

Review

Adaptive changes induced by noble-metal nanostructures *in vitro* and *in vivo*

Qianqian Huang^{1,2,3}, Jinchao Zhang⁴, Yuanyuan Zhang⁵, Peter Timashev⁶, Xiaowei Ma^{1,2}✉, Xing-Jie Liang^{1,2,3}✉

1. CAS Key Laboratory for Biomedical Effects of Nanomaterials and Nanosafety, CAS Center for Excellence in Nanoscience, National Center for Nanoscience and Technology of China, Beijing 100190, China
2. University of Chinese Academy of Sciences, Beijing 100049, China
3. Sino-Danish Center for Education and Research, Sino-Danish College of University of Chinese Academy of Sciences, Beijing, 100049, China
4. Key Laboratory of Medicinal Chemistry and Molecular Diagnosis of the Ministry of Education, College of Chemistry & Environmental Science, Hebei University, Baoding 071002, China
5. Wake Forest Institute for Regenerative Medicine, Wake Forest University School of Medicine, Winston-Salem, North Carolina, USA
6. Institute for Regenerative Medicine, Sechenov University, Moscow, Russia

✉ Corresponding authors: Xiaowei Ma: maxw@nanocr.cn; Xing-Jie Liang: liangxj@nanocr.cn

© The author(s). This is an open access article distributed under the terms of the Creative Commons Attribution License (<https://creativecommons.org/licenses/by/4.0/>). See <http://ivyspring.com/terms> for full terms and conditions.

Received: 2019.11.29; Accepted: 2020.03.01; Published: 2020.04.27

Abstract

The unique features of noble-metal nanostructures (NMNs) are leading to unprecedented expansion of research and exploration of their application in therapeutics, diagnostics and bioimaging fields. With the ever-growing applications of NMNs, both therapeutic and environmental NMNs are likely to be exposed to tissues and organs, requiring careful studies towards their biological effects *in vitro* and *in vivo*. Upon NMNs exposure, tissues and cells may undergo a series of adaptive changes both in morphology and function. At the cellular level, the accumulation of NMNs in various subcellular organelles including lysosomes, endoplasmic reticulum, Golgi apparatus, mitochondria, and nucleus may interfere with their functions, causing changes in a variety of cellular functions, such as digestion, protein synthesis and secretion, energy metabolism, mitochondrial respiration, and proliferation. In animals, retention of NMNs in metabolic-, respiratory-, immune-related, and other organs can trigger significant physiological and pathological changes to these organs and influence their functions. Exploring how NMNs interact with tissues and cells and the underlying mechanisms are of vital importance for their future applications. Here, we illustrate the characteristics of NMNs-induced adaptive changes both *in vitro* and *in vivo*. Potential strategies in the design of NMNs are also discussed to take advantage of beneficial adaptive changes and avoid unfavorable changes for the proper implementation of these nanoplatforms.

Key words: noble-metal nanostructures (NMNs), applications, exposure, adaptive changes, *in vitro* and *in vivo*, proper implementation.

Introduction

The integration of nanoscience and biotechnology has spawned an increasing field of research which possesses immense potential to positively impact public health. In this amazing arena, the technological innovation of synthesizing materials at nanoscale level has provided great opportunities to achieve potential clinical applications encompassing healthcare treatment, diagnosis and imaging [1, 2]. NMNs such as gold, silver and platinum are particularly attractive for theranostics due to their

unique and promising features. Generally, noble metals refer to the metallic chemical elements in groups VIIIb, VIII and 1b of the second and third transition series of the periodic table, including rhodium (Rh), ruthenium (Ru), palladium (Pd), silver (Ag), osmium (Os), iridium (Ir), platinum (Pt), and gold (Au), in order of atomic number [3]. The most important characteristics of noble metals are their outstanding resistance to a wide range of corrosive attack and their stability even under conditions where

base metals are oxidized rapidly. The history of noble metals dates back to as early as the Egyptian First Dynasty [4]. They belong to a class of precious elements that have been found with a wide range of applications such as electronic devices, aerospace engineering, and most importantly, the health care system [3]. Most NMNs are eclectic nontoxic and biocompatible agents. Table 1 shows a brief comparison of different types of NMNs and their main applications in academic studies. Among all NMNs, Au and Ag nanoparticles are the most extensively studied as they could be easily synthesized and are relatively safe. The intriguing physicochemical properties of NMNs such as plasmonic, optical, photothermal, and catalytic abilities allow them to act as efficient biosensing and imaging labels [5]. By means of strong metal-thiolate bonds, they can be conveniently functionalized with biomolecules and exert multiple biological functions [6, 7]. Furthermore, recent investigations have demonstrated broad application prospects of NMNs as self-therapeutic agents [8, 9].

The specific physicochemical properties of NMNs will result in increased activities within biological systems. With the increasing application of NMNs, both therapeutic and environmental NMNs are likely to be exposed to tissues and cells. Inert NMNs with excellent stability enable them to persist in complex biological systems for a long time. In general, NMNs that do not cause serious acute changes in cells or tissues are often considered to be biocompatible. How cells response to NMNs is of vital importance for their biological changes and survival.

Tissue, organs and cells may undergo a series of adaptive changes upon NMNs exposure. Basic understanding of how NMNs interact with cells, organs, tissues and the underlying mechanisms are crucial for their various applications. Through in-depth study of these adaptive changes, we can skillfully take advantage of favorable adaptive changes and avoid unfavorable changes in order to obtain fantastic nanosystems [10]. In this review, we give a summary of the characteristics of NMNs-induced adaptive changes and provide detailed understanding of these changes both *in vitro* and *in vivo*. Moreover, we discuss the potential usage of NMNs-induced beneficial adaptive changes for proper implementation of exquisite nanoplatforms.

Characteristics of NMNs-induced adaptive changes *in vitro* and *in vivo*

NMNs can enter the body through inhalation, skin exposure, gastrointestinal absorption and biomedical applications. NMNs entering vascular system through various pathways will tend to be further distributed to peripheral tissues, organs, and cells, or be absorbed in the spleen and bone marrow [3]. After reaching different tissues and organs, NMNs will interact with histocyte immediately and continuously. Cells will gradually evolve adaptive changes to achieve tolerance to NMNs and new cellular homeostasis, which is considered as basic condition for their survival in response to external stimulus and stressors.

Table 1. Different types of NMNs and their main applications in academic studies.

| Type of NMNs | Shape | Properties | Applications | References |
|------------------|--------------------------|---|---|------------|
| Au nanoparticles | Nanostars | Large absorption of near-infrared light | PTT/ PDT agents for cancer treatment | [11] |
| | Nanoshell | Hyperthermia | Disrupt overactive sebaceous glands | [12] |
| | Spherical | Antibacterial | Inhibition of vancomycin-resistant enterococci, <i>E. coli</i> and <i>S. aureus</i> | [13] |
| | Spherical | Antiviral | Inhibition of HIV-1 | [14] |
| | Spherical | SERS | <i>In vivo</i> multiplex molecular imaging | [15] |
| Ag nanoparticles | Spherical | Osteoinductive | Osteoinductive agent for implant dentistry | [16] |
| | Nanocrystalline | Anti-angiogenic | Combat multidrug-resistant cancer | [17] |
| | Spherical | Antibacterial | Prevention of tuberculosis | [18] |
| | Spherical | Antifungal | Inhibitory action against <i>Candida albicans</i> , <i>C. tropicalis</i> , <i>trichophyton mentaprophytes</i> , <i>C. glabrata</i> and <i>C. krusei</i> | [19] |
| Pt nanoparticles | Crystalline | Antiprotozoal | Inhibitory action against <i>cryptosporidium parvum</i> in water purification | [20] |
| | Spherical | Sturdy and durable | Dental resin filler composite | [21] |
| | Spherical | Strong affinity with dopamine | Dopamine sensors | [22] |
| | Spherical | Antioxidant | ROS scavenge | [23] |
| | Spherical, cubic, flower | Antibacterial | Inhibitory activity against <i>P. aeruginosa</i> | [24] |
| Pd nanoparticles | Spherical | Electrocatalytic | Detection of hydrogen peroxide in cells | [25] |
| | Spherical | Catalytic properties | Cancer treatment | [26] |
| Ru nanoparticles | Sheet | LSPR/photothermal ablation | As PTT agents for cancer treatment | [27] |
| | Spherical | Antibacterial | Inhibition of Gram-positive and Gram-negative bacteria | [28] |
| Rh nanoparticles | Spherical | Promote osteogenic differentiation | Modulate the behavior of stem cells | [29] |
| | Shell, frame, plate | Photothermal | Cancer phototherapy | [30] |
| Ir nanoparticles | Spherical | Photosensitive | Enhanced photodynamic performance | [31] |
| | Spherical | Hydrophobicity | Drug delivery | [32] |
| | Spherical | Charge transfer | Bioimaging | [33] |

The chemotherapy drug tolerance, which is described as the corresponding mechanisms evolved by cells to lose sensitivity to chemotherapeutics, is an example of cells gaining tolerance to improve survival [34]. Similarly, with extensive exposure of cells to NMNs, a series of adaptive changes both *in vitro* and *in vivo* will be seen on cells for better survival. When subjected to repeated NMNs stimulation, cells may even lose sensitivity to NMNs, leading to reduced cellular accumulation and change of tolerance-related genes of NMNs. For example, Priscila *et al.* found a single administration of Au nanoparticles caused long-term genetic changes in cells, and these genes were involved in oxidative damage, cell cycle regulation, and inflammation [35].

Current research of biological effects of NMNs mainly focused on the short-term interaction between nanomaterials and biological systems and subsequent consequences. At the cellular level, researchers focused on the influences of NMNs on organelles, cell morphology, cell cycle, cell proliferation, and genes, etc. Most changes induced by NMNs are related to reactive oxygen species (ROS), known as oxidative stress. ROS possess multi-functions in cellular biology such as cellular proliferation, differentiation and death, with ROS production a key factor in NMNs-induced changes [36]. NMNs-related stress is due to their smallness, notably large surface area per unit mass and high surface reactivity. When internalized into cells, NMNs will interact with intracellular nicotinamide adenine dinucleotide phosphate oxidase. The unique physicochemical reactivities of NMNs are correlated with the ability to trigger increased expression of pro-inflammatory and fibrotic cytokines and activation of inflammatory cells like macrophages and neutrophils. Cells show potent antioxidant defenses capable of overcoming oxidative stress and restoring redox balance when exposed to low concentrations of NMNs. By contrast, exposure to high concentrations of NMNs overwhelms cellular antioxidant systems and results in a sequence of pathological events including inflammation, fibrosis, genotoxicity, and carcinogenesis [37]. The sizes of nanomaterials are comparable to those of viruses or bacteria for which our immune system has elaborated sophisticated responses. Specific rather than random oxidative reactions are elicited by cells to raise a protective response different from cell death mechanism when exposed to NMNs, which is critical to NMNs intracellular fate and effects [38]. Cellular changes such as organelle functions, cell growth and apoptosis are often reported. Lysosomes and endosomes are demonstrated to have the most significant changes in cells since they are in direct contact with NMNs [39]. In the absence of

membrane-penetrating peptides, internalized NMNs mainly accumulate in lysosomes without entering the cytosol. Nevertheless, with the pursuit of precision medicine, subcellular targeting has become one of the biggest breakthroughs in achieving optimum therapeutic effect with minimal dosing. With the assistance of organelle-targeted molecules, internalized NMNs will eventually find their way to specific organelles and subcellular structures [40]. Lysosomes, endoplasmic reticulum (ER), Golgi apparatus, mitochondria, and nucleus are the primary targets for various therapeutics as their functions are intimately bounded up with cell proliferation, cell growth, differentiation, and cell death [41]. The accumulation of NMNs in these core organelles will interfere with their functions and cause changes in a variety of cellular functions including cellular digestion, protein synthesis and secretion, cellular respiration, and cellular replication. To maintain the homeostasis of cellular degradation system, cells will synthesis more lysosomes to compensate the missed degradative function after lysosome swelling. The alternation in protein processing allows cells to be adaptive to nanomaterials stimulation by activating specific signaling pathway, secreting various chemical substances, and regulating functional protein synthesis. Even when intercellular oxygen is sufficient, cells may initiate glycolysis to satisfy their energy demand. In animals, there are heightened interest in the physiological and pathological effects of NMNs on tissues and organs. Superior biocompatibility and immune-compatibility are the main points of *in vivo* evaluation of nanomaterials. Scientists are concerned with the distribution of NMNs in tissues and organs and their biocompatibility, which is related to their safe applications. A plethora of studies have shown accumulation of NMNs in liver, kidney, lung, heart, spleen, and brain [42]. Although they have various exposure pathways, NMNs are mainly stored in reticular endothelial system through first-pass metabolism. Due to their inertness and good biocompatibility, NMNs can withstand various influences of the body and remain relatively stable in tissues and organs. Internalized NMNs may store in these tissues and organs at different time points, resulting in unforeseen physiological and pathological effects on metabolism, respiration, consciousness and immunity.

With the widespread application of NMNs, both therapeutic and environmental NMNs are likely to be exposed to tissues, organs and cells for a long time. Research on long-term exposure of NMNs, although relatively rare, has been proved to be of vital importance to study adaptive changes induced by

NMNs [43]. When exposed to a certain amount of NMNs for a long time, cells may undergo a series of morphological and functional changes. Cellular response to NMNs is an attempt to try to ensure their survival. The adaptive changes caused by long-term NMNs exposure are different from short-term exposure, and long-term exposure causes relatively less cytotoxicity. *Kristen et al.* implemented a chronic *in vitro* model treated with low dosage of Ag nanoparticles. Chronic exposure at low dosages of Ag nanoparticles did not cause cytotoxic changes, but instead activated sustained stress and signaling responses. Long-term Ag nanoparticles treated cells operated normally with augmented stress response and improved cellular function compared to acutely treated cells, indicating that cells gradually adapted to long-term stimulation of NMNs [43]. Similar to chemotherapeutic drug tolerance, cells become resistance to NMNs under long-term stimulation, thus losing sensitivity and inhibiting uptake of NMNs. Continuous exposure of Au nanoparticles at low doses results in rapid intracellular accumulation and increased ER stress. However, when the concentration of Au nanoparticles exceeds the intracellular concentration threshold, further uptake of Au nanoparticles is inhibited, thereby reducing ER stress [44]. Chronic and low dosage of NMNs exposure may lead to prolonged changes in cell physiology, which are direct indicators of cellular responses to long-term stimulus. Acute Au nanoparticles exposed cells showed significant cell area decrease, but after long-term incubation, the decreased cell area recovered [35]. All these results suggest that cells turn to change under both acute and chronic incubation of NMNs, and long-term exposed cells increase their adaptability to the stimulation for better survival.

NMN-induced changes in cell morphology and functions

Cellular morphology change by NMNs exposure

Cellular and subcellular morphology are responsible for intercellular communication, cell homeostasis and functions. Recent advances in high-resolution imaging and biophysical technology have made it feasible to directly evaluate detailed changes of cellular and subcellular morphology in response to NMNs exposure [45]. When in contact with cells, nanomaterials are typically ingested by endocytosis, and are trapped inside endosomes or lysosomes without entering cytoplasm [46]. Once entering the cells, NMNs would have profound effects on cellular structures and morphologically

related functions. The alterations of cytoskeleton and organelle structure caused by NMNs exposure have been reported in multiple studies.

