Supplementary Materials for

- **Pathological analysis revealed hypoxia inducible factor 3α prevents COPD by inhibiting alveolar epithelial cells ferroptosis via HIF-3α-GPx4 axis**

Junchao Jiang *et al.*

This PDF file includes:

Supplementary
Figs: S1 to S9

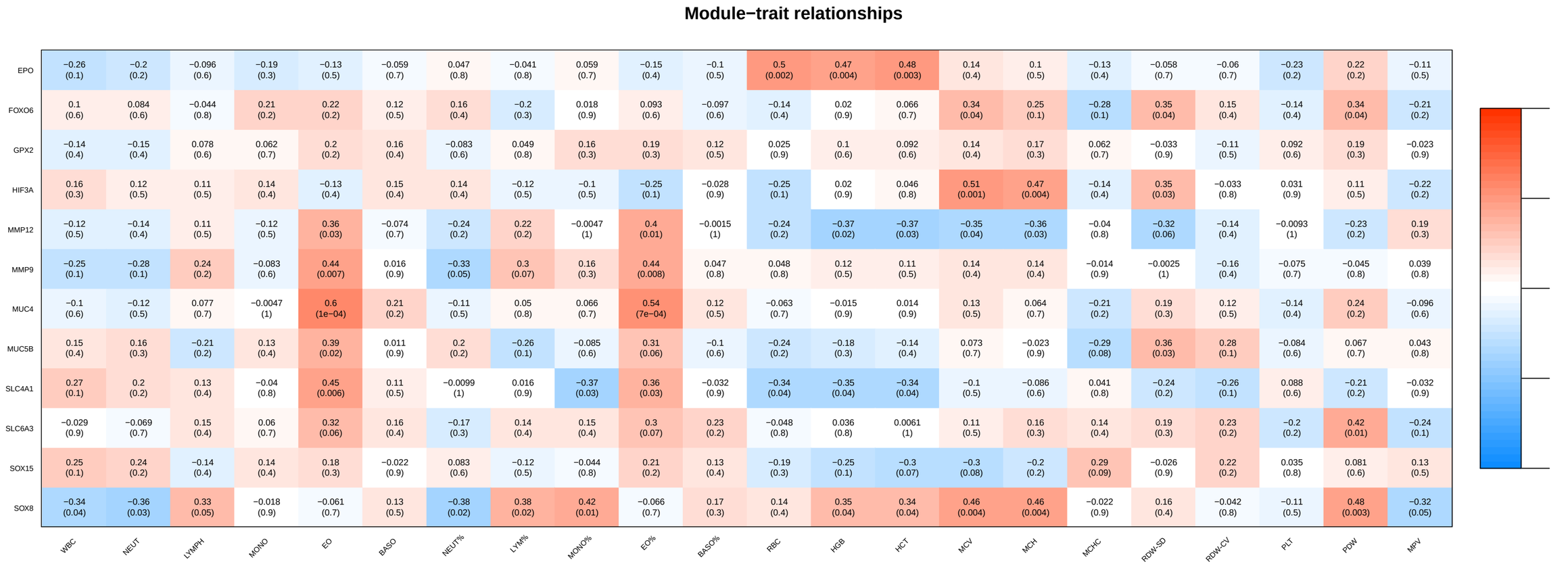

Fig S2. Core gene and complete blood count (CBC) correlation analysis. Sample matched mRNA value for each core gene and CBC results were used for correlation analysis, color in red represent positive correlation, in blue represent negative correlation.

