

Figure S1. *Pcyt2* is expressed mainly in hepatocytes in the mouse liver and was overexpressed by PCYT2-expressing AAVs. (A) The single-cell sequencing database Tabula Muris shows the expression pattern of Pcyt2 in diverse liver cells. (B) Hepatocytes and non-parenchymal cells (NPC) were isolated from mouse livers. Western blot analysis of the protein levels of PCYT2. n = 4 mice per group. (C) Bubble diagram of PE levels in isolated hepatocytes at 1 day after surgery. n = 8 mice per group. (D) Eight-week-old mice were administered AAV-Pcyt2-flags bearing the TBG promoter or control AAV through tail vein injection. Western blot analysis of the protein levels of PCYT2 in the liver. β-Actin was used as the internal control. n = 6 mice per group. Data present the mean  $\pm$  SEM. \*p < 0.05. Exo, exogenous; Endo, endogenous.

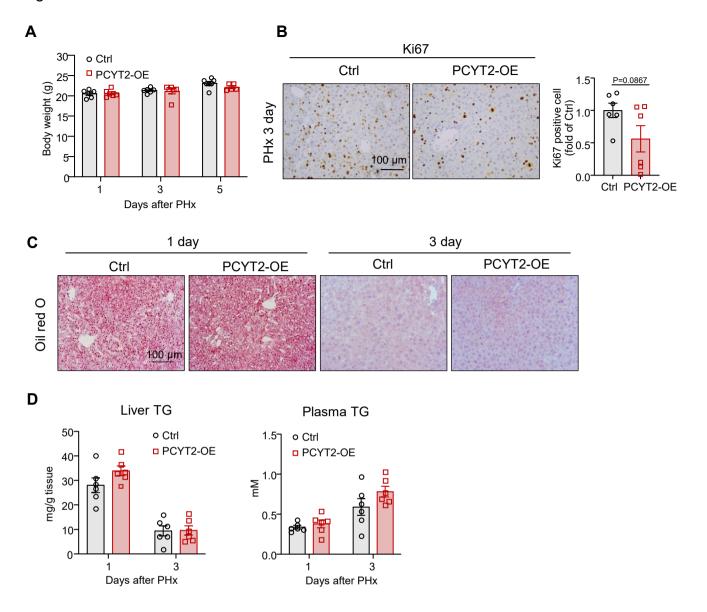


Figure S2. Effects of PCYT2 overexpression on Ki67-positive cell counts and lipid accumulation in the liver after 70% partial hepatectomy (PHx). Eight-week-old mice were administered AAV-PCYT2-flags bearing the TBG promoter or control AAV through tail vein injection. After 10 days, the mice were subjected to 70% PHx, and the tissues were harvested for analysis at the indicated time points. (A) Body weights of the mice. (B) Immunohistochemical staining of Ki67 in liver sections from mice at 3 days (scale bar =  $100 \mu m$ ). (C) Oil red O staining of the liver (scale bar =  $100 \mu m$ ). (D) Plasma and liver triglyceride (TG) contents. n = 5-7 mice per group. Data present the mean  $\pm$  SEM. \*p < 0.05. TG, triglyceride.

Figure S3

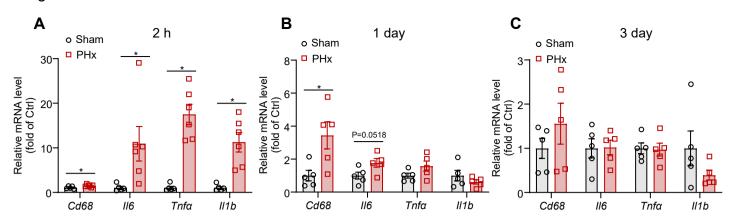


Figure S3. The expression of inflammatory factors in the liver after partial hepatectomy (PHx). Eight-week-old mice were subjected to 70% PHx or sham surgery. qPCR analysis of the mRNA levels of inflammatory factors in mouse livers at 2 h (A), 1 day (B) or 3 days postoperatively (C). n = 5-6 mice per group. Data present the mean  $\pm$  SEM. \*p < 0.05.

## Figure S4

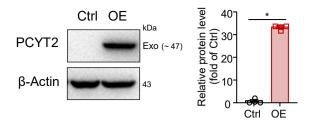


Figure S4. PCYT2 was overexpressed in primary mouse hepatocytes via adenovirus.

Primary mouse hepatocytes were treated with Ad-PCYT2 or Ad-Ctrl for 48 h. Western blot analysis of the protein levels of PCYT2.  $\beta$ -Actin was used as the internal control. n = 4 independent experiments. Data present the mean  $\pm$  SEM. \*p < 0.05. OE, PCYT2 overexpression. Exo, exogenous.