Cell cytoskeleton is the center part for maintaining cell morphology. *Pernodet et al.* found that the presence of Au nanoparticles affected cell morphology in a concentration-dependent manner. Upon Au nanoparticles exposure, multi parameters of actin stress fibers in human dermal fibroblasts, such as diameter, density, stretching state changed, consequently affecting cell shape, growth and viability [47]. A similar study demonstrated concentration-dependent effects of Au nanoparticles on actin and tubulin. Cellular morphology change was observed at concentrations above 50nM [48]. Additional research focused on the relationship between nanoparticles morphology change and alternation of cytoskeleton. Different sized nanoparticles induced different extent of cytoskeletal filament disruption [49]. Another study about the effects of nanoparticles modification on cellular uptake and cytotoxicity showed that naked nanoparticles caused changes in cell morphology, while high concentrations of PEGylated Au spheres did not cause such changes in cells [50]. The aggregation states of Au nanoparticles also cause various cellular responses in cell cytoskeleton. After uptake of aggregated and non-aggregated cationic Au nanoparticles, human dermal fibroblast cells exhibited varying degrees of F-actin fiber disruption with the appearance of actin dots [51]. Studies on the influence of NMN on cell morphology are mainly focused on short-term effect. Some new findings also demonstrated the impact of acute Ag nanoparticles treatment on cellular membrane and cytoskeleton reconstruction [52, 53]. Recently, small but significant morphology change was found in cells at each tested time point under long time exposure (20 weeks) of Au nanoparticles. For all nonchronic nanoparticles-exposed cells, cell area decreased significantly, which reflected the contraction of cytoskeleton, while after long-term culture, the decrease in cell area recovered [35]. Our group further checked the alternation of cellular and subcellular morphology induced by NMNs (Figure 1). Cells undergo organelles structures change, alterations in cytoskeleton and reduction of focal adhesion contact area after Au nanoparticles treatment. The effects of NMNs on cellular morphology can be observed at concentrations lower than that of on cell viability, giving an important quantitative indicator to evaluate the effects of NMNs [54].

The impact of NMNs on cytoskeleton may influence the behaviors associated with cytoskeleton reconstruction, including cell junction, division and

movement, since the components of cytoskeleton are known for their roles in these behaviors [55]. *Lin et al.* observed the intercellular tight junctions were loosed by different sized Au nanoparticles [56]. Our previous work demonstrated the focal adhesion contact area and the number of filopodia were reduced after Au nanoparticles treatment [54]. The disruption of tight junction by NMNs was also observed by *Xu et al.* They demonstrated that Ag nanoparticles could induce destruction of tight junctions and cross through the blood brain barrier (BBB) [57]. Furthermore, cell division is subject to change, as well as cell migration. *Lee et al.* mentioned the regulation of cytoskeletal structure by Au nanoparticles repressed cell division. This could be further explained by the transformation of crosstalk between actin and β -tubulins after nanoparticles incubation [58]. *Rafailovich et al.* analyzed cellular effect of Au nanoparticles on human adipose-derived stromal cells. Cells exhibited concentration-dependent increase in population doubling times, decrease in cell motility, and contraction of cell-mediated collagen following Au nanoparticles exposure [59].

It is clear that the uptake of NMNs has great impacts on cellular morphology. Cytoskeleton plays an important role on attaching and arranging organelles inside cells. The effect of NMNs on

cytoskeleton in turn affects organelles that bind to it. cellular and subcellular morphology changes occur not just due to direct aggregation of NMNs inside cells. Complex cellular network lets us take the indirect interactions among different intracellular organelles into account. Under the treatment of NMNs, cells turn to change their morphology followed by altering morphologically related cellular functions, which increase their adaptability to NMNs stimulation.

Alteration of core organelles-related cell functions by NMNs exposure

After entering cells, nanoparticles firstly stay inside acidic lysosomes, or with the assist of organelle targeting agents, nanoparticles will then accumulate in targeted organelles like mitochondria, ER, nucleus, etc. [60]. Upon NMNs exposure, organelle morphologies are altered either through direct nanoparticles accumulation or indirect subcellular interaction. Moreover, the adaptive changes of subcellular morphologies induced by NMNs will result in associated adjustment of cell behaviors and organelle-related cell functions. These alternations make cells adaptable in response to environmental changes (Figure 2).

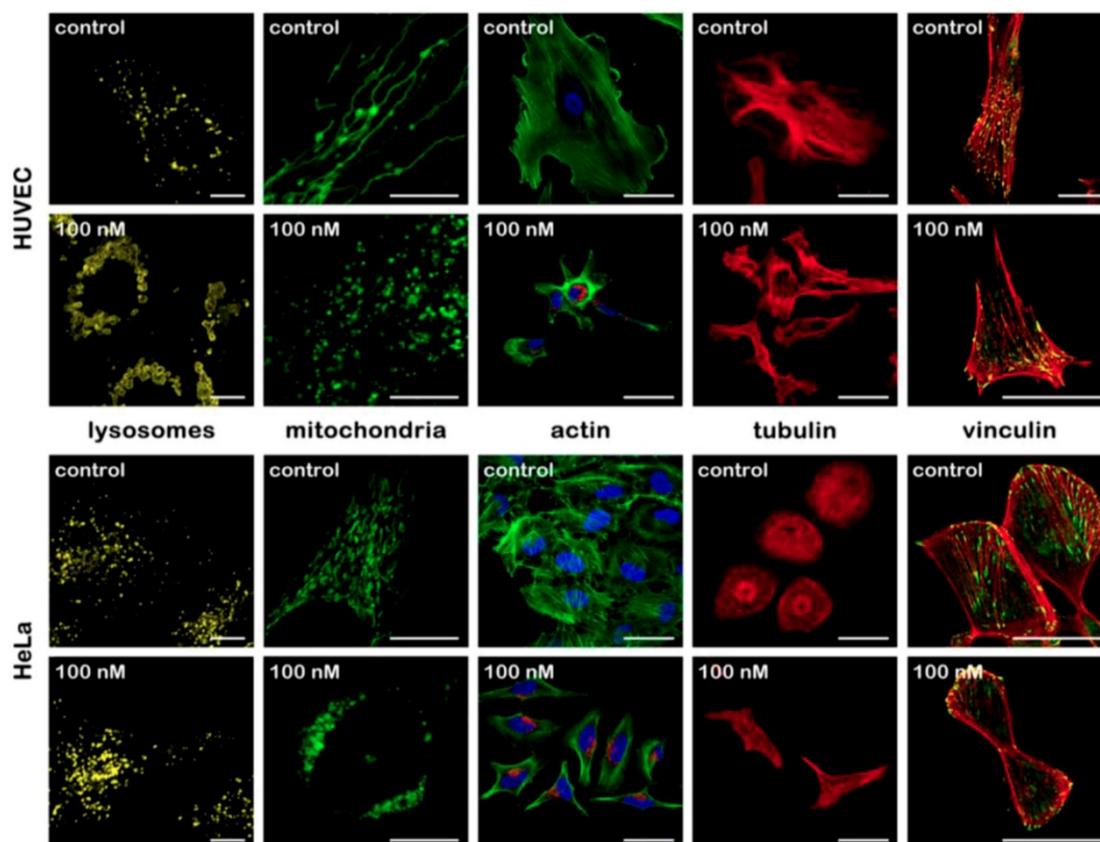


Figure 1. Morphological changes of different cellular structures without and after Au nanoparticles treatment in HUVECs and HeLa cells. Reproduced with permission from ref [54]. Copyright 2017 American Chemical Society.

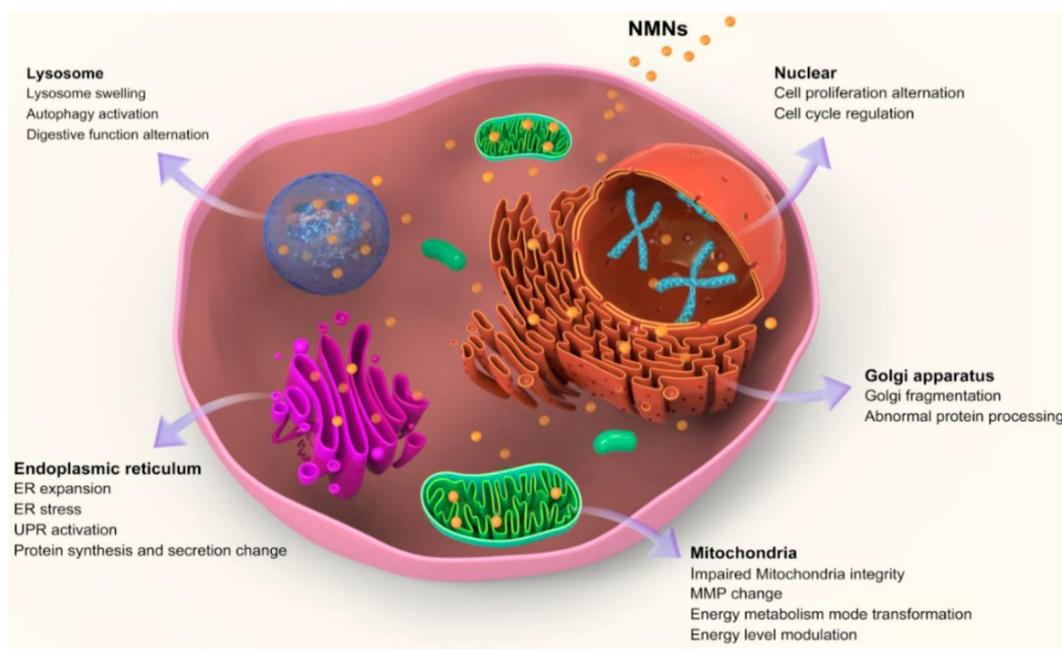


Figure 2. Schematic illustrating alterations of core sub-organelles and related cell functions by NMNs exposure.

Cellular digestion

Nanoparticles are ingested by cells mainly through endocytic pathway and will stay inside acidic lysosomes firstly. Our previous work found Au nanoparticles were ingested by cells in a size-dependent manner. Large number of Au nanoparticles accumulated in lysosomes led to lysosomes enlargement followed by lysosome alkalization, which affected autophagy [61]. Remarkable lysosome swelling due to nanoparticles accumulation was also identified by biological transmission electron microscope [62]. Lysosomes is crucial for maintaining cell homeostasis and is considered to be the center of sophisticated signal transmission and metabolism [63]. It the end point of endocytic pathway and the digestive organelle of both intracellular and exogenous cargos. Superior accumulation of NMNs in lysosomes greatly influences cellular digestion [39].

Autophagy is a lysosome-mediated degradative process to achieve the metabolic needs of cells and the renewal of certain organelles. During autophagy, self-cytoplasmic proteins or organelles are coated into vesicles and fused with lysosomes to form autolysosomes, then the engulfed contents will be degraded [64]. There has been increasing interest in autophagy since it was first discovered in 1963. Numerous studies have reported that various NMNs are capable of inducing autophagy. *Zhang et al.* proposed that autophagy is a common cellular response induced by NMNs. HeLa cells treated with Pd nanoparticles at low concentrations induced

autophagy through mTOR signaling pathway in a size-dependent manner. With increased particle size of incubated Pd nanoparticles, the lysosome degradation capability decreased [65]. As evidenced by transmission electron microscopy, Ag nanoparticles activated autophagy and induced autophagosome formation [66]. Au nanomaterials are widely used in biomedical field due to their low toxicity and good biocompatibility. Au nanoparticles promoted the expression of autophagic proteins and proved to be a novel kind of autophagy activator [62, 67]. Our group further explored the underlying mechanism. Different sized Au nanoparticles entered cells in a size dependent manner and eventually accumulated in acid lysosomes. Accumulation of Au nanoparticles in lysosomes leads to lysosomal alkalization. By monitoring the degradation of autophagy substrate p62, we provided that the accumulation of autophagosome after Au nanoparticles exposure was owing to the blockade of autophagic flux rather than autophagy induction [61]. Similarly, another work based on Ag nanoparticles engulfed by A549 cells also showed autophagosome accumulation was caused by autophagy flux blockade via lysosomal alkalization [68].

The aforementioned results give us detailed information about NMNs induced autophagy. The activation of autophagy is a cellular stress response to intracellular foreign bodies, which is essential for cellular self-protection, as well as the recycle of damaged proteins and organelles to sustain metabolism homeostasis [69]. The presence of nanomaterials inflicts lysosomes alkalization, which

causes partial loss of endolysosome function. In order to accommodate these changes, cells will synthesis more lysosomes and initiate autophagy to make up the missed degradative function. The induced autophagy thus is considered as a protective mechanism against NMNs stimulation [70]. *Li et al.* analyzed the role of Au nanoparticles in inducing autophagy both *in vitro* and *in vivo*. Ag nanoparticles treatment induced autophagy via calmodulin-dependent protein kinase kinase β and adenosine 5'-monophosphate-activated protein kinase pathway. Inhibition of autophagy significantly enhanced cytotoxicity of Ag nanoparticles, suggesting autophagy played a protective role in Ag nanoparticles-induced cell apoptosis [71]. Moreover, a key step in cell digestion is to take up the digested material into cells. The exposure of NMNs can influence micropinocytosis and thus may inhibit cell digestion. Long-time exposure of Au nanoparticles above intracellular concentration threshold inhibited micropinocytosis mechanism to cease further nanoparticles uptake, which adaptively reduced ER stress [44].

Protein synthesis and secretion

ER is the largest organelle in cells. ER and Golgi apparatus are responsible for membrane interaction and protein synthesis. They are central for regulating various signaling pathways and cell response to stress [72]. With the pursuit of precision therapy, accumulating efforts have been made by researchers to target ER and Golgi apparatus with nanotechnology [73]. *Xu et al.* observed ER expansion in astrocytes after Ag nanoparticles incubation [57]. As the major organelle for protein folding, ER complete the folding process of incompletely folded proteins with the help of molecular chaperones and folding enzymes [74]. NMNs exposure may impede protein folding capacity and thus lead to the accumulation of misfolded proteins in ER, named ER stress. It was shown that PEGylated nanogels containing Au nanoparticles increased the expression of ER stress-related proteins [75]. With the accumulation of misfolded proteins in ER, the equilibrium between ER protein load and folding capacity under normal conditions will be damaged. In order to restore ER homeostasis to better withstand nanoparticles stimulation, an adaptive signaling pathway termed as unfolded protein response (UPR) will be activated, which result in protein synthesis and secretion changes [76]. For example, Ag nanoparticles treatment led to accumulation and aggregation of misfolded proteins in cells which caused ER stress and activated UPR. Through this process, downregulation of Mcl-1 and χ IAP protein

expression as well as a processing of PARP protein were observed [77].

Golgi apparatus is the organelle for final processing and packaging of proteins. Vesicles from ER fuse with Golgi membrane and deliver the contents into Golgi cavity where newly synthesized proteins continue to be modified and packaged [78]. Due to structural and functional continuity of ER and Golgi apparatus, NMNs-induced ER stress can also cause Golgi apparatus stress by affecting normal protein transportation. Alternation of Golgi apparatus by NMNs will ultimately affect protein transport and secretion. Recently, we first discovered Au nanoparticles increased cytoplasmic calcium and induced Golgi fragmentation, thereby impaired normal Golgi functions, while cell viability was not compromised. The fragmentation of Golgi apparatus induced by Au nanoparticles hampered normal Golgi functions, which led to abnormal protein processing and cellular adhesion change [79].

Energy level and cell fate

Mitochondrial respiration modulation by NMNs

Mitochondria are best known to be the powerhouse of cells, which are in charge of many important cellular functions. On one hand, mitochondria decompose carbohydrates to produce ATP through electron transport and oxidative phosphorylation, providing most of the energy needed for metabolism and movement of organisms. On the other hand, mitochondria are involved in various metabolic pathways and apoptosis [80]. Hence, mitochondria determine cellular energy level and cellular fate and are attractive targets to treat mitochondria-associated diseases [73]. Multiple studies have revealed NMNs-induced mitochondria morphology change. For example, *Pan et al.* reported Au nanoparticles with diameter of 1.4 nm compromised mitochondrial integrity, potential, and its substrate reduction [81]. Localization of ammonium-modified Au nanoparticles in cellular mitochondria caused mitochondria swelling and enlargement [82]. *Xu et al.* observed severe mitochondrial shrinkage after Ag nanoparticles treatment [57]. Mitochondria are sensitive to external stimuli and are key regulators of cell survival and death [83]. Previous research demonstrated that changes in mitochondria occurred at the lowest Au nanoparticles concentration compared with that of other organelles [54]. When NMNs enter the cell, whether through direct mitochondrial aggregation or indirect interaction with the membrane, nanoparticles would trigger mitochondrial oxidative stress, affect mitochondrial respiration, and change cellular energy levels.

