Supplementary tables and figures

Topical application of TAK1 inhibitor encapsulated by gelatin particle alleviates corneal neovascularization

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Supplementary tables

Table S1 Key Resources table

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Antibodies		•
IrBa (I 3545) mouse mAb	Cell Signaling	4814
	Technology	1011
Phospho-NFkB p65 (Ser536) (93H1) rabbit	Cell Signaling	3033
mAb	Technology	
NFκB p65 (C22B4) rabbit mAb	Cell Signaling	4764
Dhamba $p44/42$ MADK (Erk1/2)	Call Signaling	
(Thr 202/Tyr 204) rabbit nAb	Technology	9101
	Cell Signaling	
p44/42 MAPK (Erk1/2) (137F5) rabbit mAb	Technology	4695
Phospho-SAPK/JNK (Thr183/Tyr185) (G9)	Cell Signaling	0055
mouse mAb	Technology	9255
SADV/INIV white Ah	Cell Signaling	0252
SAPK/JNK raddil pAd	Technology	9232
Phospho-p38 MAPK (Thr180/Tyr182)	Cell Signaling	0211
rabbit pAb	Technology	9211
$n38 MAPK (D13E1) XP^{\mathbb{R}}$ rabbit mAb	Cell Signaling	8690
	Technology	0070
Actin (clone C4) mouse mAb	Merck Millipore	MAB1501
Goat anti-mouse IgG HRP-conjugated	Life Technologies	31430
secondary Ab	Australia	51150
Goat anti-rabbit IgG HRP-conjugated	Life Technologies	656120
secondary Ab	Australia	
Chemicals	[1 //
5Z-7-Oxozeaenol	Tocris Bioscience	3604/1
Recombinant Human TNFα protein	Gibco TM	PHC3015
Recombinant Human VEGF165 Protein	R&D Systems	293-VE-010
Type A gelatin (bloom 175)	Sigma-Aldrich	G2625
DMSO	JT Baker	9224-01
Glutaraldehyde	Sigma-Aldrich	G6257
Acetone	Seedchem Company	AC0061
Uranium acetate	Thermo Fisher Scientific	541-09-3
Acetonitrile	Aencore Chemical	AE0627
5-Carboxytetramethylrhodamine,	Thermo Fisher Scientific	C2211
succinimidyl ester $(5-1 \text{AMRA})$	X 7' 1	
	Virbac	n/a
Kompun	Bayer	n/a
Alcaine	Alcon	n/a 1500
Silver Nitrate Applicators 6 inch	Graico	1090
	Generon	40043
Critical Commercial Assays		G25006
CYQUANT [®] NF cell proliferation assay kit	Lite Technologies	C35006

	Australia	
Quick-RNA MiniPrep kit	Zymo Research	R1055
High-capacity cDNA reverse transcription Kit	Life Technologies Australia	4368814
TaqMan [™] Fast Advanced Master Mix	Life Technologies Australia	4444553
Pierce [™] BCA assay kit	Life Technologies Australia	23227
Amersham ECL Prime Western Blotting Detection Kit	GE Healthcare	RPN2232
MycoAlert TM Mycoplasma Detection Kit	Lonza	LT07
Experimental Models: Cell Lines		
Telomerase-immortalized human microvascular endothelium cell (TIME)* *Cell was derived from a primary culture of neonatal foreskin microvascular endothelial cells of the dermis.	ATCC	CRL-4025™
Human Umbilical Vein Endothelial Cell (HUVEC)* *Cells was derived from the endothelium of veins from the umbilical cord.	Life Technologies Australia	C0035C
Experimental Models: Organisms/Strains		
C57BL/6 mouse	Taipei Medical University	LAC-2015-0328 and LAC-2019-063
qPCR Probes		
qPCR Probes Human VEGFA TaqMan probe	Applied Biosystems	Hs00900055_m1
qPCR ProbesHuman VEGFA TaqMan probeHuman ICAM1 TaqMan probe	Applied Biosystems Applied Biosystems	Hs00900055_m1 Hs00164932_m1
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	Australia	
Protease inhibitor cocktail	Roche Diagnostics	11697498001
Bolt [™] 4-12% Bis-Tris Plus Gel	Invitrogen	NW04120BOX
Immuno-Blot® PVDF Membrane	Bio-Rad Laboratories	1620177
EBM [™] Plus Basal Medium supplied with EGM [™] Plus SingleQuots [™] supplements	Lonza	CC-5035
Modified Davidson's fluid	CIS-Bio, Taiwan	D476
10 % neutral buffered formalin solution	CIS-Bio, Taiwan	D468
FSC22 Frozen section media	Leica	3801480
Fluoromount TM aqueous mounting medium	Sigma-Aldrich	F-4680
Incucyte® 96-well WoundMaker Tool	Incucyte	4563
TrypLE Express Enzyme (1X)	Life Technologies Australia	12605028
Spectra/Por, Float-A-Lyzer G2 Dialysis Device (MWCO 20 kDa)	Sigma-Aldrich	Z726710-12EA

