

Supplementary Information

Inactivating IL34 promotes regenerating muscle stem cell expansion and attenuates Duchenne muscular dystrophy in mouse

Yang Su^{1,2,3#}, Yuxin Cao^{1#}, Chang Liu¹, Qing Xu¹, Na Li¹, Miaomiao Lan¹, Lei Li¹, Kun Wang¹, Zeyu Zhang³, and Qingyong Meng^{1,4,✉}

1. State Key Laboratories for Agrobiotechnology, College of Biological Sciences, China Agricultural University, Yuanmingyuan West Road No. 2, Haidian District, Beijing 100193, China

2. Department of Cell Biology, Third Military Medical University (Army Medical University), Gaotanyan Road No. 30, Shapingba District, Chongqing 400038, China

3. State Key Lab of Animal Nutrition, College of Animal Science and Technology, China Agricultural University, Yuanmingyuan West Road No. 2, Haidian District, Beijing 100193, China

4. Beijing Advanced Innovation Center for Food Nutrition and Human Health, College of Biological Sciences, China Agricultural University, Yuanmingyuan West Road No. 2, Haidian District, Beijing 100193, China

✉ Corresponding authors: qymeng@cau.edu.cn

Yang Su and Yuxin Cao contributed equally to this work.

Figure S1

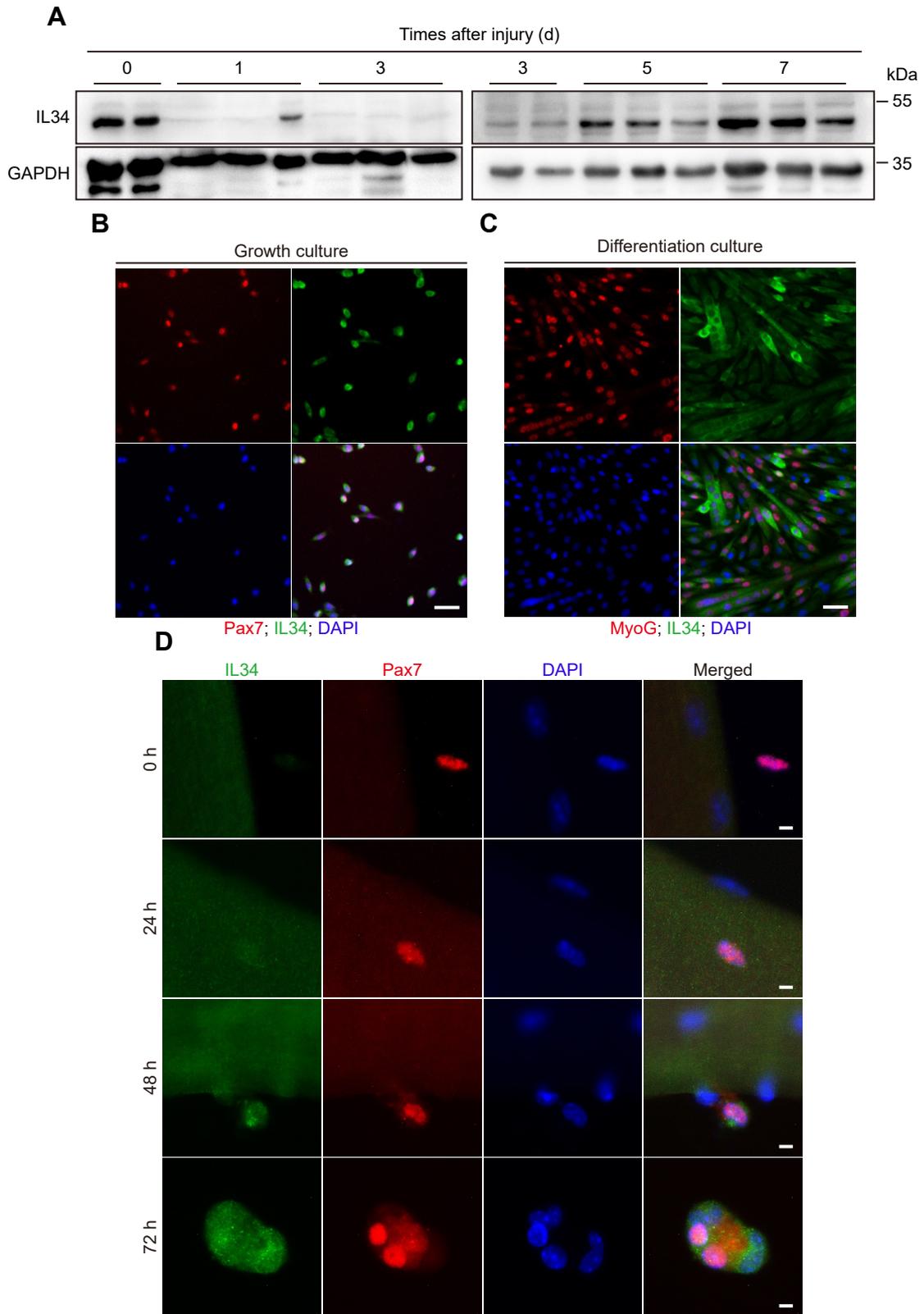


Figure S1. Expression patterns of IL34 during in vitro myogenesis and muscle regeneration. (A) Western blot showing the protein levels of IL34 and unrelated β -tubulin at various stages of skeletal muscle regeneration. (B) Cells undergoing growth culture were fixed and labeled with anti-Pax7 and anti-IL34 antibodies. DAPI was used to identify nuclei. Representative individual and overlaid images of WT cultures after labeling with Pax7, IL34 and DAPI. Scale bars: 30 μ m. (C) Cells induced to differentiate were fixed and labeled with anti-MyoG and anti-IL34 antibodies. DAPI was used to identify nuclei. Representative individual and overlaid images of WT cultures after staining for MyoG, IL34 and DAPI. Scale bars: 30 μ m. (D) IL34 immunofluorescence in Pax7⁺ SCs attached to freshly isolated EDL myofibers (0 h) or after culture for 24-72 h. Scale bar: 5 μ m.

