antibodies facilitated targeting to glioblastoma spheroids in multi-cellular *in vitro* models, with 40.5% *vs.* 8.1% for non-target nanovectors. The combined treatment with magnetothermal hyperthermia was able to efficiently disintegrate the GBM spheroids and induce significant tumor cell death. Under magnetic fields, the nanovector induced mild hyperthermia (43°C) and enhanced anticancer effect against U87 MG cells.

Besides the lipid nanovectors, many synthetic polymeric nanosystems were also exploited as delivery vectors for combined treatments. Yin et al. exploited a highly magnetic zinc-doped iron oxide nanoparticle ($ZnFe_2O_4$) decorated with branched polyethyleneimine (PEI) to deliver lethal-7a miRNA (let-7a) (MNP-PEI/miRNA/PEI complexes), which was a tumor suppressor that inhibited malignant growth by targeting the downstream effectors of heat shock proteins (HSPs) (Figure 12) [145]. The nanosystem MNP-PEI/miRNA/PEI exhibited obvious apoptosis of brain cancer cells and led to caspase-3 mediated apoptosis by combining with magnetic hyperthermia.

Wen-Chia Huang et al. reported a tumortropic adipose-derived stem cells (ADSCs) encapsulating the nanotherapeutics (SPNPs) for chemo-thermotherapy [146]. The nanotherapeutics were co-assembled with poly(γ -glutamic acid-co-distearyl γ -glutamate), poly(lactic-co-glycolic acid), the anticancer drug paclitaxel (PTX) and oleic acid-coated superparamagnetic iron oxide NPs. With the guide of ADSCs to tumor sites,

hyperthermia was activated by the high frequency magnetic field (HFMF) and triggered drug release in tumor cells. The data *in vivo* demonstrated that the combined therapy extended the lifespan of the brain tumor-bearing mice almost 3 folds, from 11 to 31 days, in comparison with the PBS control group. Whereas, the ADSCs without HFMF activation was only 15 days.

Although the combination of magnetothermal therapy with chemotherapy showed better inhibition against the GBM than single therapy, the combined strategy had not reached the required threshold for efficient clinical applications in GBM treatments. The reasons for this were as follows. Firstly, the magnetothermal efficiency of the magnetic systems was insufficient for clinical applications. The amount and characteristics of magnetic materials and their intratumoral distribution were the key factors for therapeutic effects. Secondly, the timing of the magnetothermal therapy and chemotherapy should greatly influence the overall results of treatments, which needed further investigations. Finally, the design of the delivery systems for the combination was very important, since these would directly affect the anti-tumor effects. Albeit these challenges, such combinations offered multimodal therapeutic strategies in a single treatment. The thermo-agents in the magnetothermal therapy, like the iron oxide nanoparticles, could also exhibit magnetic resonance imaging for tracing the combined nanosystems and monitoring the drug accumulation in *in vivo* distribution.

Combined phototherapy with immunotherapy

Near-infrared phototherapy (NIR-PIT) was a recently emerging therapy for cancers [147]. It employed a conjugate, which consisted of a near-infrared photosensitizer and a target-specific antibody. They could selectively kill the cancer cells and activate the body's antitumor immune responses. And few of the photo-immunoconjugates displayed toxicity until the activation by the light with specific wavelength. The NIR-PIT was applied for clinical trials against inoperable head and neck cancers by using cetuximab-IR700 (RM1929), and they exhibited promising results [148, 149].

Utilizing the unique character of NIR-PIT, Jing et al. chose the AC133 monoclonal antibody to conjugate with the photosensitizer IR700 (AC133-IR700) for glioblastoma treatments [150]. The AC133 could recognize the CD133 on human cancer stem cells, and enhance the active targeting to orthotopic AC133⁺ brain tumor. The treated group extended the median survival time to 33.5 days in comparison with non-irradiated control group (with 15 days of survival time). The epidermal growth factor receptor (EGFR) was the general genetic mutation in GBM. Hence, an EGFR-specific antibody (Z_{EGFR-03115}) was conjugated to the photosensitizer IR700DX for glioblastoma treatments [151]. The tumor distribution of the Z_{EGFR-03115}-IR700DX was 6-fold higher than that of the control group Z_{Taq}-IR700DX. And they exhibited a significant inhibition of tumor growth in U87-MGvIII bearing mice over a period of 18 days.

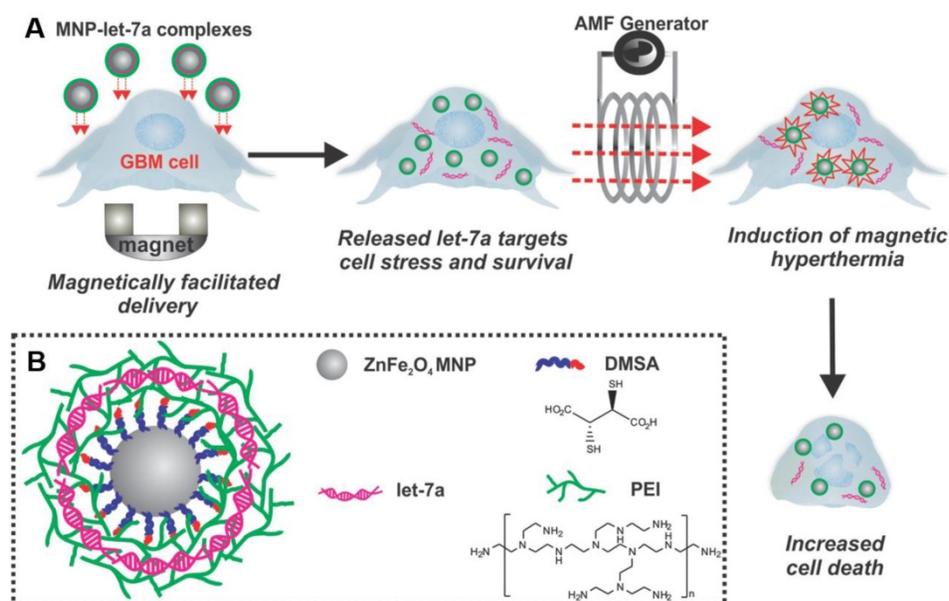


Figure 12. The MNP-PEI/miRNA/PEI complexes enhanced the treatment against brain cancer. A) The MNP complexes were first delivered to GBM cells, which was enhanced by magnetofection. Once inside the cells, let-7a miRNA was released for targeting downstream effectors of HSPs. This sensitized the cancer cells to subsequent magnetic hyperthermia for enhanced apoptosis. B) MNPs were complexed with let-7a miRNA branched PEI via a layer-by-layer approach. Adapted with permission from [145]. Copyright 2014 WILEY-VCH Verlag GmbH & Co.