	Dve	Average ra	Average rate of change		Difference to free dye			Difference to first 10 mins		
	Dyc	%/minute	(95% CI)	%/minute	(95% CI)	p-value*	%/minute	(95% CI)	p-value*	
0-10 mins	Free dye	-5.4	(-6.2,-4.7)							
	TAMRA-GNPs	-4.5	(-5.2,-3.7)	0.9	(-0.1,2.0)	0.083				
	TAMRA-GNPs-Oxo	-0.8	(-1.6,-0.1)	4.6	(3.5,5.7)	< 0.001				
>10-60 mins	Free dye	-0.7	(-1.1,-0.4)				4.7	(3.9,5.5)	< 0.001	
	TAMRA-GNPs	0.0	(-0.3,0.3)	0.7	(0.3, 1.2)	0.002	4.5	(3.7,5.3)	< 0.001	
	TAMRA-GNPs-Oxo	-0.5	(-0.9,-0.2)	0.2	(-0.2,0.7)	0.380	0.3	(-0.5,1.1)	0.505	
* Derived from	n a piecewise linear mixe	d-effects mod	el with rando	m intercepts a	nd slopes					

Table S2 Estimated rate of change in fluorescent intensity from a piecewise linear mixed-effects model

Table S3 Observed values at specific time points from Figure 4F

Time	Duo	Fluorescent intensity (%)	Difference free dye (%)	
i mie Dye -		Average (SD)	Average	p-value*
10 mins	Free dye	40.4 (13.1)		
	TAMRA-GNPs	57.1 (9.8)	16.7 (9.5)	0.153
	TAMRA-GNPs-Oxo	95.3 (6.9)	54.9 (8.5)	0.003
60 mins	Free dye	24.2 (5.4)		
	TAMRA-GNPs	54.1 (9.2)	29.9 (6.1)	0.008
	TAMRA-GNPs-Oxo	75.3 (24.4)	51.2 (14.4)	0.024

Supplementary figures



Figure S1

Figure S1 (A) Principal component analysis of RNA-seq data of control, 0.2 μ M and 1 μ M 5Z-7-oxozeaenol treated human telomerase-immortalized human microvascular endothelial (TIME) cells. Each dot represents an experimental replicate. **(B)** Venn diagram shows the overlapped dysregulated genes between dysregulated genes identified in 0.2 μ M and 1 μ M 5Z-7-oxozeaenol treated TIME cells.

Figure S2



Figure S2 Heat map generated from unsupervised clustering of genes from gene set of 'cell cycle' (Reactome Pathway Database) of 0.2 μ M and 1 μ M 5Z-7-oxozeaenol treated TIME cells and normal controls.

Figure S3



Figure S3 Heat map generated from unsupervised clustering of genes from gene set of 'DNA Replication' (Reactome Pathway Database) of 0.2 μ M and 1 μ M 5Z-7-oxozeaenol treated TIME cells and normal controls.



KEGG_ Cell Cycle: Control vs. 1 µM Oxo





600

0.2uN_Oxo 1uN_Oxo condition ch

KEGG_ Cell Cycle: Control vs. 1 µM Oxo

2500

0.2uM_Oxo 1uM_Oxo condition

0.2uM_0xu

1uM_0xp Cirl condition 0.2uM_Oxo 1uM_Oxo condition



KEGG_ Cell Cycle: Control vs. 1 µM Oxo



KEGG_ Cell Cycle: Control vs. 1 µM Oxo

Figure S4 (A) Circular plot of KEGG pathway (cell cycle and DNA replication) enrichment analysis. (B) Bar charts show the normalized read count of each gene in the gene set of 'Cell Cycle' identified by GSEA study in TIME cells treated with 1 μ M 5Z-7-oxozeaenol. (C) Bar charts show the normalized read count of each gene in the gene set of 'DNA Replication' identified by GSEA study in TIME cells treated with 1 μ M 5Z-7-oxozeaenol.





Figure S5 5Z-7-oxozeaenol suppresses inflammatory cytokine-mediated MAPK signaling. (A) Western blot characterization of the TNF α -induced phosphorylation of JNK, p38, and ERK proteins in TIME cell stimulated by TNF α (10 ng/ml) for 10 minutes. (B-D) Phosphorylation of JNK, p38, and ERK proteins in MAPK signaling was decreased in cells treated with 5Z-7-oxozeaenol in a dose-dependent manner (n = 3 from three experiments). Statistical analysis was conducted by one-way ANOVA and Tukey's multiple comparison test. ***P < 0.001, ****P < 0.0001.





Figure S6 Characterization of gelatin nanoparticle-encapsulated 5Z-7-oxozeaenol. Representative images from dynamic light scattering (DLS) analysis revealed the particle polydispersity index (PDI), size, and zeta potential of GNP and GNP-Oxo particles. Each sample was tested in triplicates, shown as red, green, and blue lines.





Figure S7 The distribution of nanoparticles conjugated with fluorescence dye (TAMRA) in HUVECs. Cells displayed clearer staining in the cytoplasm and even near nucleus 30 minutes post-treatment with GNPs-Oxo and much more staining intracellularly 2 hours post-treatment with GNPs-Oxo. Red and blue represent fluorescence dye (TAMRA) and DAPI staining, respectively. Scale bar: 100 μm.



A 2 days post-injury

Figure S8 Gelatin nanoparticle-encapsulated 5Z-7-oxozeaenol reduces inflammatory responses developed in the mouse model of CoNV. (**A and B**) IL-1 β and TNF α expression was significantly increased 2- and 7-days post-injury in mice, while GNP-Oxo treatment moderately reduced the IL-1 β and TNF α expression (n = 5-8 corneas per treatment group). (**C**) Quantitative analysis of corneal blisters after chemical cauterization showed no difference among groups (see methods about the grading; n = 14 animals per treatment group from three experiments). Statistical analysis was conducted by one-way ANOVA and Tukey's multiple comparison test. **P* < 0.05, ****P* < 0.001.

Uncropped WB images Figure 3A











Figure S5A





