Supporting Information

An AIM2 inflammasome biomimetic mineralization inhibitor for vascular dementia therapy

Yueqi Zhang^{a,d}[‡], Lixian Jiang^{b,c}[‡], Rongrong Wu^b[‡], Wei Gao^c*, Xiaojie Zhang^{a,d}, Lan Liu^{a,d}, Yaxuan Zhang^{a,d}, Jin Lu^e, Yuanyi Zheng^{b,c}*, Xiaojun Cai^{b,c}*, Jianliang Fu^{a,d}*

^aDepartment of Neurology, Shanghai Sixth People's Hospital Affiliated to Shanghai Jiao Tong University School of Medicine, Shanghai 200233, P. R. China

^bShanghai Key Laboratory of Neuro-Ultrasound for Diagnosis and Treatment, Shanghai 200233, P. R. China

^cDepartment of Ultrasound in Medicine, Shanghai Sixth People's Hospital Affiliated to Shanghai Jiao Tong University School of Medicine, Shanghai 200233, P. R. China

^dShanghai Neurological Rare Disease Biobank and Precision Diagnostic Technical Service Platform, Shanghai 200233, P. R. China

^eDepartment of Pharmacy, Shanghai Sixth People's Hospital Affiliated to Shanghai Jiao Tong University School of Medicine, Shanghai 200233, P. R. China

Corresponding author:

Jianliang Fu: <u>fujianliang@163.com</u>; Xiaojun Cai: <u>c1x2j34@163.com</u> or <u>caixiaojun00@sjtu.edu.cn</u>; Yuanyi Zheng: <u>zhengyuanyi@sjtu.edu.cn</u> ; Wei Gao: <u>1033452945@qq.com</u>



Figure S1. Characterization HMPB. A) Representative Transmission electron microscopy (TEM) images of HMPB. Scale bar: 100nm. B) X-ray diffraction spectrum (XRD) of HMPB.
C) Fourier transform infrared spectroscopy (FTIR) of HMPB. D) X-ray photoelectron spectroscopy (XPS) binding energy peaks of Fe and Mn in HMPB.



Figure S2. The element mapping of HMPB (C, O, N, K, Fe, Mn, and S). Scale bar: 500 nm.



Figure S3. Characteristics of M2 macrophage-derived exosomes (M2 exosomes). A) Representative TEM image of M2 exosome. Scale bar: 0.2 μm. **B**) Nanoparticle tracking analysis (NTA) of M2 exosome. C) Western blot analysis of exosomal markers (CD9, CD63, and CD81).



Figure S4. Physicochemical properties of M2exo@HMPB. A) Intensity size distribution profiles of HMPB and M2exo@HMPB by dynamic light scattering (DLS). **B)** Hydrodynamic diameter of HMPB and M2exo@HMPB by DLS. **C)** Zeta potentials of HMPB and M2exo@HMPB.



Figure S5. *In vitro* and *in vivo* biodistribution of M2exo@HMPB. A) Blood-brain barrier (BBB) translocation efficiency of M2exo@HMPB *in vitro*. n=3 per group. B) Comparative brain Mn levels 24 h post intravenous (*iv*) versus intracerebroventricular (*ic*) administration. n = 3-6 per group. Data: mean \pm SEM. *P < 0.05; **P < 0.01; ***P < 0.001.



Figure S6. Validation of chronic cerebral hypoperfusion (CCH) rat model via bilateral common carotid artery occlusion (BCCAO). A) Schematic of laser speckle imaging (LSI) detection region. B) Quantification of cerebral blood flow (CBF) before and after BCCAO surgery. The CBF decreased significantly to approximately 50% of the baseline. n = 3 per group. Data: mean \pm SEM. *P <0.05, **P < 0.01, ***P < 0.001.



Figure S7. Long-term *in vivo* **metabolic profile of M2exo@HMPB.** Data: mean ± SEM. n=3-6/group.



Figure S8. Hypoglycemia/hypoxia (HH)-induced microglial damage. A) Time-dependent LDH release under HH conditions. n = 4 per group. Data: mean \pm SEM. *P < 0.05, **P < 0.01, ***P < 0.001. B) Live/dead staining of primary microglia (calcein AM [green]/propidium iodide [red]) post-HH treatments. n = 3 per group. Scale bar: 50 µm.



Figure S9. M2exo@HMPB mitigates prolonged HH-induced microglial death. Live/dead staining of primary microglia post-treatments. Scale bar: 50 μ m. n = 3 per group.



Figure S10. Experimental design for microglia and neuron co-culture.



Figure S11. AIM2 inflammasome modulation by M2exo@HMPB. A) AIM2 (red)/Iba-1 (green) co-staining in microglia. Scale bar: $50\mu m$, n = 3 for each group. B) IL-1 β and C) IL-18 levels in supernatant by ELISA. n = 4. Data: mean \pm SEM. *P < 0.05; **P < 0.01; ***P < 0.001. ns: not significant.



Figure S12. Intracerebroventricular M2exo@HMPB improves cognitive deficits in CCH rats. A) Representative Morris water maze swim paths (probe trial). B) Escape latency during hidden platform training. C) Platform crossings and D) Target quadrant occupancy in probe trial. E) Swimming speeds. n = 8-10 per group. Data: mean \pm SEM. *P < 0.05, **P < 0.01, ***P < 0.001 vs. CCH; #P < 0.05, ##P < 0.01, ###P < 0.001 vs. sham; ns: not significant.



Figure S13. Effect of intracerebroventricular M2exo@HMPB on the levels of glial activation, white-matter integrity, and hippocampal neuronal density in the hippocampus in CCH rats. A) Representative H&E staining of the rat hippocampus. Scale bar: 500 μ m (left panel) or 50 μ m (right panel), n = 3 per group. B-C) Nissl staining and neuronal quantification in CA1/CA2/CA3 regions. Scale bar: 500 μ m (upper panel) or 50 μ m (lower panels), n = 4 per group. D-E) Immunostaining of Iba-1 and Iba-1⁺ microglia quantification. Scale bars: 200 μ m

(left panel) or 20 μ m (right panel), n = 4 per group. Representative Luxol fast blue staining F) and quantification of white-matter integrity G) in the corpus callosum (paramedian), corpus callosum (medial), caudoputamen, internal capsule, and optic tract in rats. Scale bars: 1mm (left panel) or 50 μ m (right panels), n = 4 per group. Data: mean ± SEM. *P < 0.05; **P < 0.01; ***P < 0.001 vs. CCH. ns: not significant.



Figure S14. *In vivo* biosafety assessment of nanoparticles. A) H&E-staining major organs. Scale bars = 50 μ m. B) Serum liver function biochemical markers, alanine aminotransferase (ALT), and C) aspartic acid transferase (AST). D) Serum kidney function biochemical markers, Creatinine (Cre) and E) blood urea nitrogen (BUN). Data: mean ± SEM, n = 3 per group.



Figure S15. Transcriptomic profiling of CCH vs M2exo@HMPB-treated hippocampi. A) Principal component analysis (PCA) plot of differentially expressed genes (DEGs). **B)** Volcano plot of DEGs (CCH vs M2exo@HMPB).

Supplementary Movies

Movie S1: MD of AIM2^{HIN} domain with manganese ferrocyanide interface. **Movie S2:** MD of AIM2^{PYD} domain with manganese ferrocyanide interface.