Hexosamine biosynthetic pathway promotes the antiviral activity of SAMHD1 by enhancing O-GlcNAc transferase-mediated protein

O-GIcNAcylation

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Supplementary Figure Legends

Figure S1. HBV infection enhances HBP activity and upregulates GLUT1 expression. (A) KEGG pathway enrichment analysis of metabolites detected using metabolomics assay in HepG2 cells infected with AdHBV1.3 or AdGFP for 72 h. (B-C) Fold change in UDP-GlcNAc (B) and glucose (C) levels in HBV1.3-infected HepG2 cells was determined using LC-MS/MS targeted metabolomics assay, n = 6. (D) Immunoblot of total O-GlcNAc from PHH cells treated for the indicated periods. (E-F) RT-qPCR quantification (E) and immunofluorescence staining (F) of GLUT1 in HepG2 cells, DAPI (blue) was used to counterstain nuclei; n = 3. Scale bar, 10 µm. (G-H) Immunoblots of OGA, OGT, p-GFPT1, and GFPT1 from HepG2, HepAD38 (G), and HepG2-NTCP (H) cells. Data are expressed as the mean \pm SD. *P* values were derived from unpaired, two-tailed Student's *t*-test in B-C and E (***P* < 0.01).

Figure S2. Role of O-GlcNAc modification in regulating HBV replication in HepG2 cells. (A-C) Immunoblot of total O-GlcNAc from HBV1.3-infected HepG2 cells treated with or without GLUT1 inhibitor WZB117 (50 μ M) (A), GFPT1 inhibitor DON (30 μ M) (B), or OGT inhibitor ST04 (100 μ M) (C) for 72 h. (D-F) Quantification of HBV core DNA levels in HBV1.3-infected HepG2 cells treated as indicated using qPCR, n = 3. (G) Immunoblot of total O-GlcNAc from HBV1.3-infected HepG2 cells treated with or without OGA inhibitor TMG (100 μ M) for 72 h. (H) Quantification of HBV core DNA levels in HBV1.3-infected HepG2 cells treated as in (G) using qPCR, n = 3. Data are expressed as the mean \pm SD. *P* values were derived from unpaired, two-tailed Student's *t*-test in D-F and H; (***P* < 0.01, ****P* < 0.001).

Figure S3. O-GlcNAc modification regulates HBV replication in

HepG2-NTCP cells and PHH cells. (A-E) Immunoblot of total O-GlcNAc from HBV-infected cells treated as indicated. (F-J) Quantification of HBV core DNA levels in HBV-infected cells treated as indicated using qPCR, n = 3. (K-L) Immunoblot of total O-GlcNAc from HBV-infected cells treated with or without TMG (100 μ M) for 72 h. (M-N) qPCR quantification of HBV core DNA levels in HBV-infected cells treated with or without TMG, n = 3. Data are expressed as the mean \pm SD. *P* values were derived from unpaired, two-tailed Student's *t*-test in F-J and M-N (***P* < 0.01, ****P* < 0.001).

Figure S4. Inhibition of protein O-GlcNAcylation enhances HBV replication. (A-B) Immunoblot of total O-GlcNAc from HBV1.3-infected HepG2 cells following shRNA-mediated GFPT1 or OGT knockdown. (C-D) Quantification of HBV core DNA levels in HBV1.3-infected HepG2 cells treated as above using qPCR, n = 3. (E) Immunoblot of total O-GlcNAc from HBV1.3-infected OGA knockdown HepG2 cells. (F) Quantification of HBV core DNA levels in HBV1.3-infected HepG2 cells treated as in (E) using qPCR, n =3. Data are expressed as the mean \pm SD. *P* values were derived from one-way

ANOVA in C-D and F; (***P* < 0.01, ****P* < 0.001).

Figure S5. HBV infection promotes SAMHD1 O-GlcNAcylation. (A)

IP-LC-MS/MS analysis of O-GlcNAc modified proteins. HepAD38 cell lysates were immunoprecipitated using an anti-O-GlcNAc antibody. (B) HEK293T cells, (C) Tetracycline-inducible HepAD38 cells, (D) HepG2-NTCP cells were transfected with the Flag-SAMHD1 construct or the control vector for 48 h, and treated with 100 µM TMG for 12 h (B), Cells lysates were purified using sWGA-conjugated agarose beads and probed with an anti-Flag or anti-O-GlcNAc antibody. (E) PHH cells were infected with HBV for 72 h. After cell lysis, O-GlcNAc-modified proteins were purified using the sWGA-conjugated agarose beads and probed with an anti-SAMHD1 antibody. The immunoprecipitated and input proteins were probed with an anti-O-GlcNAc or anti-SAMHD1 antibody. (F-G) Southern blot assay in stable HBV-expressing HepAD38 and SAMHD1-KO HepAD38 cells. rc DNA, relaxed circular DNA; ds DNA, double-stranded DNA; ss DNA, single-stranded DNA. (H) SAMHD1-KO HepG2-NTCP cells were transfected with either empty vector, Flag-tagged SAMHD1 WT or the S93A mutant, and treated with 100 µM PUGNAc for 12 h. Cells lysates were purified using sWGA-conjugated agarose beads and probed with an anti-Flag or anti-O-GlcNAc antibody.