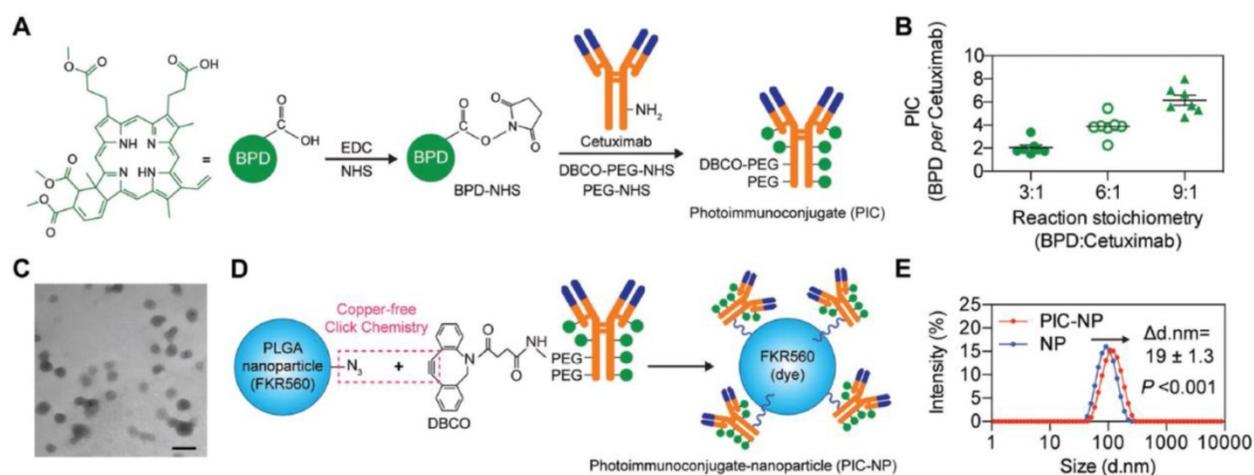


Figure 13. The synthesis route of photo-immunoconjugate nanoparticles (PIC-NPs). A) The illustration of PIC synthesis. Benzoporphyrin derivative (BPD) photosensitizers were conjugated to PEGylated cetuximab via carbodiimide crosslinker chemistry. B) The varied stoichiometry of BPD reacted with cetuximab. C) Transmission electron microscopy (TEM) image of poly(ethylene glycol)-poly(lactic-co-glycolic acid) (PEG-PLGA) polymeric nanoparticles prepared via nanoprecipitation method. Scale bar 100 nm. D) Schematic depiction of PIC-NP synthesis via copper-free click chemistry. Azide-containing FKR560 dye-loaded PLGA nanoparticles were reacted with the dibenzocyclooctyne (DBCO)-containing PICs to form PIC-NPs. E) Covalent conjugation of PICs onto 80 nm PEG-PLGA NPs to form monodispersed PIC-NPs around 100 nm in diameter. Adapted with permission from [152]. Copyright 2018 WILEY-VCH Verlag GmbH & Co.

However, due to the limited ratio of the photosensitizer-to-antibody, the amount of photo-immunoconjugates delivered to tumor cells was not sufficient. Hence, Huang et al. adopted poly(lactic-co-glycolic acid) (PLGA) nanoparticles to overcome the obstacles [152]. The photo-immunoconjugates benzoporphyrin derivative (BPD)-cetuximab was bound to PLGA nanoparticles by click coupling (Figure 13). The photo-immunoconjugate nanoparticle (PIC-NP) could significantly enhance the photosensitizers to cancer cells, and increase light-activated cytotoxicity in EGFR-overexpressing U87 cells. The nanotechnology improved the overall treatment outcomes comparing with the photo-immunoconjugates.

Compared to classic photodynamic therapy, the photoimmunotherapy utilized the specific antibodies that could facilitate targeting the tumor cells without damaging the normal cells. It also enhanced photosensitizer delivery into tumors and increased the light-activated cytotoxicity. Despite these promising advances, the limited ratio of photosensitizer-to-antibody restricted the amount of photosensitizers delivered by photo-immunoconjugates. Hence, the nanodelivery systems provided the larger loading capacity and increased accumulation in the tumors. They efficiently maximized the photo-toxicity to the targeted tissues. However, the photoimmunotherapy for glioblastoma was still in its infancy. The immune activation was not studied well for these anti-glioma treatments.

Combined gene therapy with immunotherapy

Gene therapy had emerged as a novel treatment for various human diseases including cancers. It could specifically regulate the oncogenes in the anti-tumor

treatments [153, 154]. The adenovirus-mediated gene therapy with sitimagene ceradenovec and ganciclovir after resection increased survival time of patients with newly diagnosed glioblastoma multiforme [155]. Hence, the gene therapy was usually proposed as a useful adjuvant for the current glioblastoma treatments [156]. The combination of gene therapy with immunotherapy should increase the whole outcomes of glioblastoma treatments [157, 158]. For example, Maria-Carmela Speranza et al. utilized the nonreplicating adenovirus containing the HSV TK gene (AdV-tk) to potentiate the anti-PD-1 efficacy in syngeneic glioblastoma mouse models. The AdV-tk could upregulate the IFN signaling and increase the PD-L1 levels. And cytotoxic CD8⁺ T cells were induced to accumulate in tumors. The combination treatment significantly increased the percentage of long-term survival (LTS ≥ 100 days) animals from 30-50% (single agents) to 88% [159].

Despite the potentials of gene therapy, the intrinsic characteristics of the gene limited the efficient delivery to tumor sites and hindered their progress to the clinical applications [26, 160, 161]. Especially the crossing through the BBB exhibited an extra challenge for anti-glioma treatments. Nanoparticles could compensate the shortages of the gene and safely deliver gene and immunotherapeutic agents to brain tumors [162]. For instance, Gulsah Erel-Akbaba et al. developed a cyclic peptide iRGD (CCRGDKGPDC)-decorated solid lipid nanoparticle (SLN) to deliver siRNA against both the epidermal growth factor receptor (EGFR) and programmed cell death ligand-1 (PD-L1) for combined therapy (Figure 14) [163]. The targeting nanoparticles f(SLN)-iRGD: siRNA led to 54.7% and 58.6% decrease for EGFR and

PD-L1, respectively. Moreover, the median survival of the mice treated with f(SLN)-iRGD:siRNA with radiation increased to 38 days. These three combinations largely improved the survival of the GL261-bearing mice.

Evidences exhibited that the gene therapy was usually applied to regulate the immunosuppressive signals and enhance the systemic therapy in glioma treatments [164]. These could synergistically augment the immune responses with immunotherapy. But the reported studies on this strategy were less. The gene therapy and immunotherapy usually served as adjuvant treatments in existing anti-glioma therapies. Moreover, the lack of specific genes and antigens was still the critical barriers for satisfied treatments.

Conclusion and Perspective

Glioblastoma is a complex and deadly disease in central nervous system. The high infiltration, heterogeneity, rapid growth rate and regeneration ability of the GBM stem cells result in poor prognosis and high recurrence. Furthermore, the existence of the blood-brain barrier (BBB) greatly hinders adequate accumulation of the therapeutic drugs in tumor sites, which also induces drug-resistance to therapy. With the deep understanding of glioblastoma, improved therapeutic strategies are exploited to enhance the efficacy against glioblastoma. They mainly include chemotherapy, gene therapy, radiotherapy, immunotherapy, phototherapy and magnetothermal therapy. Considering the intricate characteristics of GBM, combined-therapeutic strategies are developed for synergistic anti-glioma effects and reduced drug-resistance [165]. And the pros and cons of the combined therapies are summarized and illustrated in Table 1.

Table 1. The pros and cons of the combined treatments against glioblastoma.

Combined treatments	Advantages	Disadvantages
Chemotherapies	Reduced drug resistance; Multiple mechanisms	Non-specificity to tumor cells
Chemotherapy and radiotherapy	Increased tumor sensitivity to chemotherapeutic drugs	Hard to control radiation margin; Timing of radiotherapy
Chemotherapy and phototherapy	Multiple mechanisms	Limited tissue penetration of light; Drug to light interval
Chemotherapy and immunotherapy	Multiple mechanisms; Reduced metastasis and recurrence	Lack of specific antigens; Timing of immunotherapy
Chemotherapy and magnetothermal therapy	Multimodality; Theranostics	Thermal efficiency; Hard to focus heating on tumor sites
Phototherapy and immunotherapy	Enhanced targeting; Reduced metastasis and recurrence	Restricted ratio of photosensitizer to antibody;
Gene therapy and immunotherapy	Indirect anti-tumor effect; Reduced drug resistance	Lack of specific genes and antigens

