Supplementary Materials for

• FAK signaling suppression by OCT4-ITGA6 mediates the effectively removal of residual pluripotent stem cells and enhances application safety

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Figure S1. Identification of PSCs. A. Bright-field images of iPSCs-001-5, EPSCs-001-5, iPSCs-006-1, and EPSCs-006-1. Scale bars, 100 μ m; B. qPCR analysis showing the differential expression of pluripotency genes (OCT4, SOX2, Nanog, Klf17, Dppa3) between PSCs and the negative control HUVECs. Data are from three independent experiments (n = 3), presented as mean \pm SEM, and statistically analyzed using one-way ANOVA. Comparison with HUVECs: *p<0.05; **p<0.01; ***p<0.001; C. Crystal violet staining images of iPSCs-001-5, EPSCs-001-5, iPSCs-006-1, and EPSCs-006-1; D. Fluorescence images showing the protein expression

of pluripotency markers in the four PSC lines. Scale bars, 200 μ m; E. Schematic representation of the construction of EGFP-iPSCs; F. Fluorescence images displaying the expression of EGFP, OCT4, and SOX2 in EGFP-iPSCs. Scale bars, 100 μ m; G. Bright-field images of EBs formed by iPSCs. Scale bars, 200 μ m; H. Fluorescence images showing the expression of three germ layer protein markers in EBs after attachment. Scale bars, 200 μ m; I. qPCR analysis of pluripotency genes and three germ layer differentiation marker genes in iPSCs and iPSC-derived EBs. Data are from three independent experiments (n = 3), presented as mean \pm SEM, and statistically analyzed using Student's t-test. *p<0.05; **p<0.01; ***p<0.001.



Figure S2. BSS (Ca-Mg-) treatment causes detachment of multiple PSCs. A-B. Bright-field images of iPSCs-006-1 before and after treatment with BSS (Ca-Mg-) at 37° C and room temperature. Scale bars, 50 μ m; C-D. Bright-field images of EPSCs-006-1 before and after treatment with BSS (Ca-Mg-) at 37° C and room temperature. Scale bars, 50 μ m; E-F. Bright-field images of EPSCs-001-5 before and after treatment with BSS (Ca-Mg-) at 37° C and room

temperature. Scale bars, 200 μ m; G. Crystal violet staining image of iPSCs-006-1 after BSS (Ca-Mg-) treatment; H. Crystal violet staining image of EPSCs-006-1 after BSS (Ca-Mg-) treatment; I. Crystal violet staining image of EPSCs-001-5 after BSS (Ca-Mg-) treatment.



Figure S3. Establishment of iPSCs and iDC co-culture systems and DPBS (Ca-Mg-)

treatment. A. Schematic diagram of the co-culture system of iPSCs with iFCs, iOBs, and iEPCs, and the subsequent DPBS (Ca-Mg-) treatment; B. Bright-field images of iOBs, iFCs, and iEPCs before and after DPBS (Ca-Mg-) treatment (37° C, 30 min). White arrows indicate iPSCs, and blue arrows indicate iDCs. Scale bars, $200 \mu m$; C. Bright-field images of the iFCs, iOBs, and iEPCs co-culture system and after DPBS (Ca-Mg-) treatment. Scale bars, $200 \mu m$; D. Flow cytometry analysis showing the percentage of EGFP-positive cells among adherent cells before and after DPBS (Ca-Mg-) treatment.