Figure S6. SAMHD1 protein expression, purification, and dNTPase

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activity. (A-B) His-tagged pET28a-SAMHD1 constructs were expressed in Escherichia coli BL21 (DE3) cells and induced with 0.2 mM IPTG. Cell lysates were purified using Ni-NTA affinity column. The purified target protein was stained with Coomassie Brilliant Blue dye. Lane 1, molecular-mass markers; Lane 2, cell supernatant after ultrasonic crash; Lane 3, Ni–NTA affinity column flow through fraction; Lane 4-6, eluted target protein with 20 to 400 mM imidazole; Lane 7, eluted protein following hyperfiltration. (C-D) HPLC assay to determine SAMHD1 dNTPase activity. SAMHD1-WT (C) and SAMHD1-S93A (D) constructs were purified using Ni-NTA affinity and size-exclusion chromatography. Nucleotide hydrolysis reactions were performed in SAMHD1 buffer containing 500 µM dNTPs and 500 µM SAMHD1-WT, the peak height in Y-axis(mAU) refers to the maximum concentration of 'dG', X-axis(min) refers to retention time. The reactions were terminated using 10 mM EDTA. Protein was separated using an Amicon Ultra 0.5-mL 10-kDa filter and analyzed using a Venusil MP-C18 column on an HPLC system.























Name	Sequence (5'-3')		
Primer sequences for quantitative RT-PCR.			
HBV	Forward: TGCGGCGTTTTATCATATTCC		
	Reverse: ATACCTTGGTAGTCCAGAAGAACCA		
GLUT1	Forward: ACCGCAACGAGGAGAACCG		
	Reverse: CCACAAACAGCGACACGACAG		
OGA	Forward: AGCAGCCTTGAGTGGTGAGC		
	Reverse: TGGGGATTTTGATTCAGCTATG		
OGT	Forward: AGAGGCAGTTCGCTTGTATCG		
	Reverse: AGTAGGCATCAGCAAAGGTAGG		
GFPT1	Forward: CCAGTCCTGTCAATAGCCACC		
	Reverse: CACAAGTGCAAAAGCACCTTC		
SAMHD1	Forward: TGCCCGTGTCTGTGAAGTAG		
	Reverse: ATTGCTGTAGAAATGCGATA		
GAPDH	Forward: GAAATCCCATCACCATCTTCCAGG		
	Reverse: GAGCCCCAGCCTTCTCCATG		
Primer sequences for knockdown.			
shOGA	Forward1:		
	TGGACAATTCTTTATGACATTTCAAGAGAATGTCAT		
	AAAGAATTGTCCTTTTTTC		
	Reverse1:		
	TCGAGAAAAAAGGACAATTCTTTATGACATTCTCT		
	TGAAATGTCATAAAGAATTGTCCA		
	Forward2:		
	TGCCTTTGTACACTGCGGAATTCAAGAGTTCCGC		
	AGTGTACAAAGGCTTTTTTC		
	Reverse2:		

Table S1. Sequence of Oligonucleotides.