In review of the intensive studies for GBM, some problems should be settled for optimizing GBM treatment. Firstly, the choices of the therapeutic agents in combined strategies are very important for overall treatment outcomes. The therapeutic mechanisms of the agents should not counteract the pharmaceutical effects for each other. Some of the combinations in clinical trials do not obtain the satisfied results compared with monotherapies [40, 166]. It has been reported that the combination of erlotinib and sorafenib in phase II study exhibits significant pharmacokinetic interactions, which may display negative impacts on the therapeutic efficacy [167]. Moreover, the recent therapeutic agents selected for therapy can not cover the critical points in the progression of GBM. Therefore, multi-modal combined strategies should be tested in anti-glioma treatments. Secondly, the timing of the therapeutic strategies in combination needs more investigations. For instance, the administration timing of

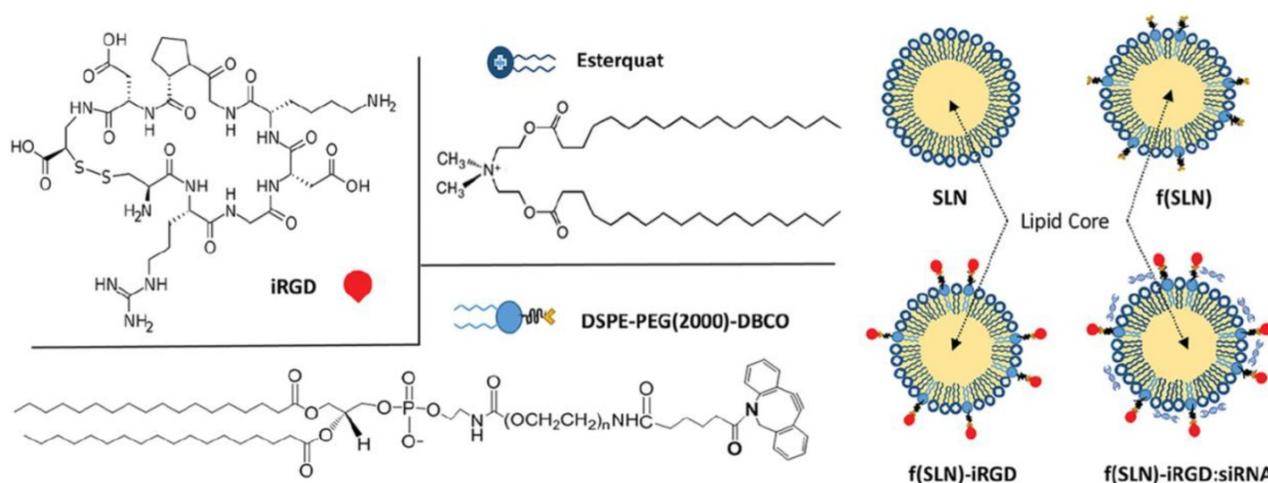


Figure 14. The chemical structures of the DSPE-PEG(2000)-DBCO, Esterquat and iRGD peptide, and the construction of the nanoparticles SLN, f(SLN), f(SLN)-iRGD and f(SLN)-iRGD:siRNA. Adapted with permission from [163]. Copyright 2019 American Chemical Society.

chemotherapy in combinations obviously affects the efficacy of immunotherapy [127]. Hence, alternative therapeutic strategies are required to attempt for maximizing the therapeutic effects. Finally, the suitable delivery systems are demanded for effective drug delivery. Increasing studies demonstrate that nanosystems perform preferable therapeutic results in comparison with free therapeutic agents. The delivery platforms possess active brain targeting and diverse stimuli-responses [168, 169]. Therefore, these strategies significantly promote the loading agents to cross BBB and accumulate in GBM cells. And they extend the overall survival time of GBM in animal experimental studies. However, glioblastoma remains a complicated cancer. The current reported targets are not specific for the endothelial cells of BBB and tumor cells. The further and better understanding of the intricate tumor microenvironment is still required. Hence, the specific targets and the suitable stimuli-responses of the GBM are required to exploit, which will facilitate the diverse agents to release in the right places and ensure sufficient therapeutic agents in GBM cells. Moreover, most of the biomaterials in studies have not been approved by FDA, which aggravate the difficulties for clinical translation.

Although the current combined-therapeutic strategies have enhanced the efficacy of GBM treatments, great efforts are still required on the understanding the complicated mechanisms of the growth, progression and invasion process of GBM. Multidisciplinary knowledge of GBM will guide the development of optimal therapeutic strategies and rational nanosystems for satisfied outcomes, and direct the investigation for other malignant diseases in central nerves system.

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Competing Interests

The authors have declared that no competing interest exists.