Mitochondria are more complex compared with other organelles because of the double-layer membranes. Most of ATP used by cells is synthesized by mitochondrial respiration via oxidative phosphorylation. This process causes a potential difference between mitochondrial inner and outer membranes. Although the double-layer membranes block many substances, harmful or therapeutic, from entering into mitochondria, mitochondrial membrane potential (MMP) makes it possible to transport bioactive molecules into mitochondria with assist of nanomaterials [41]. A growing number of reports have revealed the modulation of mitochondrial respiration by NMNs, among which the most widely reported are changes in MMP. *Chen et al.* compared the effect of Au nanorods on cancer and normal cells. Selective accumulation and long-term retention of Au nanorods in cancer cell mitochondria led to destruction of MMP and increase of ROS. This may be used for mitochondria-targeted cancer therapy [84]. They also analyzed the bioeffect of Ag nanoparticles. Electron transfer impairment in mitochondria was found after Ag nanoparticles incubation in their study, as well as impairment of ATP synthesis, which triggered oxidative stress [85]. In addition to *in vitro* studies, *in vivo* analysis also revealed mitochondrial alteration after NMNs exposure. Prolonged exposure to a low dose of Ag nanoparticles was sufficient to cause alteration in hepatic mitochondrial function [86]. Since the significant change of MMP after NMNs exposure, a chapter in *nanotoxicity* proposed an overview of nanomaterial-induced loss of MMP and presented a detailed method to accurately quantify the loss of MMP upon nanomaterials treatment [87].

Under normal conditions, energy homeostasis is maintained within cells through concerted signaling and metabolic pathways. Mitochondria are the main organelle responsible for energy homeostasis in mammalian cells. Alternation of mitochondrial integrity or mitochondrial dysfunction may lead to severe impairments of energy metabolism, causing associated diseases. Therefore, the influences of NMNs on mitochondria require cells and organisms to maintain internal energy homeostasis to accommodate NMNs stimuli. Transformation between different energy metabolism modes is a cellular adaptive change to satisfy the energy demand for cell survival. For example, energy in normal cells is primarily produced by the metabolism of oxidative phosphorylation of reactive mitochondrial respiration, whereas in tumor cells, tumor cells initiate glycolysis-based metabolism for energy production simultaneously even in the presence of high oxygen concentration [88]. When cells are exposed to NMNs, cells tend to shift energy metabolism mode to satisfy

energy demand for their survival. *Chen et al.* studied Ag nanoparticles-mediated effects on energy metabolism in cells under sublethal exposure. Ag nanoparticles treatment at nontoxic concentrations resulted in compromised MMP and electron transfer along respiratory chain. Decline of ATP synthesis and attenuation of respiratory chain function led to initiation of glycolysis to compensate ATP decrease and satisfy basal energy demand for cell survival [89]. As illustrated in Figure 3, this schematic diagram deciphers the mechanisms by which Ag nanoparticles incur an adaptive change of energy metabolism mode to glycolysis under sublethal exposure in both tumor and normal cells. Alterations of several energy metabolism related genes are involved. All these changes force cells to reprogram energy metabolism by switching from oxidative phosphorylation-based aerobic metabolism to anaerobic glycolysis. This adaption process makes up the energy decrease in response to NMNs exposure to satisfy energy demand for cell survival.

Nucleus change: cell proliferation and growth

Nucleus contains the vast majority of genetic material of cells and controls gene expression. It is surrounded by a bilayered nucleus membrane that separates genome from cytoplasm to ensure accurate DNA replication and gene expression. There are thousands of protein machines, namely nucleus pore complexes (NPCs), on nucleus to maintain material exchange and information transmission between nucleus and cytoplasm [90]. Although the small-sized and highly selective NPCs are complicated biological barriers for exogenously applied macromolecules, some ultrasmall metal nanoparticles might still diffuse into nucleus [91]. In addition, with the aid of nucleus-targeting ligands, various surface modifications of NMNs allow them to be accurately delivered to nucleus [40, 92]. As the most important organelle of eukaryotic cells, nucleus performs central regulation function of cell proliferation, metabolism, and cell cycle. There is heightened interest in the influence of NMNs on nucleus since it is critical for cell survival. The direct and indirect interactions between NMNs and nucleus have significance effects on cellular nucleus, which may result in cell growth, proliferation, mitosis, and apoptosis alternation.

Among all kinds of NMNs, Au nanoparticles are most widely used for nucleus targeting due to their controllable ultrasmall size and tunable surface modification. In the research of *Mostafa et al.*, Au nanoparticles localized at nucleus broke DNA double-strand and induced cell cycle arrest in cancer cells. Nucleus change upon acute Au nanoparticles exposure led to the failure of complete cell mitosis

[93]. The size, shape, and even surface modification of Au nanoparticles may change multiple nucleus parameters significantly. Small Au nanospheres and nanoflowers altered nucleus morphology, organization and function of MCF-7 cells, but big Au nanospheres did not. The changes of nucleus organization and function greatly influenced cell survival [94]. Another research focused on the interactions between cells and Au nanoparticles modified with different PEG layer properties. Au nanoparticles with all PEG grafting densities caused significant nucleus damage of cancer cells. The alternation of nucleus led to cell cycle arrest without apoptosis [95]. The long-term effect of Au nanoparticles on nucleus also demonstrated modulation of cell cycle, and acute NMNs exposure was more harmful to cells compared with chronic treatment [35]. In mouse embryonic fibroblasts, after treatment with low doses of cobalt nanoparticles for three months, in addition to changes in cell morphology, characteristics similar to tumor cells such as increased secretion of metalloproteinases, enhanced anchorage-independent cell proliferation were observed [96]. NMNs and NMNs-induced oxidative stress will also influence nucleus components such as nuclear lamins and histones. *Zhao et al.* reported that Ag nanoparticles induced phosphorylation of histone H2AX followed by DNA double-strand breaks, which was synergistically enhanced by ultraviolet A exposure [97]. *Jennifer et al.* indicated that Ag nanoparticles treatment reduced nucleus lamin B1, which resulted in the up-regulation

of telomeric repeat binding factor protein expression and maintenance of telomere length [98]. Stress-induced premature senescence (SIPS) or stress-associated premature senescence (SAPS) is a type of senescence appears after exposure of cells to stress by chemical, physical or biological agents, such as oxidants, ultraviolet radiation, and viruses, among which nanomaterials exposure is a typical example [99]. After NMNs stimulation, chronic or acute, stressed cells developed a “senescence-like” phenotype. *Jennifer et al.* demonstrated for the first time that nanoparticles may stimulate SIPS. P53 and p21 levels were elevated and SIPS was observed in cell treated with Ag nanoparticles, suggesting that nanoparticles promote a telomere-focused cell adaptive response [98].

Under the stimulation of NMNs, cells not only undergo changes in nucleus morphology, but also in nucleus function and related cell behaviors. Cellular response to NMNs is an attempt by cells to ensure survival. The observed effects of NMNs on nucleus indicate that cells will become more like cancer cells to accommodate NMNs stimulation, such as enhanced cell proliferation and cell viability. These characters enable cells to better cope with damage caused by NMNs. Moreover, chronic treatment of NMNs may lead to prolonged changes in cell physiology. In other words, cells can adapt to long-term NMNs exposure, which can be considered as part of adaptive responses to long-term effects of external stimulus [35]. However, when NMN-induced nucleus change is beyond their self-repair, cell death will appear [100].

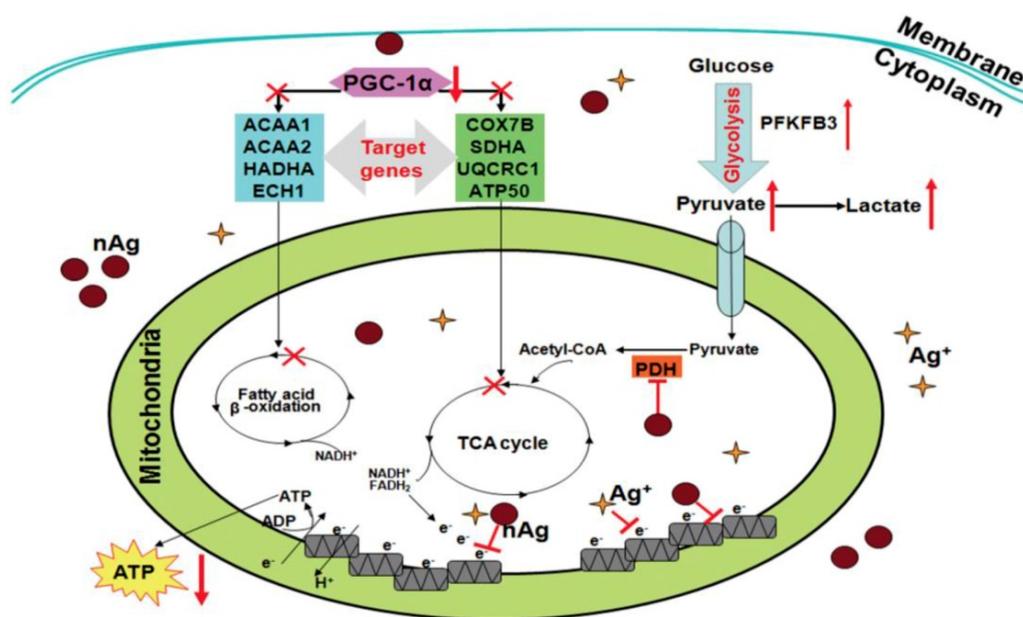


Figure 3. Schematic diagram showing the molecular bases underlying Ag nanoparticles-conducted reprogramming of energy metabolism under sublethal exposure. Reproduced with permission from ref [89]. Copyright 2014 American Chemical Society.

Genetic alteration: DNA structure and gene expression

Genetic alteration testing is of vital importance in the safety assessment of NMs as it may have implications to induce chronic diseases such as cardiovascular disease and mental disorders, or even cancer. The impact of NMs on genes is affected by NMs properties, such as the composition, size, shape, surface properties, and also affected by exposure time, cell type, and treatment regime [101]. Genetic assessments of different NMs have been performed, including their impacts on DNA structure, chromosome, gene expression, potential mutagenesis. Confocal microscopy images showed that localization of Au nanoparticles at nucleus of cancer cells broke DNA double-strand structure, which resulted in binucleate cell formation after mitosis [102]. Li *et al.* identified the interaction energy between different types of nanoparticles and DNA. NMs with high binding affinity to DNA like Ag and Au nanoparticles had greater potential to interfere with DNA replication [103]. Chromosome abnormalities are a direct consequence of DNA structure changes. NMs-induced DNA structure damage such as double strand breaks and misrepair of strand breaks will influence chromosome morphology, resulting in chromosome rearrangement [104]. However, in the research of Xia *et al.*, although significant DNA breaks were observed after Au nanoparticles treatment, there was no significant difference in the incidence of chromosomal aberrations between cells treated with/without Au nanoparticles, indicating that cells were trying to repair DNA damage during this process [105]. The impacts of NMs on genetic structure will appear as changes in protein and gene expression. It is therefore not surprising to find up-regulation of genes associated with DNA repair after NMs exposure [106]. Many other genes associated with cell cycle [81], anti-apoptotic [104], stress response [107] also changed in various cell lines after NMs exposure.

Published data indicate that there are direct and indirect mechanisms of NMs-induced genetic alterations as summarized in Figure 4. Direct genetic changes in NMs-treated cells occur as direct interactions of NMs or their released ions with either nuclear DNA or mitochondrial DNA. The indirect mechanisms are that NMs indirectly affect the molecules and intermediates involved in normal genome function or cell division. ROS production is considered to be the key factor causing NMs-induced gene alteration [101]. Production of ROS can cause DNA injury as well as changes in lipids, proteins, and other cellular components. Induced epigenetic alterations mediate chromosomal

instability or locus specific gene expression changes, which possibly affect multiple cellular pathways and targets. Another mechanism of NM-induced genetic alternation is mediated by extracellular ROS accumulation caused by inflammatory response. Activation of inflammatory responses of macrophages and neutrophils leads to production of ROS, which can also induce genetic alternations in nearby cells [108]. In addition, some other molecular mechanisms have emerged recently that could explain the genetic changes caused by NMs. For example, when NMs accumulated inside endosomes/lysosomes abundantly, acidic conditions of endosomes/lysosomes will change accordingly and promote the release of metal ions or induce cascading intracellular gene toxicity [109]. In these cases, cells initiate various responses to repair DNA damage and strive to adapt the stimulation of NMs. Nevertheless, when NMs-induced genetic alterations are beyond their repair capability or intracellular antioxidants are depleted, cells may undergo cell death via apoptosis or necrosis [104].

Physiological and pathological effects of NMs on tissues and organs

The diverse applications of NMs have led to increasing attention about their possible impact on human beings and the environment. Human beings are easily exposed to NMs via ingestion, inhalation, and skin contact. Lately, the emergence of therapeutic drugs based on NMs makes us more concerned about their fate after administration [42]. More and more imaging modalities for quick and reliable mapping of NMs distribution *in vivo* have been developed to optimize their physicochemical properties for various theranostic applications [110]. Reports suggest that typical *in vivo* fate of NMs depends on multi parameters such as size, shape, charge. These materials tend to accumulate within organs and are slowly degraded or excreted from the body [111]. Due to non-biodegradable nature and prolonged circulation time, NMs have high probability of accumulating in human body and interacting with tissues and organs. The long-time retention of NMs in human body will have adverse effects on individual development and longevity by inducing ROS-mediated stress responses. For example, shortened lifespan and compromised tolerance to oxidative stress were observed when *Drosophila melanogaster* was exposed to sublethal doses of Ag nanoparticles [112]. The life span of *Caenorhabditis elegans* was also reduced when the concentration of Ag nanoparticles exceeded 10 $\mu\text{g}/\text{mL}$ [113]. The clearance of NMs is a big challenge. A certain number of NMs were found in

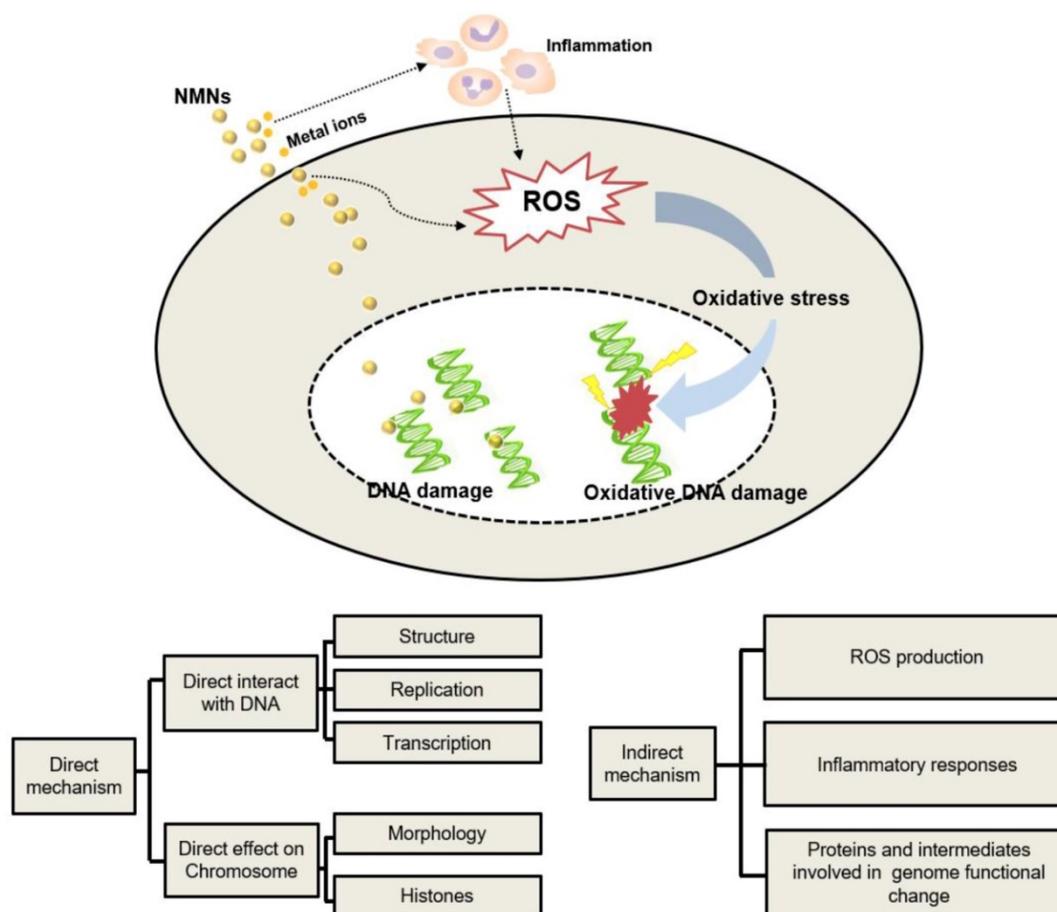


Figure 4. The mechanisms of NMNs-induced genetic alteration.

the liver, spleen, kidney, bone marrow, the central nervous system (CNS) and lymph nodes after inhalation, oral or intravenous injection [114, 115]. The retention of NMNs in these tissues and organs is associated with metabolism, respiration, consciousness and immunity and can result in significant physiological and pathological effects, as shown in Figure 5. Human placenta is a multifunctional organ of particular interest because it controls material exchange between maternal and fetal tissues. The effect of NMNs on embryogenesis requires additional consideration. Increasing evidence has shown that transplacental transfer of nanomaterials also pose effects on embryogenesis [116]. The results of *in situ* hybridization indicated that Ag nanoparticles caused a dose-dependent effect on embryonic development. Ag nanoparticles treatment led to delayed embryo development [117]. Table 2 lists the summary of effects of NMNs on different tissues and organs.