Figure S2

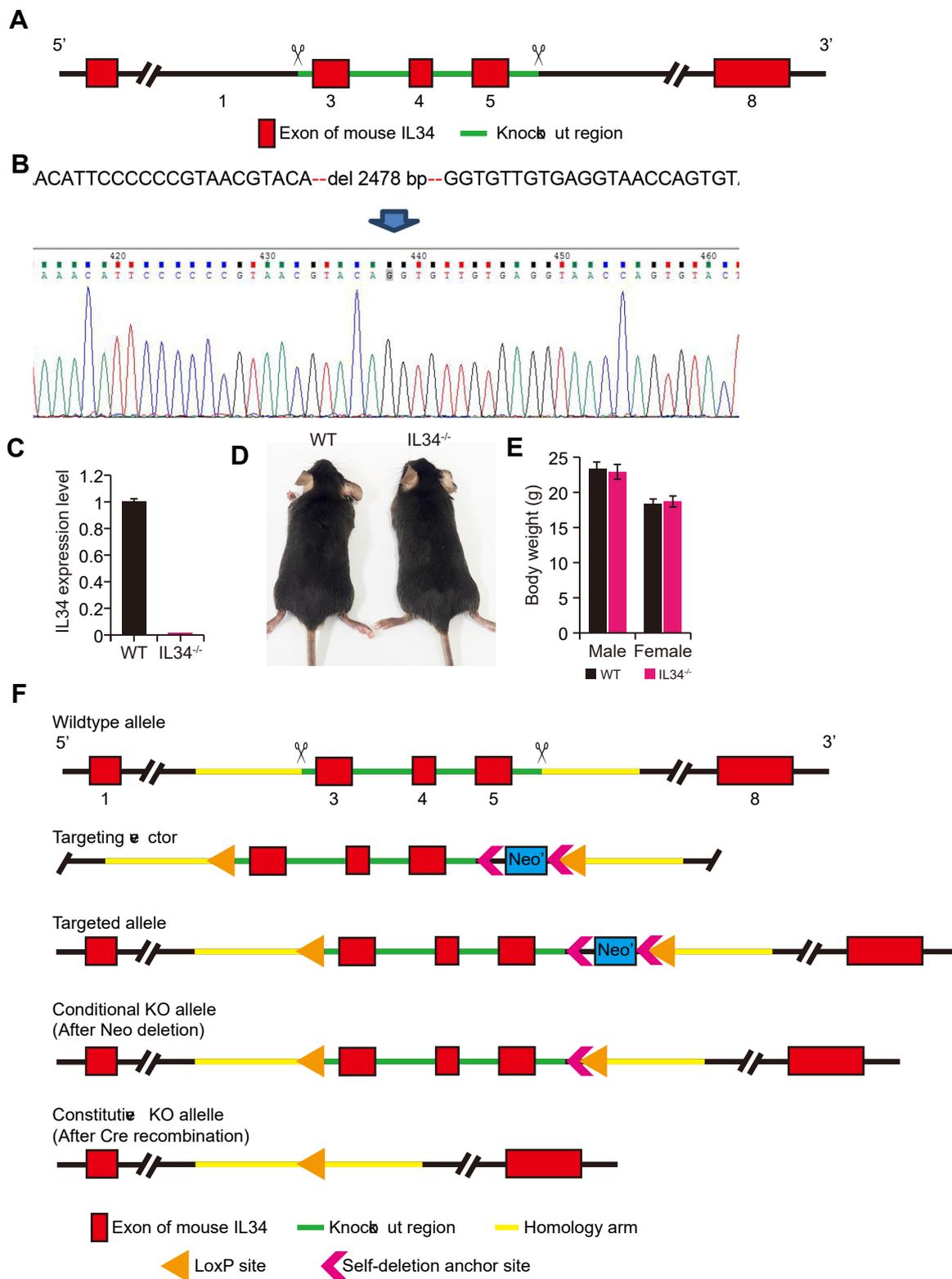


Figure S2. Generation of IL34 gene knockout mice. (A) Gene-targeting strategy for the generation of IL34 constitutive knockout mice. (B) DNA sequencing detecting the IL34 gene fragment that was effectively depleted in the mouse genome. (C) IL34 expression level in adult WT mice and IL34 gene deletion mice. (D) Comparison of adult WT mice and IL34^{-/-} mice. (E) Measurement of body weight in WT and IL34^{-/-} mice. N=5 mice in each group. (F) Gene-targeting strategy for the generation of IL34 gene conditional knockout mice.