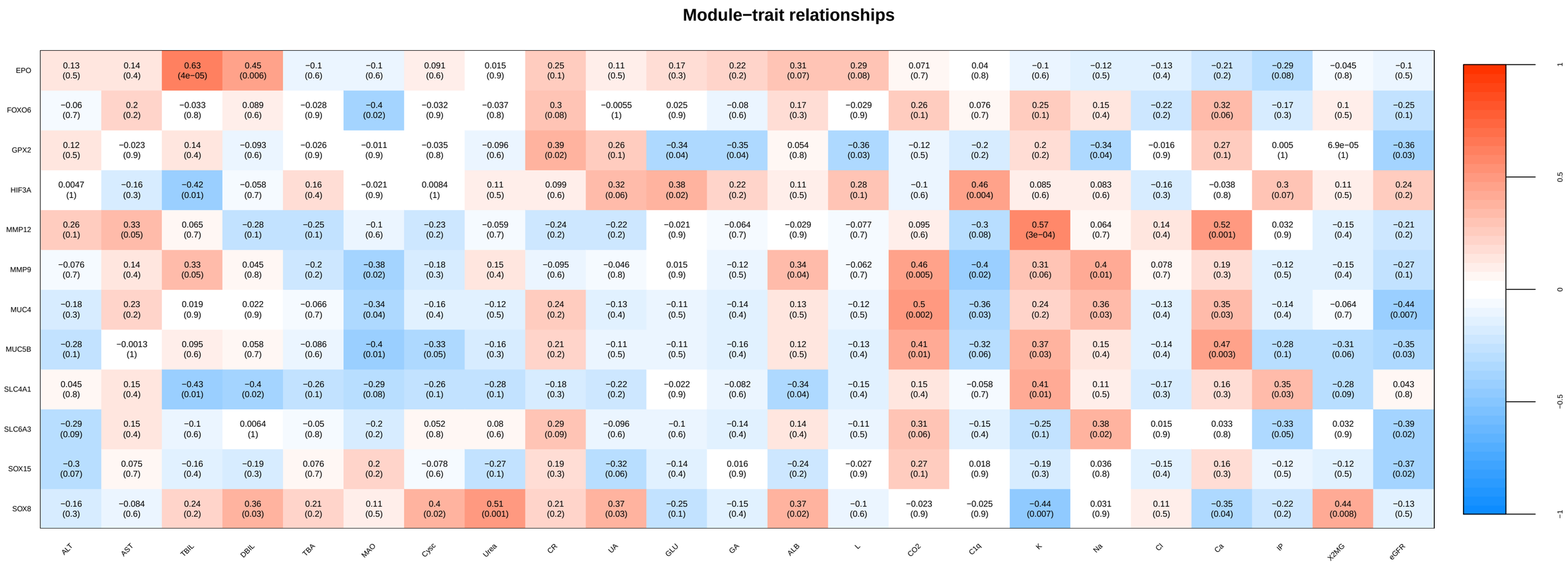

Fig S3. Core gene and blood biochemistry test correlation analysis. Sample matched mRNA value for each core gene and blood biochemistry test results were used for correlation analysis, color in red represent positive correlation, in blue represent negative correlation.