Effects of NMNs on metabolic-related tissues and organs

The gastrointestinal tract is in direct constant contact with the external environment. NMNs are

found in various types of foods or food packaging materials [118]. They can easily enter into human digestive system and be trapped in gastrointestinal tract [119]. It is demonstrated that smaller nanomaterials pass through gastrointestinal tract more easily [120]. After entering vascular circulation, NMNs can be efficiently excreted through the kidney along with the urine. Kidney is the core organ of metabolism and is involved in a range of crucial biological functions [121]. As the most effective way of NMNs excretion, renal clearance of NMNs is favorable due to reduced possible toxic effects because it requires minimal interaction with the body [122]. The emergence of renal clearable NMNs has made it possible toward addressing challenge of nonspecific accumulation of therapeutic NMNs [123]. However, larger NMNs (larger than 6-8 nm) cannot be efficiently cleared via kidney. They may undergo biliary excretion and be eliminated via gastrointestinal tract [42]. This process is governed by the liver, the metabolic center of the body [124]. The liver converts molecules into waste products that can then be excreted through the kidney, representing the primary route for the remove of nanomaterials that cannot be directly cleared by the kidney [125].

Table 2. Summary of the effects of NMNs on different tissues/organs.

| Tissues and organs | and | Type of NMNs | Model | Physiological and pathological effects | References |
|--------------------|-----|------------------|--------------------|---|------------|
| Liver | | Ag nanoparticles | Rats | Liver cytoplasmic vacuolization, no effect in hematology and biochemical parameters | [176] |
| | | Au nanoparticles | Mice | Significant change in genes, but histological analysis did not show any pathological changes | [131] |
| Kidney | | Au nanoparticles | Mice | No pathomorphological changes | [126] |
| | | Au nanoparticles | Rats | Lactate dehydrogenase release and glucuronidase induction, pro-inflammatory effect | [133] |
| | | Pt nanoparticles | Mice | Necrosis of tubular epithelial cells and urinary casts, dose-dependent elevation of blood urea nitrogen | [130] |
| | | Ag nanoparticles | Rats | Dose-dependent effect on alkaline phosphatase and cholesterol. More accumulation in kidneys of female than male | [177] |
| Stomach | | Au nanoparticles | Mice | Renal fibrosis | [79] |
| Intestine | | Ag nanoparticles | 3D-organoid models | Form complexes with <i>Helicobacter pylori</i> , attenuate <i>H. pylori</i> infection | [178] |
| Spleen | | Ag nanoparticles | Mice | Loss of microvilli, reduced intestinal epithelial absorption and reduced weight | [129] |
| | | Ag nanoparticles | Rats | Immune cells in the spleen increased sharply as well as the level of antibody, Spleen weight increased | [173] |
| Lung | | Au nanoparticles | Mice | Distorted lymphoid architecture, minimized lymphoid follicles, diffused white pulp | [175] |
| | | Au nanoparticles | Rats | Significant change in genes related to defensive reaction | [127] |
| | | Au nanoparticles | Mice | Only star-like Au nanoparticles are able to accumulate in the lung, actual penetration into parenchyma | [179] |
| Heart | | Ag nanoparticles | Rats | Yellow discoloration of the lung, but no haematological and histopathological change | [143] |
| | | Ag nanoparticles | Rats | Increases in alveolar inflammation and small granulomatous lesions | [180] |
| | | Ag nanowires | Rats | Lung inflammation at day 1, but disappeared by day 21 | [145] |
| Brain | | Pt nanoparticles | Zebrafish | A concentration dependent drop of heart rate, touch response and axis curvature | [104] |
| | | Ag nanoparticles | Zebrafish | Decrease in heart rate | [146] |
| Neuron | | Ag nanoparticles | Mice | Induce oxidative stress and altered gene expression | [160] |
| | | Ag nanoparticles | Mice | Inflammation | [161] |
| | | Au nanoparticles | Zebrafish | Hypo-locomotor activity and abnormal behavioral activity | [164] |
| | | Au nanoparticles | Mice | Hippocampus functional change, impairment of learning and memory in a size dependent manner | [157] |
| Pancreas | | Ag nanoparticles | Zebrafish | Altered neurological development and result in small heads | [163] |
| Reproduction | | Au nanoparticles | Diabetic rats | Increase relatively ROS factors | [181] |
| | | Au nanoparticles | Mice | Inhibit matrix deposition, enhance angiogenesis | [8] |
| Reproduction | | Ag nanoparticles | Rats | Altered testicular histology and sperm morphology abnormalities | [182] |
| | | Ag nanoparticles | Mice | Atrophy of seminiferous tubules and absence of spermatids | [183] |
| | | Ag nanoparticles | Mice | Microgranulomas in endometrium | [184] |

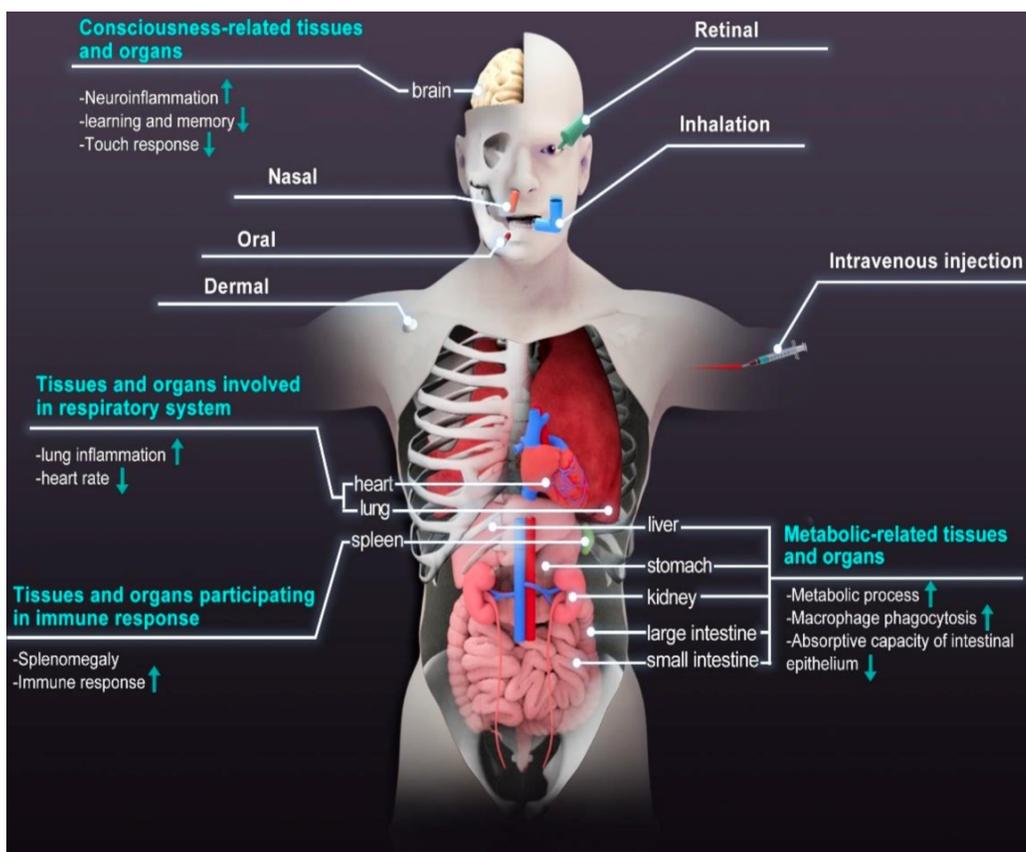


Figure 5. Routes of exposure commonly used for NMNs administration and physiological and pathological effects of NMNs on core tissues and organs.

Compared with other organs, the liver and the kidney, which are in charge of NMNs removal, receive and accumulate external nanomaterials at much higher volumes. As a model of NMNs, metallic Au nanoparticles have been widely reported in various studies to evaluate the biological effects of nanomaterials. They can be easily synthesized with different shapes (balls, rods, dots) and sizes. Au nanoparticles after intravenous administration are easily detected in liver and most of the administered dose can be retained in liver for a long time [126]. A bio-distribution study of Au nanoparticles in rats via intravenous injection showed rapid accumulation of Au nanoparticles in the liver [127]. Even for intratracheal instillation of Au nanoparticles, the uptake and transport of ultrasmall Au nanoparticles resulted in accumulation within the liver [124]. By using quantitative imaging based on laser ablation inductively coupled plasma mass spectrometry, *Elci et al.* demonstrated that surface charge determined suborgan distributions of intravenously injected Au nanoparticles in the kidney, liver and spleen. Positively charged nanoparticles accumulated extensively in the kidney, especially in the glomeruli [128]. All these studies have shown that NMNs are capable of distributing from exposure site (e.g. blood, gut) to the secondary organs (liver, kidney). Since clearance of NMNs may occur over a timescale from several minutes to even a few months, internalized NMNs can persist within the body for a long time, trapped in the kidney, liver, and reticuloendothelial system, causing great effects on these metabolic-related organs.

By interacting with cells of sub-organs, NMNs may affect the function and histopathology of metabolic-related organs. Changes in epithelial cell microvilli as well as intestinal glands were observed in the study of *in vivo* effects of Ag nanoparticles. The loss of microvilli reduced the absorptive capacity of intestinal epithelium and weight loss in mice occurred after Ag nanoparticles treatment [129]. A study evaluated biological effects of Pt nanoparticles less than 1nm in size. The research showed that Pt nanoparticles did not cause obvious side effects in the lung, spleen, and heart. However, histological analysis revealed necrosis of tubular epithelial cells and urinary casts in the kidney. The mice treated with Pt nanoparticles exhibited dose-dependent elevation of blood urea nitrogen. But these toxic effects were not observed in mice injected with 8nm Pt nanoparticles, suggesting the toxicity of Pt nanoparticles may be reduced by increasing nanoparticles size [130]. In analyzing the existing results from the perspective of NMNs liver toxicity, it was found that NMNs did not induce significance damage to the liver after entry

into the body. *Wallin et al.* studied the fate of Au nanoparticles in mice by tail vein injection. The mice were killed at different exposure times and any pathomorphological changes in the liver were observed [126]. *Cho et al.* found PEG-coated 4- and 100-nm Au nanoparticles after intravenous administration in mice shared 67.1% and 50.9% of the significantly changed genes, respectively, whereas histological analysis of the liver tissues did not indicate any pathological changes in all treatment groups [131].

The accumulation of NMNs in metabolic-related organs can be seen as a protective mechanism of the body. The liver is the main detoxifying organ of the body. Liver storage may reduce systemic toxicity of NMNs. The fact showed that some intact NMNs were detected throughout the cytosol in a smaller size than originally administered. This was confirmed by the detection of low-molecular Au nanoparticles with different sizes in the liver [132]. NMNs tend to be digested or metabolized biologically or chemically in liver before being neutralized and stored in the body in an attempt to reduce toxicity. The degradation process was mainly based on phagocytic activity of liver Kupffer cells. Au nanoparticles were identified in almost all Kupffer cells several hours after injection. Although Au nanoparticles were distributed in liver quickly, hepatocytes did not assimilate Au nanoparticles. Transmission electron microscopic analysis revealed that they accumulated inside vesicular structures of macrophages [126]. The same result was described by other authors [133]. Enzyme inside macrophage lysosomes would digest the accumulated NMNs, which somehow reduced systemic toxicity of NMNs. Furthermore, in the study of nanoparticles toxicity assessed on rat liver slices, although Au nanoparticles were rapidly intrahepatic collected, no overt signs of cytotoxicity were observed due to lactate dehydrogenase release, methylthiazolylidiphenyl tetrazolium bromide and glutathione level reduction [133]. These morphological or functional changes increase the body's tolerance to NMNs.

Alteration of tissues and organs involved in respiratory system

Inhalation is considered to be the most important route of NMNs exposure, especially in occupational settings [134]. After inhalation of particle laden air, particles will enter into respiratory tract following airstream. Some inhaled particles tend to travel along the original path to reach deep into the lung, while other particles will be adsorbed on airway surface due to sudden directional change or airway bifurcation [135]. Generally, large particles are likely to be

deposited in nasopharyngeal region. Those particles that fail to be captured in the nasopharyngeal region will retain in tracheobronchial region and further removed by mucociliary clearance. However, the remaining ultrasmall NMNs (<100 nm) may penetrate deep into the lung, translocate beyond epithelial barrier into interstitium where they can escape the clearance mechanism and accumulate in the lung for a long time [136, 137]. The lung is the first port of entry for inhaled nanomaterials into the body. Many types of NMNs-based products, such as sprays, are likely to be inhaled by consumers during respiration and accumulate in the lung. Moreover, NMNs exposures by other pathways were also found in the lung [42, 138]. Since NMNs are nondegradable, they can stay in the lung for a long time once entering respiratory tract. The accumulated NMNs in the lung will enter into different compartments of the cell. Due to particle-cell interactions, the small-sized NMNs may induce detectable alternations in the lung. Importantly, internalized NMNs will be potentially further transported into systemic circulation, accumulate in other organs involved in respiratory system such as the heart, thereby inducing direct effects on these respiratory related organs.

The most commonly used method to evaluate the short-term or long-term effect of NMNs to the lung is by using intratracheal instillation. Many studies have reported the induction of pulmonary inflammation after NMNs exposure [139, 140]. As a crucial indicator of inflammatory reaction, lactate dehydrogenase index is useful to indicate pneumocyte change [141]. For example, *Francesca et al.* tested lactate dehydrogenase and glucuronidase activities of Ag nanoparticles treated alveolar macrophages. Significant lactate dehydrogenase release and glucuronidase induction indicated pro-inflammatory effect of Ag nanoparticles to the lung. Although such obvious changes occurred at cellular level, there were no significant alternation in histology and pathology [142]. The same conclusion was drawn by *Hong et al.* In their repeated-dose toxicity study of Ag nanoparticles, no death was observed in any group. Yellow discoloration of the lungs was observed in a few rats, but the haematology, serum biochemical test and histopathological analysis between control and Ag treated groups revealed no statistically significant differences [143]. For seeking of hazard ranking of nanomaterials, the acute lung, liver and systemic responses were assessed in C57BL/6N mice for three different nanomaterials. Exposure to low dose of Ag nanoparticles was also not accompanied by obvious pulmonary change or cytotoxicity [144]. Pulmonary inflammation is initially a defense response to exotic nanomaterials. When

NMNs are deposited in the lung, the body tries to recruit more macrophages through inflammatory reaction to remove nanomaterials. Most of NMNs are phagocytosed and digested by pulmonary macrophages [124]. Quantitative analysis of both short and long Ag nanowires inside the lung showed a small but significant reduction of Ag nanowires lengths. For both Ag nanowires, there was lung inflammation at day 1, but disappeared at day 21 [145]. The lung tries to inactivate and digests internalized NMNs. However, the durability of nondegradable NMNs in the lung might finally cause pulmonary fibrosis, which eventually affects respiratory function.

