Supplementary Materials

An organolutetium nanosensitizer synergizes with PARP inhibition to unleash STING-mediated immunity for low-dose radioimmunotherapy

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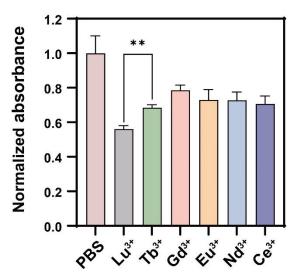


Figure S1. Normalized absorbance of DPPH radicals in the presence of various lanthanide ions following 6 Gy X-ray irradiation (n = 3). Data are presented as mean \pm SD; **P < 0.01.

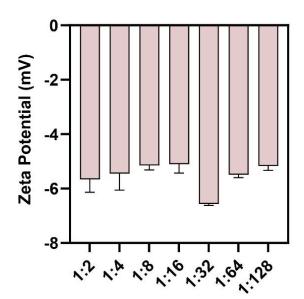


Figure S2. Zeta potential of LSP nanoparticles prepared at varying molar ratios of Lu³⁺/Sal⁻ (n = 3). Data are presented as mean \pm SD.

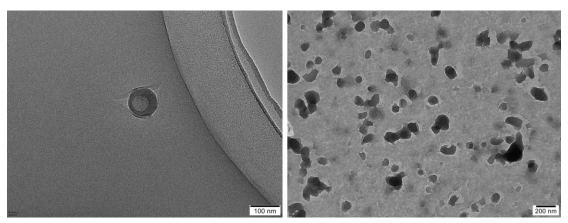


Figure S3. Representative TEM images of LSP nanoparticles.

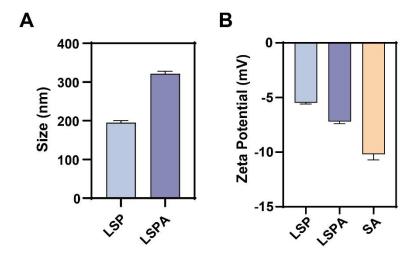


Figure S4. (A) Hydrodynamic size distribution and **(B)** zeta potential of LSP and LSPA (n = 3). Data are presented as mean \pm SD.

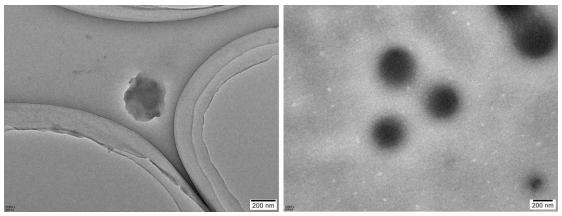


Figure S5. Representative TEM images of LSPA nanoparticles.

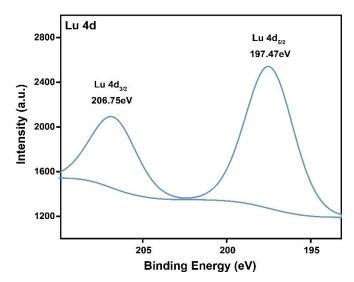


Figure S6. High-resolution XPS spectrum of Lu³⁺ in LSPA nanoparticles.

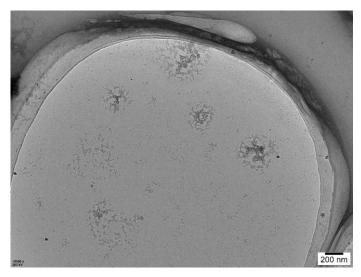


Figure S7. Representative TEM images of LSPA nanoparticle disassembly under pH 4.8 conditions.

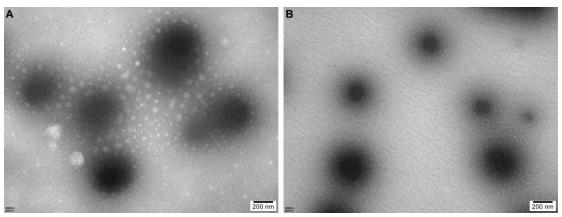


Figure S8. Representative TEM images of LSPA nanoparticles after incubation for 24 h in cell culture media containing 10% FBS (A) or 10 μ g mL⁻¹ heparin (B).