TCGAGAAAAAAGCCTTTGTACACTGCGGAATCTC

	TTGAATTCCGCAGTGTACAAAGGCA		
shOGT	Forward1: TGCATGTTATTTGAAAGCAATTCAAGAG		
	ATTGCTTTCAAATAACATGCTTTTTTC		
	Reverse1: TCGAGAAAAAAGCATGTTATTTGAAAGC		
	AATCTCTTGAATTGCTTTCAAATAACATGCA		
	Forward2: TGCCCTAAGTTTGAGTCCAATTCAAGAG		
	ATTGGACTCAAACTTAGGGCTTTTTTC		
	Reverse2: TCGAGAAAAAAGCCCTAAGTTTGAGTC		
	CAATCTCTTGAATTGGACTCAAACTTAGGGCA		
shGFTP1	Forward1: TGGAGAGAGTTATCCAACAATTCAAGAG		
	ATTGTTGGATAACTCTCTCCTTTTTC		
	Reverse1: TCGAGAAAAAAGGAGAGAGTTATCCAA		
	CAATCTCTTGAATTGTTGGATAACTCTCTCCA		
	Forward2: TGGAAGTACTGAGCATGGATTTCAAGAG		
	AATCCATGCTCAGTACTTCCTTTTTC		
	Reverse2: TCGAGAAAAAAGGAAGTACTGAGCATG		
	GATTCTCTTGAAATCCATGCTCAGTACTTCCA		
Primer sequences for molecular cloning.			
pSEB-Flag-OGT	Forward: ACGCGTCGACACCATGGCGTCTTCCGTGGGC		
	Reverse: CGCGGATCCTGCTGACTCAGTGACTTCAACAGG		
pBud-HA-OGT	Forward: ATAGCGGCCGCACCATGGCGTCTTCCGTGGGC		
	Reverse: CGCGGATCCTGCTGACTCAGTGACTTCAACAGG		
pET28a- SAMHD1	Forward: TGCGGATCCCAGCGAGCCGATTCCGAGC		
	Reverse: CCGCTCGAGTCACATTGGGTCATCTTTAA		
pSEB-FlagSAMHD1	Forward: TGAAGATCTACCATGGGCCAGCGAGCCGATTCCG		
	Reverse: CGCGTCGACCATTGGGTCATCTTTA AAAAGC		
Primer sequences for	or mutagenesis.		
pSEB-Flag-	Forward: ATCTTGGAGTAAGTGCCTTGGGGGGAGA		
SAMHD1 (S93A)	Reverse: TCTCCCCCAAGGCACTTACTCCAAGAT		

pSEB-Flag-	Forward:		
SAMHD1 (T592E)	GGATGGCGATGTTATAGCCCCACTCATAGAACCTCAAAAA		
	Reverse:		
	TTTTTGAGGTTCTATGAGTGGGGCTATAACATCGCCATCC		
pSEB-Flag-	Forward: TGAAGATCTACCATGGGCCAGCGAGCCGATTCCG		
SAMHD1 (1-150)	Reverse: CGCGTCGACCAGCTGTTTGATGTATCGAAGAC		
pSEB-Flag-	Forward: TGAAGATCTACCATG		
SAMHD1(151-328)	GGAGGTGGTTACTATGTTTTTCC		
	Reverse: CGCGTCGACATTATTTTGGATTCCAAGATGATG		
pSEB-Flag-	Forward: TGAAGATCTACCATG		
SAMHD1(329-626)	GGCTTTGATTACAAGCGCTTTATTA		
	Reverse: CGCGTCGACCATTGGGTCATCTTTAAAAAGC		
OGT, SAMHD1-targeted single gRNAs.			
SAMHD1- sgRNA	Forward: CACCGCTTAGTTATATCCAGCGAT		
	Reverse: AAACATCGCTGGATATAACTAAGC		
OGT-sgRNA	Forward: CACCGCACATCGAGAATATCAGGC		
	Reverse: AAACGCCTGATATTCTCGATGTGC		
lentiCRISPR-v2	Forward: GGTTTATTACAGGGACAGCAG		
	Reverse: ACACGACATCACTTTCCCAG		

Table S2. Reagent or resource.

REAGENT or RESOURCE Antibodies	SOURCE	IDENTIFIER
Mouse anti-O-GlcNAc	Abcam	Cat#ab2739
Rabbit anti-OGT	Abcam	Cat#ab177941
Rabbit anti-OGA/MGEA5	Abcam	Cat#ab124807
Rabbit anti-GFPT1/GFAT1	Abcam	Cat#ab125069
Mouse monoclonal anti-SAMHD1	Abcam	Cat#ab67820
Rabbit anti-GLUT1	Abcam	Cat#ab115730
Rabbit anti-K48	Abcam	Cat#ab140601
Goat anti-Rabbit IgG	Abcam	Cat# ab6721
Goat anti-Mouse IgG	Abcam	Cat# ab6789
Mouse monoclonal anti-FLAG M2-peroxidase (HRP)	Sigma-Aldrich	Cat#A8592
ANTI-FLAG M2 Affinity Gel	Sigma-Aldrich	Cat#A2220
Mouse anti-GAPDH	Beyotime	Cat#AF0006
Mouse monoclonal anti-HA	Thermo	Cat#26183-HRP
Mouse anti-β-actin	ZSGB-BIO	Cat#TA-09
Rabbit polyclonal anti-HBcAg	Dako	Cat#B0586
Rabbit anti-p-GFPT1/GFAT1(Ser243)	Immuno-Biological Laboratories	Cat# JP28123
Goat anti-rabbit IgG/TRITC, secondary	ZSGB-BIO	Cat#ZF-0316
Goat anti-mouse IgG/TRITC, secondary	ZSGB-BIO	Cat#ZF-0313
Goat anti-rabbit IgG/FITC, secondary	ZSGB-BIO	Cat#ZF-0311
Goat anti-mouse IgG/ FITC, secondary	ZSGB-BIO	Cat#ZF-0312
Bacterial and Virus Strains		
E.coli BL21	Lab stock	
DH10B Chemically Competent E. coli	Lab stock	