The small-sized NMNs may also cause alternations of other respiratory-related organs, among which the heart has attracted more attention than other organs. Comparison of toxicity of Ag, Au and Pt nanoparticles in developing zebrafish embryos revealed that both Ag and Pt nanoparticles induced a concentration dependent drop of heart rate, touch response and axis curvature. Ag nanoparticles even induced cardiac morphology change, while Au nanoparticles treatment did not cause any variation of zebrafish heart [104]. The decrease in heart rate induced by NMNs was also reported by *Lee et al* [146]. Cardiovascular system changes upon inhaled NMNs exposure can be explained by the following mechanisms. On one hand, the small size characteristics of NMNs allow them enter blood circulation by passing through the thin alveolar-capillary wall and influence cardiovascular system directly. On the other hand, long-term and amplified inflammation responses lead to overflow of inflammatory factors into the blood, which provides signals to the CNS to regulate heart activity. Furthermore, the deposited NMNs in the lung may stimulate alveolar sensory receptors to further activate cardiovascular system [147]. The autonomic regulation of nervous system activity will eventually manifest as alternation of cardiac function.

Alteration of consciousness-related tissues and organs

The CNS is the most important part of controlling regular physiological functions of animals. As one of the tightest barriers in human body, the BBB protects the CNS from toxins and large, hydrophilic molecules diffusing into cerebrospinal fluid [148]. In order to protect against invading substances or organisms, the BBB plays a key role in guarding the brain parenchyma and keeping a stable environment for normal neuron activities [149]. NMNs exhibit extraordinary properties and are considered as superior carriers for achieving the delivery of a broad

range of therapeutics into the brain which is well protected by the BBB [150]. More and more studies have demonstrated that NMNs could cross the BBB and further access the CNS via several different routes [151, 152]. NMNs can access to the blood or lymph nodes and translocate to the brain, or direct access to the brain from nasal cavity via nose-brain transport [153]. After crossing the BBB, nanomaterials have a tendency to accumulate in specific regions of the brain including olfactory bulb, hippocampus, cerebral cortex, and striatum [154, 155]. The accumulated NMNs in brain may gain access to neural cells, such as astrocytes, neurons, and microglia [156]. The interactions between NMNs and those cells could greatly affect the structure, function, or chemistry of the CNS, followed by a variety of cognitive changes [157].

After entering the brain, NMNs can cause direct alteration of the structure and activity of the neural system or the follow-up effects. The effects of NMNs on CNS are various, including oxidative stress, cell apoptosis and autophagy, immune responses, and neuroinflammation. The most commonly reported are the extensive production of ROS, which leads to oxidative stress and inflammation [151]. The brain oxygen consumption accounts for nearly a quarter of the whole body's oxygen consumption. Hence, the brain is more sensitive to hypoxic injury than other tissues. Upon NMNs treatment, cytokines release in the brain may cause neuroinflammation, and then neuronal damage, which will finally affect associated brain functions [153]. Many commonly used NMNs have been reported as potentially neurotoxic materials [158]. Researchers have analyzed changes in brain tissue and related brain functions after NMNs treatment, and histological alterations of the brain were observed. Deposition of NMNs in neural tissues, cerebral cortex or hippocampus can affect associated brain functions such as attention, memory, and judgement [159].

Rahman et al. found intraperitoneal injection of Ag nanoparticles in mice induced oxidative stress and altered gene expression in the brain [160]. Inflammation- and oxidative stress-related brain alterations in adult mice were also observed after intravenous administration of Ag nanoparticles [161]. Since the morphology of the brain may change under long-term NMNs exposure, *El-Drieny et al.* studied the histology of adult rat brain after long-term Au nanoparticles treatment. Au nanoparticles mainly deposited in the neurons of cerebral cortex and hippocampus as well as the epithelium of choroid plexus [162]. The study of influence of Ag nanoparticles on neurological development showed Ag nanoparticles altered neurological development of

zebrafish and resulted in small heads [163]. These changes may finally lead to alternations of brain functions. *Truong et al.* found Au nanoparticles treated zebrafish exhibited hypo-locomotor and abnormal behavioral activities [164]. Another example showed invasion of different sized Au nanoparticles to brain hippocampus caused impairment of learning and memory in a size dependent manner, which was associated with hippocampus functional change [165]. Adaptive change in the brain such as early inflammatory warn the body to protect it from more serious damage, but long-term stimulation will cause irreversible effects like morphological change in the brain. All these changes in consciousness-related tissues and organs allow us to better understand both beneficial and negative impacts of NMNs on the brain.

Modulation of tissues and organs participating in immune response

The immune system is in charge of defending against foreign substances in order to protect the host. It is divided into innate immunity and adaptive immunity which are considered as two equally crucial aspects of the immune system. The innate immunity is nonspecific and the first line of defense of human body. It plays an essential role in early recognition of foreign substances and subsequent defense responses. Unlike innate immune immunity, the adaptive immune system is antigen-specific and reacts with organisms that induce the response [166]. The innate immunity and adaptive immunity together form a sensitive target for xenobiotic induced toxicity. When exposed to the body, most NMNs are recognized as foreign antigens and thus elicit immune response. Although proteins in biological environment may adsorb to NMNs "foreign" surface and result in poor recognition by immune cells, however, most NMNs cannot escape immune recognition [167]. Since innate defense system is the first in contact with foreign materials entering into the body, most studies of immunotoxicology induced by NMNs have focused on changes in innate immune system [168]. There are enriched innate immune systems particularly in tissues that come into contact with external environment such as skin, respiratory mucosa, and gastrointestinal tract. However, NMNs may transport and penetrate through epithelial barriers to the blood due to ultrasmall size. Inside the body, NMNs will be endocytosed by immune cells, accumulate in tissues and organs of immune system, and interact with those organs [169]. These direct interactions have profound effects on tissues and organs participating in immune response, which may be shown as activation or inhibition of immune function [170].

The most frequently mentioned immunological organ affected by NMNs is the spleen. The spleen is warehouse of the blood and is responsible for filtering the blood. As the largest lymphoid organ, the spleen is important both in innate and adaptive immune responses [171]. NMNs can easily accumulate in the spleen after being phagocytosed, causing adaptive changes of the spleen both in morphology and function. The intravenously injected Ag nanoparticles with large particle size (80, 110 nm) had the highest concentration in the spleen compared to that of small particle size (20 nm). Larger NMNs are more likely to be recognized by macrophages than smaller sized one and are more likely to be trapped in the spleen [172]. Moreover, particle size may influence their distribution in the spleen. Accumulation of Ag nanoparticles were found in the spleen, mesenteric lymph nodes, and axillary lymph nodes. 20 nm Ag nanoparticles was only distributed in the red pulp of the spleen, while 100 nm Ag nanoparticles is distributed in both the red and white pulp of the spleen [173]. The widespread distribution of NMNs in the spleen is a manifestation of the body trying to remove foreign particles, which may cause morphology change of the spleen. Splenomegaly is often reported after nanoparticles deposition. In attempt to clear internalized NMNs, immune response will be activated to reduce toxicity. Lymphocyte proliferation is an important phase in immune response. It was shown in the research of *Wim* and co-workers that the number of T cells, B cells, and NK cells in the spleen increased sharply as well as the level of antibody IgM and IgE after Ag nanoparticles exposure, resulting in significant increase of spleen weight [173]. This suggested that NMNs are able to regulate immune response by activating lymphocyte. Based on this conclusion, *Marc et al.* analyzed whether tumor presence would influence nanoparticles circulation since the immune system is altered when tumor exist. Due to an increase of M2-like macrophages, tumor presence significantly increased the clearance of nanoparticles from blood circulation, resulting in increased accumulation in the liver and spleen [174].

The immune response is an adaptive response involving multiple cell types and mediators. After entering human body, NMNs are recognized by “detectives” in the body and trigger a series of immune responses to try to get rid of them. Their relationship is like a hunter and a killer. The modulation of tissues and organs participating in immune system upon NMNs exposure enables the body react quickly and become adaptive to NMNs stimulus in a short time. However, this may be a double-edged sword. The adaptive changes induced

by NMNs under a long period of time may cause pathological changes of immune-related tissues and organs, which lead to damage of immunological functions. The mice spleen showed obvious pathological changes under constant stimulation of Au nanoparticles, which were observed in form of distorted lymphoid architectures, minimized lymphoid follicles, diffused white pulp, and increased giant macrophages [175]. Long term retention of NMNs in the spleen even lead to potential genetic changes. *Suresh et al.* examined gene expression in rat liver and spleen after intravenous administration of Au nanoparticles. Biodistribution of Au nanoparticles in the spleen at 1 day, 1 week, 1 month and 2 months were observed. The spleen showed significant changes in genes related to defensive reaction. Multiple genes involved in external stimuli, wound healing, and coagulation were downregulated, suggesting reduced spleen response external stimuli after long time Au nanoparticles treatment [127].

Conclusion and perspectives

The ever-growing application of NMNs in therapeutics, diagnostics and bioimaging fields have yielded significant positive impacts toward the betterment of human health. The popularization of diverse NMNs-based products makes us more concern about their fate after entering human body. Inside the body, NMNs will distribute to specific tissues and organs, target cells and eventually interact with subcellular components. In order to better accommodate NMNs stimulation, cells and tissues will undergo various adaptive changes both *in vitro* and *in vivo* for better survival. Cellular and subcellular morphology changes are the first and most easily observed. Cells tend to become more rounded from stretched state to reduce their exposure of nanomaterials [185]. The accumulation of NMNs in different subcellular components and their interaction with core organelles such as lysosomes, ER, Golgi apparatus, mitochondria, and nucleus will interfere their functions and cause influence on a variety of cellular functions. Cells try to adjust their digestion, protein synthesis and secretion, energy metabolism, mitochondrial respiration, and proliferation in response to intracellular foreign bodies. For example, the induction of autophagy is a defense and stress regulation mechanism of cells to try to make up the missed degradative function after lysosome swelling [70, 186]. Changes in protein processing and cell adhesion are the result of cells to better withstand NMNs stimulation [76]. Even under conditions where oxygen is sufficient, cells treated with NMNs might shift energy homeostasis as an adaptive process to satisfy energy demand for their survival [89].

Moreover, adaptive changes of cells induced by long-term NMNs are different from that induced by short-term exposure, and long-term exposed cells increase their adaptability to NMNs stimulation [35, 43]. When it comes to a cluster of cells, these cumulative effects are manifested as adaptive changes in tissues and organs. The chronic effects of NMNs on human body are of great interest since most MNs are nondegradable and stay in tissues and organs for a long time. Accumulation of NMNs in metabolic-, respiratory-, consciousness- and immune-related tissues and organs seems to trigger significant physiological and pathological effects on core bodily functions. For instance, the liver is a detoxifying organ. Before being neutralized or stored in the body, NMNs tend to be digested or metabolized in the liver to reduce toxicity. When NMNs are deposited in the lung, pulmonary inflammation is initially a defense response to exotic nanomaterials. Changes in the brain such as early inflammatory warn the body to protect it from more serious damage. The body tries to recruit more macrophages through inflammatory reaction to remove nanomaterials. The immune system is in charge of defending against foreign substances in order to protect the host cells. For the body, to trigger a series of immune responses is a typical example to protect the body against NMNs exposure [167, 187]. Definitely, under long-term stimulation of NMNs, there may be many adverse and even irreversible effects. Multi body functions will be affected, like energy supply, respiratory behavior, and even cognitive level. Premature senescence will appear under long-term stress induced by NMNs [98]. When these adverse changes are beyond self-repair ability, cell death may occur via apoptosis or necrosis [104]. The long-time retention of NMNs will have adverse effects on individual development and life span [112]. Moreover, NMNs can also cross the placental barrier and pose effect on embryogenesis, which requires additional consideration [117].

Detailed investigation of interactions between NMNs and human body provides more comprehensive references for their applications. Ideally, these adaptive changes induced by NMNs can be exploited to create fantastic nanosystems. Through in-depth study of these adaptive changes, on one hand, we can take advantages of favorable adaptive changes, on the other hand, unfavorable changes can be avoided as much as possible to maximize the expected effects. The growing incidence of cancer is the biggest culprit of human death. At the cellular level, rapid tumor growth and proliferation depend on normal function of cancer cells, which is strongly associated with various roles of cellular and

subcellular components. Thus, the adaptive changes followed by NMNs treatment can be designed to affect metabolism of cancer cells and enhance cancer cells killing. By affecting different cellular processes like cellular digestion, metabolism, respiration, proliferation as well as cell morphology, tailored NMNs provide great potential to increase sensitivity of cancer cells [10]. For example, cellular morphology integrity is crucial for tumor invasion and metastasis [55]. Alterations in cytoskeleton, organelles and other subcellular structures by NMNs treatment make cancer cells more susceptible to chemotherapy [188]. Mitochondria respiration is directly associated with a variety of signal pathways that involved in apoptosis and necrosis. This highlights the usage of NMNs in drug resistant reversal through disruption of mitochondria energy metabolism [89]. Also, NMN-induced autophagy, DNA dysfunction, ER stress and cell cycle redistribution have huge potential for smart therapeutic nanosystem design [189-191]. In animal, the immune response initiated by NMNs as a protective mechanism can be well utilized with tumor immunotherapy to amplify immune efficacy [192]. However, not all adaptive changes induced by NMNs are beneficial. For example, the accumulation of NMNs in detoxifying organs is seen as protective responses to reduce systemic toxicity of NMNs. But for therapeutic proposes, we hope that drug loaded NMNs circulate in the body for a long time and accumulate in specific lesions. Thus, various modifications of NMNs have been used to limit their side-effects. The size of nanomedicine is controlled to minimize their accumulation in metabolic organs and prolong their blood circulation time [193]. The surface charge and shape of specific NMNs are also important to their *in vitro* and *in vivo* fate. For example, spherical NMNs are most commonly used drug carriers since renal clearable NMNs are generally spherical. Nanostructures with high aspect ratios but diameters smaller than kidney filtration threshold are also found to be renal clearable and therefore nonspecific accumulation in healthy tissues/organs is reduced [123]. Surface charge affects *in vivo* fate of NMNs by influencing opsonization process. The density of surface charge impacts plasma protein adsorption both qualitatively and quantitatively, which, subsequently, has critical impacts on blood circulation and biodistribution of NMNs. High surface charge densities usually result in faster blood clearance while neutral charged NMNs contribute to extended blood circulation and reduced reticuloendothelial system clearance [194]. Multiple factors control the circulation and organ clearance of NMNs. Structure design and surface modifications of NMNs are crucial to improve the biocompatibility, *in vivo* metabolism

and adverse changes induced by NMNs. The size, shape, surface charge, and functional group of NMNs as well as tissue microenvironment could be monitored according to different purposes to reduce their side effect, change their blood circulation time, and regulate specific retention profiles within organs. Glutathione-mediated biotransformation is a well-known detoxification process in the liver to eliminate small xenobiotics. By designing a glutathione-coated Au nanoprobe that can bind to serum protein and be transported to the liver, Jiang *et al.* used detoxification process in the liver as a bridge toward maximizing targeting and minimizing nanotoxicity [195]. Excessive immune response can damage the normal function of the body. The adaptive change of immune system under long-term NMNs stimulation may cause immunological function damage. Moreover, rapid identification by immune system results in rapid elimination of nanomedicine in human body. In order to avoid these problems, surface group modified and cell membrane coated NMNs are commonly used to escape immune recognition [196, 197]. The investigation of interactions between NMNs and cells allows us to screen the most suitable NMNs to achieve maximum benefits, thereby taking advantage of favorable adaptive changes induced by NMNs and limiting their side-effects through surface modifications.

