## В Α С 1.00 0.75 10 Percent of cells Component2 Cluste **ISNE2** 6 9 11 0.50 0.25 0.00 ΡТ LM U tSNE1 Component1 D Ε ALLMARK\_ESTROGEN\_RESPONSE\_EARLY ACIA BGI CALD CCN COLIA COLIA COLIA DCI FLN, IGFBP-INHB, JUI LGALS MMP DGFR SPAR TGFE THBS TGFE HALLMARK\_COAGULATION HALLMARK NOTCH SIGNALING ALLMARK\_XENOBIOTIC\_METABOLISM HALLMARK IL6 JAK STAT3 SIGNALING HALLMARK\_APICAL\_SURFACE -0.5 HALLMARK KRAS SIGNALING DN ALLMARK\_PANCREAS\_BETA\_CELLS HALLMARK HEDGEHOG SIGNALING ALLMARK\_WNT\_BETA\_CATENIN\_SIGNALING HALLMARK MYOGENESIS HALLMARK\_INFLAMMATORY\_RESPONSE HALLMARK\_IL2\_STAT5\_SIGNALING HALLMARK\_OXIDATIVE\_PHOSPHORYLATION HALLMARK\_TGF\_BETA\_SIGNALING ALLMARK UNFOLDED PROTEIN RESPONSE HALLMARK\_MYC\_TARGETS\_V2 HALLMARK\_EPITHELIAL\_MESENCHYMAL\_TRANSITION HALLMARK\_REACTIVE\_OXYGEN\_SPECIES\_PATHWAY HALLMARK\_PI3K\_AKT\_MTOR\_SIGNALING F G Н 10 MIF staining s

## **Supplementary Figures and Figure Legends**

Figure S1. Characterization of ductal cells related to Figure 2. (A) The cell differentiation dynamics of ductal cell subpopulations during evolutionary trajectories. (B) UMAP plot of subpopulations 6, 9, and 11 derived from the the ductal cells clusters in Figure 2A. (C) Bar plots showing the cell proportion of subpopulations 6, 9, and 11 from PT and LM. (D) Heatmap showing the expression levels of metastasis-related genes in subpopulations 6, 9, and 11. (E) GSVA enrichment analysis of biological functions and relevant signaling pathways in subpopulations 6, 9, and 11 based on Hallmark gene sets. (F) GO enrichment analysis of differentially expressed genes contrasting subpopulation 6 with subpopulations 9 and 11. (G) Immunohistochemistry detection of MIF expression in pancreatic adenocarcinoma primary tumors and liver metastatic lesions. Scale bars, 50  $\mu$ m. (H) Quantified staining scores of MIF in (G), with statistical testing by t-test (\*\*\*\*, p < 0.0001).



Figure S2. Single-cell transcriptomic landscape of PDAC and transcriptional signatures of ductal cell subpopulations. (A) UMAP plots of high-quality cells from primary tumor (PT) and liver metastatic tumors (LM), showing cell types, colour-coded by inferred cell types. (B) UMAP plots of the cell atlas showing their sample origins, colour-coded according to their tissues. (C) Dotplot showing the expression levels of major differentially expressed genes in the 9 cell types. (D) UMAP plot of the ductal cells landscape. Cells were coloured according to their clusters (left) or tissues (right). (E) Box plots showing EMT scores for each ductal cell subcluster. (F) Monocle pseudotime trajectory analysis of ductal cells clusters during the progression process. (G) Feature plots of LUM, FSTL1 and CITED4 along evolutionary trajectories. (I) Hallmark pathway enrichment analysis of differentially expressed genes between subpopulation 11 and other ductal cell subclusters. (J) Spatial feature plots of subpopulation 11 in PDAC spatial transcriptomic samples.



Figure S3. Monocle pseudotime trajectory analysis of T cells, spatial features and intercellular ligand-receptor prediction between ductal cells and lymphoid cells. (A) Monocle pseudotime trajectory analysis of CD8 cells related to Figure 3D. (B) Monocle pseudotime trajectory analysis of CD4 cells related to Figure 3E. (C) SPOTLight-based spatial deconvolution of metastatic dissemination cells in PDAC spatial transcriptomics datasets. (D) Hierarchical plot showing the strength of the CCL (left) and IL16 (right) signalling pathway between metastatic dissemination cells and lymphoid cells. (E) Scatter plot showing the relative strength of the outgoing and incoming signalling pathways in non-metastatic dissemination cells and lymphoid cells. (G) Bubble plots showing the specific ligand-receptor pairs of outgoing and incoming signalling pathways in non-metastatic dissemination cells and lymphoid cells. (H) Hierarchical plot showing the strength of the GALECTIN (left) and GDF (right) signalling pathway between non-metastatic dissemination cells and lymphoid cells.