Figure S3

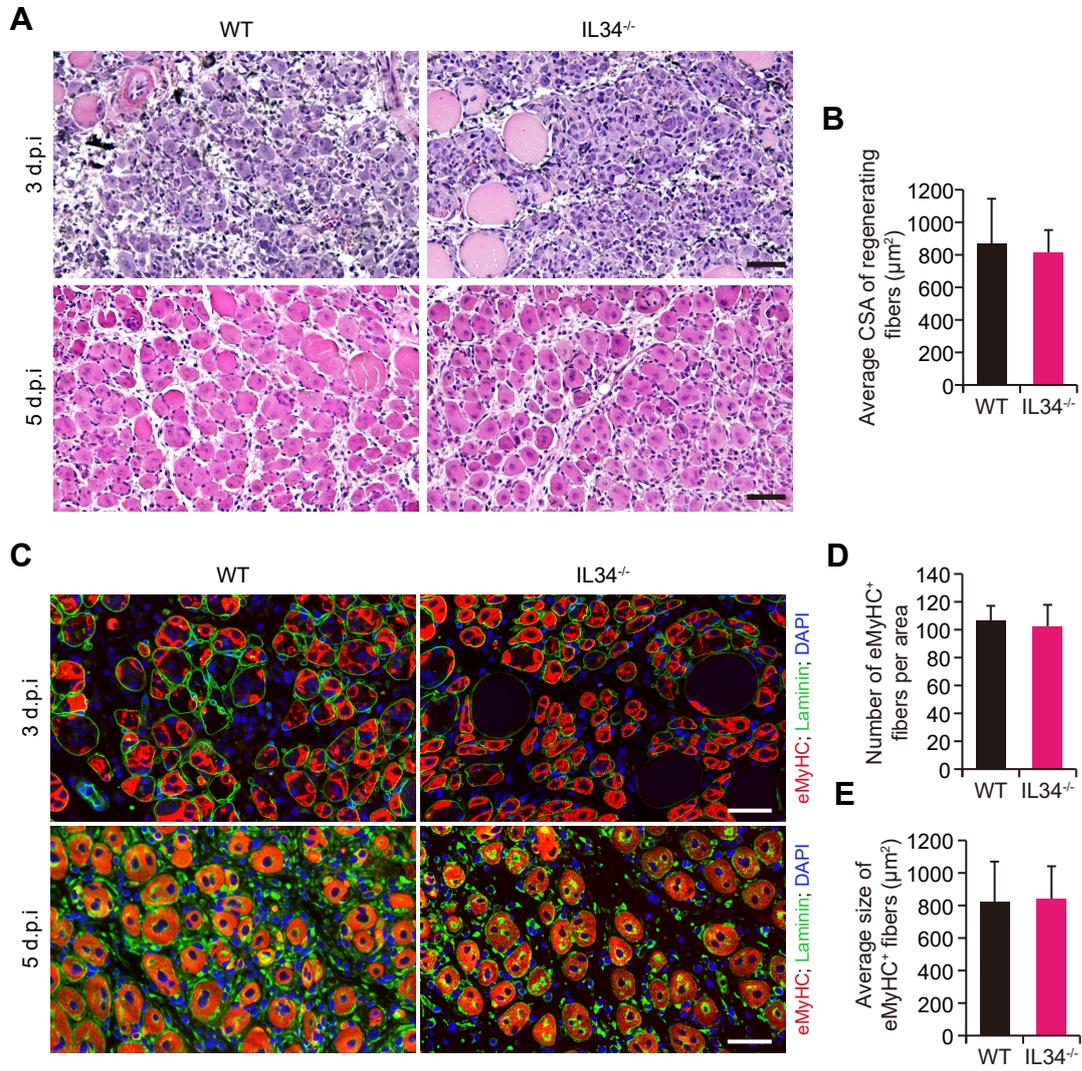


Figure S3. IL34 has weak effects on the early stage of skeletal muscle regeneration.

(A) H&E staining of transverse sections of TA muscle 3 and 5 days after injury. Scale bar, 50 μm . **(B)** Quantification of the average CSA of nuclei-centralized myofibers in 5-day postinjury TA muscle from WT and IL34 KO mice. **(C)** Representative merged photomicrographs of transverse sections of TA muscle 3 and 5 days after injury immunostained for eMyHC, laminin and DAPI. Scale bar, 50 μm . **(D)** Quantification of the number of eMyHC⁺ cells per area in 3-day postinjury transverse sections of TA muscle. **(E)** Measurement of the average CSA of eMyHC⁺ cells in transverse sections of the TA muscle 5 days postinjury.