Here, an in-depth understanding of interactions among NMNs and cells, tissues, and organs are provided. Systematic study of adaptive changes induced by NMNs *in vitro* and *in vivo* enables us to better utilize NMNs for smart nanosystem design. Still, there is a lot of work to be done before a comprehensive understanding of NMNs-related adaptive changes is achieved. Apart from aforementioned observations, little is known about the fate of NMNs in human body. Detailed comparative studies on long-term and short-term effects of NMNs are lacking. Sometimes there is no obvious boundaries between adaptive changes and toxicities caused by NMNs. Most importantly, the choice of animal model is also a problem which needs to be considered carefully since many animal models we used now cannot fully reflect the complex physiological environment of human body. To elucidate the interplay of NMNs with human body, further in-depth research of *in vivo* tracking of NMNs is expected. Moreover, complete analysis of NMNs kinetics after administration needs to be conducted. We hope that the adaptive effects induced by NMNs will attract more and more interests for the design of smart nanopharmaceuticals, and it is possible that in the near future, advances in nanobiotechnology could enhance our capacity to make full use of these

adaptive changes for improving the quality of human life.

Abbreviations

Au: gold; Ag: silver; BBB: blood brain barrier; CNS: central nervous system; CT: computed tomography; ER: endoplasmic reticulum; Ir: iridium; LSPR: localized surface plasmon resonance; MMP: mitochondrial membrane potential; NMNs: noble-metal nanostructures; Os: osmium; Pd: palladium; Pt: platinum; PEG: polyethylene glycol; PTT: photothermal therapy; PDT: photodynamic therapy; ROS: reactive oxygen species; Rh: rhodium; Ru: ruthenium; SERS: surface-enhanced Raman spectroscopy; UPR: unfolded protein response; VEGF: vascular endothelial growth factor.

Acknowledgements

This work was supported Chinese National Natural Science Foundation project (Grant No. 31971312), and Beijing Natural Science Foundation project (Grant No. 7192151). This work was partly supported by the National Natural Science Foundation of China (NSFC) key project (grant No. 31630027) and NSFC-German Research Foundation (DFG) project (grant No. 31761133013). The authors also appreciate the support by the “Ten Thousand Elite Plan” (grant No. Y9E21Z11) and CAS international collaboration plan (grant No. E0632911ZX) as well as National Key Research & Development Program of China (grant No. 2018YFE0117800).

Competing Interests

The authors have declared that no competing interest exists.

References

1. Yu MK, Park J, Jon S. Targeting strategies for multifunctional nanoparticles in cancer imaging and therapy. *Theranostics*. 2012; 2: 3-44.
2. Singh AP, Biswas A, Shukla A, Maiti P. Targeted therapy in chronic diseases using nanomaterial-based drug delivery vehicles. *Signal Transduct Target Ther*. 2019; 4: 1-21.
3. Azharuddin M, Zhu GH, Das D, Ozgur E, Uzun L, Turner APF, et al. A repertoire of biomedical applications of noble metal nanoparticles. *Chem Commun*. 2019; 55: 6964-96.
4. Arvizo RR, Bhattacharyya S, Kudgus RA, Giri K, Bhattacharya R, Mukherjee P. Intrinsic therapeutic applications of noble metal nanoparticles: past, present and future. *Chem Soc Rev*. 2012; 41: 2943-70.
5. Loynachan CN, Soleimany AP, Dudani JS, Lin YY, Najer A, Bekdemir A, et al. Renal clearable catalytic gold nanoclusters for *in vivo* disease monitoring. *Nat Nanotechnol*. 2019; 14: 883-890.
6. Gong NQ, Ma XW, Ye XX, Zhou QF, Chen XA, Tan XL, et al. Carbon-dot-supported atomically dispersed gold as a mitochondrial oxidative stress amplifier for cancer treatment. *Nat Nanotechnol*. 2019; 14: 379-387.
7. Huo S, Gong N, Jiang Y, Chen F, Guo H, Gan Y, et al. Gold-DNA nanosunflowers for efficient gene silencing with controllable transformation. *Sci Adv*. 2019; 5: eaaw6264.
8. Saha S, Xiong X, Chakraborty PK, Shameer K, Arvizo RR, Kudgus RA, et al. Gold nanoparticle reprograms pancreatic tumor microenvironment and inhibits tumor growth. *ACS Nano*. 2016; 10: 10636-51.
9. Rastinehad AR, Anastos H, Wajswol E, Winoker JS, Sfakianos JP, Doppalapudi SK, et al. Gold nanoshell-localized photothermal ablation of prostate tumors in a clinical pilot device study. *Proc Natl Acad Sci U S A*. 2019; 116: 18590-6.

10. Wang YF, Liu J, Ma XW, Liang XJ. Nanomaterial-assisted sensitization of oncotherapy. *Nano Res.* 2018; 11: 2932-50.
11. Zhang L, Yang XQ, Wei JS, Li X, Wang H, Zhao YD. Intelligent gold nanostars for *in vivo* CT imaging and catalase-enhanced synergistic photodynamic & photothermal tumor therapy. *Theranostics.* 2019; 9: 5424-42.
12. Paithankar D, Hwang BH, Munavalli G, Kauvar A, Lloyd J, Blomgren R, et al. Ultrasonic delivery of silica-gold nanoshells for photothermolysis of sebaceous glands in humans: Nanotechnology from the bench to clinic. *J Control Release.* 2015; 206: 30-6.
13. Li X, Robinson SM, Gupta A, Saha K, Jiang Z, Moyano DF, et al. Functional gold nanoparticles as potent antimicrobial agents against multi-drug-resistant bacteria. *ACS Nano.* 2014; 8: 10682-6.
14. Bowman MC, Ballard TE, Ackerson CJ, Feldheim DL, Margolis DM, Melander C. Inhibition of HIV fusion with multivalent gold nanoparticles. *J Am Chem Soc.* 2008; 130: 6896-7.
15. Noonan J, Asiala SM, Grassia G, MacRitchie N, Gracie K, Carson J, et al. *In vivo* multiplex molecular imaging of vascular inflammation using surface-enhanced Raman spectroscopy. *Theranostics.* 2018; 8: 6195-209.
16. Jadhav K, Hr R, Deshpande S, Jagwani S, Dhamecha D, Jalalpure S, et al. Phytosynthesis of gold nanoparticles: Characterization, biocompatibility, and evaluation of its osteoinductive potential for application in implant dentistry. *Mater Sci Eng C Mater Biol Appl.* 2018; 93: 664-70.
17. Liu J, Zhao Y, Guo Q, Wang Z, Wang H, Yang Y, et al. TAT-modified nanosilver for combating multidrug-resistant cancer. *Biomaterials.* 2012; 33: 6155-61.
18. Amarnath Praphakar R, Jeyaraj M, Ahmed M, Suresh Kumar S, Rajan M. Silver nanoparticle functionalized CS-g-(CA-MA-PZA) carrier for sustainable anti-tuberculosis drug delivery. *Int J Biol Macromol.* 2018; 118: 1627-38.
19. Rai M, Kon K, Ingle A, Duran N, Galdiero S, Galdiero M. Broad-spectrum bioactivities of silver nanoparticles: the emerging trends and future prospects. *Appl Microbiol Biotechnol.* 2014; 98: 1951-61.
20. Abebe LS, Su YH, Guerrant RL, Swami NS, Smith JA. Point-of-Use removal of cryptosporidium parvum from water: independent effects of disinfection by silver nanoparticles and silver ions and by physical filtration in ceramic porous media. *Environ Sci Technol.* 2015; 49: 12958-67.
21. Dias HB, Bernardi MIB, Bauab TM, Hernandez AC, de Souza Rastelli AN. Titanium dioxide and modified titanium dioxide by silver nanoparticles as an anti biofilm filler content for composite resins. *Dent Mater.* 2019; 35: e36-e46.
22. Lee JS, Oh J, Kim SG, Jang J. Highly sensitive and selective field-effect-transistor nonEnzyme dopamine sensors based on Pt/conducting polymer hybrid nanoparticles. *Small.* 2015; 11: 2399-406.
23. Zhang L, Laug L, Munchgesang W, Pippel E, Gosele U, Brandsch M, et al. Reducing stress on cells with apoferritin-encapsulated platinum nanoparticles. *Nano Lett.* 2010; 10: 219-23.
24. Gopal J, Hasan N, Manikandan M, Wu HF. Bacterial toxicity/compatibility of platinum nanospheres, nanocuboids and nanoflowers. *Sci Rep.* 2013; 3: 1260.
25. Bai Z, Li G, Liang J, Su J, Zhang Y, Chen H, et al. Non-enzymatic electrochemical biosensor based on Pt NPs/RGO-CS-Fc nano-hybrids for the detection of hydrogen peroxide in living cells. *Biosens Bioelectron.* 2016; 82: 185-94.
26. Weiss JT, Dawson JC, Macleod KG, Rybski W, Fraser C, Torres-Sanchez C, et al. Extracellular palladium-catalysed dealkylation of 5-fluoro-1-propargyl-uracil as a bioorthogonally activated prodrug approach. *Nat Commun.* 2014; 5: 3277.
27. Tang S, Chen M, Zheng N. Sub-10-nm Pd nanosheets with renal clearance for efficient near-infrared photothermal cancer therapy. *Small.* 2014; 10: 3139-44.
28. Huang N, Chen X, Zhu X, Xu M, Liu J. Ruthenium complexes/polypeptide self-assembled nanoparticles for identification of bacterial infection and targeted antibacterial research. *Biomaterials.* 2017; 141: 296-313.
29. Liu Y, Liu Y, Zheng C, Huang N, Chen X, Zhu X, et al. Ru nanoparticles coated with gamma-Fe₂O₃ promoting and monitoring the differentiation of human mesenchymal stem cells via MRI tracking. *Colloids Surf B Biointerfaces.* 2018; 170: 701-11.
30. Kang S, Shin W, Choi MH, Ahn M, Kim YK, Kim S, et al. Morphology-controlled synthesis of rhodium nanoparticles for cancer phototherapy. *ACS Nano.* 2018; 12: 6997-7008.
31. Zhang L, Li Y, Che W, Zhu D, Li G, Xie Z, et al. AIE multinuclear Ir(III) complexes for biocompatible organic nanoparticles with highly enhanced photodynamic performance. *Adv Sci.* 2019; 6: 1802050.
32. Fan Y, Li C, Li F, Chen D. pH-activated size reduction of large compound nanoparticles for *in vivo* nucleus-targeted drug delivery. *Biomaterials.* 2016; 85: 30-9.
33. Wu Y, Shi M, Zhao L, Feng W, Li F, Huang C. Visible-light-excited and europium-emissive nanoparticles for highly-luminescent bioimaging *in vivo*. *Biomaterials.* 2014; 35: 5830-9.
34. Vasan N, Baselga J, Hyman DM. A view on drug resistance in cancer. *Nature.* 2019; 575: 299-309.
35. Falagan-Lotsch P, Grzincic EM, Murphy CJ. One low-dose exposure of gold nanoparticles induces long-term changes in human cells. *Proc Natl Acad Sci U S A.* 2016; 113: 13318-23.
36. Waris G, Ahsan H. Reactive oxygen species: role in the development of cancer and various chronic conditions. *J Carcinog.* 2006; 5: 14.
37. Abdal Dayem A, Hossain MK, Lee SB, Kim K, Saha SK, Yang GM, et al. The role of reactive oxygen species (ROS) in the biological activities of metallic nanoparticles. *Int J Mol Sci.* 2017; 18.
38. Shvedova AA, Pietroiusti A, Fadeel BL, Kagan VE. Mechanisms of carbon nanotube-induced toxicity: focus on oxidative stress. *Toxicol Appl Pharmacol.* 2012; 261: 121-33.
39. Maysinger D, Gran ER, Bertorelle F, Fakhouri H, Antoine R, Kaul ES, et al. Gold nanoclusters elicit homeostatic perturbations in glioblastoma cells and adaptive changes of lysosomes. *Theranostics.* 2020; 10: 1633-48.
40. Chen WH, Luo GF, Zhang XZ. Recent advances in subcellular targeted cancer therapy based on functional materials. *Adv Mater.* 2019; 31: e1802725.
41. Ma XW, Gong NQ, Zhong L, Sun JD, Liang XJ. Future of nanotherapeutics: Targeting the cellular sub-organelles. *Biomaterials.* 2016; 97: 10-21.
42. Bourquin J, Milosevic A, Hauser D, Lehner R, Blank F, Petri-Fink A, et al. Biodistribution, clearance, and long-term fate of clinically relevant nanomaterials. *Adv Mater.* 2018; 30: e1704307.
43. Comfort KK, Braydich-Stolle LK, Maurer EJ, Hussain SM. Less is more: long-term *in vitro* exposure to low levels of silver nanoparticles provides new insights for nanomaterial evaluation. *ACS Nano.* 2014; 8: 3260-71.
44. Gunduz N, Ceylan H, Guler MO, Tekinay AB. Intracellular accumulation of gold nanoparticles leads to inhibition of macropinocytosis to reduce the endoplasmic reticulum stress. *Sci Rep.* 2017; 7: 40493.
45. Park Y, Depeursinge C, Popescu G. Quantitative phase imaging in biomedicine. *Nat Photonics.* 2018; 12: 578-89.
46. Cheng LC, Jiang XM, Wang J, Chen CY, Liu RS. Nano-bio effects: interaction of nanomaterials with cells. *Nanoscale.* 2013; 5: 3547-69.
47. Pernodet N, Fang XH, Sun Y, Bakhtina A, Ramakrishnan A, Sokolov J, et al. Adverse effects of citrate/gold nanoparticles on human dermal fibroblasts. *Small.* 2006; 2: 766-73.
48. Soenen SJ, Manshian B, Montenegro JM, Amin F, Meermann B, Thiron T, et al. Cytotoxic effects of gold nanoparticles: a multiparametric study. *ACS Nano.* 2012; 6: 5767-83.
49. Mironava T, Hadjiargyrou M, Simon M, Jurukovski V, Rafailovich MH. Gold nanoparticles cellular toxicity and recovery: effect of size, concentration and exposure time. *Nanotoxicology.* 2010; 4: 120-37.
50. Soenen SJ, Manshian BB, Abdelmonem AM, Montenegro JM, Tan S, Balcaen L, et al. The cellular interactions of PEGylated gold nanoparticles: effect of PEGylation on cellular uptake and cytotoxicity. *Part Part Syst Charact.* 2014; 31: 794-800.
51. Yang JA, Lohse SE, Murphy CJ. Tuning cellular response to nanoparticles via surface chemistry and aggregation. *Small.* 2014; 10: 1642-51.
52. Montano E, Vivo M, Guarino AM, di Martino O, Di Luccia B, Calabro V, et al. Colloidal silver induces cytoskeleton reorganization and E-cadherin recruitment at cell-cell contacts in HaCaT cells. *Pharmaceuticals.* 2019; 12: 72.
53. De Matteis V, Cascione M, Toma CC, Leporati S. Morphomechanical and organelle perturbation induced by silver nanoparticle exposure. *J Nanopart Res.* 2018; 20: 273.
54. Ma XW, Hartmann R, de Aberasturi DJ, Yang F, Soenen SJH, Manshian BB, et al. Colloidal gold nanoparticles induce changes in cellular and subcellular Morphology. *ACS Nano.* 2017; 11: 7807-20.
55. Dogterom M, Koenderink GH. Actin-microtubule crosstalk in cell biology. *Nat Rev Mol Cell Bio.* 2019; 20: 38-54.
56. Lin IC, Liang MT, Liu TY, Ziora ZM, Monteiro MJ, Toth I. Interaction of densely polymer-coated gold nanoparticles with epithelial caco-2 monolayers. *Biomacromolecules.* 2011; 12: 1339-48.
57. Xu LM, Dan M, Shao AL, Cheng X, Zhang CP, Yokel RA, et al. Silver nanoparticles induce tight junction disruption and astrocyte neurotoxicity in a rat blood-brain barrier primary triple coculture model. *Int J Nanomedicine.* 2015; 10: 6105-19.
58. Lee E, Jeon H, Lee M, Ryu J, Kang C, Kim S, et al. Molecular origin of AuNPs-induced cytotoxicity and mechanistic study. *Sci Rep.* 2019; 9: 2494.
59. Mironava T, Hadjiargyrou M, Simon M, Rafailovich MH. Gold nanoparticles cellular toxicity and recovery: Adipose Derived Stromal cells. *Nanotoxicology.* 2014; 8: 189-201.
60. Ma X, Gong N, Zhong L, Sun J, Liang XJ. Future of nanotherapeutics: Targeting the cellular sub-organelles. *Biomaterials.* 2016; 97: 10-21.
61. Ma X, Wu Y, Jin S, Tian Y, Zhang X, Zhao Y, et al. Gold nanoparticles induce autophagosome accumulation through size-dependent nanoparticle uptake and lysosome impairment. *ACS Nano.* 2011; 5: 8629-39.
62. Zhou HL, Gong XQ, Lin HY, Chen HM, Huang DT, Li D, et al. Gold nanoparticles impair autophagy flux through shape-dependent endocytosis and lysosomal dysfunction. *J Mater Chem B.* 2018; 6: 8127-36.
63. Lawrence RE, Zoncu R. The lysosome as a cellular centre for signalling, metabolism and quality control. *Nat Cell Biol.* 2019; 21: 133-42.
64. Xie ZP, Klionsky DJ. Autophagosome formation: Core machinery and adaptations. *Nat Cell Biol.* 2007; 9: 1102-9.
65. Zhang L, Chen XR, Wu JZ, Ding SP, Wang X, Lei QF, et al. Palladium nanoparticles induce autophagy and autophagic flux blockade in HeLa cells. *RSC Adv.* 2018; 8: 4130-41.
66. Lee YH, Cheng FY, Chiu HW, Tsai JC, Fang CY, Chen CW, et al. Cytotoxicity, oxidative stress, apoptosis and the autophagic effects of silver nanoparticles in mouse embryonic fibroblasts. *Biomaterials.* 2014; 35: 4706-15.
67. Li JJ, Hartono D, Ong CN, Bay BH, Yung LYL. Autophagy and oxidative stress associated with gold nanoparticles. *Biomaterials.* 2010; 31: 5996-6003.
68. Sipsos A, Kim KJ, Sioutas C, Crandall ED. Evidence for nanoparticle-induced lysosomal dysfunction in lung adenocarcinoma (A549) cells. *Int J Mol Sci.* 2019; 20: 5253.

