

Supplementary Information for

**Endothelial cell-derived SDF-1 α elicits stemness traits of glioblastoma
via dual-regulation of GLI1**

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Running title: Stemness traits regulation by endothelial cells

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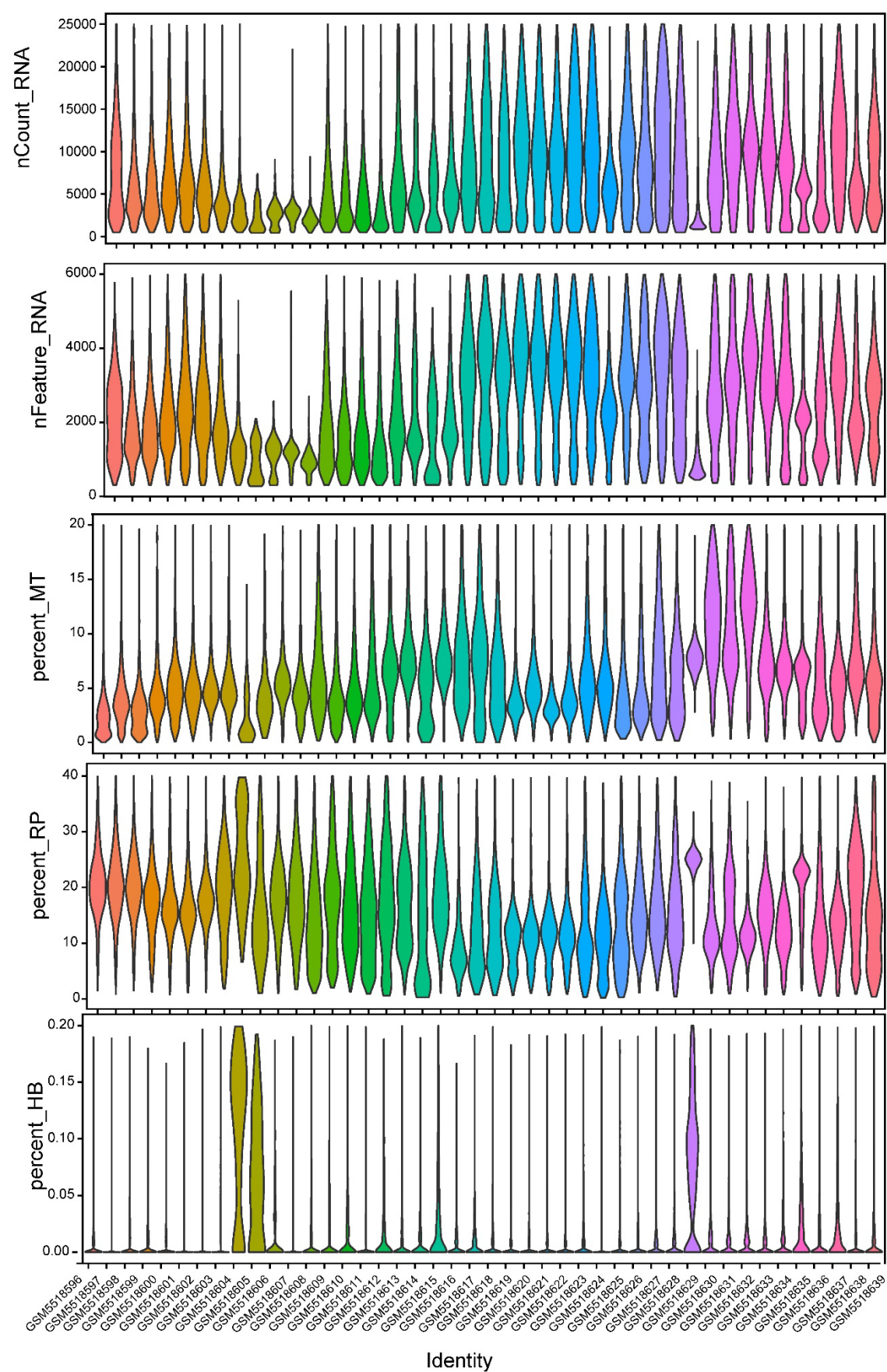


Figure S1 Filtering of low-quality cells.