Figure S4

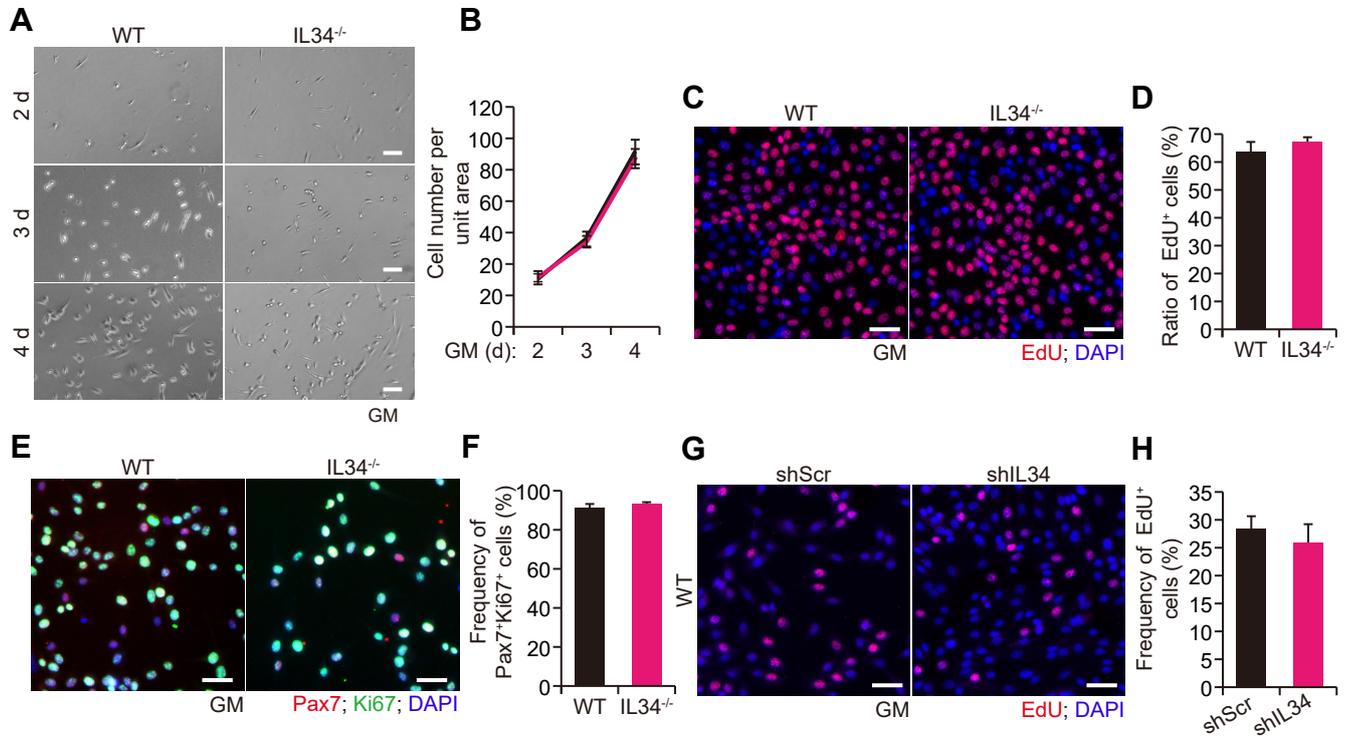


Figure S4. Proliferation assays of WT and IL34-KO SCs. (A) Approximately 4×10^4 FACS-isolated WT and IL34-KO SCs were plated in 12-well plates precoated with Matrigel and cultured in growth medium. Representative photomicrographs of SCs grown for 2 days, 3 days, and 4 days. Scale bar, 100 μm . (B) Quantification of the growth rate of WT and IL34-KO SCs in growth medium. (C) SCs were incubated in growth medium. Representative overlaid images of expanded WT and IL34-KO cultures after labeling with EdU. Nuclei were costained with DAPI. Scale bar: 30 μm . (D) Quantification of the frequency of EdU⁺ cells in WT and IL34-KO myoblasts. (E) Representative merged images of WT and IL34-KO myoblasts after immunostaining with Pax7, Ki67 and DAPI. Scale bar: 30 μm . (F) Quantification ratio of Pax7⁺Ki67⁺ cells in WT and IL34-KO SCs cultured in growth medium. (G) Representative merged images of WT myoblasts infected with lentivirus expressing shIL34 or shScr after labeling with EdU and DAPI. Scale bar: 30 μm . (H) Quantification ratio of EdU⁺ cells in WT myoblasts interfering with IL34.