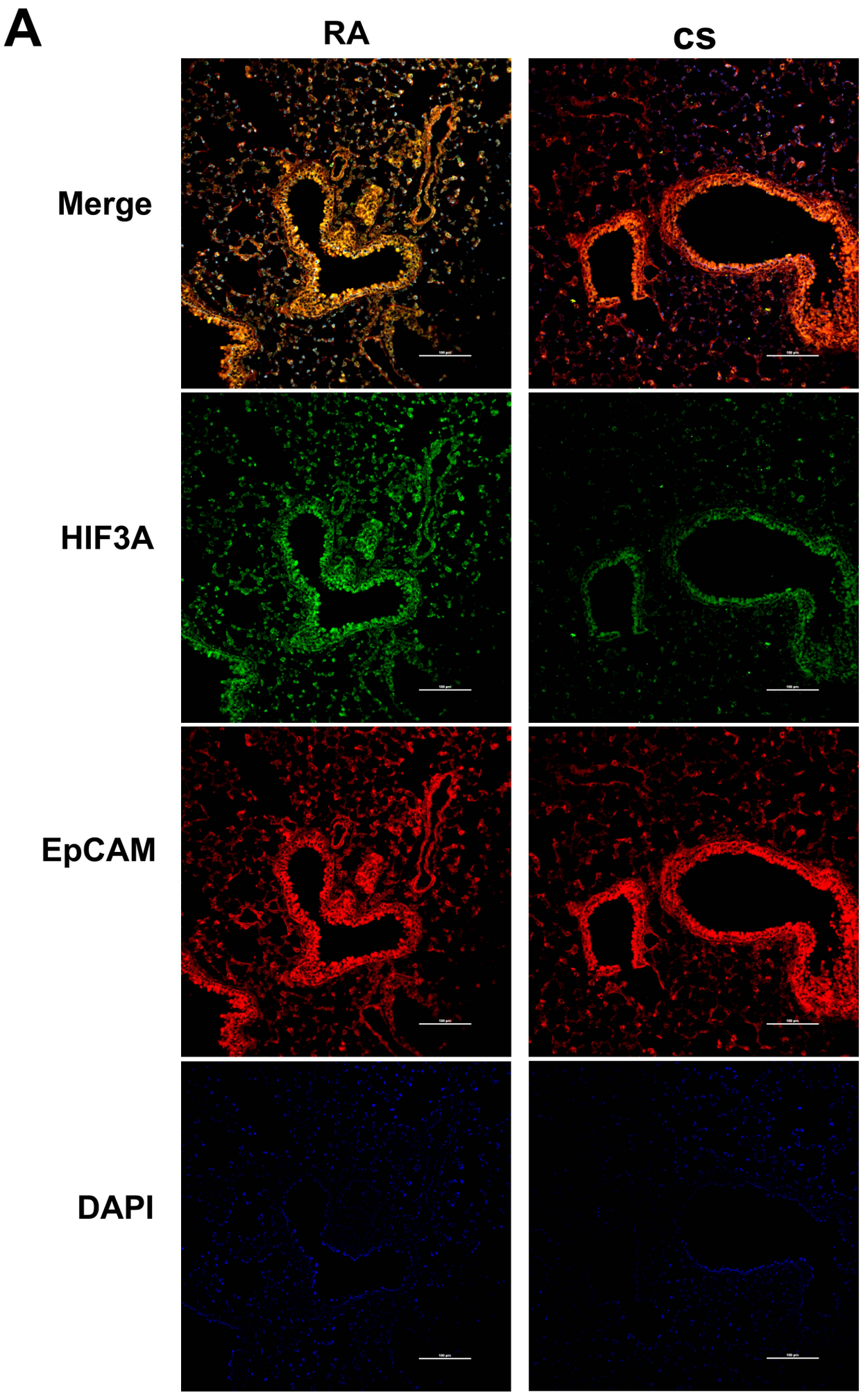

Fig S4. Co-localization analysis of HIF-3α and EpCAM in mice lungs. A) Colocalization of HIF-3α and EpCAM in mouse lung samples; RA stands for room air group; CS stands for cigarette smoking group. Original magnification ×10. Scale bars: 100 μm.