Figure S4. DPBS (Ca-Mg-) short-term treatment does not affect iMSCs characteristics. A. Schematic diagram of iMSCs characteristic detection before and after DPBS (Ca-Mg-) treatment; B. Bright-field images of iMSCs cultured with MSCM (37° C) after DPBS (Ca-Mg-) treatment at 37° C and room temperature. Scale bars, 200 µm; C. Flow cytometry analysis of surface markers (CD44, CD73, CD90, CD105, CD166, HLA-DR, CD34, and CD45) expression in iMSCs before and after DPBS (Ca-Mg-) treatment; D. Alcian blue staining to assess the chondrogenic differentiation ability of iMSCs before and after DPBS (Ca-Mg-) treatment. Scale bars, 200 µm; E. Oil Red O staining to assess the adipogenic differentiation ability of iMSCs before and after DPBS (Ca-Mg-) treatment. Scale bars, 50 µm; F-G. ALP staining and alizarin red staining to assess the osteogenic differentiation ability of iMSCs before and after DPBS (Ca-Mg-) treatment. Scale bars, 200 µm; F-G. ALP staining and alizarin red staining to assess the osteogenic differentiation ability of iMSCs before and after DPBS (Ca-Mg-) treatment. Scale bars, 200 µm; F-G. ALP staining and alizarin red staining to assess the osteogenic differentiation ability of iMSCs before and after DPBS (Ca-Mg-) treatment. Scale bars, 200 µm; H-I. Wound healing assay and subsequent quantification to

evaluate the cell migration ability of iMSCs before and after DPBS (Ca-Mg-) treatment. Data from three independent experiments (n = 3) are presented as mean \pm SEM, and statistical analysis was performed using two-way ANOVA. ns p>0.05, *p<0.05; **p<0.01; ***p<0.001. J-K. Colony formation assay and subsequent quantification to evaluate the self-renewal ability of iMSCs before and after DPBS (Ca-Mg-) treatment. Data from three independent experiments (n = 3) are presented as mean \pm SEM, and statistical analysis was performed using Student's t-test. ns p>0.05, *p<0.05; **p<0.01; ***p<0.001; L. CCK8 assay to assess the cell proliferation ability of iMSCs before and after DPBS (Ca-Mg-) treatment. Data (n = 6) are presented as mean \pm SEM, and statistical analysis was performed. NovA. ns p>0.05; **p<0.05; **p<0.05; **p<0.05; **p<0.01; ***p<0.01; *



Figure S5. A. Violin plot comparing the expression levels of common integrins in PSCs and iMSCs. The p-values from the Wilcoxon test are shown in the figure.



Figure S6. Altered OCT4, ITGA6, and FAK signaling in iPSCs after differentiation into neuroectoderm and mesendoderm. A. Schematic diagram showing the differentiation of iPSCs into neuroectoderm and mesendoderm; B. qPCR analysis showing changes in differentiation markers and pluripotency markers after iPSCs differentiation into neuroectoderm and mesendoderm; C. Fluorescence images showing the protein expression of OCT4 and ITGA6

after iPSCs differentiation into neuroectoderm and mesendoderm. Scale bars, 200 μ m; D. Fluorescence images showing the expression of differentiation markers after iPSCs differentiation into neuroectoderm and mesendoderm. Scale bars, 100 μ m; E-F. Fluorescence images showing the expression and quantification of p-FAK and FAK after iPSCs differentiation into neuroectoderm and mesendoderm. Scale bars, 200 μ m. Data (n = 5) are presented as mean ± SEM and analyzed using one-way ANOVA. *p<0.05; **p<0.01; ***p<0.001; G-H. Western blot results and quantification of p-FAK, and ITGA6 after iPSCs differentiation into neuroectoderm and mesendoderm. Data (n = 3) are presented as mean ± SEM and analyzed using one-way ANOVA. *p<0.01; ***p<0.001; ***



Figure S7. Inhibition of FAK signaling upregulated OCT4 expression in iPSCs. A. Schematic diagram illustrating the strategy for inhibiting FAK signaling in iMSCs using a PF-562271; B. qPCR analysis of the expression levels of OCT4 in FAKi-treated iPSCs. Data are presented as mean \pm SEM from three independent experiments (n = 3), with statistical significance determined using one-way ANOVA (*p < 0.05; **p < 0.01; ***p < 0.001); C-D. Immunofluorescence images and quantitative analysis of p-FAK and FAK in iPSCs treated with FAKi. Quantification is based on five independent experiments (n = 5), with data presented as mean \pm SEM and analyzed using one-way ANOVA, *p < 0.05; **p < 0.01; ***p < 0.001. Scale bars, 100 µm; E. Schematic diagram of regulation between OCT4, ITGA6 and FAK signaling in iPSCs.