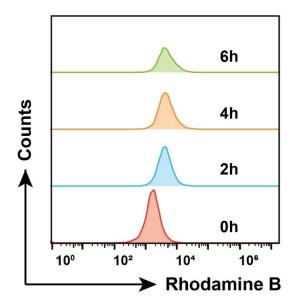


Figure S9. Flow cytometric analysis of 4T1 cells incubated with Rhodamine B-labeled LSPA nanoparticles (600 μg mL⁻¹) at various time points.

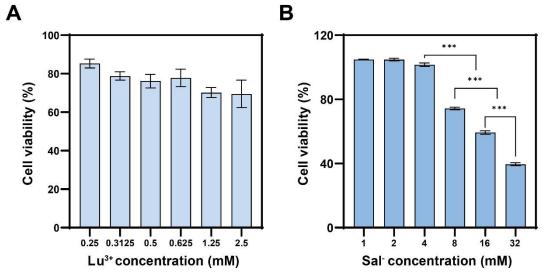


Figure S10. Cytotoxicity of 4T1 cells treated with (**A**) Lu³⁺ (0.25–2.5 mM) and (**B**) Sal⁻ (1–32 mM) (n = 3). Data are presented as mean \pm SD; ***P < 0.001.

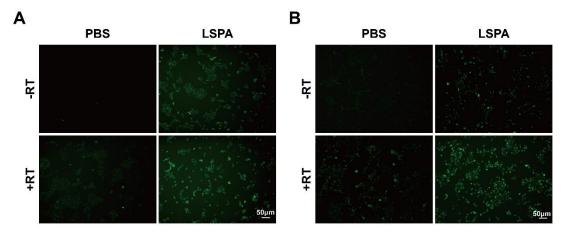


Figure S11. Confocal laser scanning microscopy (CLSM) images of 4T1 cells stained with **(A)** BBoxiProbe O22 probe (for 1O_2 detection) and **(B)** BBoxiProbe O27 probe (for •OH detection) after treatment with PBS (control) or LSPA, with or without 6 Gy X-ray irradiation (RT). Scale bar: 50 μm.

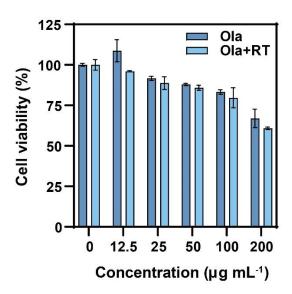


Figure S12. Cytotoxicity of 4T1 cells treated with increasing concentrations of Olaparib (Ola), with or without 6 Gy X-ray irradiation (RT, n = 3). Data are presented as mean \pm SD.

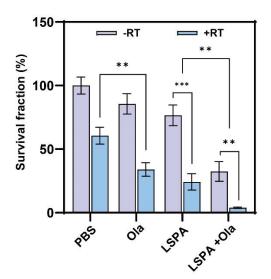


Figure S1. Clonogenic survival fraction (%) of 4T1 cells following the indicated treatments (n = 3). RT: 6 Gy X-ray irradiation; Ola: Olaparib. Data are presented as mean \pm SD; **P < 0.01; ***P < 0.001.

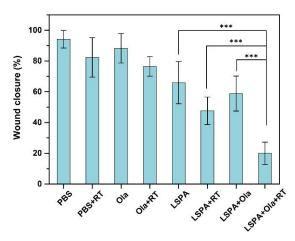


Figure S2. Percentages of wound closure in 4T1 cells following the indicated treatments (n = 3). RT: 6 Gy X-ray irradiation; Ola: Olaparib. Data are presented as mean \pm SD; ***P < 0.001.

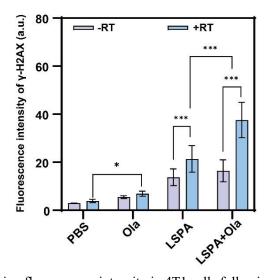


Figure S3. γ -H2AX staining fluorescence intensity in 4T1 cells following the indicated treatments (n = 3, fluorescence intensity per nucleus). RT: 6 Gy X-ray irradiation; Ola: Olaparib. Data are presented as mean \pm SD; *P < 0.05; ***P < 0.001.