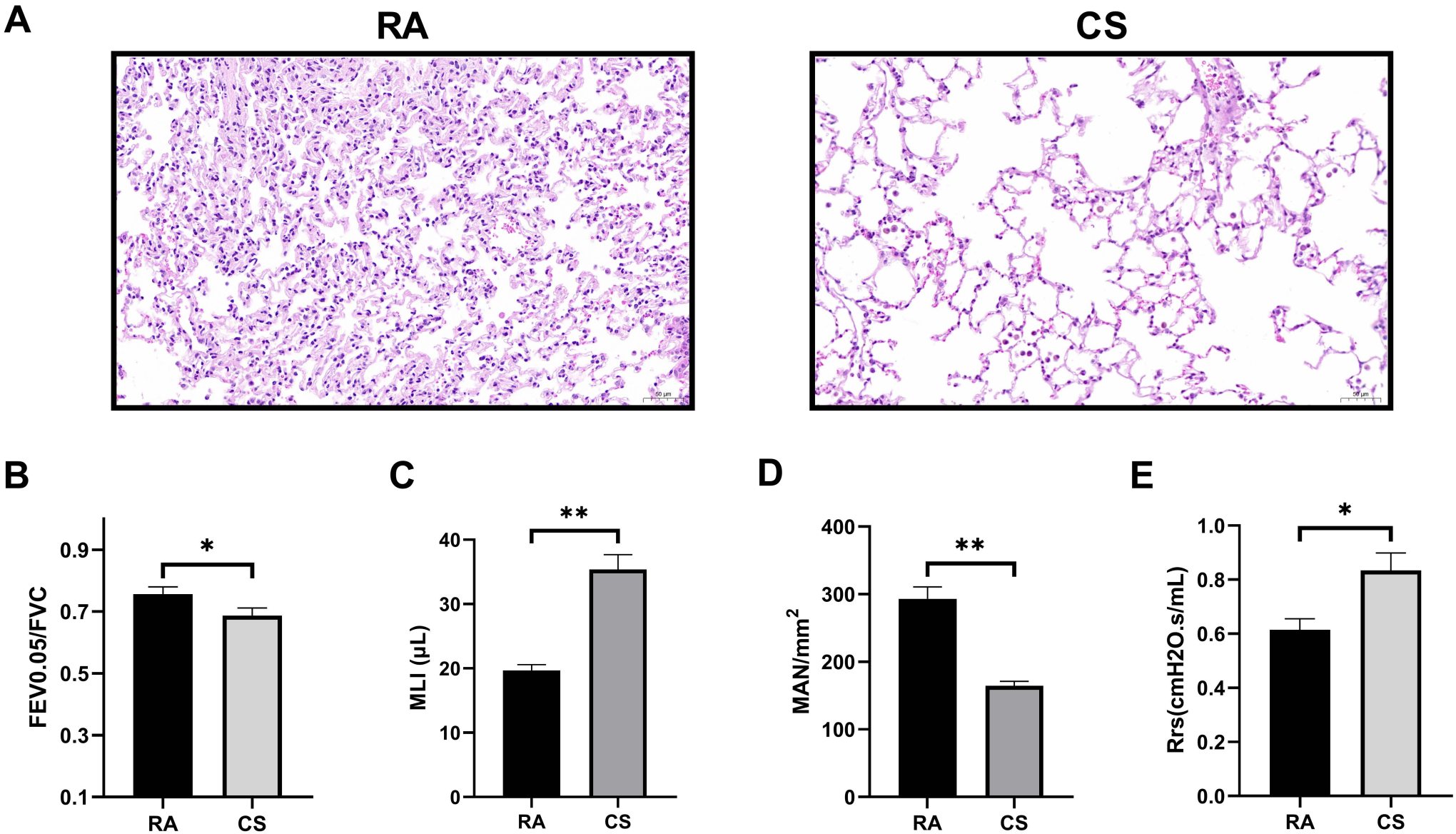

Fig S5. COPD mice model establishment and evaluation. "COPD" mice model was established as described in Method section. **A)** Result of H&E staining in mice model lung specimens were shown, RA stands for room air group, CS stands for cigarette smoking group. We further evaluated the **C)** mean linear intercept (MLI) and **D)** Mean Alveolar Number (MAN) in each group. Using a FlexiVent system, we tested the **B)** $FEV_{0.05}/FVC$, **E)** airway total resistance (Rrs) for each mice at least three times. *: $p<0.05$; **: $p<0.01$; ***: $p<0.005$; ****: $p<0.001$, ns: $p>0.05$ (nonsignificant).