Gene Symbol	Sequence (5'->3')	
OCT4	Forward primer	GTGTTCAGCCAAAAGACCATCT
	Reverse primer	GGCCTGCATGAGGGTTTCT
SOX2	Forward primer	GGGAAATGGGAGGGGTGCAAAAGAGG
	Reverse primer	TTGCGTGAGTGTGGATGGGATTGGTG
Nanog	Forward primer	TTTGTGGGCCTGAAGAAAACT
	Reverse primer	AGGGCTGTCCTGAATAAGCAG
KLF17	Forward primer	GCTGCCCAGGATAACGAGAAC
	Reverse primer	ATCTCTGCGCTGTGAGGAAAG
Dppa3	Forward primer	TTAATCCAACCTACATCCCAGGG
	Reverse primer	AGGGGAAACAGATTCGCTACTA
ITGA1	Forward primer	GGTTACCCTGTGCTGTACCCAA
	Reverse primer	TGCCTCGTTTGAGATGGTCA
ITGA2	Forward primer	TCTGAGACTGCCAAGGTCTTCA
	Reverse primer	CAGCTGGTATTTGTCGGACATC
ITGA3	Forward primer	GAACCCCTTCAAACGGAACC
	Reverse primer	ACCTCAAAGGCGATGAGCAG
ITGA5	Forward primer	AGATCCTGAAATGCCCGGA
	Reverse primer	CAGACTCGGAAATGCAACTGC
ITGA6	Forward primer	GTTTGATAACGATGCTGACCCC
	Reverse primer	TGAGCACATGTCACGACCTTG
ITGA7	Forward primer	AACCTGGAAGAACCCAAGCAC
	Reverse primer	TGACATTTTCCTGGAGCTGGA
ITGAV	Forward primer	TGACATTTTCCTGGAGCTGGA
	Reverse primer	TCTCTGACTGCTGGTGCACACT
ITGB1	Forward primer	ATGCCATCATGCAAGTTGCA
	Reverse primer	CCCATCTCCAGCAAAGTGAAAC
ITGB5	Forward primer	ACCAAGAGAGATTGCGTCGAGT
	Reverse primer	CAGCCTCCTGGTCATCTTTCA
GAPDH	Forward primer	GGAGCGAGATCCCTCCAAAAT
	Reverse primer	GGCTGTTGTCATACTTCTCATGG

Table S1. The primer sequences applied in this study.

Antibody	Item number		Company	Location
Anti-OCT4 antibody	ab19857		Abcam	Cambridge, UK
Anti-SOX2 antibody	ab97959		Abcam	Cambridge, UK
Anti-Nanog antibody	ab218524		Abcam	Cambridge, UK
Anti-TRA-1-60 antibody	ab16288		Abcam	Cambridge, UK
Anti-SSEA-4 antibody	MC-813-70		Invitrogen	California, USA
Anti- α SMA antibody	ab7817		Abcam	Cambridge, UK
Anti-GATA4 antibody	WL01293		VANCL Biotech	Jiangsu, China
Anti-PDGFR α antibody	Ab203491		Abcam	Cambridge, UK
Anti-βIII-Tubulin antibody	ab18207		Abcam	Cambridge, UK
Anti-PAX6 antibody	ab5790		Abcam	Cambridge, UK
Anti-NESTIN antibody	ab22035		Abcam	Cambridge, UK
Anti-FAK (phospho Y397) antibody	ab81298		Abcam	Cambridge, UK
Anti-FAK antibody	66258-1-Ig		Proteintech	Illinois, USA
Anti-ITGA6 antibody	27189-1-AP		Proteintech	Illinois, USA
Anti-CD44 antibody		338808	Biolegend	California, USA
Anti-CD73 antibody		344016	Biolegend	California, USA
Anti-CD90 antibody		328108	Biolegend	California, USA
Anti-CD105 antibody		560819	BD Biosciences	New Jersey, USA
Anti-CD166 antibody		343904	Biolegend	California, USA
Anti-CD34 antibody		555821	BD Biosciences	New Jersey, USA
Anti-CD45 antibody		555482	BD Biosciences	New Jersey, USA
Anti-HLA-DR antibody		555811	BD Biosciences	New Jersey, USA

 Table S2. The primary antibody applied in this study.