References

- Reifenberger G, Wirsching H-G, Knobbe-Thomsen CB, Weller M. Advances in the molecular genetics of gliomas - implications for classification and therapy. *Nat Rev Clin Oncol*. 2017; 14: 434-52.
- Shergalis A, Armand Bankhead I, Luesakul U, Muangsin N, Neamati N. Current challenges and opportunities in treating glioblastoma. *Pharmacol Rev*. 2018; 70: 412-45.
- Glas M, Happold C, Rieger J, Wiewrodt D, Bähr O, Wick JPS, et al. Long-term survival of patients with glioblastoma treated with radiotherapy and lomustine plus temozolomide. *J Clin Oncol*. 2009; 27: 1257-61.
- Alifieris C, Trafalis DT. Glioblastoma multiforme: pathogenesis and treatment. *Pharmacol Ther*. 2015; 152: 63-82.
- Kaneko S, Eljamel MS. Fluorescence image-guided neurosurgery. *Future Oncol*. 2017; 13: 2341-8.
- Li Y, Rey-Dios R, Roberts DW, Valdés PA, Cohen-Gadol AA. Intraoperative fluorescence-guided resection of high-grade gliomas: a comparison of the present techniques and evolution of future strategies. *World Neurosurg*. 2014; 82: 175-85.
- D'Amico RS, Englander ZK, Canoll P, Bruce JN. Extent of resection in glioma-A review of the cutting edge. *World Neurosurg*. 2017; 103: 538-49.
- Lieberman FS, Wang M, Robins HI, Tsien CI, Walter J, Curran J, Werner-Wasik M, et al. Phase 2 study of radiation therapy plus low-dose temozolomide followed by temozolomide and irinotecan for glioblastoma: NRG oncology RTOG trial 0420. *Int J Radiat Oncol Biol Phys*. 2019; 103: 878-86.
- Bruhn H, Strandéus M, Milos P, Hallbeck M, Vrethem M, Lind J. Improved survival of Swedish glioblastoma patients treated according to Stupp. *Acta Neurol Scand*. 2018; 138: 332-7.
- Warren KT, Liu L, Liu Y, Milano MT, Walter KA. The impact of timing of concurrent chemoradiation in patients with high-grade glioma in the era of the Stupp protocol. *Front Oncol*. 2019; 9: 186.
- Lim M, Xia Y, Bettgeowda C, Weller M. Current state of immunotherapy for glioblastoma. *Nat Rev Clin Oncol*. 2018; 15: 422-42.
- Barani IJ, Larson DA. Radiation therapy of glioblastoma. USA: Springer; 2015.
- Clarke J, Butowski N, Chang S. Recent advances in therapy for glioblastoma. *Arch Neurol*. 2010; 67: 279-83.
- Stupp R, Hegi ME, Gilbert MR, Chakravarti A. Chemoradiotherapy in malignant glioma: Standard of care and future directions. *J Clin Oncol*. 2007; 25: 4127-36.
- Tsai Y-C, Vijayaraghavan P, Chiang W-H, Chen H-H, Liu T-I, Shen M-Y, et al. Targeted delivery of functionalized upconversion nanoparticles for externally triggered photothermal/photodynamic therapies of brain glioblastoma. *Theranostics*. 2018; 8: 1435-48.
- Jin X, Kim LJY, Wu Q, Wallace LC, Prager BC, Sanvoranart T, et al. Targeting glioma stem cells through combined BMI1 and EZH2 inhibition. *Nat Med*. 2017; 23: 1352-61.
- Osuka S, Meir EGV. Overcoming therapeutic resistance in glioblastoma: the way forward. *J Clin Invest*. 2017; 127: 415-56.
- Sun X, Wang G, Zhang H, Hu S, Liu X, Tang J, et al. The blood clearance kinetics and pathway of polymeric micelles in cancer drug delivery. *ACS Nano*. 2018; 12: 6179-92.
- Parrish KE, Cen L, Murray J, Calligaris D, Kizilbash S, Mittapalli RK, et al. Efficacy of PARP inhibitor rucaparib in orthotopic glioblastoma xenografts is limited by ineffective drug penetration into the central nervous system. *Mol Cancer Ther*. 2015; 14: 2735-43.
- Sun X, Yan X, Jacobson O, Sun W, Wang Z, Tong X, et al. Improved tumor uptake by optimizing liposome based RES blockade strategy. *Theranostics*. 2017; 7: 319-28.
- Sun Q, Zhou Z, Qiu N, Shen Y. Rational design of cancer nanomedicine: nanoproperty integration and synchronization. *Adv Mater*. 2017; 29: 1606628.
- Kingwell K. Drug delivery: new targets for drug delivery across the BBB. *Nat Rev Drug Discov*. 2016; 15: 84-5.
- Banks WA. From blood-brain barrier to blood-brain interface: new opportunities for CNS drug delivery. *Nat Rev Drug Discov*. 2016; 15: 275-92.
- Kreuter J. Nanoparticulate systems for brain delivery of drugs. *Adv Drug Deliv Rev*. 2001; 47: 65-81.
- Zuchero YJY, Chen X, Bien-Ly N, Bumbaca D, K.Tong R, Gao X, et al. Discovery of novel blood-brain barrier targets to enhance brain uptake of therapeutic antibodies. *Neuron*. 2016; 89: 70-82.
- Cheng Y, Morshed R, Auffinger B, Tobias AL, Lesniak MS. Multifunctional nanoparticles for brain tumor imaging and therapy. *Adv Drug Deliv Rev*. 2014; 66: 42-57.
- Jensen SA, Day ES, Ko CH, Hurley LA, Luciano JP, Kouri FM, et al. Spherical nucleic acid nanoparticle conjugates as an RNAi-based therapy for glioblastoma. *Sci Transl Med*. 2013; 5: 209ra152.
- Tang W, Fan W, Lau J, Deng L, Shen Z, Chen X. Emerging blood-brain-barrier-crossing nanotechnology for brain cancer theranostics. *Chem Soc Rev*. 2019; 48: 2967-3014.
- Jiang X, Wang C, Fitch S, Yang F. Targeting tumor hypoxia using nanoparticle-engineered CXCR4-overexpressing adipose-derived stem cells. *Theranostics*. 2018; 8: 1350-60.
- Gao H, Zhang S, Cao S, Yang Z, Pang Z, Jiang X. Angiopep-2 and activatable cell-penetrating peptide dual-functionalized nanoparticles for systemic glioma-targeting delivery. *Mol Pharm*. 2014; 11: 2755-63.
- Wanjale MV, Kumar GSV. Peptides as a therapeutic avenue for nanocarrier-aided targeting of glioma. *Expert Opin Drug Deliv*. 2017; 14: 811-24.
- Huang R, Harmsen S, Samii JM, Karabeber H, Pitter KL, Holland EC, et al. High precision imaging of microscopic spread of glioblastoma with a targeted ultrasensitive SERRS molecular imaging probe. *Theranostics*. 2016; 6: 1075-84.
- Khan AR, Yang X, Fu M, Zhai G. Recent progress of drug nanoformulations targeting to brain. *J Control Release*. 2018; 291: 37-64.