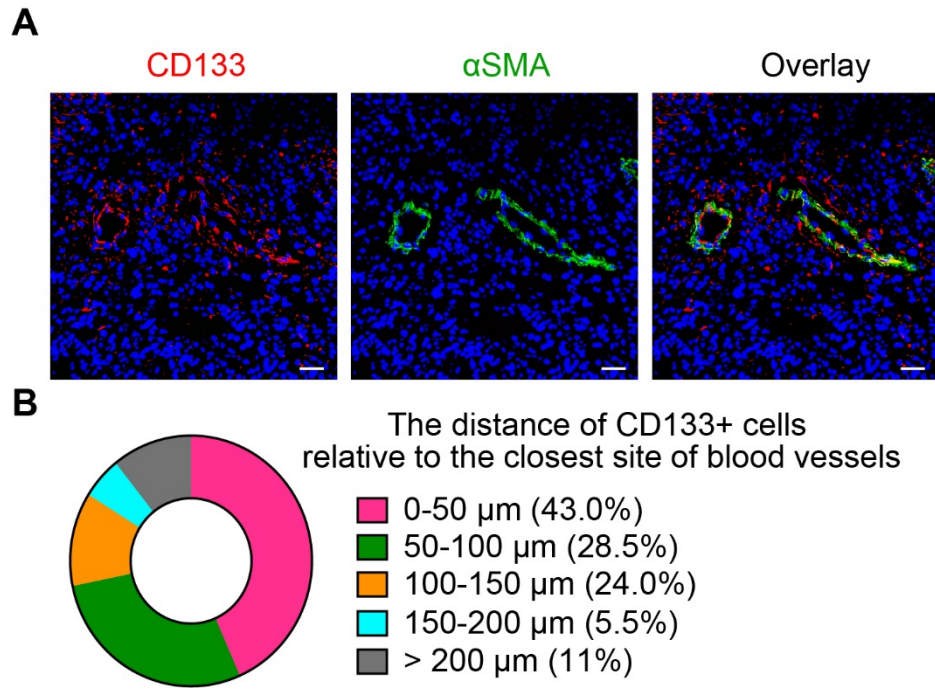


Figure S2 Spatial tissue localization of GSCs relative to microvessels.

(A) Immunofluorescent staining image illustrating co-localization of GSC marker CD133 (red) and vascular pericyte marker α SMA (green) in human GBM tissue. Scale bar = 50 μ m. (B) The overall perivascular distribution of GSCs was defined by calculating the distances between CD133⁺ cells and the closest sites in the blood vessel ring, data were collected from analyzing 200 GSCs.