Figure	Statistical methods	Groups	p-value
Fig. 2E	Two-way ANOVA	iPSC:iMSC 1:3 -DPBS(Ca-Mg-) vs. iPSC:iMSC 1:3 +DPBS(Ca-Mg-)	p < 0.0001
		iPSC:iMSC 3:1 -DPBS(Ca-Mg-) vs. iPSC:iMSC 3:1 +DPBS(Ca-Mg-)	p < 0.0001
Fig. 2F	Two-way ANOVA	iPSCs vs. iMSCs	p=0.0001
		iPSCs vs. iPSC:iMSC 1:3 -DPBS(Ca-Mg-)	p=0.1439
		iPSCs vs. iPSC:iMSC 1:3 +DPBS(Ca-Mg-)	p=0.0001
		iPSCs vs. iPSC:iMSC 3:1 -DPBS(Ca-Mg-)	p=0.0009
		iPSCs vs. iPSC:iMSC 3:1 +DPBS(Ca-Mg-)	p=0.0001
		iMSCs vs. iPSC:iMSC 1:3 -DPBS(Ca-Mg-)	p=0.0062
		iMSCs vs. iPSC:iMSC 1:3 +DPBS(Ca-Mg-)	p>0.9999
		iMSCs vs. iPSC:iMSC 3:1 -DPBS(Ca-Mg-)	p=0.6932
		iMSCs vs. iPSC:iMSC 3:1 +DPBS(Ca-Mg-)	p>0.9999
Fig. 2G	One-way ANOVA	OCT4: iPSCs vs. iMSCs	p<0.0001
		OCT4: iPSCs vs. iPSC:iMSC 1:3 -DPBS(Ca-Mg-)	p<0.0001
		OCT4: iPSCs vs. iPSC:iMSC 3:1 -DPBS(Ca-Mg-)	p=0.0052
		OCT4: iPSCs vs. iPSC:iMSC 1:3 +DPBS(Ca-Mg-)	p<0.0001
		OCT4: iPSCs vs. iPSC:iMSC 3:1 +DPBS(Ca-Mg-)	p<0.0001
		OCT4: iMSCs vs. iPSC:iMSC 1:3 +DPBS(Ca-Mg-)	p=0.8844
		OCT4: iMSCs vs. iPSC:iMSC 3:1 +DPBS(Ca-Mg-)	p=0.0641
		SOX2: iPSCs vs. iMSCs	p<0.0001
		SOX2: iPSCs vs. iPSC:iMSC 1:3 -DPBS(Ca-Mg-)	p<0.0001
		SOX2: iPSCs vs. iPSC:iMSC 3:1 -DPBS(Ca-Mg-)	p=0.0002
		SOX2: iPSCs vs. iPSC:iMSC 1:3 +DPBS(Ca-Mg-)	p<0.0001
		SOX2: iPSCs vs. iPSC:iMSC 3:1 +DPBS(Ca-Mg-)	p<0.0001
		SOX2: iMSCs vs. iPSC:iMSC 1:3 +DPBS(Ca-Mg-)	p>0.9999
		SOX2: iMSCs vs. iPSC:iMSC 3:1 +DPBS(Ca-Mg-)	p=0.9973
		Nanog: iPSCs vs. iMSCs	p<0.0001
		Nanog: iPSCs vs. iPSC:iMSC 1:3 -DPBS(Ca-Mg-)	p<0.0001
		Nanog: iPSCs vs. iPSC:iMSC 3:1 -DPBS(Ca-Mg-)	p<0.0001
		Nanog: iPSCs vs. iPSC:iMSC 1:3 +DPBS(Ca-Mg-)	p<0.0001
		Nanog: iPSCs vs. iPSC:iMSC 3:1 +DPBS(Ca-Mg-)	p<0.0001
		Nanog: iMSCs vs. iPSC:iMSC 1:3 +DPBS(Ca-Mg-)	p>0.9999
		Nanog: iMSCs vs. iPSC:iMSC 3:1 +DPBS(Ca-Mg-)	p=0.9944
Fig. 5A	Student's t-test	OCT4: iPSCs vs. iMSCs	p=0.000028
		SOX2: iPSCs vs. iMSCs	p=0.000014