69. Mizushima N, Levine B, Cuervo AM, Klionsky DJ. Autophagy fights disease through cellular self-digestion. *Nature*. 2008; 451: 1069-75.
70. Nakashima A, Higashisakaz K, Kusabiraki T, Aoki A, Ushijima A, Ono Y, et al. Autophagy is a new protective mechanism against the cytotoxicity of platinum nanoparticles in human trophoblasts. *Sci Rep*. 2019; 9: 5478.
71. Li L, Li L, Zhou X, Yu Y, Li Z, Zuo D, et al. Silver nanoparticles induce protective autophagy via Ca(2+)/CaMKKbeta/AMPK/mTOR pathway in SH-SY5Y cells and rat brains. *Nanotoxicology*. 2019; 13: 369-91.
72. Phillips MJ, Voeltz GK. Structure and function of ER membrane contact sites with other organelles. *Nat Rev Mol Cell Bio*. 2016; 17: 69-82.
73. Gao P, Pan W, Li N, Tang B. Boosting cancer therapy with organelle-targeted nanomaterials. *ACS Appl Mater Interfaces*. 2019; 11: 26529-58.
74. Hou CC, Tsai TL, Su WP, Hsieh HP, Yeh CS, Shieh DB, et al. Pronounced induction of endoplasmic reticulum stress and tumor suppression by surfactant-free poly(lactic-co-glycolic acid) nanoparticles via modulation of the PI3K signaling pathway. *Int J Nanomedicine*. 2013; 8: 2689-707.
75. Yasui H, Takeuchi R, Nagane M, Meike S, Nakamura Y, Yamamori T, et al. Radiosensitization of tumor cells through endoplasmic reticulum stress induced by PEGylated nanogel containing gold nanoparticles. *Cancer Lett*. 2014; 347: 151-8.
76. Huo LL, Chen R, Zhao L, Shi XF, Bai R, Long DX, et al. Silver nanoparticles activate endoplasmic reticulum stress signaling pathway in cell and mouse models: The role in toxicity evaluation. *Biomaterials*. 2015; 61: 307-15.
77. Simard JC, Durocher I, Girard D. Silver nanoparticles induce irremediable endoplasmic reticulum stress leading to unfolded protein response dependent apoptosis in breast cancer cells. *Apoptosis*. 2016; 21: 1279-90.
78. Solis GP, Bilousov O, Koval A, Luchtenborg AM, Lin C, Katanaev VL. Golgi-resident galphao promotes protrusive membrane dynamics. *Cell*. 2017; 170: 939-55.
79. Ma X, Sun J, Zhong L, Wang Y, Huang Q, Liu X, et al. Evaluation of turning-sized gold nanoparticles on cellular adhesion by Golgi disruption *in vitro* and *in vivo*. *Nano Lett*. 2019; 19: 8476-87.
80. Friedman JR, Nunnari J. Mitochondrial form and function. *Nature*. 2014; 505: 335-43.
81. Pan Y, Leifert A, Ruau D, Neuss S, Bornemann J, Schmid G, et al. Gold nanoparticles of diameter 1.4 nm trigger necrosis by oxidative stress and mitochondrial damage. *Small*. 2009; 5: 2067-76.
82. Gallud A, Kloditz K, Ytterberg J, Ostberg N, Katayama S, Skoog T, et al. Cationic gold nanoparticles elicit mitochondrial dysfunction: a multi-omics study. *Sci Rep*. 2019; 9: 4366.
83. Danial NN, Korsmeyer SJ. Cell death: critical control points. *Cell*. 2004; 116: 205-19.
84. Wang LM, Liu Y, Li W, Jiang XM, Ji YL, Wu XC, et al. Selective targeting of gold nanorods at the mitochondria of cancer cells: implications for cancer therapy. *Nano Lett*. 2011; 11: 772-80.
85. Zhang T, Wang L, Chen Q, Chen C. Cytotoxic potential of silver nanoparticles. *Yonsei Med J*. 2014; 55: 283-91.
86. Teodoro JS, Silva R, Varela AT, Duarte FV, Rolo AP, Hussain S, et al. Low-dose, subchronic exposure to silver nanoparticles causes mitochondrial alterations in Sprague-Dawley rats. *Nanomedicine*. 2016; 11: 1359-75.
87. Hussain S. Measurement of Nanoparticle-induced mitochondrial membrane potential alterations. *Methods Mol Biol*. 2019; 1894: 123-31.
88. Zhang Y, Yang JM. Altered energy metabolism in cancer: a unique opportunity for therapeutic intervention. *Cancer Biol Ther*. 2013; 14: 81-9.
89. Chen Y, Wang Z, Xu M, Wang X, Liu R, Liu Q, et al. Nanosilver incurs an adaptive shunt of energy metabolism mode to glycolysis in tumor and nontumor cells. *ACS Nano*. 2014; 8: 5813-25.
90. Sakiyama Y, Mazur A, Kapinos LE, Lim RYH. Spatiotemporal dynamics of the nuclear pore complex transport barrier resolved by high-speed atomic force microscopy. *Nat Nanotechnol*. 2016; 11: 719-723.
91. Huo SD, Jin SB, Ma XW, Xue XD, Yang KN, Kumar A, et al. Ultrasmall gold nanoparticles as carriers for nucleus-based gene therapy due to size-dependent nuclear entry. *ACS Nano*. 2014; 8: 5852-62.
92. Suh J, Wirtz D, Hanes J. Efficient active transport of gene nanocarriers to the cell nucleus. *Proc Natl Acad Sci U S A*. 2003; 100: 3878-82.
93. Kang B, Mackey MA, El-Sayed MA. Nuclear targeting of gold nanoparticles in cancer cells induces DNA damage, causing cytokinesis arrest and apoptosis. *J Am Chem Soc*. 2010; 132: 1517-1519.
94. Kodhiha M, Hutter E, Boridy S, Juhás M, Maysinger D, Stochaj U. Gold nanoparticles induce nuclear damage in breast cancer cells, which is further amplified by hyperthermia. *Cell Mol Life Sci*. 2014; 71: 4259-73.
95. Uz M, Bulmus V, Altinkaya SA. Effect of PEG grafting density and hydrodynamic volume on gold nanoparticle-cell interactions: an investigation on cell cycle, Apoptosis, and DNA Damage. *Langmuir*. 2016; 32: 5997-6009.
96. Annangi B, Bach J, Vales G, Rubio L, Marcos R, Hernandez A. Long-term exposures to low doses of cobalt nanoparticles induce cell transformation enhanced by oxidative damage. *Nanotoxicology*. 2015; 9: 138-47.
97. Zhao X, Takabayashi F, Ibuki Y. Coexposure to silver nanoparticles and ultraviolet A synergistically enhances the phosphorylation of histone H2AX. *J Photochem Photobiol B*. 2016; 162: 213-22.
98. Mytych J, Pacyk K, Pepek M, Zebrowski J, Lewinska A, Wnuk M. Nanoparticle-mediated decrease of lamin B1 pools promotes a TRF protein-based adaptive response in cultured cells. *Biomaterials*. 2015; 53: 107-16.
99. Sikora E, Arendt T, Bennett M, Narita M. Impact of cellular senescence signature on ageing research. *Ageing Res Rev*. 2011; 10: 146-52.
100. Sun H, Jia J, Jiang C, Zhai S. Gold Nanoparticle-induced cell death and potential applications in nanomedicine. *Int J Mol Sci*. 2018; 19: 754.
101. Magdolenova Z, Collins A, Kumar A, Dhawan A, Stone V, Dusinska M. Mechanisms of genotoxicity. A review of *in vitro* and *in vivo* studies with engineered nanoparticles. *Nanotoxicology*. 2014; 8: 233-78.
102. Kang B, Mackey MA, El-Sayed MA. Nuclear targeting of gold nanoparticles in cancer cells induces DNA damage, causing cytokinesis arrest and apoptosis. *J Am Chem Soc*. 2010; 132: 1517-9.
103. Li K, Zhao X, B KH, Du S, Chen Y. Nanoparticles inhibit DNA replication by binding to DNA: modeling and experimental validation. *ACS Nano*. 2013; 7: 9664-74.
104. Teow Y, Asharani PV, Hande MP, Valiyaveetil S. Health impact and safety of engineered nanomaterials. *Chem Commun*. 2011; 47: 7025-38.
105. Xia Q, Li H, Liu Y, Zhang S, Feng Q, Xiao K. The effect of particle size on the genotoxicity of gold nanoparticles. *J Biomed Mater Res A*. 2017; 105: 710-9.
106. Singh N, Nelson BC, Scanlan LD, Coskun E, Jaruga P, Doak SH. Exposure to engineered nanomaterials: impact on DNA repair pathways. *Int J Mol Sci*. 2017; 18: 1515.
107. Asharani PV, Hande MP, Valiyaveetil S. Anti-proliferative activity of silver nanoparticles. *BMC Cell Biol*. 2009; 10: 65.
108. Gonzalez L, Kirsch-Volders M. Biomonitoring of genotoxic effects for human exposure to nanomaterials: The challenge ahead. *Mutat Res Rev Mutat Res*. 2016; 768: 14-26.
109. Sabella S, Carney RP, Brunetti V, Malvindi MA, Al-Juffali N, Vecchio G, et al. A general mechanism for intracellular toxicity of metal-containing nanoparticles. *Nanoscale*. 2014; 6: 7052-61.
110. Wang Y, Liu Y, Luehmann H, Xia X, Brown P, Jarreau C, et al. Evaluating the pharmacokinetics and *in vivo* cancer targeting capability of Au nanocages by positron emission tomography imaging. *ACS Nano*. 2012; 6: 5880-8.
111. Park SM, Aalipour A, Vermesh O, Yu JH, Gambhir SS. Towards clinically translatable *in vivo* nanodiagnosics. *Nat Rev Mater*. 2017; 2: 17014.
112. Mao BH, Chen ZY, Wang YJ, Yan SJ. Silver nanoparticles have lethal and sublethal adverse effects on development and longevity by inducing ROS-mediated stress responses. *Sci Rep*. 2018; 8: 2445.
113. Moon J, Kwak JI, Kim SW, An YJ. Multigenerational effects of gold nanoparticles in *Caenorhabditis elegans*: continuous versus intermittent exposures. *Environ Pollut*. 2017; 220: 46-52.
114. Khlebtsov N, Dykman L. Biodistribution and toxicity of engineered gold nanoparticles: a review of *in vitro* and *in vivo* studies. *Chem Soc Rev*. 2011; 40: 1647-71.
115. Huang XL, Li LL, Liu TL, Hao NJ, Liu HY, Chen D, et al. The shape effect of mesoporous silica nanoparticles on biodistribution, clearance, and biocompatibility *in vivo*. *ACS Nano*. 2011; 5: 5390-9.
116. Muoth C, Aengenheister L, Kucki M, Wick P, Buerki-Thurnherr T. Nanoparticle transport across the placental barrier: pushing the field forward! *Nanomedicine*. 2016; 11: 941-57.
117. Xia G, Liu T, Wang Z, Hou Y, Dong L, Zhu J, et al. The effect of silver nanoparticles on zebrafish embryonic development and toxicology. *Artif Cells Nanomed Biotechnol*. 2016; 44: 1116-21.
118. Eleftheriadou M, Pyrgiotakis G, Demokritou P. Nanotechnology to the rescue: using nano-enabled approaches in microbiological food safety and quality. *Curr Opin Biotechnol*. 2017; 44: 87-93.
119. McClements DJ, Xiao H, Demokritou P. Physicochemical and colloidal aspects of food matrix effects on gastrointestinal fate of ingested inorganic nanoparticles. *Adv Colloid Interface Sci*. 2017; 246: 165-80.
120. Hillyer JF, Albrecht RM. Gastrointestinal persorption and tissue distribution of differently sized colloidal gold nanoparticles. *J Pharm Sci*. 2001; 90: 1927-36.
121. Tryggvason K, Wartiovaara J. How does the kidney filter plasma? *Physiology*. 2005; 20: 96-101.
122. Yu MX, Zheng J. Clearance pathways and tumor targeting of imaging nanoparticles. *ACS Nano*. 2015; 9: 6655-74.
123. Liu J, Yu M, Zhou C, Zheng JJMT. Renal clearable inorganic nanoparticles: a new frontier of bionanotechnology. *Mater Today*. 2013; 16: 477-86.
124. Sadauskas E, Jacobsen NR, Danscher G, Stoltenberg M, Vogel U, Larsen A, et al. Biodistribution of gold nanoparticles in mouse lung following intratracheal instillation. *Chem Cent J*. 2009; 3: 16.
125. Tsoi KM, MacParland SA, Ma XZ, Spetzler VN, Echeverri J, Ouyang B, et al. Mechanism of hard-nanomaterial clearance by the liver. *Nat Mater*. 2016; 15: 1212-21.
126. Sadauskas E, Danscher G, Stoltenberg M, Vogel U, Larsen A, Wallin H. Protracted elimination of gold nanoparticles from mouse liver. *Nanomedicine*. 2009; 5: 162-9.
127. Balasubramanian SK, Jittiwat J, Manikandan J, Ong CN, Yu LE, Ong WY. Biodistribution of gold nanoparticles and gene expression changes in the liver and spleen after intravenous administration in rats. *Biomaterials*. 2010; 31: 2034-42.
128. Elci SG, Jiang Y, Yan B, Kim ST, Saha K, Moyano DF, et al. Surface charge controls the suborgan biodistributions of gold nanoparticles. *ACS Nano*. 2016; 10: 5536-42.
129. Shahare B, Yashpal M, Singh G. Toxic effects of repeated oral exposure of silver nanoparticles on small intestine mucosa of mice. *Toxicol Mech Method*. 2013; 23: 161-7.