34. Jiang Y, Yang W, Zhang J, Meng F, Zhong Z. Protein toxin chaperoned by LRP-1-targeted virus-mimicking vesicles induces high-efficiency glioblastoma therapy *in vivo*. *Adv Mater*. 2018; 30: 1800316.
35. Sita TL, Kouri FM, Hurley LA, Merkel TJ, Chalastanis A, May JL, et al. Dual bioluminescence and near-infrared fluorescence monitoring to evaluate spherical nucleic acid nanoconjugate activity *in vivo*. *Proc Natl Acad Sci USA*. 2017; 114: 4129-34.
36. Mangraviti A, Tzeng SY, Kozielski KL, Wang Y, Jin Y, Gullotti D, et al. Polymeric nanoparticles for nonviral gene therapy extend brain tumor survival *in vivo*. *ACS Nano*. 2015; 9: 1236-49.
37. Taal W, Oosterkamp HM, Walenkamp AME, Dubbink HJ, Beerepoot LV, Hanse MCJ, et al. Single-agent bevacizumab or lomustine *versus* a combination of bevacizumab plus lomustine in patients with recurrent glioblastoma (BELOB trial): a randomised controlled phase 2 trial. *Lancet Oncol*. 2014; 15: 943-53.
38. Lun X, ConnorWells J, Grinshtein N, King JC, Hao X, Dang N-H, et al. Disulfiram when combined with copper enhances the therapeutic effects of temozolomide for the treatment of glioblastoma. *Clin Cancer Res*. 2016; 22: 3860-75.
39. Wick W, Gorlia T, Bendszus M, Taphoorn M, Sahm F, Harting I, et al. Lomustine and bevacizumab in progressive glioblastoma. *N Engl J Med*. 2017; 377: 1954-63.
40. Galanis E, Anderson SK, Lafky JM, Uhm JH, Giannini C, Kumar SK, et al. Phase II study of bevacizumab in combination with sorafenib in recurrent glioblastoma (N0776): a north central cancer treatment group trial. *Clin Cancer Res*. 2013; 19: 4816-23.
41. Lee EQ, Reardon DA, Schiff D, Drappatz J, Muzikansky A, Grimm SA, et al. Phase II study of panobinostat in combination with bevacizumab for recurrent glioblastoma and anaplastic glioma. *Neuro Oncol*. 2015; 17: 862-7.
42. Verreault M, Wehbe M, Strutt D, Masin D, Anantha M, Walker D, et al. Determination of an optimal dosing schedule for combining Irinophore CTM and temozolomide in an orthotopic model of glioblastoma. *J Control Release*. 2015; 220: 348-57.
43. Lundy DJ, Lee K-J, Peng I-C, Hsu C-H, Lin J-H, Chen K-H, et al. Inducing a transient increase in blood-brain barrier permeability for improved liposomal drug therapy of glioblastoma multiforme. *ACS Nano*. 2019; 13: 97-113.
44. Guo L, Fan L, Pang Z, Ren J, Ren Y, Li J, et al. TRAIL and doxorubicin combination enhances anti-glioblastoma effect based on passive tumor targeting of liposomes. *J Control Release*. 2011; 154: 93-102.
45. Sharpe MA, Marcano DC, Berlin JM, Widmayer MA, Baskin DS, Tour JM. Antibody-targeted nanovectors for the treatment of brain cancers. *ACS Nano*. 2012; 6: 3114-20.
46. Lam FC, Morton SW, Wyckoff J, Han T-LV, Hwang MK, Maffa A, et al. Enhanced efficacy of combined temozolomide and bromodomain inhibitor therapy for gliomas using targeted nanoparticles. *Nat Commun*. 2018; 9: 1991.
47. Jiang Y, Zhang J, Meng F, Zhong Z. Apolipoprotein E peptide-directed chimeric polymersomes mediate an ultrahigh-efficiency targeted protein therapy for glioblastoma. *ACS Nano*. 2018; 12: 11070-9.
48. Locatelli E, Naddaka M, Uboldi C, Loudos G, Fragogeorgi E, Molinari V, et al. Targeted delivery of silver nanoparticles and alisertib: *in vitro* and *in vivo* synergistic effect against glioblastoma. *Nanomedicine* 2014; 9: 839-49.
49. Johannessen T-CA, Bjerkvig R, Tysnes BB. DNA repair and cancer stem-like cells-potential partners in glioma drug resistance? *Cancer Treat Rev*. 2008; 34: 558-67.
50. Batista LFF, Roos WP, Christmann M, Menck CFM, Kaina B. Differential sensitivity of malignant glioma cells to methylating and chloroethylating anticancer drugs: p53 determines the switch by regulating xpc, ddb2, and DNA double-strand breaks. *Cancer Res*. 2007; 67: 11886-95.
51. Ma J, Murphy M, O'Dwyer PJ, Berman E, Reed K, Gallo JM. Biochemical changes associated with a multidrug-resistant phenotype of a human glioma cell line with temozolomide-acquired resistance. *Biochem Pharmacol*. 2002; 63: 1219-28.
52. Murphy SF, Varghese RT, Lamouille S, Guo S, Pridham KJ, Kanabur P, et al. Connexin 43 inhibition sensitizes chemoresistant glioblastoma cells to temozolomide. *Cancer Res*. 2016; 76: 139-49.
53. Adair JE, Johnston SK, Mrugala MM, Beard BC, Guyman LA, Baldock AL, et al. Gene therapy enhances chemotherapy tolerance and efficacy in glioblastoma patients. *J Clin Invest*. 2014; 124: 4082-92.
54. Mrugala MM, Chamberlain MC. Mechanisms of disease: temozolomide and glioblastoma - look to the future. *Nat Clin Pract Oncol*. 2008; 5: 476-86.
55. Kim S-S, Rait A, Kim E, Pirollo KF, Nishida M, Farkas N, et al. A nanoparticle carrying the p53 gene targets tumors including cancer stem cells, sensitizes glioblastoma to chemotherapy and improves survival. *ACS Nano*. 2014; 8: 5494-514.
56. Zhang W, Hu X, Shen Q, Xing D. Mitochondria-specific drug release and reactive oxygen species burst induced by polyprodrug nanoreactors can enhance chemotherapy. *Nat Commun*. 2019; 10: 1704.
57. Mura S, Nicolas J, Couvreur P. Stimuli-responsive nanocarriers for drug delivery. *Nat Mater*. 2013; 12: 991-1003.
58. Pan Y-J, Chen Y-Y, Wang D-R, Wei C, Guo J, Lu D-R, et al. Redox/pH dual stimuli-responsive biodegradable nanohydrogels with varying responses to dithiothreitol and glutathione for controlled drug release. *Biomaterials*. 2012; 33: 6570-9.
59. Wang Y, Zhou K, Huang G, Hensley C, Huang X, Ma X, et al. A nanoparticle-based strategy for the imaging of a broad range of tumours by nonlinear amplification of microenvironment signals. *Nat Mater*. 2014; 13: 204-12.
60. Liu S, Guo Y, Huang R, Li J, Huang S, Kuang Y, et al. Gene and doxorubicin co-delivery system for targeting therapy of glioma. *Biomaterials*. 2012; 33: 4907-16.
61. Li J, Guo Y, Kuang Y, An S, Ma H, Jiang C. Choline transporter-targeting and co-delivery system for glioma therapy. *Biomaterials*. 2013; 34: 9142-8.
62. Zheng M, Liu Y, Wang Y, Zhang D, Zou Y, Ruan W, et al. ROS-responsive polymeric siRNA nanomedicine stabilized by triple interactions for the robust glioblastoma combinational RNAi therapy. *Adv Mater*. 2019: 1903277.
63. Zhou J, Patel TR, Sirianni RW, Strohhahn G, Zheng M-Q, Duong N, et al. Highly penetrative, drug-loaded nanocarriers improve treatment of glioblastoma. *Proc Natl Acad Sci USA*. 2013; 110: 11751-6.
64. Esposito CL, Nuzzo S, Kumar SA, Rienzo A, Lawrence CL, Pallini R, et al. A combined microRNA-based targeted therapeutic approach to eradicate glioblastoma stem-like cells. *J Control Release*. 2016; 238: 43-57.
65. Chen J, Li Y, Yu T-S, McKay RM, Burns DK, Kernie SG, et al. A restricted cell population propagates glioblastoma growth after chemotherapy. *Nature*. 2012; 488: 522-6.
66. Lopez-Bertoni H, Kozielski KL, Rui Y, Lal B, Vaughan H, Wilson DR, et al. Bioreducible polymeric nanoparticles containing multiplexed cancer stem cell regulating miRNAs inhibit glioblastoma growth and prolong survival. *Nano Lett*. 2018; 18: 4086-94.
67. Lu Z, Li Y, Shi Y, Li Y, Xiao Z, Zhang X. Traceable nanoparticles with spatiotemporally controlled release ability for synergistic glioblastoma multimodal treatment. *Adv Funct Mater*. 2017; 27: 1703967.
68. Fang RH, Kroll AV, Gao W, Zhang L. Cell membrane coating nanotechnology. *Adv Mater*. 2018; 30: 1706759.
69. Hu Q, Sun W, Qian C, Wang C, Bomba HN, Gu Z. Anticancer platelet-mimicking nanovehicles. *Adv Mater*. 2015; 27: 7043-50.
70. Pang H-H, Chen P-Y, Wei K-C, Huang C-W, Shiu Y-L, Huang C-Y, et al. Convection-enhanced delivery of a virus-like nanotherapeutic agent with dual-modal imaging for besiegement and eradication of brain tumors. *Theranostics*. 2019; 9: 1752-63.
71. Alessandri G, Coccè V, Pastorino F, Paroni R, Cas MD, Restelli F, et al. Microfragmented human fat tissue is a natural scaffold for drug delivery: potential application in cancer chemotherapy. *J Control Release*. 2019; 302: 2-18.
72. Zhu Q, Ling X, Yang Y, Zhang J, Li Q, Niu X, et al. Embryonic stem cells-derived exosomes endowed with targeting properties as chemotherapeutics delivery vehicles for glioblastoma therapy. *Adv Sci*. 2019; 6: 1801899.
73. Xue J, Zhao Z, Zhang L, Xue L, Shen S, Wen Y, et al. Neutrophil-mediated anticancer drug delivery for suppression of postoperative malignant glioma recurrence. *Nat Nanotechnol*. 2017; 12: 692-700.
74. Zou Y, Liu Y, Yang Z, Zhang D, Lu Y, Zheng M, et al. Effective and targeted human orthotopic glioblastoma xenograft therapy *via* a multifunctional biomimetic nanomedicine. *Adv Mater*. 2018; 30: 1803717.
75. Stafford JH, Hirai T, Deng L, Chernikova SB, Urata K, West B, et al. Colony stimulating factor 1 receptor inhibition delays recurrence of glioblastoma after radiation by altering myeloid cell recruitment and polarization. *Neuro Oncol*. 2016; 18: 797-806.
76. Amsbaugh MJ, Yusuf MB, Gaskins J, Burton EC, Y.Woo S. Patterns of care and predictors of adjuvant therapies in elderly patients with glioblastoma: an analysis of the national cancer data base. *Cancer*. 2017; 123: 3277-84.
77. Perry JR, Laperriere N, O'Callaghan CJ, Brandes AA, Menten J, Phillips C, et al. Short-course radiation plus temozolomide in elderly patients with glioblastoma. *N Engl J Med*. 2017; 376: 1027-37.
78. Stupp R, Hegi ME, Mason WP, Bent MJvd, Taphoorn MJB, Janzer RC, et al. Effects of radiotherapy with concomitant and adjuvant temozolomide *versus* radiotherapy alone on survival in glioblastoma in a randomised phase III study: 5-year analysis of the EORTC-NCIC trial. *Lancet Oncol*. 2009; 10: 459-66.
79. Chinot OL, Wick W, Mason W, Henriksson R, Saran F, Nishikawa R, et al. Bevacizumab plus radiotherapy-temozolomide for newly diagnosed glioblastoma. *N Engl J Med*. 2014; 370: 709-22.
80. Shi J, Fan D, Dong C, Liu H, Jia B, Zhao H, et al. Anti-tumor effect of integrin targeted ¹⁷⁷Lu-3PRGD₂ and combined therapy with endostar. *Theranostics*. 2014; 4: 256-66.
81. Tamborini M, Locatelli E, Rasile M, Monaco I, Rodighiero S, Corradini I, et al. A combined approach employing chlorotoxin-nanovectors and low dose radiation to reach infiltrating tumor niches in glioblastoma. *ACS Nano*. 2016; 10: 2509-20.
82. Hottinger AF, Aissa AB, Espeli V, Squiban D, Dunkel N, Vargas MI, et al. Phase I study of sorafenib combined with radiation therapy and temozolomide as first-line treatment of high-grade glioma. *Br J Cancer*. 2014; 110: 2655-61.
83. Chatterjee DK, Fong LS, Zhang Y. Nanoparticles in photodynamic therapy: an emerging paradigm. *Adv Drug Deliv Rev*. 2008; 60: 1627-37.
84. Agostinis P, Berg K, Cengel KA, Foster TH, Girotti AW, Gollnick SO, et al. Photodynamic therapy of cancer: an update. *CA Cancer J Clin*. 2011; 61: 250-81.
85. Benachour H, Aymeric Sève3, Bastogne T, Frochot C, Vanderesse R, Jasniowski J, et al. Multifunctional peptide-conjugated hybrid silica