		ITGA1: iPSCs vs. iMSCs	p=0.000835
		ITGA2: iPSCs vs. iMSCs	p=0.001296
		ITGA3: iPSCs vs. iMSCs	p=0.00277
		ITGA5: iPSCs vs. iMSCs	p=0.00057
		ITGA6: iPSCs vs. iMSCs	p=0.005573
		ITGA7: iPSCs vs. iMSCs	p=0.003685
		ITGAV: iPSCs vs. iMSCs	p=0.001086
		ITGB1: iPSCs vs. iMSCs	p=0.009233
		ITGB5: iPSCs vs. iMSCs	p=0.245077
Fig. 5D	One-way ANOVA	pFAK: iMSCs vs. iMSCs+DPBS(Ca-Mg-)	p<0.0001
		pFAK: iMSCs vs. iPSCs	p=0.0001
		pFAK: iMSCs vs. iPSCs+DPBS(Ca-Mg-)	p<0.0001
		pFAK: iMSCs+DPBS(Ca-Mg-) vs. iPSCs	p=0.9961
		pFAK:iMSCs+DPBS(Ca-Mg-) vs. iPSCs+DPBS(Ca-Mg-)	p=0.0003
		pFAK: iPSCs vs. iPSCs+DPBS(Ca-Mg-)	p=0.0002
		FAK: iMSCs vs. iMSCs+DPBS(Ca-Mg-)	p=0.9867
		FAK: iMSCs vs. iPSCs	p<0.0001
		FAK: iMSCs vs. iPSCs+DPBS(Ca-Mg-)	p<0.0001
		FAK: iMSCs+DPBS(Ca-Mg-) vs. iPSCs	p<0.0001
		FAK: iMSCs+DPBS(Ca-Mg-) vs. iPSCs+DPBS(Ca-Mg-)	p=0.0002
		FAK: iPSCs vs. iPSCs+DPBS(Ca-Mg-)	p=0.5067
		p-FAK/FAK: iMSCs vs. iMSCs+DPBS(Ca-Mg-)	p=0.0005
		p-FAK/FAK: iMSCs vs. iPSCs	p<0.0001
		p-FAK/FAK: iMSCs vs. iPSCs+DPBS(Ca-Mg-)	p<0.0001
		p-FAK/FAK: iMSCs+DPBS(Ca-Mg-) vs. iPSCs	p=0.0054
		p-FAK/FAK: iMSCs+DPBS(Ca-Mg-) vs. iPSCs+DPBS(Ca-Mg-)	p<0.0001
		p-FAK/FAK: iPSCs vs. iPSCs+DPBS(Ca-Mg-)	p=0.0255
Fig. 5F	Student's t-test	ITGA6: iPSCs vs. iMSCs	p<0.0001
	One-way ANOVA	p-FAK: iPSCs:NC vs. iPSCs:15min	p=0.9146
		p-FAK: iPSCs:NC vs. iPSCs:30min	p=0.0005
		p-FAK: iPSCs:NC vs. iPSCs:60min	p=0.0003
		p-FAK: iPSCs:NC vs. iPSCs:120min	p<0.0001
		p-FAK: iPSCs:NC vs. iMSCs:NC	p<0.0001
		p-FAK: iPSCs:30min vs. iMSCs:30min	p=0.0276
		p-FAK: iMSCs:NC vs. iMSCs:15min	p=0.0002
		p-FAK: iMSCs:NC vs. iMSCs:30min	p<0.0001

