1 Supplementary Figures







Figure S2. EE exposure induces chromatin modifications in the peri-infarct area. (A) Experimental design for (C). (B) Representative immunoblots and bar graph showing the time course of acetyl-H4 levels in the peri-infarct cortex. Mice were exposed to SH or EE 5 days after stroke. One-way ANOVA followed by post hoc Scheffe test, $F_{(4,25)} = 7.81$, *p = 0.048, #p = 0.025. (C) Representative immunofluorescence images showing acetyl-H4 in the peri-infarct cortex. Scale bar, 20 µm. EE, environmental enrichment; SH, standard housing.



Figure S3. EE exposure primes a transcriptional program for the expression of 33 BDNF, which regulates trafficking of GAT-1. 34

(A) Bar graph showing changes in histone acetylation in the promoter regions of 35 BDNF genes in peri-infarct tissue. Fragmented chromatin was immunoprecipitated 36 with antibody recognizing acetyl-H4 and quantified with real-time polymerase chain 37 reaction. One-way ANOVA followed by post hoc Scheffe test. For Bdnf-p1, $F_{(2,9)} =$ 38 9.44, *p = 0.047, #p = 0.007; for *Bdnf-p4*, $F_{(2,9)} = 7.01$, #p = 0.015. (B) Confirmation 39 of the membrane protein fractions extraction method. Representative immunoblots 40 showing that β-actin was exclusively detected in cytosol fractions, while E-cadherin 41 was exclusively detected in membrane fractions. (C) Representative immunoblots and 42

43	bar graph showing GAT-1 content in peri-infarct cortex. Two-tailed <i>t</i> -test, $F_{(1,10)} =$
44	2.82. (D) Representative immunoblots and bar graph showing GAT-1 content in
45	membrane fraction. Two-tailed <i>t</i> -test, $F_{(1,10)} = 25.96$, *** $p < 0.001$. (E) Representative
46	immunoblots and bar graph showing GAT-1 content in cytosol fractions. Two-tailed
47	<i>t</i> -test, $F_{(1,10)} = 20.58$, ^{**} $p = 0.001$. BDNF, brain-derived neurotrophic factor.
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66 Figure S4. TSA reverses OGD-induced internalization of GAT-1 in
 67 hESCs-derived neurons.

(A) Representative fluorescence image and bar graph showing the percentage of
neurons and astrocytes in hESCs-derived cells. Scale bar, 40 μm. (B) Representative
whole-cell patch clamp trace illustrating the number of APs in hESCs-derived neurons
evoked by current injection. (C) Representative voltage clamp (-70 mV) traces
showing hESCs-derived neurons displaying spontaneous postsynaptic currents. (D)
hESCs-derived neurons were exposed to 3 h of OGD. TSA (0.5 μM) was applied for
24 h, starting 24 h after OGD. Representative fluorescence images showing Tuj-1 and

75	GAT-1 in hESCs-derived neurons at 48 h after OGD. Scale bar, 20 µm. AP, action
76	potential; hESC, human embryonic stem cell; OGD, oxygen glucose deprivation; TSA,
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99 Figure S5. OGD and TSA treatment have no effect on the expression of GAT-1 in 100 astrocytes.

(A) Representative fluorescence image showing GFAP (an astrocyte marker) and GAT-1 in the peri-infarct area. Arrows indicate GAT-1 co-localized with astrocytes. Scale bar, 20 µm. (B) Cultured astrocytes were exposed to 3 h of OGD. TSA (0.5 µM) was applied for 24, 48, and 72 h, starting 24 h after OGD. Immunoblots showing the time course of GAT-1 levels in cultured astrocytes after OGD exposure and TSA treatment. Bar graph showing the time course of GAT-1 levels in astrocytes after OGD exposure and TSA treatment. One-way ANOVA followed by post hoc Scheffe test, $F_{(4,25)} = 0.49$. OGD, oxygen glucose deprivation; TSA, trichostatin A.





116 Figure S6. HDAC2 but not HDAC1 negatively regulates GAT-1 expression.

(A) Representative immunoblots and bar graph showing HDAC1 content in the 117 peri-infarct cortex 5 days after stroke. One-way ANOVA followed by post hoc Scheffe 118 test, $F_{(2,12)} = 10.18$, $^{\#}p = 0.008$. (B) Representative immunoblots and bar graph 119 120 showing GAT-1 content in the peri-infarct cortex 5 days after stroke. One-way ANOVA followed by post hoc Scheffe test, $F_{(2,12)} = 1$. (C) Representative 121 immunoblots and bar graph showing HDAC2 content in the peri-infarct cortex 5 days 122 after stroke. One-way ANOVA followed by post hoc Scheffe test, $F_{(2,12)} = 52.41$, ***p123 < 0.001, ^{###}p < 0.001. (D) Representative immunoblots and bar graph showing GAT-1 124 content in the peri-infarct cortex 5 days after stroke. One-way ANOVA followed by 125 post hoc Scheffe test, $F_{(2,12)} = 15.75$, **p = 0.001, ##p = 0.003. HDAC, histone 126 deacetylase. 127

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Figure S7. GAT-1 is critical for the TSA-mediated increase in dendritic spine 132 density and synaptogenesis after hypoxia. 133