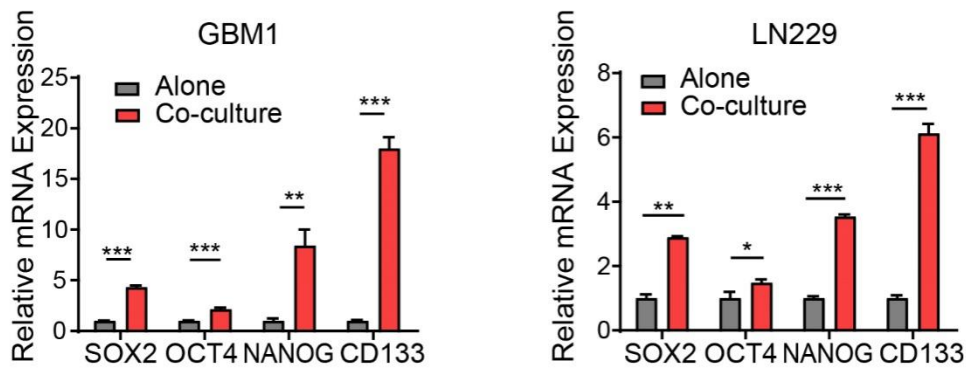
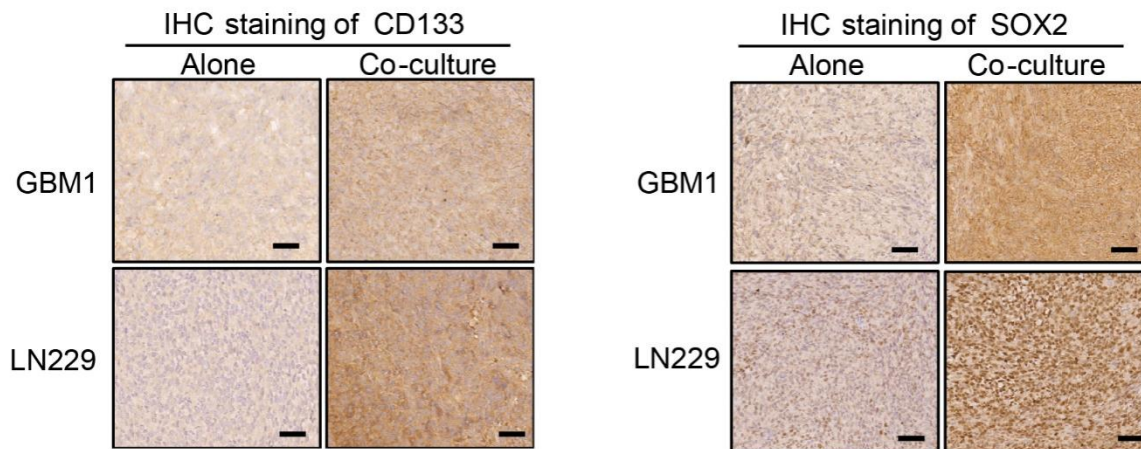
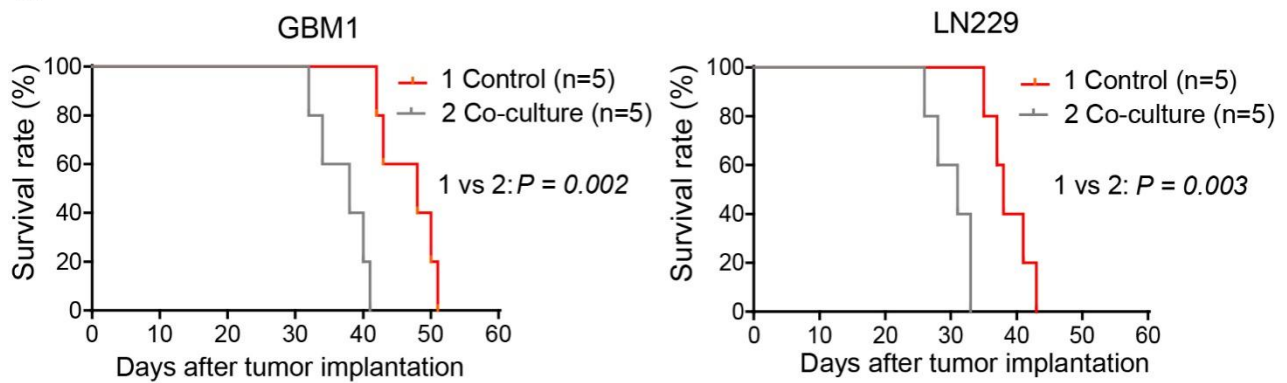
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Figure S3 ECs induce the expression of stemness markers and traits in co-cultured non-GSC populations.

(A) qRT-PCR assay tested the expression of SOX2, OCT4, NANOG, and CD133 in ECs co-cultured and control GBM cells. (B) Representative images depicting IHC staining of CD133 and SOX2 in xenografts from mice bearing GBM cells, both co-cultured with ECs and in control conditions. Scale

bar = 40 μ m. (C) Kaplan-Meier survival analysis of the mice bearing xenograft tumors of GBM cells, comparing those with prior co-culture with ECs to those with GBM cells alone. Data are means \pm SD. *, $P < 0.05$; **, $P < 0.01$; ***, $P < 0.001$.