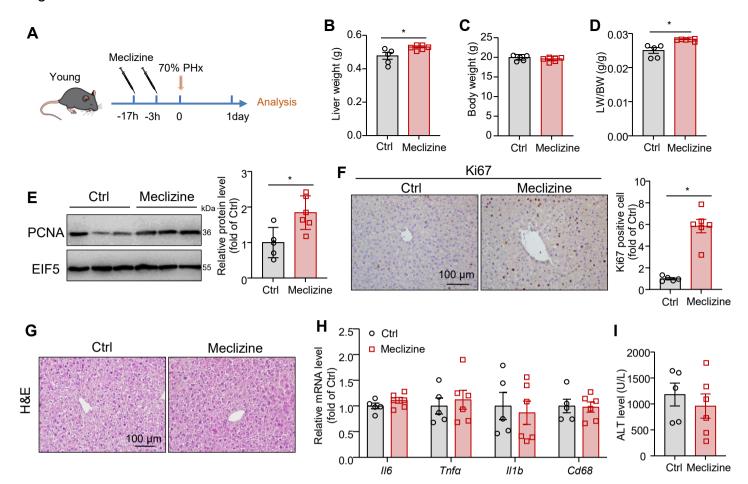


Figure S5. Inhibition of PCYT2 by meclizine promoted liver regeneration in eight-week-old mice. Eight-week-old mice were administered 60 mg/kg meclizine or vehicle 17 h and 3 h before 70% PHx, and then the tissues were harvested for analysis at 1 day after surgery (A). (B) Liver weights, (C) body weights, (D) and liver-to-body weight ratios of the mice. (E) Western blot analysis of the protein levels of PCNA in the liver. EIF5 was used as the internal control. (F) Immunohistochemical staining of Ki67 in liver sections (scale bar =  $100 \,\mu\text{m}$ ) and quantification of Ki67-positive cells. (G) H&E staining of liver sections (scale bar =  $100 \,\mu\text{m}$ ). (H) qPCR analysis of the mRNA levels of inflammatory factors in the liver. (I) Plasma ALT levels. n = 5 in the control group, n = 6 in the meclizine group. Data present the mean  $\pm$  SEM. \*p < 0.05. LW/BW, liver-to-body weight ratios.

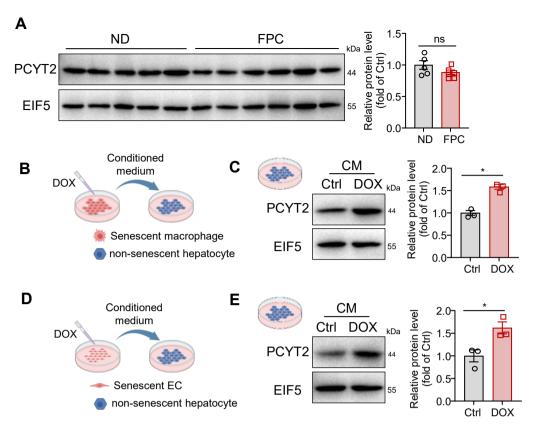


Figure S6. The protein level of PCYT2 in MASH mouse livers and in hepatocytes

treated with conditioned medium from senescent cells. (A) Eight-week-old mice were fed a diet rich in fructose, palmitate, and cholesterol (FPC) or a normal diet (ND) for 24 weeks. Western blot analysis of the protein levels of PCYT2. EIF5 was used as the internal control. n = 5–6 mice per group. (B–C) Macrophages (RAW264.7 cells) were treated with doxorubicin (DOX) to induce senescence. The conditioned medium was collected to treat non-senescent HepG2 cells. The protein levels of PCYT2 in non-senescent hepatocytes treated with the conditioned medium; EIF5 was used as the internal control. (D–E) Endothelial cells (EA.hy926 cells) were treated with DOX to induce senescence. The conditioned medium was collected to treat non-senescent HepG2 cells. The protein levels of PCYT2 in non-senescent hepatocytes treated with conditioned medium; EIF5 was used as the internal control. (C and E) n = 3 independent experiments. Data present the mean ± SEM. \*p < 0.05. EC, Endothelial cells.