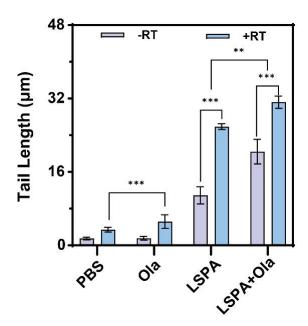


Figure S4. Quantification of DNA damage by comet tail length in 4T1 cells following the indicated treatments (n = 3). RT: 6 Gy X-ray irradiation; Ola: Olaparib. Data are presented as mean \pm SD; **P < 0.01; ***P < 0.001.

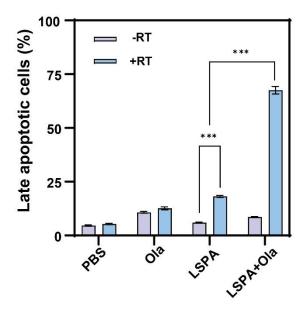


Figure S5. Late apoptosis (Annexin V⁺ PI⁺) quantification in 4T1 cells following the indicated treatments (n = 3). RT: 6 Gy X-ray irradiation; Ola: Olaparib. Data are presented as mean \pm SD; ***P < 0.001.

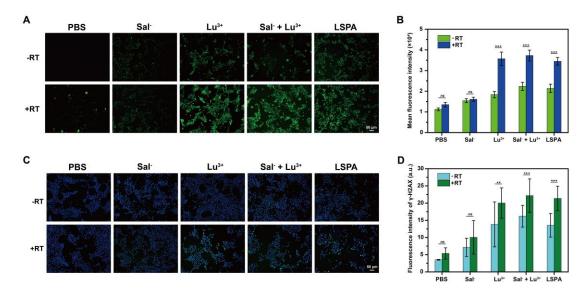


Figure S18. Verification of the *in vitro* radiosensitization effect of LSPA nanoparticles. **(A)** CLSM images of DCFH-DA-stained 4T1 cells (ROS imaging) following the indicated treatments with or without RT. Scale bar: 50 μm. **(B)** Corresponding fluorescence intensities of DCFH-DA-stained 4T1 cells treated with various treatments with or without RT (n = 3). **(C)** γ-H2AX immunofluorescence staining (a marker for DSBs) of 4T1 cells following the indicated treatments (DAPI counterstaining). Scale bar: 50 μm. **(D)** Corresponding γ-H2AX staining fluorescence intensities in 4T1 cells following the indicated treatments (n = 3, fluorescence intensity per nucleus). RT: 6 Gy X-ray irradiation. Data are presented as mean ± SD; ns: no significance; **P < 0.01; ***P < 0.001.

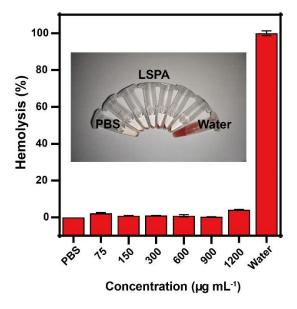


Figure S6. Hemocompatibility assessment of LSPA. Hemolysis rates (%) of red blood cells (RBCs) incubated with LSPA (75–1200 μ g mL⁻¹) at 4 °C for 3 h (n = 3). Data are presented as mean \pm SD.

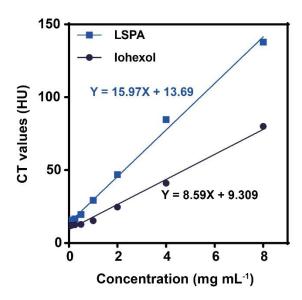


Figure S20. Contrast performance of LSPA versus Iohexol. CT signal enhancement as a function of concentration for LSPA and Iohexol using monoenergetic image reconstruction at 100 keV.