		p-FAK: iMSCs:NC vs. iMSCs:60min	p<0.0001
		p-FAK: iMSCs:NC vs. iMSCs:120min	p<0.0001
		FAK: iPSCs:NC vs. iPSCs:15min	p>0.9999
		FAK: iPSCs:NC vs. iPSCs:30min	p>0.9999
		FAK: iPSCs:NC vs. iPSCs:60min	p>0.9999
		FAK: iPSCs:NC vs. iPSCs:120min	p>0.9999
		FAK: iPSCs:NC vs. iMSCs:NC	p=0.1265
		FAK: iPSCs:30min vs. iMSCs:30min	p=0.2222
		FAK: iMSCs:NC vs. iMSCs:15min	p>0.9999
		FAK: iMSCs:NC vs. iMSCs:30min	p>0.9999
		FAK: iMSCs:NC vs. iMSCs:60min	p>0.9999
		FAK: iMSCs:NC vs. iMSCs:120min	p>0.9999
		p-FAK/FAK: iPSCs:NC vs. iPSCs:15min	p>0.9999
		p-FAK/FAK: iPSCs:NC vs. iPSCs:30min	p=0.0927
		p-FAK/FAK: iPSCs:NC vs. iPSCs:60min	p=0.0487
		p-FAK/FAK: iPSCs:NC vs. iPSCs:120min	p=0.0185
		p-FAK/FAK: iPSCs:NC vs. iMSCs:NC	p<0.0001
		p-FAK/FAK: iPSCs:30min vs. iMSCs:30min	p=0.0003
		p-FAK/FAK: iMSCs:NC vs. iMSCs:15min	p=0.0499
		p-FAK/FAK: iMSCs:NC vs. iMSCs:30min	p<0.0001
		p-FAK/FAK: iMSCs:NC vs. iMSCs:60min	p<0.0001
		p-FAK/FAK: iMSCs:NC vs. iMSCs:120min	p<0.0001
Fig. 6C	Student's t-test	p-FAK: iMSCs vs. iMSCs+FAKi	p<0.0001
		FAK: iMSCs vs. iMSCs+FAKi	p<0.0001
		p-FAK/FAK: iMSCs vs. iMSCs+FAKi	p<0.0001
Fig. 6E	Student's t-test	p-FAK: iMSCs vs. iMSCs+FAKi	p=0.0001
		FAK: iMSCs vs. iMSCs+FAKi	p=0.0002
		p-FAK/FAK: iMSCs vs. iMSCs+FAKi	p=0.0016
Fig. 7B	Student's t-test	OCT4: iPSCs vs. shOCT4-iPSCs	p=0.000507
		SOX2: iPSCs vs. shOCT4-iPSCs	p=0.000537
		ITGA1: iPSCs vs. shOCT4-iPSCs	p<0.000001
		ITGA2: iPSCs vs. shOCT4-iPSCs	p=0.056988
		ITGA3: iPSCs vs. shOCT4-iPSCs	p=0.263471
		ITGA5: iPSCs vs. shOCT4-iPSCs	p=0.003651
		ITGA6: iPSCs vs. shOCT4-iPSCs	p=0.012659
		ITGA7: iPSCs vs. shOCT4-iPSCs	p=0.023149
			2
			5