Figure S7

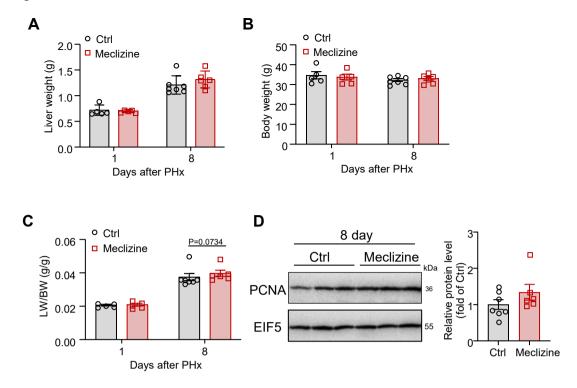


Figure S7. Liver weights of meclizine-treated middle-aged mice after 70% partial hepatectomy (PHx). Ten-month-old mice were administered 60 mg/kg meclizine or vehicle 17 h and 3 h before 70% PHx, and then the tissues were harvested for analysis at 1 day (n = 5 mice per group) or 8 days (n = 7 mice in the control group; n = 6 mice in the meclizine group) postoperatively. (A) Liver weights, (B) body weights, (C) and liver-to-body weight ratios of the mice. (D) Western blot analysis of the protein levels of PCNA in mouse livers at 8 days after surgery. EIF5 was used as the internal control. Data present the mean  $\pm$  SEM. \*p < 0.05. LW/BW, liver-to-body weight ratios.

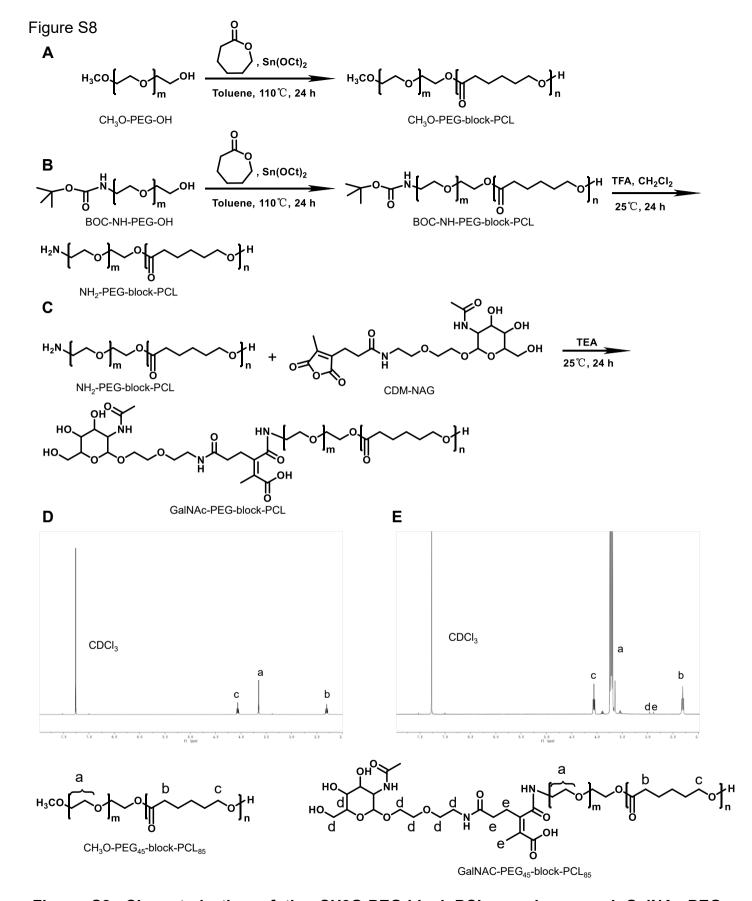


Figure S8. Characterization of the CH3O-PEG-block-PCL copolymer and GalNAc-PEG-block-PCL copolymer. Synthesis routes for the (A) CH3O-PEG-block-PCL copolymer and (B, C) GalNAc-PEG-block-PCL copolymer. (D) 1H-NMR spectrum of the CH3O-PEG-block-PCL copolymer. (E) 1H-NMR spectrum of the GalNAc-PEG-block-PCL copolymer.

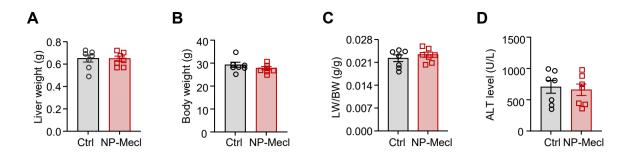


Figure S9. Liver weights of NP-meclizine-treated middle-aged mice after 70% partial hepatectomy (PHx). Ten-month-old mice were administered 150  $\mu$ g NP-meclizine (containing 25  $\mu$ g meclizine) or PBS 17 h and 6 h before 70% PHx, and then the tissues were harvested for analysis 1 day postoperatively (n = 7 mice per group). (A) Liver weights, (B) body weights, (C) and liver-to-body weight ratios. (D) Plasma ALT levels. Data present the mean  $\pm$  SEM. \*p <0.05. LW/BW, liver-to-body weight ratios; NP-Mecl, NP-meclizine.