Figure S5

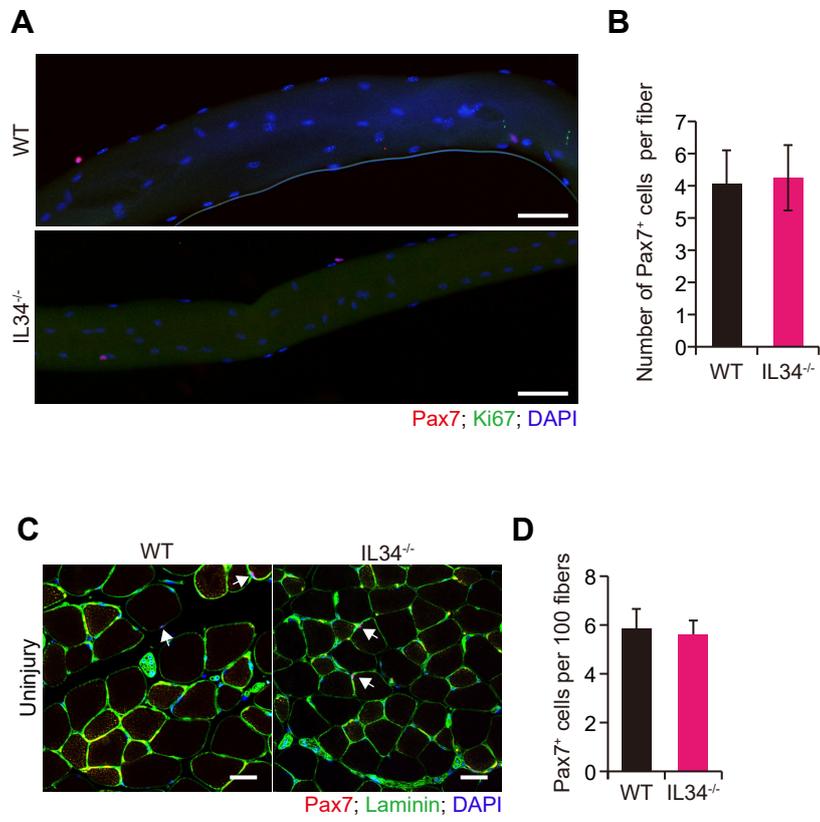


Figure S5. Inhibition of IL34 has no effects on the number of Pax7⁺ cells in freshly isolated single myofibers and transverse sections of uninjured skeletal muscle. (A) Representative merged photomicrographs of freshly isolated WT and IL34-KO single myofibers after labeling for Pax7, Ki67 and DAPI. Scale bar: 50 μ m. **(B)** Quantitative estimation of the number of Pax7⁺ cells on freshly isolated single myofibers. **(C)** Representative merged photomicrographs of normal TA muscle transverse sections labeled for Pax7, laminin and DAPI. Scale bar: 30 μ m. **(D)** Percentage of Pax7⁺ cells (normalized to myofibers) in transverse sections of uninjured TA muscle from WT and IL34-KO mice.