		ITGAV: iPSCs vs. shOCT4-iPSCs	p=0.000446
		ITGB1: iPSCs vs. shOCT4-iPSCs	p=0.001042
		ITGB5: iPSCs vs. shOCT4-iPSCs	p=0.000003
Fig. 7E	Student's t-test	p-FAK: iPSCs vs. shOCT4-iPSCs	p<0.0001
		FAK: iPSCs vs. shOCT4-iPSCs	p<0.0001
		p-FAK/FAK: iPSCs vs. shOCT4-iPSCs	p<0.0001
Fig. 7G	Student's t-test	p-FAK: iPSCs vs. shOCT4-iPSCs	p=0.0004
		FAK: iPSCs vs. shOCT4-iPSCs	p=0.0629
		p-FAK/FAK: iPSCs vs. shOCT4-iPSCs	p=0.0015
		ITGA6: iPSCs vs. shOCT4-iPSCs	p=0.0151
Fig. 8C	One-way ANOVA	ITGA6: iPSCs vs. shITGA6-1	p<0.0001
		ITGA6: iPSCs vs. shITGA6-2	p<0.0001
		ITGA6: iPSCs vs. shITGA6-3	p=0.0004
		OCT4: iPSCs vs. shITGA6-1	p=0.0002
		OCT4:iPSCs vs. shITGA6-2	p<0.0001
		OCT4:iPSCs vs. shITGA6-3	p=0.0155
Fig. 8E	One-way ANOVA	p-FAK: iPSCs vs. shITGA6-1	p=0.005
		p-FAK: iPSCs vs. shITGA6-2	p=0.0067
		p-FAK: iPSCs vs. shITGA6-3	p=0.0336
		FAK: iPSCs vs. shITGA6-1	p=0.4296
		FAK:iPSCs vs. shITGA6-2	p=0.5666
		FAK:iPSCs vs. shITGA6-3	p>0.9999
		p-FAK/FAK: iPSCs vs. shITGA6-1	p=0.0007
		p-FAK/FAK: iPSCs vs. shITGA6-2	p=0.0008
		p-FAK/FAK: iPSCs vs. shITGA6-3	p=0.0007
		ITGA6: iPSCs vs. shITGA6-1	p=0.005
		ITGA6: iPSCs vs. shITGA6-2	p=0.0045
		ITGA6: iPSCs vs. shITGA6-3	p=0.0191
Fig. 8G	One-way ANOVA	p-FAK: iPSCs vs. shITGA6-1	p<0.0001
		p-FAK: iPSCs vs. shITGA6-2	p<0.0001
		p-FAK: iPSCs vs. shITGA6-3	p<0.0001
		FAK: iPSCs vs. shITGA6-1	p<0.0001
		FAK:iPSCs vs. shITGA6-2	p<0.0001
		FAK:iPSCs vs. shITGA6-3	p<0.0001
		p-FAK/FAK: iPSCs vs. shITGA6-1	p<0.0001
		p-FAK/FAK: iPSCs vs. shITGA6-2	p<0.0001

		p-FAK/FAK: iPSCs vs. shITGA6-3	p<0.0001
Fig. 8L	Student's t-test	p-FAK: iPSCs vs. ITGA6-blocked iPSCs	p=0.0002
		FAK: iPSCs vs. ITGA6-blocked iPSCs	p=0.7572
		p-FAK/FAK: iPSCs vs. ITGA6-blocked iPSCs	p=0.0005
Supplemental Fig. S1B	One-way ANOVA	OCT4: EPSCs-006-1 vs. HUVECs	p<0.0001
		OCT4: EPSCs-001-5 vs. HUVECs	p<0.0001
		OCT4: iPSCs-006-1 vs. HUVECs	p<0.0001
		OCT4: iPSCs-001-5 vs. HUVECs	p<0.0001
		SOX2: EPSCs-006-1 vs. HUVECs	p<0.0001
		SOX2: EPSCs-001-5 vs. HUVECs	p<0.0001
		SOX2: iPSCs-006-1 vs. HUVECs	p=0.0013
		SOX2: iPSCs-001-5 vs. HUVECs	p<0.0001
		Nanog: EPSCs-006-1 vs. HUVECs	p=0.0009
		Nanog: EPSCs-001-5 vs. HUVECs	p=0.0002
		Nanog: iPSCs-006-1 vs. HUVECs	p=0.0075
		Nanog: iPSCs-001-5 vs. HUVECs	p=0.0002
		Klf17: EPSCs-006-1 vs. HUVECs	p<0.0001
		Klf17: EPSCs-001-5 vs. HUVECs	p<0.0001
		Klf17: iPSCs-006-1 vs. HUVECs	p=0.0002
		Klf17: iPSCs-001-5 vs. HUVECs	p<0.0001
		Dppa3: EPSCs-006-1 vs. HUVECs	p<0.0001
		Dppa3: EPSCs-001-5 vs. HUVECs	p<0.0001
		Dppa3: iPSCs-006-1 vs. HUVECs	p<0.0001
		Dppa3: iPSCs-001-5 vs. HUVECs	p<0.0001
Supplemental Fig. S1J	Student's t-test	DLX3: iPSCs vs. iPSCs-Ebs	p=0.001899
		PAX6: iPSCs vs. iPSCs-Ebs	p=0.000166
		Brachyury: iPSCs vs. iPSCs-Ebs	p=0.014996
		SOX17: iPSCs vs. iPSCs-Ebs	p=0.00011
		OCT4: iPSCs vs. iPSCs-Ebs	p=0.0003
		SOX2: iPSCs vs. iPSCs-Ebs	p=0.000235
Supplemental Figure			
S4I	Two-way ANOVA	3h:+DPBS(Ca-Mg-) vs. 3h:-DPBS(Ca-Mg-)	p>0.9999
		6h:+DPBS(Ca-Mg-) vs. 6h:-DPBS(Ca-Mg-)	p>0.9999
		24h:+DPBS(Ca-Mg-) vs. 24h:-DPBS(Ca-Mg-)	p>0.9999
Supplemental Figure	Student's t-test	iMSCs-DPBS (Ca-Mg-) vs. iMSCs+DPBS (Ca-Mg-)	p=0.4406