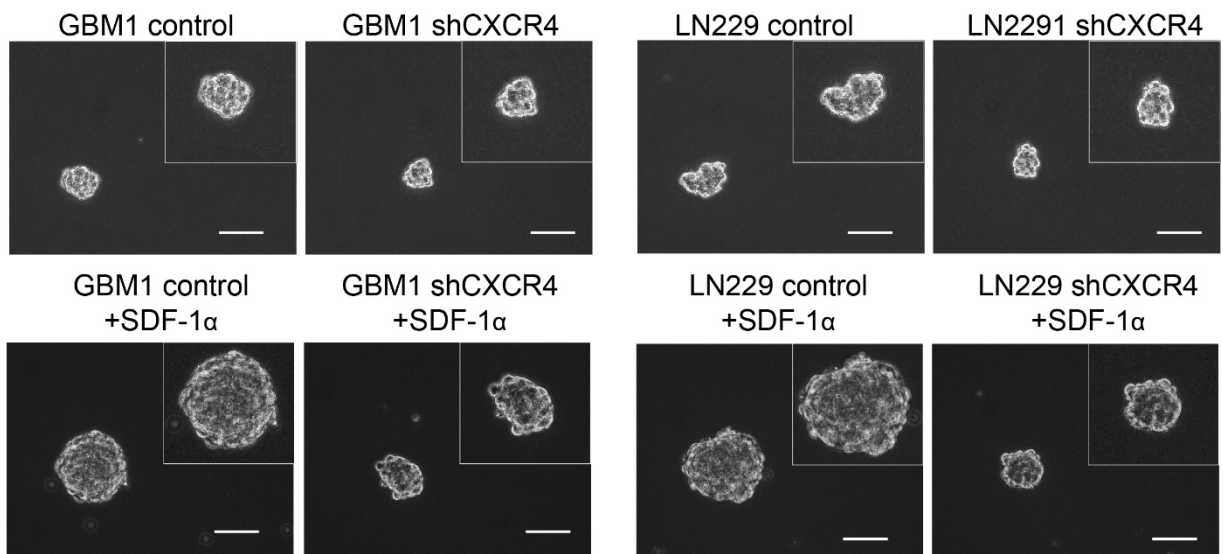
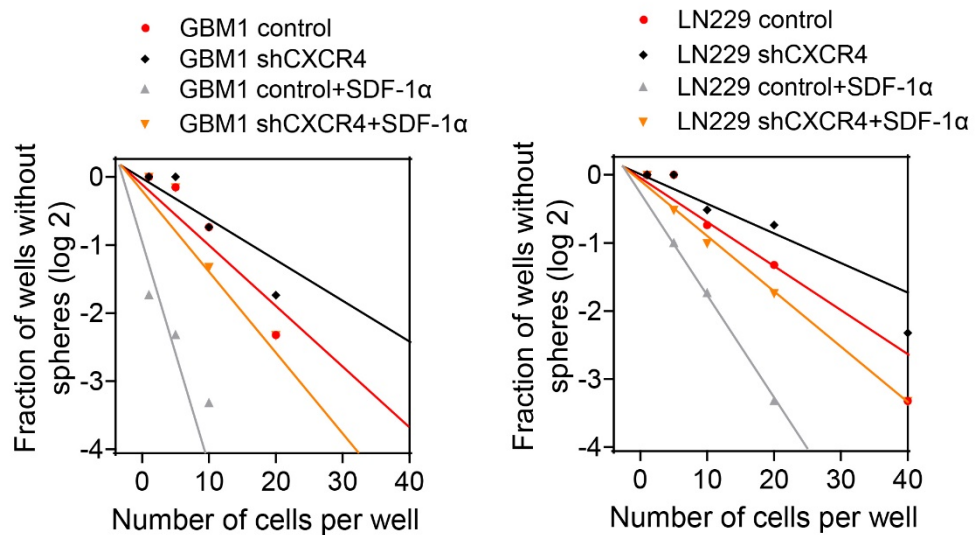
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Figure S4 Disrupting CXCR4 attenuated the SDF-1 α dependent stemness in GBM cells.

(A-B) Limiting dilution sphere-forming assays conducted on GBM1 (A) and LN229 (B) cells treated with either SDF-1 α , shCXCR4, or a combination of both, along with representative images of the resulting tumor spheres.