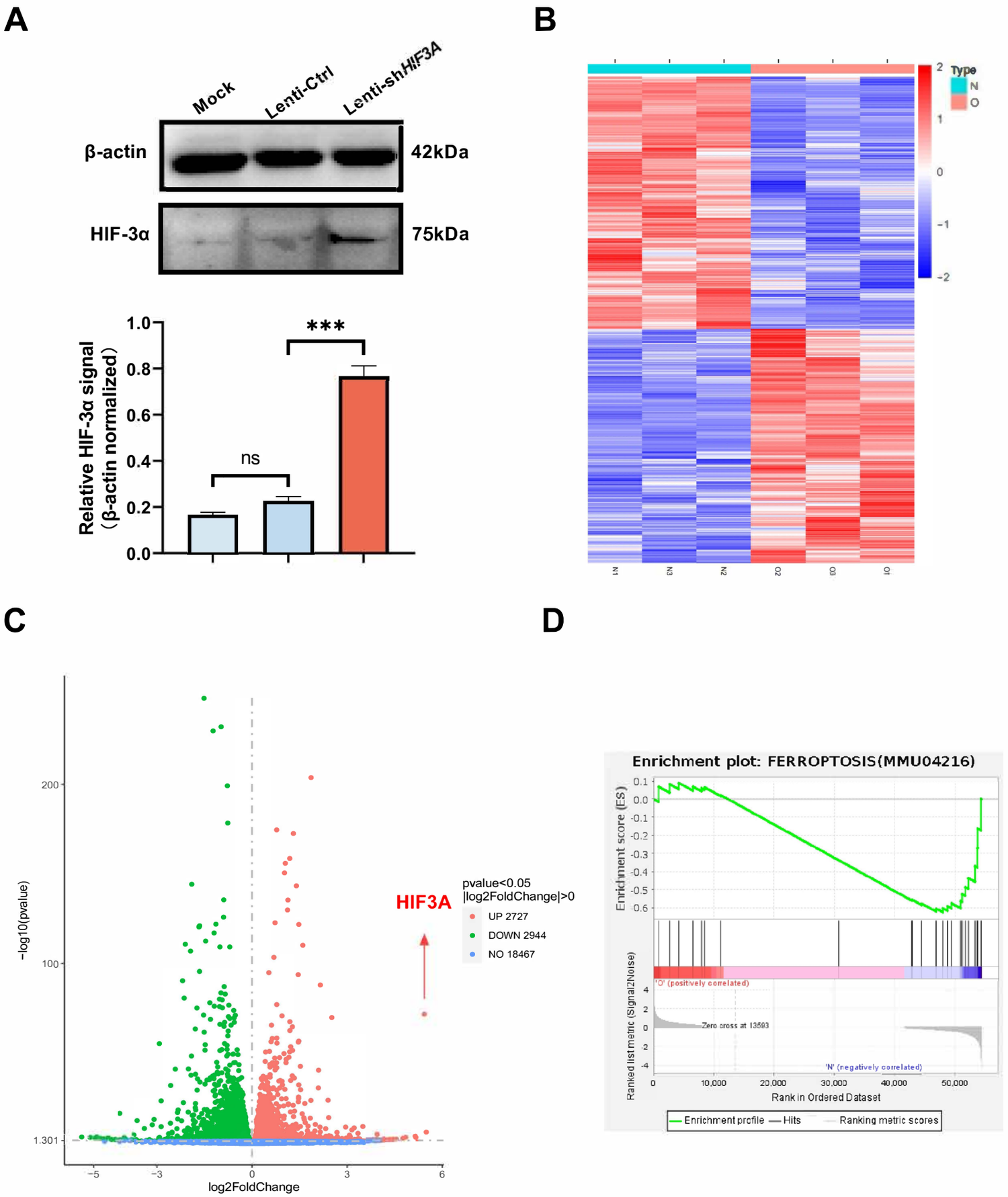

Fig S6. Bulk-RNA sequencing toward HIF-3α overexpression cells shed light on ferroptosis. **A)** MLE-12 cells were transfected with HIF-3α overexpression (OE) vector (Lenti-shHIF3A) or control vector (Lenti-Ctrl). HIF-3α protein levels were detected by Western blot and normalized to β-actin level. **B)** Heatmap for bulk-RNA sequencing, "N" stands for Lenti-ctrl group, "O" stands for Lenti-shHIF3A group. **C)** Volcano plot shows the differentially expressed genes and the log2FC value of HIF-3α. **D)** GSEA enrichment plot of ferroptosis. ***: $p<0.005$; ns: $p>0.05$ (non-significant)

