SUPPLEMENTARY MATERIALS

Wei-Jian Huang^{1,2,*}, Xu Zhou^{1,*}, Gong-Bo Fu^{1,3,*}, Min Ding^{4,*}, Hong-Ping Wu^{1,*}, Min Zeng¹, Hong-Dan Zhang⁵, Ling-Yan Xu⁶, Yi Gao^{2,#}, Hong-Yang Wang^{1,#}, and He-Xin

Yan^{1,2,5,7#}

Author affiliations

- International Cooperation Laboratory on Signal Transduction, Eastern Hepatobiliary Surgery Hospital, Second Military Medical University, Shanghai, China;
- Department of Hepatobiliary Surgery II, Guangdong Provincial Research Center for Artificial Organ and Tissue Engineering, Zhujiang Hospital, Southern Medical University ;
- Department of Medical Oncology, Jinling Hospital, First School of Clinical Medicine, Southern Medical University
- Department of Interventional Oncology, Renji Hospital, Jiaotong University School of Medicine, Shanghai, China
- 5. Shanghai Celliver Biotechnology Co. Ltd., Shanghai, China
- Department of Pharmacy, Shanghai Jiao Tong University Affiliated Sixth People's Hospital, East Campus, Shanghai, China;
- Shanghai Cancer Institute, Renji Hospital, Shanghai Jiaotong University School of Medicine, Shanghai, China.
- * These authors contributed equally to this work.

Supplementary methods

Western blot

The protein extracts were submitted to western blotting standard protocol. Briefly, livers tissues were dissected with 300µl of ice cold lysis buffer rapidly in tube and homogenized with an electric homogenizer, then centrifuged for 20 min at 12,000 rpm at 4°C in a micro centrifuge. Protein concentrations were measured using a PierceTM BCA Protein Assay Kit (Both from Thermo scientific). Then proteins were subjected to electrophoresis on 8–10% Bis-Tris protein gels and transferred to nitrocellulose membranes (GE healthcare), which were incubated with the primary antibodies, followed by a fluorescently conjugated secondary antibody. The fluorescence density on nitrocellulose membranes was measured on Odyssey CLx Western Blot Detection System (LI-COR Biosciences).

Quantitative real-time PCR (qPCR)

Total RNA of cells was extracted using TRIZOL reagent (Invitrogen) according to the manufacturer's protocols. Real-time PCR analyses were performed using a LightCycler® 96 Real-Time PCR System (Roche) and SYBR Green PCR kit (Roche). Gene transcription was evaluated using the $\Delta\Delta$ Ct method normalized to the housekeeping gene actin beta (ACTB). Primer sequences are respectively listed in Table S3. Α



HACY-CD24 vs CCL4-CD24: pvalue<0.05|log2FC|>1

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Supplementary Figure 1

(A, B) mRNA sequencing was performed to compare global gene expression profiles. Figure depicts a comparison of up- and down-regulated genes (pv<0.05 and |log2FC|>1) between HACY-CD24 cells and CCL4-CD24 cells. Heat maps for the expression of genes related to hepatic function in CD24+ progenitor cells. Α





Supplementary Figure 2

(A) The protein levels of α -SMA from models of liver fibrosis with or without HACY treatment at 40 days were detected by western blotting. GAPDH was used as an internal control.

(B) Quantification of the α-SMA levels shown in A, as determined by Image J software
(C) Growth curves of mHSCs with HGF, Y-27632, A-83-01, CHIR99021, or HACY, at different time points, as determined by CCK-8 assays. Ctrl represents a blank control.
(D) α-SMA expression in primary mHSCs with HGF, Y-27632, A-83-01, CHIR99021, or HACY on day 3 was determined by qRT-PCR analysis.

For B, D, the results are shown as mean \pm s.d. of three independent experiments, **p < 0.01, ***p < 0.001, ns represents no significance

Supplementary table1: Age and gender of the 7 patients with clinically diagnosed liver fibrosis.

Patients Number	Age	Gender
1	78	male
2	51	male
3	75	male
4	56	male
5	50	male
6	57	male
7	67	male

Supplementary tal	ble 2: Age	and gende	r of the	5 patients	with clinically	diagnosed
hepatic hemangion	ma.					

Patients Number	Age	Gender
1	49	male
2	50	male
3	63	male
4	61	male
5	32	female

Supplementary table 3: Primer list.

Gene	Forward sequence 5'->3'	Reverse sequence 5' ->3'
CD24	GTIGCACCGTITCCCGGTAA	CCCCTCTGGTGGTAGCGTTA
СК19	GTTCAGTACGCATTGGGTCAG	GAGGACGAGGTCACGAAGC
Hnf4a	ATGCGACTCTCTAAAACCCTTG	ACCTICAGATGGGGACGTGT
G6pc	CGACTCGCTATCTCCAAGTGA	GGGCGTTGTCCAAACAGAAT
Tat	TGCTGGATGTTCGCGTCAATA	CGGCTICACCTICATGTIGTC
Ttr	CTGCTGTAGACGTGGCTGTAA	CTICCAGTACGATTIGGTGTCC
Cps1	TACCCGGAAGCACTTACTGAT	GCCAGCCAGTGGTTATAGTCATT
Cyp1a2	AGTACATCTCCTTAGCCCCAG	GGGTCCGGGTGGATTCTTC
Cyp3a11	CCTGGGTGCTCCTAGCAATC	CAAGGAGAGGCGTTTGACCA
CD133	ACTGGGGCTGTGTGGAAAG	GC ATTGAAGGTATC TTGGGTC TC
Epcam	C TGGC GTC TAAATGC TTGGC	CCTIGTCGGTTCTTCGGACTC
Sox9	AGTACCCGCATCTGCACAAC	ACGAAGGGTCTCTTCTCGCT
Alb	CAAGAGTGAGATCGCCCATCG	TTACTTCCTGCACTAATTTGGCA
Actin	ATGCCACAGGATICCATACCCAAG	CTCTAGACTTCGAGCAGGAGATGG
GAPDH	GGCATGGACTGTGGTCATGAG	TGCACCACCAACTGCTIAGC

α-SMA	CCCAGACATCAGGGAGTAATGG	TCTATCGGATACTICAGCGTCA
COLLa1	ATCGGTCATGCTCTCTCCAAACCA	ACTGCAACATGGAGACAGGTCAGA
Desmin	GTTTCAGACTTGACTCAGGCAG	TCTCGCAGGTGTAGGACTGG
Nestin	C C CTGAAGTC GAGGAGCTG	CTGCTGCACCTCTAAGCGA
GFAP	TCTCGAATGACTCCTCCACTC	AAGCTCCGCCTGGTAGACAT
АСТВ	GCCTCGCTGTCCACCTTCC	TGCTGTCACCTTCACCGTTCC
H-CD24	CTCCTACCCACCGCAGATTTATTC	AGAGTGAGACCACGAAGAGAC
H-Alb	GAGACCAGAGGTTGATGTGATG	AGTTCCGGGGCATAAAAGTAAG
H-α-SMA	AAAAGACAGCTACGTGGGTGA	GCCATGTICTATCGGGTACTIC