Figure S4. Transcriptional signatures of myeloid cells identified by snRNA-seq. (A) UMAP plot of the myeloid cells landscape, coloured by subcluster. (B) Dotplot showing the expression levels of selected marker genes in the subclusters. (C) Dotplot showing the expression levels of top 10 differentially expressed genes in the TAMs subclusters. (D) MIA map of overlap between snRNA-seq-defined metastatic dissemination cells, TAMs, and ST-defined tumor, interface, and stroma regions. (E) GSVA enrichment analysis of biological functions and relevant signaling pathways in TAMs based on KEGG gene sets. (F) Monocle pseudotime trajectory analysis of TAMs during the progression process. (G) Bubble plots showing the specific ligand-receptor pairs of outgoing and incoming signalling pathways in metastatic dissemination cells and TAMs. (H) UMAP plot of the TAMs landscape. (I) Box plots showing M1 scores (left) and M2 scores (right) for each TAM subcluster. (J) Box



Figure S5. Transcriptional signatures of fibroblast cells identified by snRNA-seq. (A) UMAP plot of the fibroblast cells landscape, coloured by subcluster. (B) Dotplot showing the expression levels of selected marker genes in the subclusters. (C) Dotplot showing the expression levels of top 10 differentially expressed genes in the CAFs subclusters. (D) Monocle pseudotime trajectory analysis of CAFs during the progression process. (E) GSVA enrichment analysis of biological functions and relevant signaling pathways in CAFs based on KEGG gene sets. (F) MIA map of overlap between snRNA-seq-defined metastatic dissemination cells, CAFs, and ST-defined tumor, interface, and stroma regions. (G) Contribution of each ligand-receptor pair in metastatic dissemination cells and CAFs. (H) Bubble plots showing the specific ligand-receptor pairs of outgoing and incoming signalling pathways in metastatic dissemination cells and CAFs. (I) Hierarchical plot showing the strength of the SPP1 (left) and MIF (right) signalling pathway between metastatic dissemination cells and CAFs.



Figure S6. Transcriptional signatures of B cells identified by snRNA-seq. (A) UMAP plot of the B cells landscape, coloured by subcluster. (B) Dotplot showing the expression levels of selected marker genes in the subclusters. (C) MIA map of overlap between snRNA-seq-defined metastatic dissemination cells, B cells, and ST-defined tumor, interface, and stroma regions. (D) Monocle pseudotime trajectory analysis of B cells subclusters during the progression process. (E) Heatmap of top differentially expressed genes along evolutionary trajectories. (F) Multiplex immunofluorescence assay of pancreatic adenocarcinoma tissues stained for CD4, CD8, CD20, FOXP3, VIM, Pan-CK, and DAPI. Scale bars, 100  $\mu$ m. (G) Heatmap showing the relative strength of the outgoing and incoming signalling pathways in each cell subclusters.



Figure S7. Development of a metastatic dissemination-related prognostic signature based on disease-specific survival (DSS) analysis. (A) Feature plots showing the normalized expression of seven key MDRGs in ductal cells related to Figure 2A. (B) Univariate Cox regression analysis for MDRGs significantly associated with DSS. (C, D) LASSO regression analysis identified DSS-related candidate prognostic genes. (E) Risk score, survival status, and heatmap of the expression levels of the 7 candidate prognostic MDRGs in patients with pancreatic adenocarcinoma. (F) Kaplan-Meier survival analysis for DSS between the high- and low-risk groups. (G) ROC curves of the DSS-prognostic model for predicting 1-, 3-, and 5-year survival rates. (H) Calibration curve of the DSS-prognostic model. (J) Heatmap visualization of correlation analysis between MDRGs and immune infiltration data based on the CIBERSORT algorithm.



Figure S8. Development of a prognostic model for characterizing the immune microenvironment landscape in pancreatic adenocarcinoma. (A) Boxplot showing the xCell scores of immune cell subsets in high- and low-risk groups related to Figure 5L. (B) Immunofluorescence assay for PanCK, CD4, FOXP3, and DAPI in PT and LM samples. Scale bars, 50 µm. Statistical analysis of the number of CD4 FOXP3 T cells in PT and LM, statistical testing was performed using the t-test (\*\*, p < 0.01). (C) Kaplan-Meier curves showing DSS of patients with pancreatic adenocarcinoma according to CITED4 expression (left) or CIRBP expression (right). Grouping via P-value minimization. (D) Kaplan-Meier curves showing progress free interval (PFI) of patients with pancreatic adenocarcinoma according to CITED4 expression (left) or CIRBP expression (right). Grouping via P-value minimization. (E) Feature plots showing the normalized expression of CITED4 in cell atlas related to Figure 1B. (F) Boxplot showing the CIBERSORT scores of immune cell subsets in CITED4 highand low-expression groups. (G) Feature plots showing the ductal cells were stratified into CITED4-positive and CITED4-negative groups based on the expression status of CITED4 in ductal cells related to Figure 2A. (H) Box plots showing ICD scores for

CITED4-positive and CITED4-negative groups. (I) Boxplot showing the expression of ICOSLG and VTCN1 in high- and low-risk groups related to Figure 5L. (J) Scatter plots showing the significant negative spearman correlation of the expression of CITED4 with ICOSLG in the TCGA dataset. (K) Scatter plots showing the significant positive spearman correlation of the expression of CITED4 with VTCN1 in the TCGA dataset.