pseudotyped HIV-1 single-round luciferase virus (HIV-LUC-G)	He et al., 1995	
Lenticrispr-SAMHD1	This study	
Lenticrispr-OGT	This study	
DH10B Chemically Competent E. coli	Lab stock	
AdHBV1.3	Lab stock	
Biological Samples		
Human liver tissue with chronic hepatitis B	The Second Affiliated Hospital, Chongqing Medical University.China	
Human serum with chronic hepatitis B	The Second Affiliated Hospital, Chongqing Medical University.China	
Chemicals, Peptides, and Recombinant Proteins		
Polybrene	Solarbio	Cat#H8761
PNGase F	NEB	Cat#P0704S
Fetal Bovine Serum	Gibco	Cat#A3160801
Trypsin	Gibco	Cat#12605010
RPMI1640	Gibco	Cat#11875500BT
Opti-MEM™ I Reduced Serum Medium	Gibco	Cat#31985070
DMEM/F-12	Gibco	Cat#11330500BT
DMEM	Gibco	Cat#11995500BT
MEM	Hyclone	Cat#SH30024.01
Penicillin-Streptomycin Solution	HyClone	Cat#SV30010
Lipofectamine 2000	Invitrogen	Cat#11668019
DMSO	Sigma	Cat#D2660
Phorbol 12-myristate 13-acetate (PMA)	Sigma-Aldrich	Cat#P8139
PUGNAc	Sigma-Aldrich	Cat#A7229
Cycloheximide	MCE	Cat#HY-12320

ST045849	TimTec	Cat#MFCD03308174
Thiamet G	Selleck	Cat#S7213
WZB117	Selleck	Cat#S7927
UDP-GlcNAc	BBL Life Sciences	Cat#7512-17-6
6-diazo-5-oxo-L-norleucine(Don)	Sigma-Aldrich	Cat#D2141
SuperSignal West Pico PLUS chemiluminescent Substrates	Thermo Fisher Scientific	Cat#34577
DAPI	Roche	Cat# 10236276001
Trizol	Invitrogen	Cat# 15596026
SYBR Green PCR Master Mix	BioRad	Cat#172-5121
Critical Commercial Assays		
PrimeScript RT reagent Kit	Takara	Cat# RR047A
TIANampVirus DNA/RNA Kit	TianGen	Cat#DP315
HBsAg ELISA KIT	Kehua	Cat#S10910113
HBeAg ELISA KIT	Kehua	Cat#20163400144
DIG high prime DNA labeling and detection starter kit	Roche	Cat#11585614910
Dual-Glo® Luciferase Assay System	Promega	Cat#E2920
ALT ELISA kit	Mengbio	Cat#MBE1741
Elivision plus Polyer HRP (Mouse/Rabbit) IHC Kit	Maixin	Cat# KIT-9901
DAB kit	ZhongshanGolden Bridge	Cat# ZLI-9019
BCA protein assayKit	BEIJING DINGGUO	Cat# BCA02
Cell lysis buffer for Western and IP	Beyotime	Cat# P0013
Deposited Data		
Experimental Models: Cell Lines		
HEK293T	ATCC	
HepG2	ATCC	

HepAD38		Lab stock	
HepG2-NTCP		Lab stock	
THP-1		ATCC	
Experimental Models: Organisms	/Strains		
Mouse: C57BL/6		Experimental Animal Center of Chongqing Medical University	
HBV-transgenic (HBV-Tg) mice		Xiamen University	
Recombinant DNA			
LentiCrisprv2	Lab stock		
pAdTrack-TO4	Dr. T-C He, Un	iversity of Chicago, USA	
pBUD-HA-OGT	This paper		
pSEB-Flag-OGT	This paper		
pcDNA3-HA-SAMHD1	This paper		
pcDNA3-HA-ubiquitin	Kamitani et al.,	1997	HA-Ubiquitin, Addgene plasmid, # 18712
pSEB-Flag-SAMHD1(WT)	This paper		N/A
pSEB-Flag-SAMHD1(S93A)	This paper		N/A
pSEB-Flag-SAMHD1(T592E)	This paper		N/A
pSEB-Flag-SAMHD1(1-150)	This paper		N/A
pSEB-Flag-SAMHD1(151-328)	This paper		N/A
pSEB-Flag-SAMHD1(329-626)	This paper		N/A
Software and Algorithms			
GraphPad Prism		GraphPad Software	https://www.graphpad. com.
CorelDRAW X7		Corel Corporation	https://www.coreldraw. com/

ImageJ	Schneider et al., 2012	https://imagej.nih.gov/ij /
Image pro plus	http://www.mediacy.com/i mageproplus	N/A