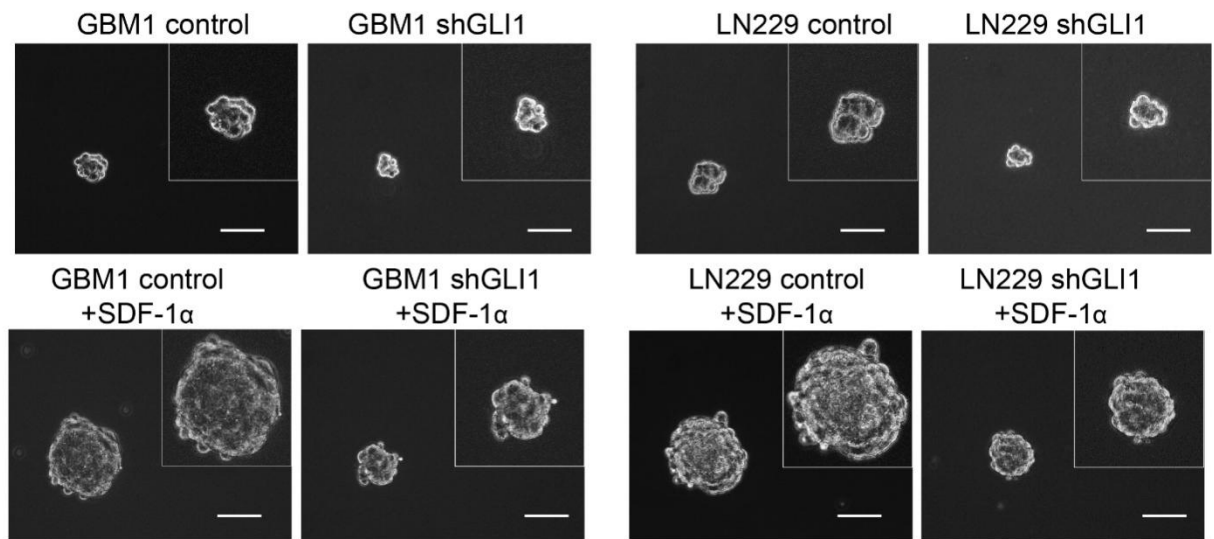
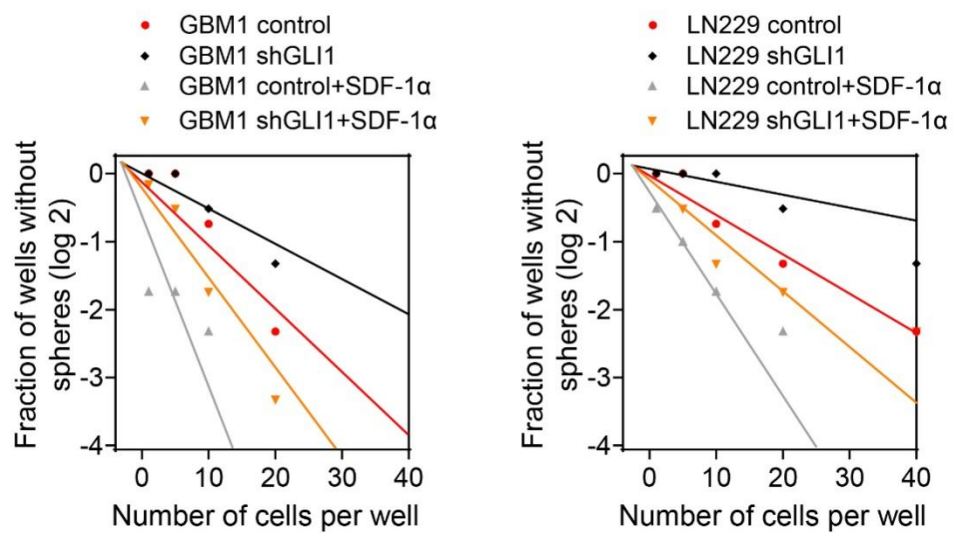
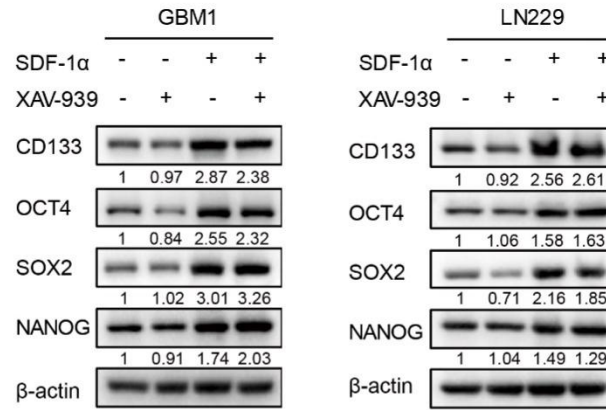
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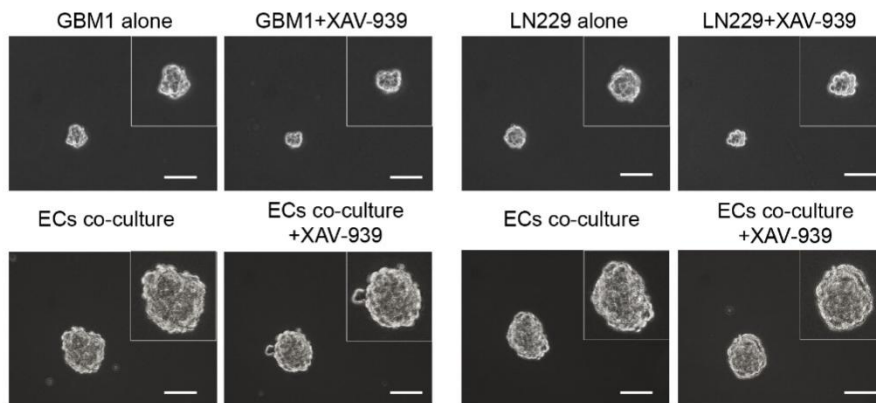
Figure S5 Disrupting GLI1 attenuates SDF-1 α dependent stemness in GBM cells.