Supplemental Figure			
S4L	Two-way ANOVA	0h:-DPBS (Ca-Mg-) vs. 0h:+DPBS (Ca-Mg-)	p>0.9999
		24h:-DPBS (Ca-Mg-) vs. 24h:+DPBS (Ca-Mg-)	p>0.9999
		48h:-DPBS (Ca-Mg-) vs. 48h:+DPBS (Ca-Mg-)	p>0.9999
		72h:-DPBS (Ca-Mg-) vs. 72h:+DPBS (Ca-Mg-)	p>0.9999
Supplemental Figure			0.000410
86B	Student's t-test	PAX6: IPSCs vs. Neuroectoderm	p=0.000418
		Brachyury: 1PSCs vs. Neuroectoderm	p<0.000001
		SOX17: iPSCs vs. Neuroectoderm	p=0.527345
		OCT4: iPSCs vs. Neuroectoderm	p=0.000033
		TTGA6: 1PSCs vs. Neuroectoderm	p=0.0017
		PAX6: 1PSCs vs. Mesendoderm	p<0.000001
		Brachyury: iPSCs vs. Mesendoderm	p=0.000265
		SOX17: iPSCs vs. Mesendoderm	p=0.00068
		OCT4: iPSCs vs. Mesendoderm	p=0.000002
		ITGA6: iPSCs vs. Mesendoderm	p=0.000051
Supplemental Figure	Two way ANOVA	nFAK: iDSCs vs. Neuroestaderm	n <0.0001
501	I wo-way ANOVA	pFAK: II SCS VS. Neuroccioucini	p = 0.0001
		FAK : iDSCs vs. Neuropertoderm	p=0.0009
		FAK: iPSCs vs. Mesendoderm	p<0.0001
		n FAK/FAK: iDSCs vs. Neuroectoderm	p≤0.0001
		p-FAK/FAK: iDSCs.vs. Mesendoderm	p = 0.0018
Supplemental Figure		p-1 AK/1 AK. II Ses vs. Wesendoderin	p=0.0018
Suppremental Figure S6H	Two-way ANOVA	pFAK: iPSCs vs. Neuroectoderm	p=0.0076
	5	pFAK: iPSCs vs. Mesendoderm	p=0.0006
		FAK: iPSCs vs. Neuroectoderm	p>0.9999
		FAK: iPSCs vs. Mesendoderm	p=0.6667
		p-FAK/FAK: iPSCs vs. Neuroectoderm	p=0.0432
		p-FAK/FAK: iPSCs vs. Mesendoderm	p=0.0132
		ITGA6: iPSCs vs. Neuroectoderm	p<0.0001
		ITGA6: iPSCs vs. Mesendoderm	p=0.0005
Supplemental Figure			1
S7B	One-way ANOVA	iPSCs vs. iPSCs+10 nM FAKi	p=0.9923
		iPSCs vs. iPSCs+100 nM FAKi	p=0.0028
		iPSCs vs. iPSCs+1 uM FAKi	p=0.0478

Supplemental Figure Student's t-test iPSCs vs. iPSCs-FAKi

p<0.0001

Table S3. Statistical methods and p-values used in this study