(A) Primary cultured mouse cortical neurons were exposed to 3 h of OGD. TSA (0.5 134 μ M) or a mixture of TSA (0.5 μ M) and NO-711 (10 μ M) was applied for 24 h, starting 135 24 h after OGD. Representative fluorescence image showing synapsin and PSD-95 in 136 primary cultured mouse cortical neurons after OGD. Each lower panel is a magnified 137

selected area from the upper iamge showing dendritic spines. Arrows indicate 138 synapsin / PSD-95 double-positive puncta. Upper scale bars, 20 µm. Lower scale bars, 139 5 µm. (B) Bar graph showing the average density of synapsin puncta in dendritic 140 segments of primary cortical neurons of the indicated groups. One-way ANOVA 141 followed by post hoc Scheffe test, $F_{(3,116)} = 46.37$, *** p < 0.001, ### p < 0.001, && p < 0.001, *** p < 0.001, 142 0.001. n indicates the number of dendrites from 3 independent experiments. (C) Bar 143 graph showing the average density of synapsin puncta in dendritic segments of 144 primary cortical neurons with the indicated treatments. One-way ANOVA followed by 145 post hoc Scheffe test, $F_{(3,116)} = 40.74$, *** p < 0.001, ###p < 0.001, && p < 0.001. (D) Bar 146 graph showing the average density of synapsin / PSD-95 colocalized puncta in 147 dendritic segments of primary cortical neurons with the indicated treatments. 148 One-way ANOVA followed by post hoc Scheffe test, $F_{(3,116)} = 82.06$, ***p < 0.001, ###p149 < 0.001, $^{\&\&\&}p < 0.001$. (E) Representative images of primary cultured cortical 150 neurons infected with LV-GFP showing the morphology at 48 h after OGD. Scale bar, 151 5 µm. (F) Bar graph showing the ratio of mushroom-like spines versus thin spines in 152 the indicated groups. One-way ANOVA followed by post hoc Scheffe test, $F_{(3,116)} =$ 153 170.22, *** p < 0.001, ### p < 0.001, && p < 0.001. n indicates the number of dendrites 154 from 3 independent experiments. OGD, oxygen glucose deprivation; TSA, trichostatin 155 156 A. 157



161 **Figure S8. Generation of GAT-1**^{*flox/flox*} **mice.**

(A) Schematic illustration of the Slc6al gene, targeting vector and the Slc6al floxed 162 locus. The Slc6al gene has 16 exons, and exon 6 was floxed to generate GAT-1^{flox/flox} 163 mice. (B) ES clones with 5' arm homologous recombination were confirmed by PCR 164 verification with a 4.3 kb band, while wild type generated a 7.6 kb band. (C) ES 165 clones with 3' arm homologous recombination were confirmed by PCR verification 166 with a 3.9 kb band, while wild type generated a 7.9 kb band. (D) Genotyping of 167 GAT-1^{flox/flox} mice. DNA was isolated for PCR with two primer pairs. The GAT-1 168 primers generated a 304 bp product in the loxP-flanked allele or a 246 bp product in 169 the wild-type allele. M, marker; PCR, polymerase chain reaction. 170





173 Figure S9. Photothrombotic stroke model induces long-lasting motor cortex
174 injury in adult mice.

(A) Time course of representative T2-TurboRARE MRI of photothrombotic stroke
mice. Arrows indicate the injured areas. (B) Representative Nissl-stained sections on
day 33 after stroke. Arrows indicate the injured areas. (C) Bar graph showing stroke

- 178 volume.





191 **(A)** GAT-3 level in the peri-infarct cortex 8 days after AAV-CAG-3Flag-T2A-mcherry 192 (AAV-null) or AAV-CAG-GAT-3-3Flag-T2A-mcherry (AAV-GAT-3) infection. 193 One-way ANOVA followed by post hoc Scheffe test, $F_{(2,12)} = 7.87$, $^{\#}p = 0.023$. **(B)** 194 Representative images showing the dendritic spines of peri-infarct neurons infected

195	with AAV-null, AAV-GAT-1 or AAV-GAT-3. For spine density analysis, only apical
196	dendrites of layer 5 pyramidal neurons in the peri-infarct cortex (within 400 μm from
197	the infarct) were included. Scale bar, 5 μ m. (C) Bar graph showing spine densities of
198	the indicated group. One-way ANOVA followed by post hoc Scheffe test, $F_{(3,113)} =$
199	83.71, $p^{***} < 0.001$, $p^{###} < 0.001$, $p^{***} < 0.001$. n indicates the number of dendrites
200	from 3 independent experiments. (D) Representative traces showing tonic inhibitory
201	currents recorded from layer 5 pyramidal neurons in AAV-null- or
202	AAV-GAT-3-infected GAT-1 knockdown mice. (E) Bar graph showing tonic current
203	density from layer 5 pyramidal neurons in AAV-null- or AAV-GAT-3-infected GAT-1
204	knockdown mice. Two-tailed <i>t</i> -test, $F_{(1,21)} = 0.02$. (F) Left, foot faults of the left
205	forelimb in the grid-walking task. Middle, foot faults of the left hindlimb in the
206	grid-walking task. Right, forelimb symmetry in the cylinder task. Two-way
207	repeated-measures ANOVA followed by post hoc Bonferroni test. Left, $F_{(1,22)} = 0.001$.
208	Middle, $F_{(1,22)} = 0.353$. Right, $F_{(1,22)} = 1.537$. AAV, adeno-associated virus; BMI,
209	bicuculline methiodide.