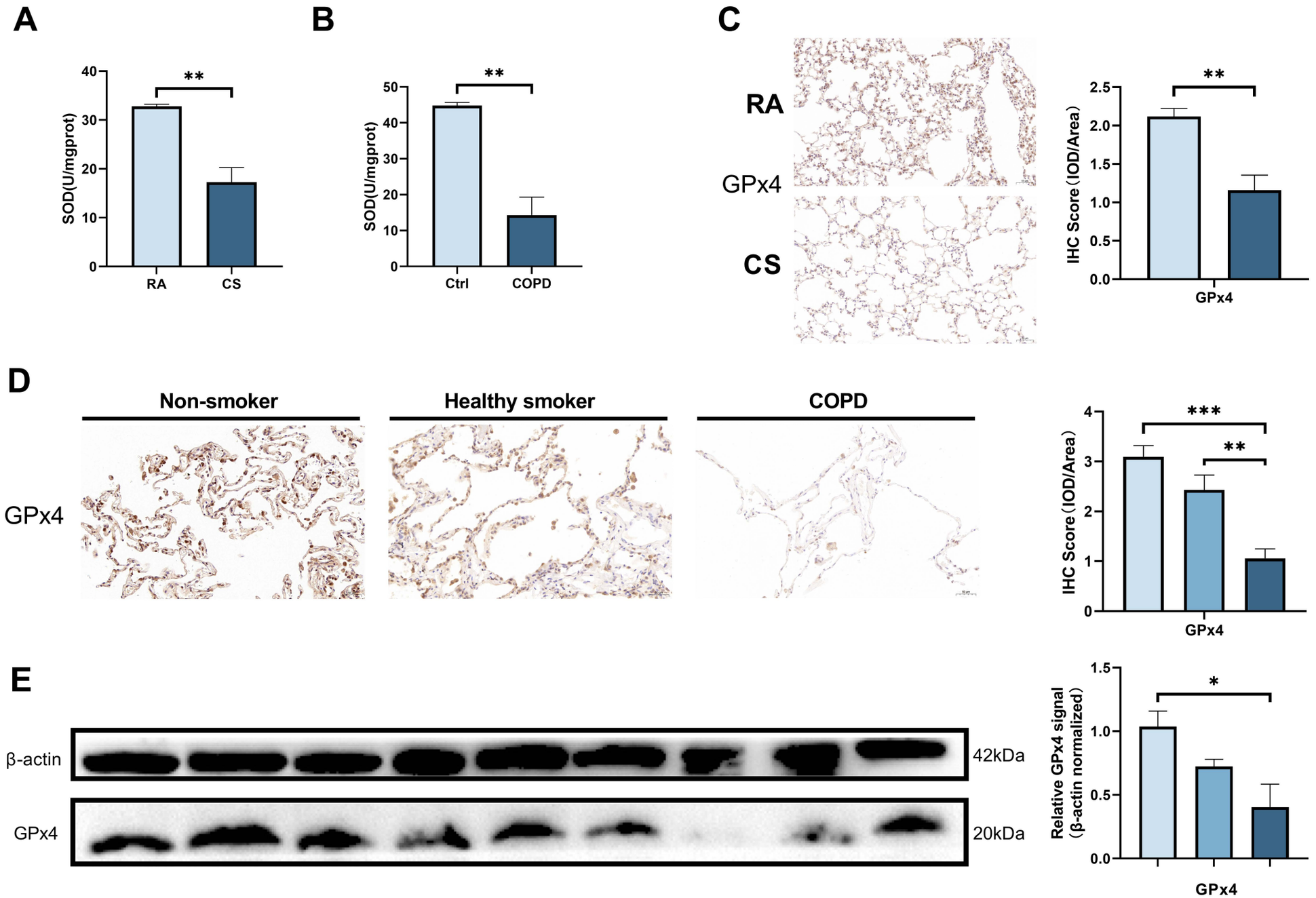

Fig S7. Disrupted GPx4 in COPD samples. Superoxide dismutase (SOD) levels in mice models **A)** humans **B)**. **C)** Immunohistochemistry showed that GPX4 was markedly lower in COPD mice models and in **D)** COPD patients. Original magnification ×20. Scale bars: 50 μm. **E)** Western blotting of human lung homogenates from COPD, non-smokers and smokers were probed with an anti-HIF-3α antibody (normalized to β-actin). IOD: integrated optical density. *: $p<0.05$; **: $p<0.01$; ***: $p<0.005$; ****: $p<0.001$. ns: $p>0.05$ (nonsignificant).