(A-B) Limiting dilution sphere-forming assays conducted on GBM1 (A) and LN229 (B) cells treated with either SDF-1 α , shGLI1, or a combination of both, along with representative images of the resulting tumor spheres.

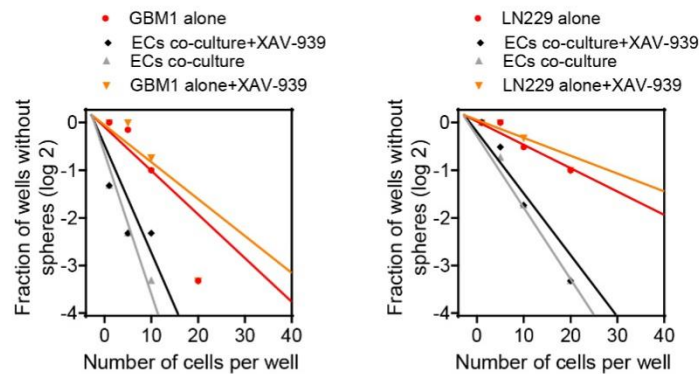
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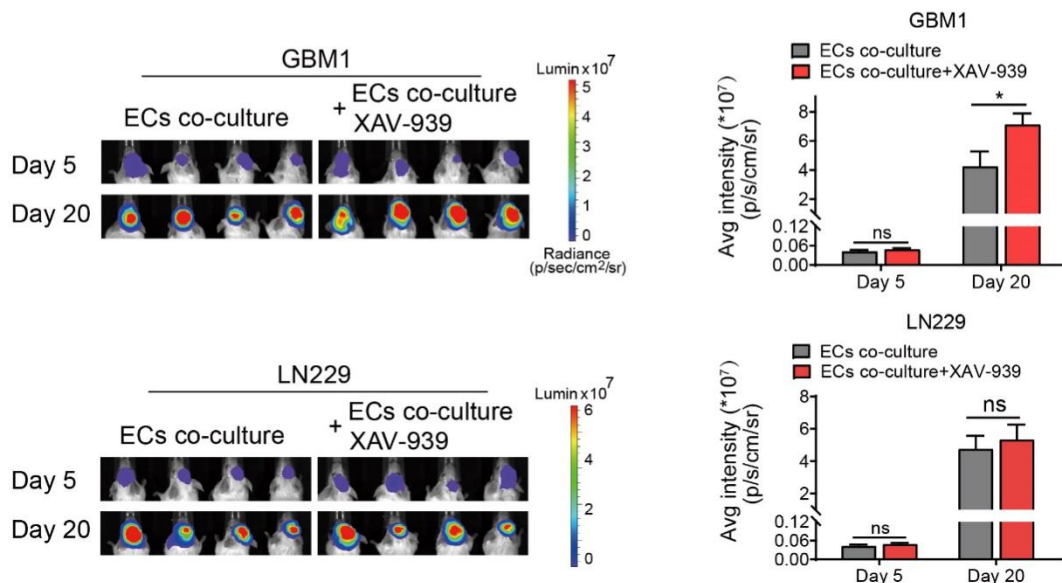