- nanoparticles for photodynamic therapy and MRI. *Theranostics*. 2012; 2: 889-904.
86. Ghosh S, Carter KA, Lovell JF. Liposomal formulations of photosensitizers. *Biomaterials*. 2019; 218: 119341.
 87. Bechet D, Couleaud P, Frochet C, Viriot M-L, Guillemin F, Barberi-Heyob M. Nanoparticles as vehicles for delivery of photodynamic therapy agents. *Trends Biotechnol*. 2008; 26: 612-21.
 88. Huang X, Zhang W, Guan G, Song G, Zou R, Hu J. Design and functionalization of the NIR-responsive photothermal semiconductor nanomaterials for cancer theranostics. *Acc Chem Res*. 2017; 50: 2529-38.
 89. Lucky SS, Soo KC, Zhang Y. Nanoparticles in photodynamic therapy. *Chem Rev*. 2015; 115: 1990-2042.
 90. Luo D, Carter KA, Miranda D, Lovell JF. Chemophototherapy: an emerging treatment option for solid tumors. *Adv Sci*. 2016; 4: 1600106.
 91. Zou L, Wang H, He B, Zeng L, Tan T, Cao H, et al. Current approaches of photothermal therapy in treating cancer metastasis with nanotherapeutics. *Theranostics*. 2016; 6: 762-72.
 92. Zhang H, Wang T, Liu H, Ren F, Qiu W, Sun Q, et al. Second near-infrared photodynamic therapy and chemotherapy of orthotopic malignant glioblastoma with ultra-small Cu₂-xSe nanoparticles. *Nanoscale*. 2019; 11: 7600-8.
 93. Liu Y, Li L, Guo Q, Wang L, Liu D, Wei Z, et al. Novel Cs-based upconversion nanoparticles as dual-modal CT and UCL imaging agents for chemo-photothermal synergistic therapy. *Theranostics*. 2016; 6: 1491-505.
 94. Yu G, Yu S, Saha ML, Zhou J, Cook TR, Yung BC, et al. A discrete organoplatinum(II) metallacage as a multimodality theranostic platform for cancer photochemotherapy. *Nat Commun*. 2018; 9: 4335.
 95. Chen Q, Wang X, Wang C, Feng L, Li Y, Liu Z. Drug-induced self-assembly of modified albumins as nanotheranostics for tumor-targeted combination therapy. *ACS Nano*. 2015; 9: 5223-33.
 96. Deng H, Zhong Y, Du M, Liu Q, Fan Z, Dai F, et al. Theranostic self-assembly structure of gold nanoparticles for NIR photothermal therapy and X-ray computed tomography imaging. *Theranostics*. 2014; 4: 904-18.
 97. Deng H, Dai F, Ma G, Zhang X. Theranostic gold nanomicelles made from biocompatible comb-like polymers for thermochemotherapy and multifunctional imaging with rapid clearance. *Adv Mater*. 2015; 27: 3645-53.
 98. Song J, Yang X, Jacobson O, Lin L, Huang P, Niu G, et al. Sequential drug release and enhanced photothermal and photoacoustic effect of hybrid reduced graphene oxide-loaded ultrasmall gold nanorod vesicles for cancer therapy. *ACS Nano*. 2015; 9: 9199-209.
 99. Quintanilla M, García I, Lázaro Id, García-Alvarez R, Henriksen-Lacey M, Vranic S, et al. Thermal monitoring during photothermia: hybrid probes for simultaneous plasmonic heating and near-infrared optical nanothermometry. *Theranostics*. 2019; 9: 7298-312.
 100. Hao Y, Zhang B, Zheng C, Ji R, Ren X, Guo F, et al. The tumor-targeting core-shell structured DTX-loaded PLGA@Au nanoparticles for chemo-photothermal therapy and X-ray imaging. *J Control Release*. 2015; 220: 545-55.
 101. Chen Z, Zhao P, Luo Z, Zheng M, Tian H, Gong P, et al. Cancer cell membrane-biomimetic nanoparticles for homologous-targeting dual-modal imaging and photothermal therapy. *ACS Nano*. 2016; 10: 10049-57.
 102. Lai J, Deng G, Sun Z, Peng X, Li J, Gong P, et al. Scaffolds biomimicking macrophages for a glioblastoma NIR-Ib imaging guided photothermal therapeutic strategy by crossing Blood-Brain Barrier. *Biomaterials*. 2019; 211: 48-56.
 103. Sun X, Wang C, Gao M, Hu A, Liu Z. Remotely controlled red blood cell carriers for cancer targeting and near-infrared light-triggered drug release in combined photothermal-chemotherapy. *Adv Funct Mater*. 2015; 25: 2386-94.
 104. Wang C, Ye Y, Hu Q, Bellotti A, Gu Z. Tailoring biomaterials for cancer immunotherapy: emerging trends and future outlook. *Adv Mater*. 2017; 29: 1606036.
 105. Ricklin D, Hajishengallis G, Yang K, Lambris JD. Complement: a key system for immune surveillance and homeostasis. *Nat Immunol*. 2010; 11: 785-97.
 106. Tanaka S, Louis DN, Curry WT, Batchelor TT, Dietrich J. Diagnostic and therapeutic avenues for glioblastoma: no longer a dead end? *Nat Rev Clin Oncol*. 2013; 10: 14-26.
 107. Paggio JCD. Immunotherapy: cancer immunotherapy and the value of cure. *Nat Rev Clin Oncol*. 2018; 15: 268-70.
 108. O'Rourke DM, Nasrallah MP, Desai A, Melenhorst JJ, Mansfield K, Morrissette JJD, et al. A single dose of peripherally infused EGFRvIII-directed CAR T cells mediates antigen loss and induces adaptive resistance in patients with recurrent glioblastoma. *Sci Transl Med*. 2017; 9: eaaa0984.
 109. Boussiotis VA, Charest A. Immunotherapies for malignant glioma. *Oncogene*. 2018; 37: 1121-41.
 110. Jackson CM, Choi J, Lim M. Mechanisms of immunotherapy resistance: lessons from glioblastoma. *Nat Immunol*. 2019; 20: 1100-9.
 111. Oh T, Sayegh ET, Fakurnejad S, Oyon D, Lamano JB, DiDomenico JD, et al. Vaccine therapies in malignant glioma. *Curr Neurol Neurosci Rep*. 2015; 15: 508.
 112. Shen Q, Yang J, Liu R, Liu L, Zhang J, Shen S, et al. Hybrid 'clusterbombs' as multifunctional nanoplatforms potentiate brain tumor immunotherapy. *Mater Horiz*. 2019; 6: 810-6.
 113. Sampson JH, Heimberger AB, Archer GE, Aldape KD, Friedman AH, Friedman HS, et al. Immunologic escape after prolonged progression-free survival with epidermal growth factor receptor variant III peptide vaccination in patients with newly diagnosed glioblastoma. *J Clin Oncol*. 2010; 28: 4722-29.