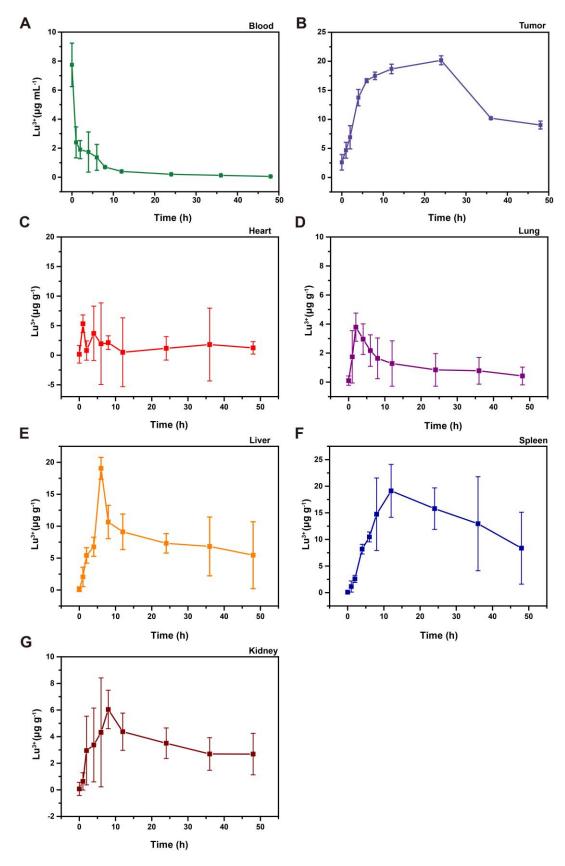


Figure S21. Time-dependent in vivo biodistribution of LSPA nanoparticles at 48 h post-injection. **(A)** blood; **(B)** tumor tissue; **(C)** heart; **(D)** lung; **(E)** liver; **(F)** spleen; and **(G)** kidney. Lu³⁺ concentrations were determined by ICP-MS (n = 3). Data are presented as mean \pm SD.

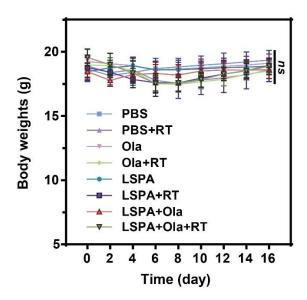


Figure S22. Body weight changes in 4T1 tumor-bearing mice over a 16-day period post-treatment (n = 5). RT: 6 Gy X-ray irradiation; Ola: Olaparib. Data are presented as mean \pm SD; ns: no significance.

Table S1. Tumor growth inhibition (TGI) values across treatment groups (n = 5).

Treatment	Primary tumor (%)		Distant tumor (%)	
Groups	Mean	SD	Mean	SD
PBS	0	6.20	0	6.89
PBS + RT	45.42	2.76	45.60	2.24
Olaparib (Ola)	-0.45*	3.00	5.85	9.19
Ola + RT	56.32	2.19	56.34	2.00
LSPA	61.28	2.80	60.75	1.46
LSPA+RT	81.60	0.60	81.25	1.16
LSPA + Ola	76.00	1.68	82.25	0.58
LSPA + Ola +	89.70	1.52	91.99	0.38

^{*} The negative TGI value observed in the Olaparib-alone group was not statistically significant compared to the control group (P > 0.5) and is attributed to normal inter-animal variation. RT: 6 Gy X-ray irradiation; Ola: Olaparib.

Table S2. Comparative analysis of nano-radiosensitizers and PARP inhibitor-enhanced radiotherapy studies.

Therapeutic	Nanosensit	Radiation	Key	Primary	Reference
Strategy	izer / Drug	Dose	Mechanism	Outcome	Reference
LSPA + Olaparib + RT	LSPA (Lubased)	6 Gy (fractionat ed)	ROS amplification; PARP inhibition; Synergistic cGAS-STING activation	Potent primary & abscopal tumor regression; Durable immune memory	This Work
Nano-radiosens itizer (classical high-Z) + RT	Gold nanoparticl es (AuNP)	30 Gy (single dose)	High-Z photoelectric effect; local dose enhancement	Improved tumor control/surviv al vs RT alone	The use of gold nanoparticles to enhance radiotherapy in mice (DOI: 10.1088/0031- 9155/49/18/N03)
Self-targeting Nano-prodrug + RT	Platinum(I V) amphiphilic prodrug nano- assembly	6 Gy (single dose)	Radiosensitiz ation (from Platinum); Synergistic chemotherapy release	Synergistic and safe chemoradiothe rapy for hepatocellular carcinoma	Self-targeting platinum(IV) amphiphilic prodrug nano- assembly as radiosensitizer for synergistic and safe chemoradiotherapy of hepatocellular carcinoma (DOI: 10.1016/j.biomaterials.2022 .121793)
Nano-radiosens itizer (Gd, clinical-stage) + RT	AGuIX (ultrasmall Gd-based nanoparticl e)	Clinical WBRT (30 Gy in 10 fractions, brain metastases)	High-Z radiosensitiza tion; MRI-visible; rapid renal clearance	Feasibility/saf ety; preliminary efficacy in brain metastases	Theranostic AGuIX nanoparticles as radiosensitizer: A phase I, dose- escalation study in patients with multiple brain metastases (NANO-RAD trial) (DOI: 10.1016/j.radonc.2021.04.0 21)
Nano-radiosens itizer (HfO ₂ , clinical-stage) + RT	NBTXR3 (HfO ₂ nanoparticl e; intratumora l)	50 Gy in 25 fractions (pre-opera tive RT, soft-tissue sarcoma)	High-Z dose enhancement (secondary electron emission)	Higher pathological response vs RT alone; acceptable safety	First-in-human study testing a new radioenhancer using nanoparticles (NBTXR3) activated by radiation therapy in patients with locally advanced soft tissue sarcomas (DOI: 10.1158/1078-0432.CCR-16-1297)