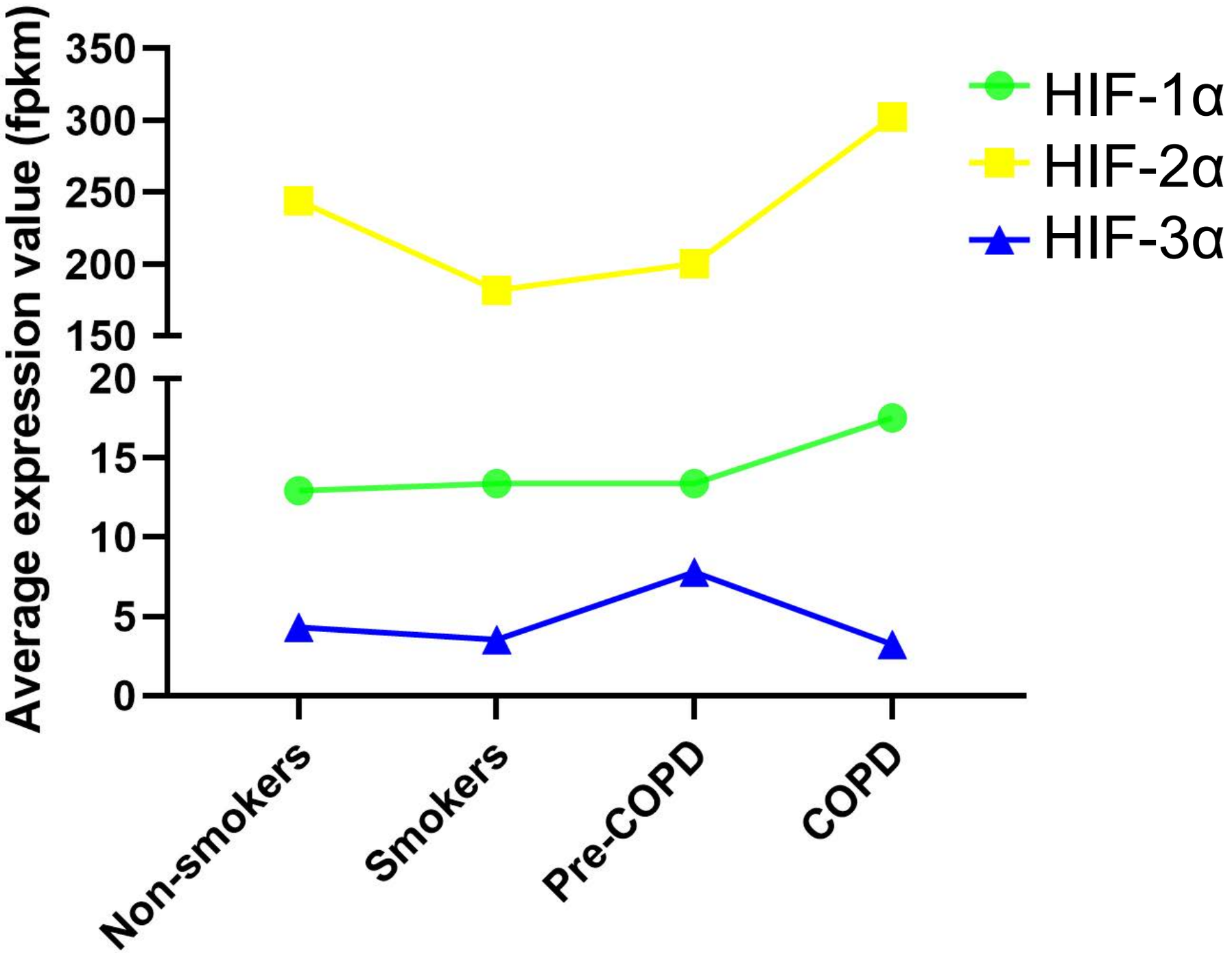

Fig S8. HIFs family expression in four groups: HIF-1α, HIF-2α and HIF-3α average expression value in four groups.