 114. Gan HK, Burgess AW, Clayton AHA, Scott AM. Targeting of a conformationally exposed, tumor-specific epitope of EGFR as a strategy for cancer therapy. *Cancer Res*. 2012; 72: 2924-30.
 115. Schuster J, Lai RK, Recht LD, Reardon DA, Paleologos NA, Groves MD, et al. A phase II, multicenter trial of rindopepimut (CDX-110) in newly diagnosed glioblastoma: the ACT III study. *Neuro Oncol*. 2015; 17: 854-61.
 116. Elsamadicy AA, Chongsathidkiet P, Desai R, Woroniecka K, Farber SH, Fecci PE, et al. Prospect of rindopepimut in the treatment of glioblastoma. *Expert Opin Biol Ther*. 2017; 17: 507-13.
 117. Weller M, Butowski N, Tran DD, Recht LD, Lim M, Hirte H, et al. Rindopepimut with temozolomide for patients with newly diagnosed, EGFRvIII-expressing glioblastoma (ACT IV): a randomised, double-blind, international phase 3 trial. *Lancet Oncol*. 2017; 18: 1373-85.
 118. Galstyan A, Markman JL, Shatalova ES, Chiechi A, Korman AJ, Patil R, et al. Blood-brain barrier permeable nano-immunoconjugates induce local immune responses for glioma therapy. *Nat Commun*. 2019; 10: 3850.
 119. Kuang J, Song W, Yin J, Zeng X, Han S, Zhao Y-P, et al. iRGD modified chemo-immunotherapeutic nanoparticles for enhanced immunotherapy against glioblastoma. *Adv Funct Mater*. 2018; 28: 1800025.
 120. Preusser M, Lim M, Hafner DA, Reardon DA, Sampson JH. Prospects of immune checkpoint modulators in the treatment of glioblastoma. *Nat Rev Neurol*. 2015; 11: 504-14.
 121. Allen E, Jabouille A, Rivera LB, Lodewijckx I, Missiaen R, Steri V, et al. Combined antiangiogenic and anti-PD-L1 therapy stimulates tumor immunity through HEV formation. *Sci Transl Med*. 2017; 9: eaak9679.
 122. Beug ST, Beauregard CE, Healy C, Sanda T, St-Jean K, Chabot J, et al. Smac mimetics synergize with immune checkpoint inhibitors to promote tumour immunity against glioblastoma. *Nat Commun*. 2017; 8: 14278.
 123. Zhao P, Wang Y, Kang X, Wu A, Yin W, Tang Y, et al. Dual-targeting biomimetic delivery for anti-glioma activity *via* remodeling the tumor microenvironment and directing macrophage mediated immunotherapy. *Chem Sci*. 2018; 9: 2674-89.
 124. Li T-F, Li K, Wang C, Liu X, Wen Y, Xu Y-H, et al. Harnessing the cross-talk between tumor cells and tumor-associated macrophages with a nano-drug for modulation of glioblastoma immune microenvironment. *J Control Release*. 2017; 268: 128-46.
 125. Kadiyala P, Li D, Nuñez FM, Althuler D, Doherty R, Kuai R, et al. High-density lipoprotein-mimicking nanodiscs for chemo-immunotherapy against glioblastoma multiforme. *ACS Nano*. 2019; 13: 1365-84.
 126. Candolfi M, Yagiz K, Wibowo M, Ahlzadeh GE, Puntel M, Ghiasi H, et al. Temozolomide does not impair gene therapy-mediated antitumor immunity in syngeneic brain tumor models. *Clin Cancer Res*. 2014; 20: 1555-65.
 127. Mathios D, Kim JE, Mangraviti A, Phallen J, Park C-K, Jackson CM, et al. Anti-PD-1 antitumor immunity is enhanced by local and abrogated by systemic chemotherapy in GBM. *Sci Transl Med*. 2016; 8: 370ra180.
 128. Qiao C, Yang J, Shen Q, Liu R, Li Y, Shi Y, et al. Traceable nanoparticles with dual targeting and ROS response for RNAi-based immunochemotherapy of intracranial glioblastoma treatment. *Adv Mater*. 2018; 30: 1705054.
 129. Fadul CE, Fisher JL, Gui J, Hampton TH, Côté AL, Ernstoff MS. Immune modulation effects of concomitant temozolomide and radiation therapy on peripheral blood mononuclear cells in patients with glioblastoma multiforme. *Neuro Oncol*. 2011; 13: 393-400.
 130. Chang D, Lim M, Goos JACM, Qiao R, Ng YY, Mansfield FM, et al. Biologically targeted magnetic hyperthermia: potential and limitations. *Front Pharmacol*. 2018; 9: 831.
 131. Thiesen B, Jordan A. Clinical applications of magnetic nanoparticles for hyperthermia. *Int J Hyperthermia*. 2008; 24: 467-74.
 132. Beik J, Abed Z, Ghoreishi FS, Hosseini-Nami S, Mehrzadi S, Shakeri-Zadeh A, et al. Nanotechnology in hyperthermia cancer therapy: From fundamental principles to advanced application. *J Control Release*. 2016; 235: 205-21.
 133. Fèvre RL, Durand-Dubief M, Chebbi I, Mandawala C, Lagroix F, Valet J-P, et al. Enhanced antitumor efficacy of biocompatible magnetosomes for the magnetic hyperthermia treatment of glioblastoma. *Theranostics*. 2017; 7: 4618-31.
 134. Wankhede M, Bouras A, Kaluzova M, Hadjipanayis CG. Magnetic nanoparticles: an emerging technology for malignant brain tumor imaging and therapy. *Expert Rev Clin Pharmacol*. 2012; 5: 173-86.
 135. Laurent S, Dutz S, Häfeli UO, Mahmoudi M. Magnetic fluid hyperthermia: focus on superparamagnetic iron oxide nanoparticles. *Adv Colloid Interface Sci*. 2011; 166: 8-23.
 136. Revia RA, Zhang M. Magnetite nanoparticles for cancer diagnosis, treatment, and treatment monitoring: recent advances. *Mater Today*. 2016; 19: 157-68.
 137. Landeghem FKv, Maier-Hauff K, Jordan A, Hoffmann K-T, Gneveckow U, Scholz R, et al. Post-mortem studies in glioblastoma patients treated with thermotherapy using magnetic nanoparticles. *Biomaterials*. 2009; 30: 52-7.
 138. Kumar CSSR, Mohammad F. Magnetic nanomaterials for hyperthermia-based therapy and controlled drug delivery. *Adv Drug Deliv Rev*. 2011; 63: 789-808.
 139. Alphand E, Idhail A, Adam C, Delattre J-Y, Schmitt C, Guyot F, et al. Development of non-pyrogenic magnetosome minerals coated with poly-L-lysine leading to full disappearance of intracranial U87-Luc glioblastoma in 100% of treated mice using magnetic hyperthermia. *Biomaterials*. 2017; 141: 210-22.