Nano-radiosens itizer (Hf-porphyrin nMOF) + RT + ICB	Hf-based nMOF	8 Gy × 3 (typical preclinical regimen)	High-Z radiosensitiza tion + radiodynamic ROS; ICD/STING activation	Enhanced local control and abscopal effect with anti-PD-L1	Nanoscale metal-organic frameworks enhance radiotherapy to potentiate checkpoint blockade immunotherapy (DOI: 10.1038/s41467-018-04703-w)
PARP Inhibitor + RT (General)	Olaparib	8 Gy × 3 or single 12 Gy (preclinica 1)	Inhibition of DNA single- strand break repair; cGAS- STING activation	Potent systemic antitumor immunity and abscopal effects	Olaparib enhances radiation-induced systemic anti-tumor effects via activating STING- chemokine signaling in hepatocellular carcinoma (DOI: 10.1016/j.canlet.2023.21650 7)

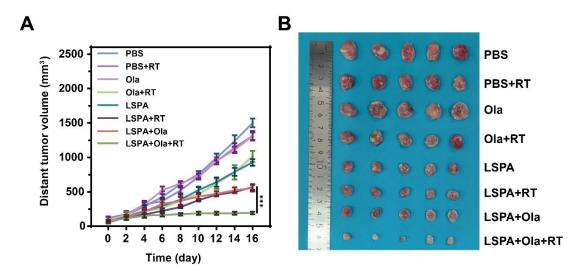


Figure S23. (A) Growth curves of distant tumors in 4T1 tumor-bearing mice over a 16-day treatment period (n = 5). (B) Representative photographs of excised distant tumors. RT: 6 Gy X-ray irradiation; Ola: Olaparib. Data are presented as mean \pm SD; ***P < 0.001.

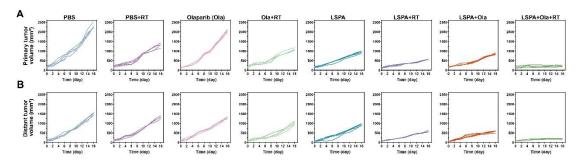


Figure S24. Individual tumor growth kinetics. **(A)** Primary and **(B)** distant tumor growth curves in 4T1 tumor-bearing mice during the 16-day treatment (n = 5). RT: 6 Gy X-ray irradiation; Ola: Olaparib. Data are presented as mean \pm SD.

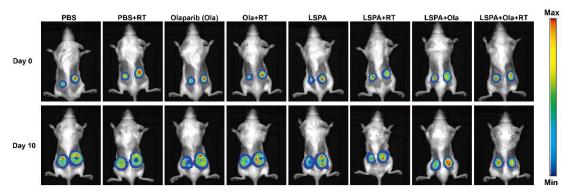


Figure S25. Representative *in vivo* bioluminescence images show primary (right) and distant (left) tumors in 4T1-Luc tumor-bearing mice following the indicated treatments. RT: 6 Gy X-ray irradiation; Ola: Olaparib.