Figure S6

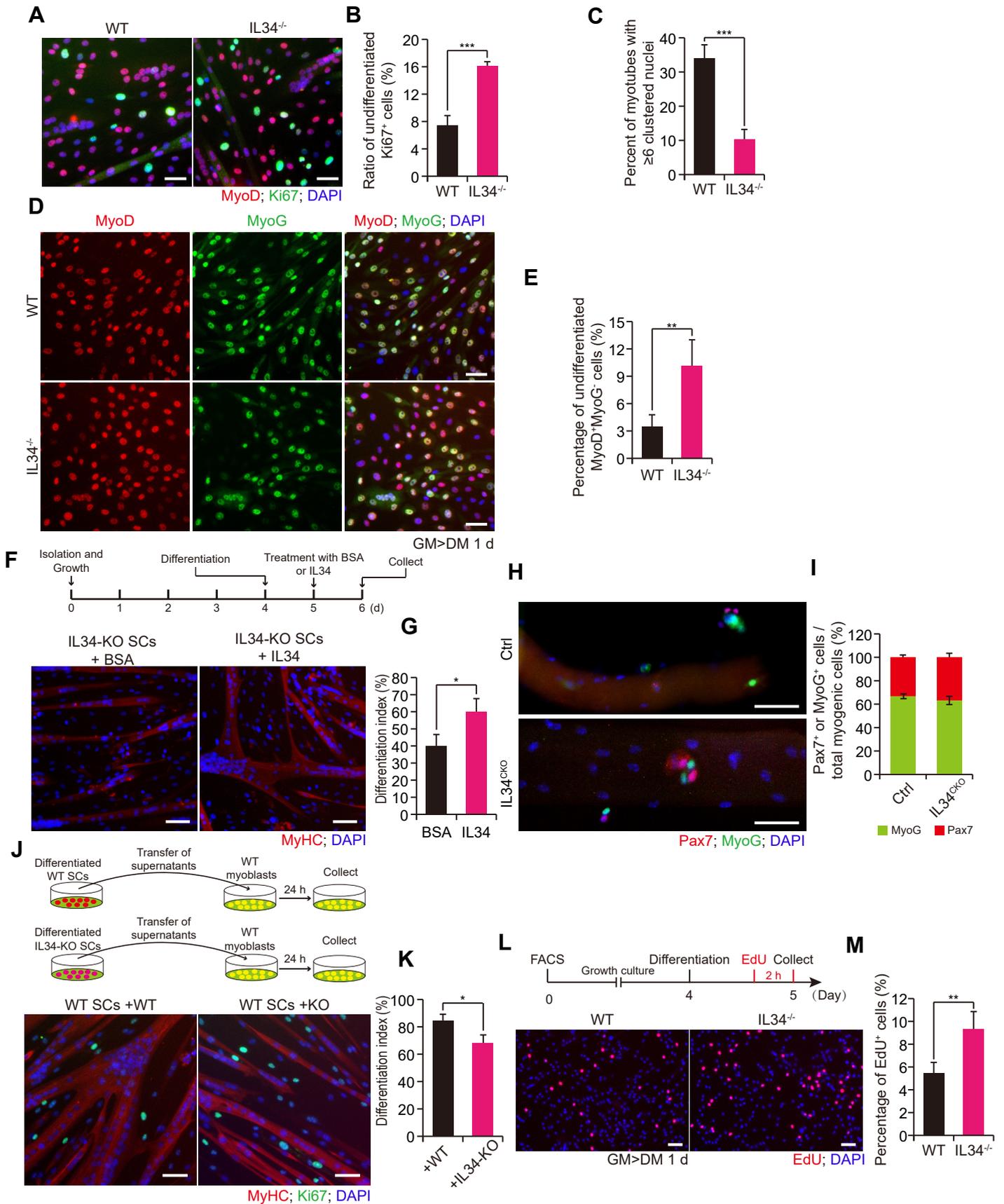


Figure S6. IL34 promotes SC differentiation. (A) WT and IL34-KO SCs were cultured in 10% horse serum medium for 3 day. The cells were then fixed and labeled with MyoD, Ki67 and DAPI. Representative merged photomicrographs of WT and IL34-KO cultures after labeling with MyoD, Ki67 and DAPI. Scale bars: 30 μ m. (B) Quantitation of the percentage of undifferentiated Ki67⁺ cells in WT and IL34-KO cultures. ***P<0.001. (C) Percentage of MyHC⁺ fibers containing \geq 6 nuclei relative to total MyHC⁺ fibers. ***p<0.001 (Related to Figure 2E and F). (D) SCs were initially incubated in growth medium, and then the growth medium was switched to differentiation medium. The cells were then fixed and labeled with anti-MyoG, anti-MyoD and DAPI. Representative overlaid images of WT and IL34-KO cultures after labeling with MyoG, MyoD and DAPI. Scale bars: 30 μ m. (E) Quantitative estimation of the percentage of undifferentiated MyoD⁺MyoG⁻ cells in WT and IL34-KO cultures. **P<0.01. (F) One-day differentiated IL34-KO SCs were treated with BSA or IL34 recombinants, and representative merged images of cultures were costained with MyHC and DAPI. Scale bar: 50 μ m. (G) Measurement of the differentiation index was calculated by the frequency of nuclei in MyHC⁺ cells relative to total nuclei. *p<0.05. (H) Representative overlapping images of 72 h cultured myofibers from the EDL muscle of Ctrl and IL34^{CKO} mice