Figure S6 XAV-939 demonstrated few or none inhibitory effect on SDF-1 α dependent stemness maintaining in GBM cells.

(A) Western blot assay tested the expression of SOX2, OCT4, NANOG, and CD133 in GBM cells treated with SDF-1 α /XAV-939 alone or together. **(B-C)** Limiting dilution sphere-forming assays were performed on ECs co-cultured with control GBM cells, with and without XAV-939 treatment, accompanied by representative images of tumor spheres. **(D)** *In vivo* bioluminescent imaging and quantification of xenograft tumor burdens were carried out in mice bearing ECs-GBM cells from the co-cultured group, with or without XAV-939 treatment. Data are means \pm SD. ns: no significance; *, $P < 0.05$.

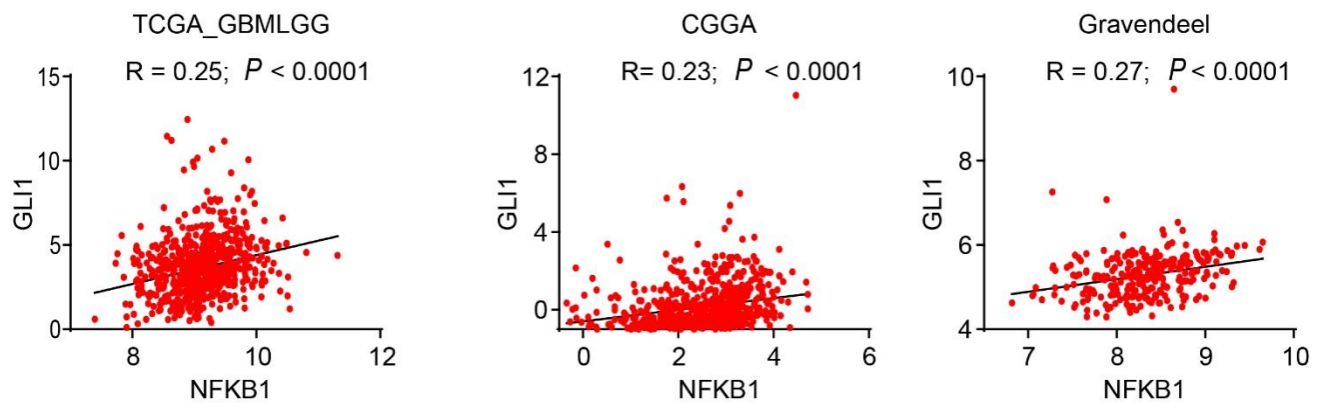


Figure S7 The correlation between expressions of NFKB1 and GLI1 in GBM databases.

The correlation plots showing relevance of NFKB1 to GLI1 from TCGA_GBMLGG, CGGA and Gravendeel datasets.