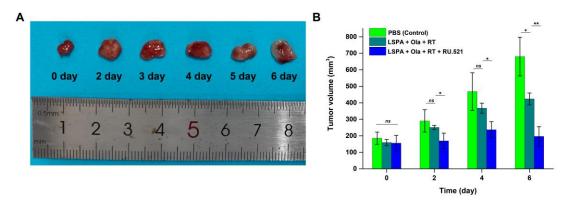
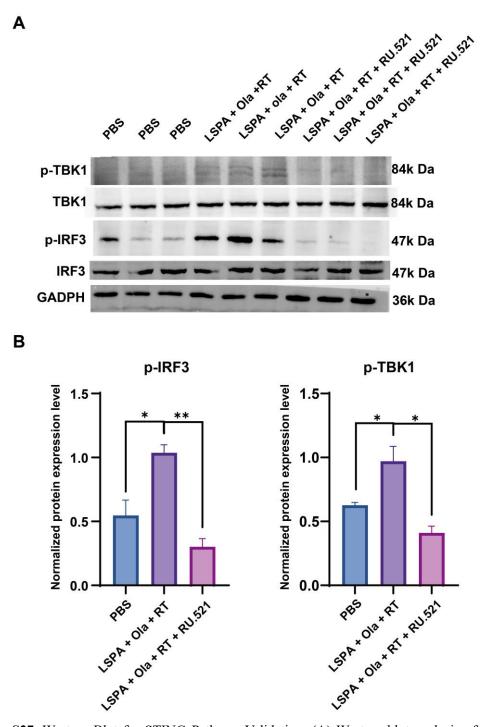


Figure S26. Verification of the therapeutic mechanism underlying cGAS-STING pathway activation. (A) Representative photograph of excised tumors following treatment within the first 6 days. (B) Corresponding tumor volumes following the indicated treatments during the first 6 days.



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Figure S27. Western Blot for STING Pathway Validation. (A) Western blot analysis of cGAS-STING pathway biomarkers (p-IRF3, p-TBK1) in tumor lysates following the indicated treatments. (B) Quantification of p-TBK1 and p-IRF3 biomarkers levels (n = 3). Ola: Olaparib; RT: 6 Gy X-ray irradiation. Data are presented as mean \pm SD. *P < 0.05; **P < 0.01.

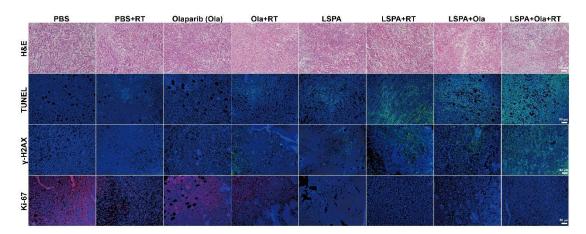


Figure S28. Immunohistochemical analysis of distant tumors reveals treatment-induced effects (H&E, TUNEL, γ -H2AX, and Ki-67 staining) following the indicated treatments. RT: 6 Gy X-ray irradiation; Ola: Olaparib. Scale bar: 50 μ m.

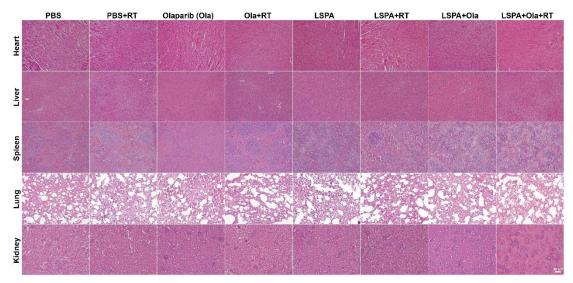


Figure S29. Histopathological evaluation (H&E staining) of major organs (heart, liver, spleen, lung, and kidney) from 4T1 tumor-bearing mice following the indicated treatments. RT: 6 Gy X-ray irradiation; Ola: Olaparib. Scale bar: 50 μm.