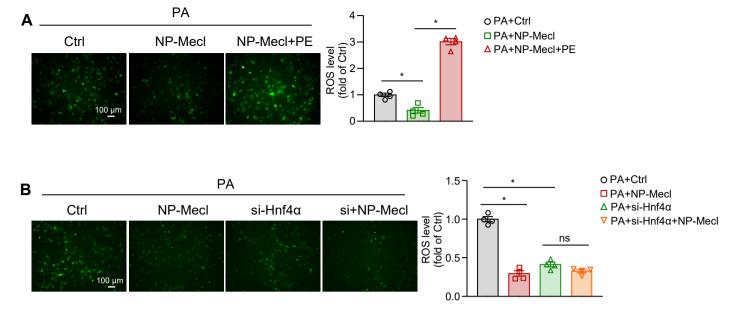
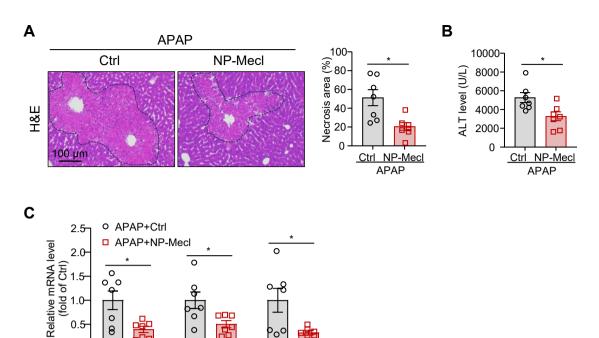


Figure S10. Influence of PE or HNF4α knockdown on the NP-meclizine-mediated ROS reduction in fatty acid-overloaded hepatocytes. (A) Primary mouse hepatocytes were treated with 3.75 μg/mL NP-meclizine (containing 1 μM meclizine) for 17 h, and then the hepatocytes were co-treated with PA (100 μM) and PE (10 μM) for another 24 h. (B) Primary mouse hepatocytes were transfected with si-Hnf4α or si-NC; 24 h later, the hepatocytes were treated with 3.75 μg/mL NP-meclizine (containing 1 μM meclizine) for 17 h, and then the hepatocytes were co-treated with PA (100 μM) for another 24 h. (A and B) Representative images and quantitative analysis of ROS in hepatocytes (scale bar = 100 μm). n = 4 independent experiments. Data present the mean  $\pm$  SEM. \*p < 0.05. NP-Mecl, NP-meclizine.

0.5 0.0

116

Tnfα



II1b

Figure S11. NP-meclizine attenuated acetaminophen (APAP)-induced liver injury.

Eight-week-old mice were administered 180 µg of NP-meclizine (containing 25 µg of meclizine) or PBS 17 h and 6 h before being treated with APAP for 24 h. (A) H&E staining of liver sections and the quantitative analysis of liver necrosis areas (scale bar = 100 µm). (B) Plasma ALT levels. (C) qPCR analysis of the mRNA levels of inflammatory factors in mouse livers. n = 7 mice in the control group. Data present the mean ± SEM. \*p < 0.05. NP-Mecl, NP-meclizine.

Table S1. List of oligonucleotide primer pairs used in qPCR

Target gene	Sense primer	Antisense primer
18S (h/m)	5'-GGAAGGGCACCACCAGGAGT-3'	5'-TGCAGCCCCGGACATCTAAG-3'
Pcyt2 (m)	5'-AGGCTGGGAGGTACAGAGAG-3'	5'-ATCTCCTGGCTGCTGATG-3'
Pemt (m)	5'-CTGTGGAGGCTTCGGCAATA-3'	5'-AACCTAGGAATGCAAGGCCC-3'
Cept (m)	5'-CAGCTTCTCTCAGGAACCGT-3'	5'-GCCACAGTACAAAGTGCAGG-3'
Pisd (m)	5'-CTAGTCACTGGGAGGTGTCC-3'	5'-AGCTTACGCCGGAAGAACTC-3'
Etnk1 (m)	5'-GTCAATCAGTTTGCGTTGGCT-3'	5'-CCGCATACCCAAGGAAGTCG-3'
Hnf4α (m)	5'-GGGGTTCCTGCAGATCACAT-3'	5'-TCCCAGAGATGGGAGAGGTG-3'
II6 (m)	5'-CACTTCACAAGTCGGAGGCT-3'	5'-GCCACTCCTTCTGTGACTCC-3'
II1b (m)	5'-CTCTGTGACTCATGGGATG-3'	5'-CTTCTTCTTTGGGTATTGC-3'
Tnfα (m)	5'-CTGTGAAGGGAATGGGTGTT-3'	5'-CAGGGAAGAATCTGGAAAGGTC-3'
Cd68 (m)	5'-CCCAAGGAACAGAGGAAG-3'	5'-GTGGCAGGGTTATGAGTG-3'
Hnf4α (h)	5'-TGCGACTCTCCAAAACCCTC-3'	5'-ATTGCCCATCGTCAACACCT-3'

m: Mus musculus; h: Homo sapiens