costained for Pax7, MyoG and DAPI. Scale bar: 50 μ m. (I) Percentage of Pax7⁺MyoG⁻ and Pax7⁻MyoG⁺ cell populations in Ctrl and IL34^{CKO} myofiber-associated SCs. (J) WT myoblasts induced to differentiation by supplementing with supernatant medium collected from differentiated WT or IL34-KO SCs, respectively. Representative merged images of cultures were costained with

MyHC, Ki67 and DAPI. Scale bar: 50 μm . **(K)** Measurement of the differentiation index was calculated by the frequency of nuclei in MyHC⁺ cells relative to total nuclei. * $p < 0.05$. **(L)** After incubation in growth medium, cells were induced to differentiate for 1 day, and EdU was added to label proliferating cells for the last 2 h. The cells were then fixed and analyzed by staining with EdU and DAPI. Representative merged photomicrographs of WT and IL34-KO cultures after staining with EdU and DAPI. Scale bars: 50 μm . **(M)** Proportion of EdU⁺ cells in WT and IL34-KO cultures. ** $p < 0.01$.

Figure S7

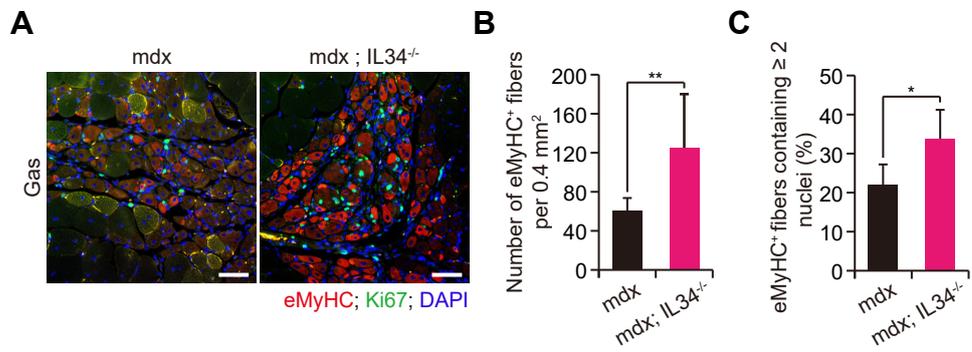


Figure S7. Genetic inactivation of IL34 in mdx mice attenuates Duchenne muscular dystrophy. (A) Representative merged images of eMyHC and Ki67 coimmunostained Gas sections from mdx and mdx::IL34^{-/-} mice at 12 weeks of age. Nuclei was labeled with DAPI. Scale bar: 30 μ m. (B) Number of eMyHC⁺ cells per field. **p<0.01. (C) Percentage of eMyHC⁺ fibers containing ≥ 2 nuclei relative to total eMyHC⁺ fibers. *p<0.05.