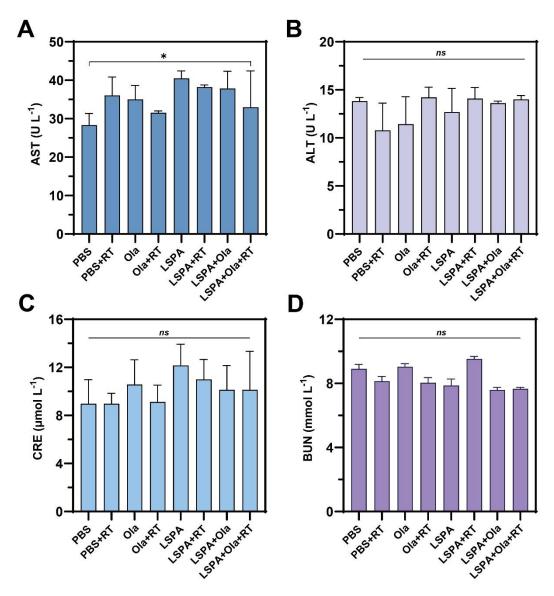


Figure S30. Serum biochemical analyses of 4T1 tumor-bearing mice after 16 days of the indicated treatments. **(A-B)** Liver function markers: aspartate aminotransferase (AST, n = 3) and alanine aminotransferase (ALT, n = 3). **(C-D)** Kidney function markers: creatinine (CRE, n = 3) and blood urea nitrogen (BUN, n = 3). RT: 6 Gy X-ray irradiation; Ola: Olaparib. Data are presented as mean \pm SD; ns: no significance. *P < 0.05.

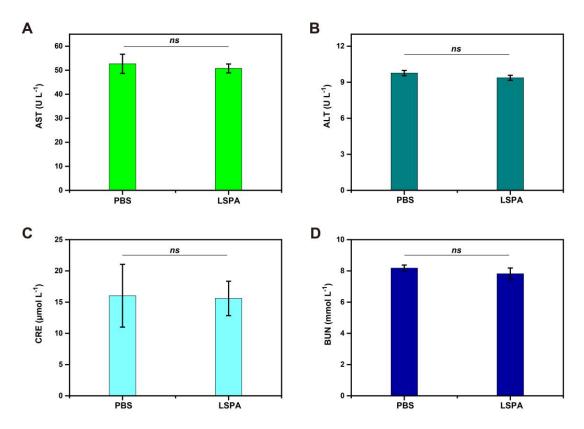


Figure S31. Long-term *in vivo* toxicity assessment of LSPA nanoparticles. Serum biochemistry analysis of healthy BALB/c mice after intravenous injection of LSPA nanoparticles or PBS every 7 days for 3 doses over a 28-day period. The levels of **(A)** aspartate aminotransferase (AST), (B) alanine aminotransferase (ALT), (C) creatinine (CRE), and (D) blood urea nitrogen (BUN) were measured. Data are presented as mean \pm SD (n = 3); ns: no significance.

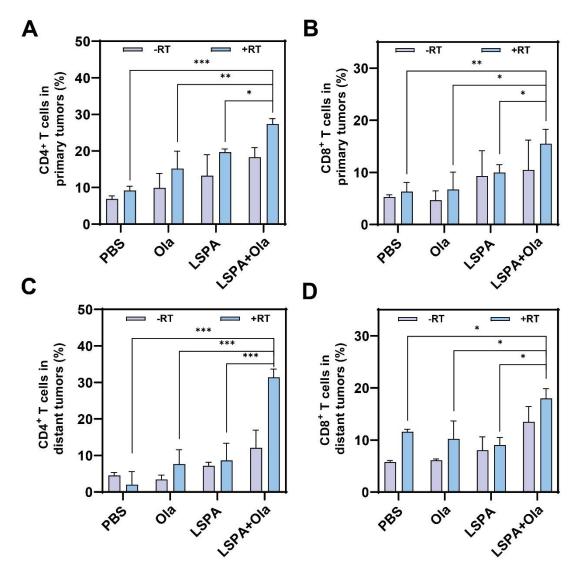


Figure S32. Immune cell infiltration analysis in tumor models. **(A)** CD4⁺ T cell infiltration in primary tumors (n = 3). **(B)** CD8⁺ T cell infiltration in primary tumors (n = 3). **(C)** CD4⁺ T cell infiltration in distant tumors (n = 3). **(D)** CD8⁺ T cell infiltration in distant tumors (n = 3). RT: 6 Gy X-ray irradiation; Ola: Olaparib. Data are presented as mean \pm SD, *P < 0.05; **P < 0.01; ****P < 0.001.