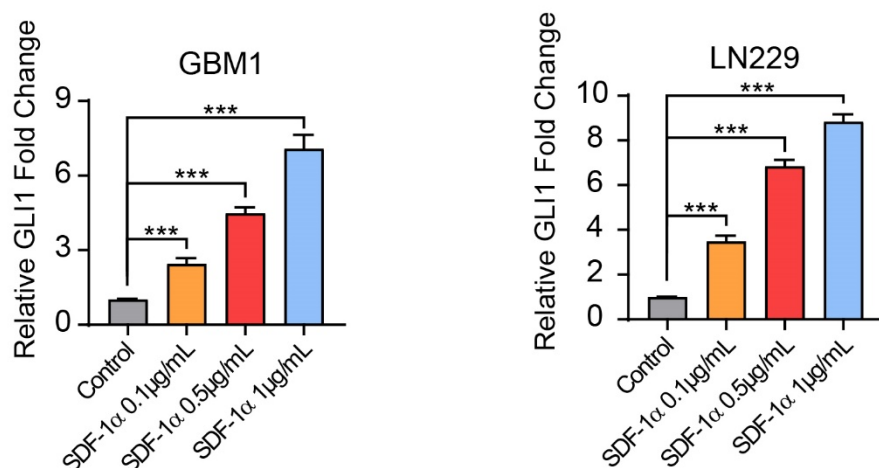
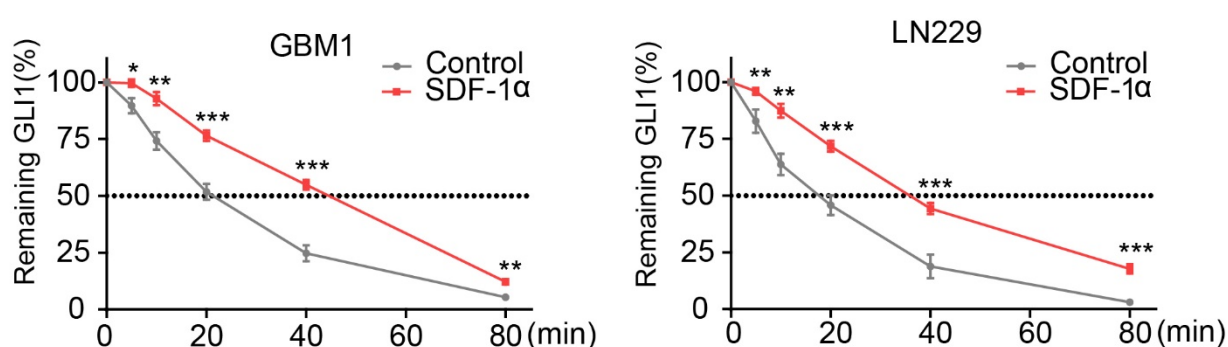
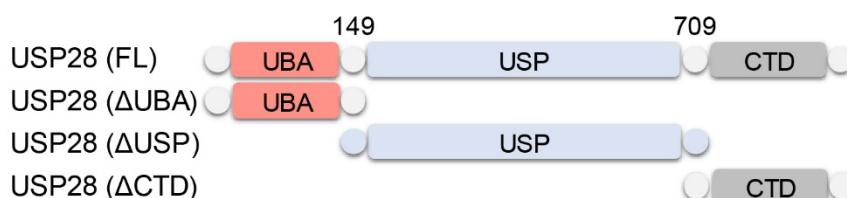
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Figure S8 SDF-1α activates USP28 to regulate the turnover of GLI1 protein.

(A) qRT-PCR assay tested the expression of GLI1 in GBM cells treated with different concentrations of SDF-1α recombinant protein. (B) The halftime of GLI1 in GBM cells treated with SDF-1α recombinant protein or not at the indicated time. (C) Schematic representation of USP28 truncation mutants. (D) Schematic representation of USP28 point mutant

Table S1 The sequences of mRNA primers used in qPCR.

Gene	Forward primer (5' to 3')	Reverse primer (5' to 3')
β -actin	AGATGTGGATCAGCAAGCA	GCGCAAGTTAGGTTTTGTCA
SOX2	ACATGAACGGCTGGAGCAACG	CTGCGAGCTGGTCATGGAGTTG
CD133	TCGCCTCAAGCCAGCCTCAG	ACTTGGTGCCTCCTGCCTCAG
OCT4	CTGGGTTGATCCTCGGACCT	CCATCGGAGTTGCTCTCCA
NANOG	TTTGTGGGCCTGAAGAAAAC	AGGGCTGTCCTGAATAAGCAG
GLI1	AGCGTGAGCCTGAATCTGTG	CAGCATGTACTGGGCTTTGAA