130. Yamagishi Y, Watari A, Hayata Y, Li XR, Kondoh M, Yoshioka Y, et al. Acute and chronic nephrotoxicity of platinum nanoparticles in mice. *Nanoscale Res Lett*. 2013; 8.
131. Cho WS, Kim S, Han BS, Son WC, Jeong J. Comparison of gene expression profiles in mice liver following intravenous injection of 4 and 100 nm-sized PEG-coated gold nanoparticles. *Toxicol Lett*. 2009; 191: 96-102.
132. Lopez-Chaves C, Soto-Alvaredo J, Montes-Bayon M, Bettmer J, Llopis J, Sanchez-Gonzalez C. Gold nanoparticles: distribution, bioaccumulation and toxicity. *In vitro* and *in vivo* studies. *Nanomedicine*. 2018; 14: 1-12.
133. Dragoni S, Franco G, Regoli M, Bracciali M, Morandi V, Sgaragli G, et al. Gold nanoparticles uptake and cytotoxicity assessed on rat liver precision-cut slices. *Toxicol Sci*. 2012; 128: 186-97.
134. Weldon BA, Faustman EM, Oberdorster G, Workman T, Griffith WC, Kneuer C, et al. Occupational exposure limit for silver nanoparticles: considerations on the derivation of a general health-based value. *Nanotoxicology*. 2016; 10: 945-56.
135. Bakand S, Hayes A, Dechsakulthorn F. Nanoparticles: a review of particle toxicology following inhalation exposure. *Inhal Toxicol*. 2012; 24: 125-35.
136. He XA, Zhang HF, Ma YH, Bai W, Zhang ZY, Lu K, et al. Lung deposition and extrapulmonary translocation of nano-ceria after intratracheal instillation. *Nanotechnology*. 2010; 21: 285103.
137. Geiser M. Update on macrophage clearance of inhaled micro- and nanoparticles. *J Aerosol Med Pulm Drug Deliv*. 2010; 23: 207-17.
138. Keramanizadeh A, Balharry D, Wallin H, Loft S, Moller P. Nanomaterial translocation-the biokinetics, tissue accumulation, toxicity and fate of materials in secondary organs-a review. *Crit Rev Toxicol*. 2015; 45: 837-72.
139. Carneiro MFH, Barbosa F. Gold nanoparticles: A critical review of therapeutic applications and toxicological aspects. *J Toxicol Environ Health B Crit Rev*. 2016; 19: 129-48.
140. Wiemann M, Vennemann A, Blaske F, Sperling M, Karst U. Silver nanoparticles in the lung: toxic effects and focal accumulation of silver in remote organs. *Nanomaterials(Basel)*. 2017; 7: 300.
141. Paranjpe M, Muller-Goymann CC. Nanoparticle-mediated pulmonary drug delivery: a review. *Int J Mol Sci*. 2014; 15: 5852-73.
142. Alessandrini F, Vennemann A, Gschwendtner S, Neumann AU, Rothballer M, Seher T, et al. Pro-inflammatory versus immunomodulatory effects of silver nanoparticles in the lung: the critical role of dose, size and surface modification. *Nanomaterials*. 2017; 7: 300.
143. Hong JS, Kim S, Lee SH, Jo E, Lee B, Yoon J, et al. Combined repeated-dose toxicity study of silver nanoparticles with the reproduction/developmental toxicity screening test. *Nanotoxicology*. 2014; 8: 349-62.
144. Gosens J, Keramanizadeh A, Jacobsen NR, Lenz AG, Bokkers B, de Jong WH, et al. Comparative hazard identification by a single doselung exposure of zinc oxide and silver nanomaterials in mice. *PLoS One*. 2015; 10: e0126934.
145. Chung KF, Seiffert J, Chen S, Theodorou IG, Goode AE, Leo BF, et al. Inactivation, clearance, and functional effects of lung-instilled short and long silver nanowires in rats. *ACS Nano*. 2017; 11: 2652-64.
146. Lee CY, Horng JL, Chen PY, Lin LY. Silver nanoparticle exposure impairs ion regulation in zebrafish embryos. *Aquat Toxicol*. 2019; 214: 105263.
147. Robertson S, Thomson AL, Carter R, Stott HR, Shaw CA, Hadoke PWF, et al. Pulmonary diesel particulate increases susceptibility to myocardial ischemia/reperfusion injury via activation of sensory TRPV1 and beta 1 adrenoceptors. *Part Fibre Toxicol*. 2014; 11: 12.
148. Grabrucker AM, Ruozzi B, Belletti D, Pederzoli F, Forni F, Vandelli MA, et al. Nanoparticle transport across the blood brain barrier. *Tissue Barriers*. 2016; 4: e1153568.
149. Daneman R, Prat A. The blood-brain barrier. *Cold Spring Harb Perspect Biol*. 2015; 7: a020412.
150. Furtado D, Bjornmalm M, Ayton S, Bush AI, Kempe K, Caruso F. Overcoming the blood-brain barrier: the role of nanomaterials in treating neurological diseases. *Adv Mater*. 2018; 30: e1801362.
151. Feng XL, Chen AJ, Zhang YL, Wang JF, Shao LQ, Wei LM. Central nervous system toxicity of metallic nanoparticles. *Int J Nanomedicine*. 2015; 10: 4321-40.
152. Wang Y, Xiong LL, Tang M. Toxicity of inhaled particulate matter on the central nervous system: neuroinflammation, neuropsychological effects and neurodegenerative disease. *J Appl Toxicol*. 2017; 37: 644-67.
153. Ge D, Du QQ, Ran BQ, Liu XY, Wang X, Ma XH, et al. The neurotoxicity induced by engineered nanomaterials. *Int J Nanomedicine*. 2019; 14: 4167-86.
154. Zhang LL, Bai R, Li B, Ge CC, Du JF, Liu Y, et al. Rutile TiO₂ particles exert size and surface coating dependent retention and lesions on the murine brain. *Toxicol Lett*. 2011; 207: 73-81.
155. Wu J, Wang C, Sun J, Xue Y. Neurotoxicity of silica nanoparticles: brain localization and dopaminergic neurons damage pathways. *ACS Nano*. 2011; 5: 4476-89.
156. Caudle WM. Occupational metal exposure and parkinsonism. *Adv Neurobiol*. 2017; 18: 143-58.
157. Chen YS, Hung YC, Lin LW, Liao I, Hong MY, Huang GS. Size-dependent impairment of cognition in mice caused by the injection of gold nanoparticles. *Nanotechnology*. 2010; 21: 485102.
158. Teleanu DM, Chircov C, Grumezescu AM, Teleanu RI. Neurotoxicity of nanomaterials: an up-to-date overview. *Nanomaterials*. 2019; 9: 96.
159. Flora S J S. The applications, neurotoxicity, and related mechanism of gold nanoparticles. In: Gao H and Jiang X, Ed. *Neurotoxicity of Nanomaterials and Nanomedicine*. Singapore. Academic Press, 2017: 179-203.
160. Rahman MF, Wang J, Patterson TA, Saini UT, Robinson BL, Newport GD, et al. Expression of genes related to oxidative stress in the mouse brain after exposure to silver-25 nanoparticles. *Toxicol Lett*. 2009; 187: 15-21.
161. Genter MB, Newman NC, Shertzer HG, Ali SF, Bolon B. Distribution and systemic effects of intranasally administered 25 nm silver nanoparticles in adult mice. *Toxicol Pathol*. 2012; 40: 1004-13.
162. El-Drieny E, Sarhan NI, Bayomy NA, Elmajied Elsherbeni SA, Momtaz R, Mohamed HE. Histological and immunohistochemical study of the effect of gold nanoparticles on the brain of adult male albino rat. *J Microsc Ultrastruct*. 2015; 3: 181-90.
163. Xin Q, Rotchell JM, Cheng JP, Yi J, Zhang Q. Silver nanoparticles affect the neural development of zebrafish embryos. *J Appl Toxicol*. 2015; 35: 1481-92.
164. Truong L, Saili KS, Miller JM, Hutchison JE, Tanguay RL. Persistent adult zebrafish behavioral deficits results from acute embryonic exposure to gold nanoparticles. *Comp Biochem Physiol C Toxicol Pharmacol*. 2012; 155: 269-74.
165. Chen YS, Hung YC, Lin LW, Liao I, Hong MY, Huang GS. Size-dependent impairment of cognition in mice caused by the injection of gold nanoparticles. *Nanotechnology*. 2010; 21: 485102.
166. Vivier E, Malissen B. Innate and adaptive immunity: specificities and signaling hierarchies revisited. *Nat Immunol*. 2005; 6: 17-21.
167. Keshavan S, Calligari P, Stella L, Fusco L, Delogu LG, Fadeel B. Nano-bio interactions: a neutrophil-centric view. *Cell Death Dis*. 2019; 10: 569.
168. Boraschi D, Italiani P, Palomba R, Decuzzi P, Duschl B, Fadeel B, et al. Nanoparticles and innate immunity: new perspectives on host defence. *Semin Immunol*. 2017; 34: 33-51.
169. Almeida JPM, Lin AY, Langsner RJ, Eckels P, Foster AE, Drezek RA. *In vivo* immune cell distribution of gold nanoparticles in naive and tumor bearing mice. *Small*. 2014; 10: 812-9.
170. Smith DM, Simon JK, Baker JR. Applications of nanotechnology for immunology. *Nat Rev Immunol*. 2013; 13: 592-605.
171. Mebius RE, Kraal G. Structure and function of the spleen. *Nat Rev Immunol*. 2005; 5: 606-16.
172. Lankveld DPK, Oomen AG, Krystek P, Neigh A, Troost-de Jong A, Noorlander CW, et al. The kinetics of the tissue distribution of silver nanoparticles of different sizes. *Biomaterials*. 2010; 31: 8350-61.
173. De Jong WH, Van Der Ven LTM, Sleijffers A, Park MVDZ, Jansen EHJM, Van Loveren H, et al. Systemic and immunotoxicity of silver nanoparticles in an intravenous 28 days repeated dose toxicity study in rats. *Biomaterials*. 2013; 34: 8333-43.
174. Kai MP, Brighton HE, Fromen CA, Shen TW, Luft JC, Luft YE, et al. Tumor presence induces global immune changes and enhances nanoparticle clearance. *ACS Nano*. 2016; 10: 861-70.
175. Ibrahim KE, Al-Mutary MG, Bakhiet AO, Khan HA. Histopathology of the liver, kidney, and spleen of mice exposed to gold nanoparticles. *Molecules*. 2018; 23: 1848.
176. Ji JH, Jung JH, Kim SS, Yoon JU, Park JD, Choi BS, et al. Twenty-eight-day inhalation toxicity study of silver nanoparticles in Sprague-Dawley rats. *Inhal Toxicol*. 2007; 19: 857-71.
177. Kim YS, Kim JS, Cho HS, Rha DS, Kim JM, Park JD, et al. Twenty-eight-day oral toxicity, genotoxicity, and gender-related tissue distribution of silver nanoparticles in Sprague-Dawley rats. *Inhal Toxicol*. 2008; 20: 575-83.
178. Westmeier D, Posselt G, Hahlbrock A, Bartfeld S, Vallet C, Abfalter C, et al. Nanoparticle binding attenuates the pathobiology of gastric cancer-associated *Helicobacter pylori*. *Nanoscale*. 2018; 10: 1453-63.
179. Talamini L, Violatto MB, Cai Q, Monopoli MP, Kantner K, Krpetic Z, et al. Influence of size and shape on the anatomical distribution of endotoxin-free gold nanoparticles. *ACS Nano*. 2017; 11: 5519-29.
180. Silva RM, Anderson DS, Peake J, Edwards PC, Patchin ES, Guo T, et al. Aerosolized silver nanoparticles in the rat lung and pulmonary responses over time. *Toxicol Pathol*. 2016; 44: 673-86.
181. Selim ME, Abd-Elhakim YM, Al-Ayadhi LY. Pancreatic response to gold nanoparticles includes decrease of oxidative stress and inflammation in autistic diabetic model. *Cell Physiol Biochem*. 2015; 35: 586-600.
182. Brohi RD, Wang L, Talpur HS, Wu D, Khan FA, Bhattarai D, et al. Toxicity of nanoparticles on the reproductive system in animal models: a review. *Front Pharmacol*. 2017; 8: 606.
183. Han JW, Jeong JK, Gurunathan S, Choi YJ, Das J, Kwon DN, et al. Male- and female-derived somatic and germ cell-specific toxicity of silver nanoparticles in mouse. *Nanotoxicology*. 2016; 10: 361-73.
184. Austin CA, Umbreit TH, Brown KM, Barber DS, Dair BJ, Francke-Carroll S, et al. Distribution of silver nanoparticles in pregnant mice and developing embryos. *Nanotoxicology*. 2012; 6: 912-22.
185. Zhao MZ, Li HY, Bu XL, Lei CN, Fang QJ, Hu ZY. Quantitative proteomic analysis of cellular resistance to the nanoparticle abraxane. *ACS Nano*. 2015; 9: 10099-112.
186. Li L, Li L, Zhou XJ, Yu Y, Li ZQ, Zuo DY, et al. Silver nanoparticles induce protective autophagy via Ca²⁺/CaMKK beta/AMPK/mTOR pathway in SH-SY5Y cells and rat brains. *Nanotoxicology*. 2019; 13: 369-91.
187. Dobrovolskaia MA, Mcneil SE. Immunological properties of engineered nanomaterials. *Nat Nanotechnol*. 2007; 2: 469-78.
188. Septiadi D, Crippa F, Moore TL, Rothen-Rutishauser B, Petri-Fink A. Nanoparticle-cell interaction: a cell mechanics perspective. *Adv Mater*. 2018; 30: e1704463.
189. Cordani M, Somoza A. Targeting autophagy using metallic nanoparticles: a promising strategy for cancer treatment. *Cell Mol Life Sci*. 2019; 76: 1215-42.

190. Her S, Cui L, Bristow RG, Allen C. Dual action enhancement of gold nanoparticle radiosensitization by pentamidine in triple negative breast cancer. *Radiat Res.* 2016; 185: 549-62.
191. Ma N, Wu FG, Zhang X, Jiang YW, Jia HR, Wang HY, et al. Shape-dependent radiosensitization effect of gold nanostructures in cancer radiotherapy: comparison of gold nanoparticles, nanopikes, and nanorods. *ACS Appl Mater Interfaces.* 2017; 9: 13037-48.
192. Ni KY, Lan GX, Chan C, Quigley B, Lu KD, Aung T, et al. Nanoscale metal-organic frameworks enhance radiotherapy to potentiate checkpoint blockade immunotherapy. *Nat Commun.* 2018; 9: 2351.
193. Du BJ, Yu MX, Zheng J. Transport and interactions of nanoparticles in the kidneys. *Nat Rev Mater.* 2018; 3: 358-74.
194. Zhao Z, Ukidve A, Krishnan V, Mitragotri S. Effect of physicochemical and surface properties on *in vivo* fate of drug nanocarriers. *Adv Drug Deliv Rev.* 2019; 143: 3-21.
195. Jiang X, Du B, Zheng J. Glutathione-mediated biotransformation in the liver modulates nanoparticle transport. *Nat Nanotechnol.* 2019; 14: 874-82.
196. Hu CMJ, Fang RH, Wang KC, Luk BT, Thamphiwatana S, Dehaini D, et al. Nanoparticle biointerfacing by platelet membrane cloaking. *Nature.* 2015; 526: 118-121.
197. Schottler S, Becker G, Winzen S, Steinbach T, Mohr K, Landfester K, et al. Protein adsorption is required for stealth effect of poly(ethylene glycol)- and poly(phosphoester)-coated nanocarriers. *Nat Nanotechnol.* 2016; 11: 372-7.