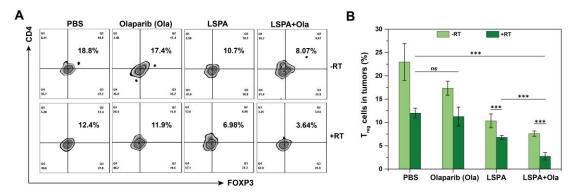


Figure S33. Flow cytometric analysis of tumor-infiltrating regulatory T (T_{reg}) cells. **(A)** Representative flow cytometry plots of T_{reg} cells (CD4⁺ FOXP3⁺). **(B)** Quantification of tumor-infiltrating T_{reg} cells (n = 3). RT: 6 Gy X-ray irradiation; Ola: Olaparib. Data are presented as mean \pm SD; ns: no significance; ***P < 0.001.

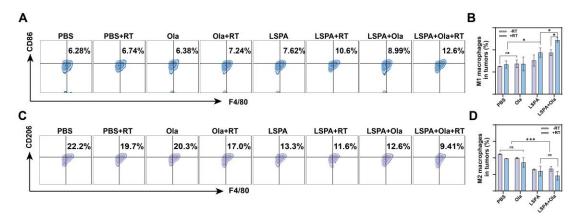


Figure S34. Polarization analysis of tumor-associated macrophages (TAMs). **(A)** Representative flow cytometry plots of M1-polarized TAMs (F4/80⁺ CD86⁺). **(B)** Quantification of M1-polarized TAMs in tumors (n = 3). **(C)** Representative flow cytometry plots of M2-polarized TAMs (F4/80⁺ CD206⁺). **(D)** Quantification of M2-polarized TAMs in tumors (n = 3). RT: 6 Gy X-ray irradiation; Ola: Olaparib. Data are presented as mean \pm SD; ns: no significance; *P < 0.05, ***P < 0.001.

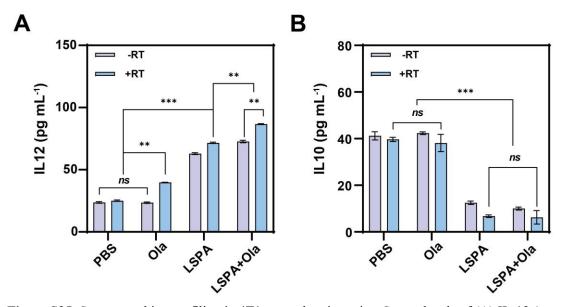


Figure S35. Serum cytokine profiling in 4T1 tumor-bearing mice. Serum levels of **(A)** IL-12 (proinflammatory) and **(B)** IL-10 (anti-inflammatory) were quantified by ELISA following the indicated treatments (n = 3). RT: 6 Gy X-ray irradiation; Ola: Olaparib. Data are presented as mean \pm SD; ns: no significance; **P < 0.01; ***P < 0.001.

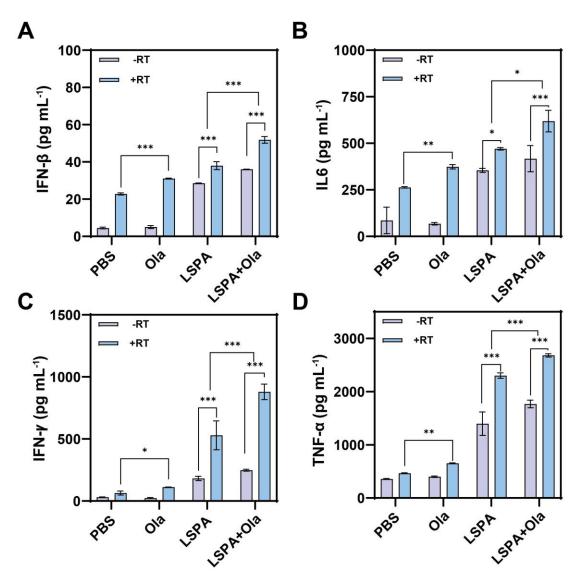


Figure S36. Serum cytokine analysis in 4T1 tumor-bearing mice. Quantification of (**A**) IFN-β (interferon-β), (**B**) IL-6 (interleukin-6), (**C**) IFN-γ (interferon-γ), and (**D**) TNF-α (tumor necrosis factor-α) by ELISA following the indicated treatments (n = 3). RT: 6 Gy X-ray irradiation; Ola: Olaparib. Data are presented as mean \pm SD. *P < 0.05; **P < 0.01; ***P < 0.001.