140. Chen L, Wu Y, Wu H, Li J, Xie J, Zang F, et al. Magnetic targeting combined with active targeting of dual-ligand iron oxide nanoparticles to promote the penetration depth in tumors for effective magnetic resonance imaging and hyperthermia. *Acta Biomater.* 2019; 96: 491-504.
141. Vverma J, Lal S, Noorden CJV. Nanoparticles for hyperthermic therapy: synthesis strategies and applications in glioblastoma. *Int J Nanomedicine.* 2014; 9: 2863-77.
142. Lu YJ, Chuang EY, Cheng YH, et al. Thermosensitive magnetic liposomes for alternating magnetic field-inducible drug delivery in dual targeted brain tumor chemotherapy. *Chem Eng J.* 2019; 373: 720-33.
143. Marino A, Camponovo A, Degl'Innocenti A, Bartolucci M, Tapeinos C, Martinelli C, et al. Multifunctional temozolomide-loaded lipid superparamagnetic nanovectors: dual targeting and disintegration of glioblastoma spheroids by synergic chemotherapy and hyperthermia treatment. *Nanoscale.* 2019, 11: 21227-48.
144. Tapeinos C, Marino A, Battaglini M, Migliorin S, Brescia R, Scarpellini A, et al. Stimuli-responsive lipid-based magnetic nanovectors increase apoptosis in glioblastoma cells through synergic intracellular hyperthermia and chemotherapy. *Nanoscale.* 2019; 11: 72-88.
145. Yin PT, Shah BP, Lee K-B. Combined magnetic nanoparticle-based microRNA and hyperthermia therapy to enhance apoptosis in brain cancer cells. *Small.* 2014; 10: 4106-12.
146. Huang W-C, Lu I-L, Chiang W-H, Lin Y-W, Tsai Y-C, Chen H-H, et al. Tumortropic adipose-derived stem cells carrying smart nanotherapeutics for targeted delivery and dual-modality therapy of orthotopic glioblastoma. *J Control Release.* 2017; 254: 119-30.
147. Mitsunaga M, Ogawa M, Kosaka N, Rosenblum LT, Choyke PL, Kobayashi H. Cancer cell-selective *in vivo* near infrared photoimmunotherapy targeting specific membrane molecules. *Nat Med.* 2011; 17: 1685-91.
148. Ogata F, Nagaya T, Nakamura Y, Sato K, Okuyama S, Maruoka Y, et al. Near-infrared photoimmunotherapy: a comparison of light dosing schedules. *Oncotarget.* 2017; 8: 35069-75.
149. Nagaya T, Okuyama S, Ogata F, Maruoka Y, Knapp DW, Karagiannis SN, et al. Near infrared photoimmunotherapy targeting bladder cancer with a canine anti-epidermal growth factor receptor (EGFR) antibody. *Oncotarget.* 2018; 9: 19026-38.
150. Jing H, Weidensteiner C, Reichardt W, Gaedicke S, Zhu X, Grosu A-L, et al. Imaging and selective elimination of glioblastoma stem cells with theranostic near-infrared-labeled CD133-specific antibodies. *Theranostics.* 2016; 6: 862-74.
151. Burley TA, Mączyńska J, Shah A, Szopa W, Harrington KJ, Boulton JKR, et al. Near-infrared photoimmunotherapy targeting EGFR-Shedding new light on glioblastoma treatment. *Int J Cancer.* 2018; 142: 2363-74.
152. Huang H-C, Pigula M, Fang Y, Hasan T. Immobilization of photo-immunoconjugates on nanoparticles leads to enhanced light-activated biological effects. *Small.* 2018; 14: 1800236.
153. Chou S-T, Patil R, Galst'yan A, Gangalum PR, Cavenee WK, Furnari FB, et al. Simultaneous blockade of interacting CK2 and EGFR pathways by tumor-targeting nanobioconjugates increases therapeutic efficacy against glioblastoma multiforme. *J Control Release.* 2016; 244: 14-23.
154. Kozielski KL, Ruiz-Valls A, Tzeng SY, Guerrero-Cázares H, Rui Y, Li Y, et al. Cancer-selective nanoparticles for combinatorial siRNA delivery to primary human GBM *in vitro* and *in vivo*. *Biomaterials.* 2019; 209: 79-87.
155. Westphal M, Ylä-Herttuala S, Martin J, Warnke P, Menei P, Eckland D, et al. Adenovirus-mediated gene therapy with sitimagene ceradenovec followed by intravenous ganciclovir for patients with operable high-grade glioma (ASPECT): a randomised, open-label, phase 3 trial. *Lancet Oncol.* 2013; 14: 823-33.
156. Lozada-Delgado EL, Grafals-Ruiz N, Vivas-Mejía PE. RNA Interference for Glioblastoma therapy: Innovation ladder from the bench to clinical trials. *Life Sci.* 2017; 188: 26-36.
157. Candolfi M, Kroeger KM, Muhammad AKMG, Yagiz K, Farrokhi C, Pechnick RN, et al. Gene therapy for brain cancer: combination therapies provide enhanced efficacy and safety. *Curr Gene Ther.* 2009; 9: 409-21.
158. S-S, Harford JB, Moghe M, Slaughter T, Doherty C, Chang EH. A tumor-targeting nanomedicine carrying the p53 gene crosses the blood-brain barrier and enhances anti-PD-1 immunotherapy in mouse models of glioblastoma. *Int J Cancer.* 2019; 145: 2535-46.
159. Speranza M-C, Passaro C, Ricklefs F, Kasai K, Klein SR, Nakashima H, et al. Preclinical investigation of combined gene-mediated cytotoxic immunotherapy and immune checkpoint blockade in glioblastoma. *Neuro Oncol.* 2018; 20: 225-35.
160. Bhaskaran V, Nowicki MO, Idriss M, Jimenez MA, Lugli G, Hayes JL, et al. The functional synergism of microRNA clustering provides therapeutically relevant epigenetic interference in glioblastoma. *Nat Commun.* 2019; 10: 442.
161. A. Hajj K, A. Whitehead K. Tools for translation: non-viral materials for therapeutic mRNA delivery. *Nat Rev Mater.* 2017; 2: 17056.
162. Schneider T, Becker A, Ringe K, Reinhold A, Firsching R, Sabel BA. Brain tumor therapy by combined vaccination and antisense oligonucleotide delivery with nanoparticles. *J Neuroimmunol.* 2008; 195: 21-7.
163. Erel-Akbaba G, Carvalho LA, Tian T, Zinter M, Akbaba H, Obeid PJ, et al. Radiation-induced targeted nanoparticle-based gene delivery for brain tumor therapy. *ACS Nano.* 2019; 13: 4028-40.
164. Samson A, Scott KJ, Taggart D, West EJ, Wilson E, Nuovo GJ, et al. Intravenous delivery of oncolytic reovirus to brain tumor patients immunologically primes for subsequent checkpoint blockade. *Sci Transl Med.* 2018; 10: aam7577.
165. Zhu C, Xia Y. Biomimetics: reconstitution of low-density lipoprotein for targeted drug delivery and related theranostic applications. *Chem Soc Rev.* 2017; 46: 7668-82.
166. Field KM, Simes J, Nowak AK, Cher L, Wheeler H, Hovey EJ, et al. Randomized phase 2 study of carboplatin and bevacizumab in recurrent glioblastoma. *Neuro Oncol.* 2015; 17: 1504-13.
167. Peereboom DM, Ahluwalia MS, Ye X, Supko JG, Hilderbrand SL, Phuphanich S, et al. NABTT 0502: a phase II and pharmacokinetic study of erlotinib and sorafenib for patients with progressive or recurrent glioblastoma multiforme. *Neuro Oncol.* 2013; 15: 490-6.
168. Gan HK, Bent Mvd, Lassman AB, Reardon DA, Scott AM. Antibody-drug conjugates in glioblastoma therapy: the right drugs to the right cells. *Nat Rev Clin Oncol.* 2017; 14: 695-707.
169. Touat M, Idbaih A, Sanson M, Ligon KL. Glioblastoma targeted therapy: updated approaches from recent biological insights. *Ann Oncol.* 2017; 28: 1457-72.