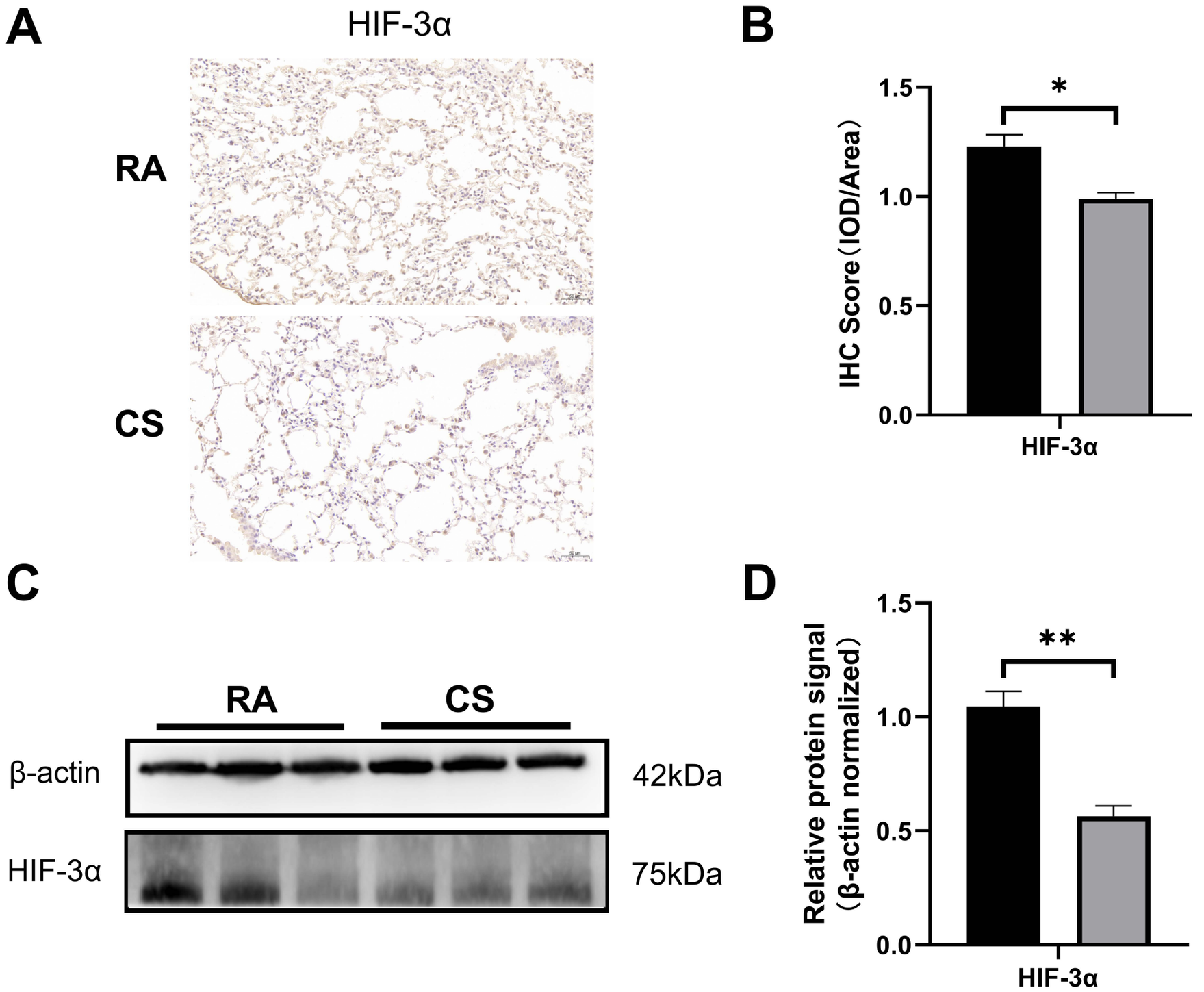

Fig S9. HIF-3α was downregulated in COPD mouse lung. A-B) Immunohistochemistry showed that HIF-3α was significantly lower in COPD mice group than RA group. Original magnification ×20. Scale bars: 50 μm. **C-D)** Western blotting of mice lung homogenates from COPD and RA groups that were probed with an anti-HIF-3α antibody (normalized to β-actin). *: $p<0.05$; **: $p<0.01$.