

Supplementary figures and materials legends

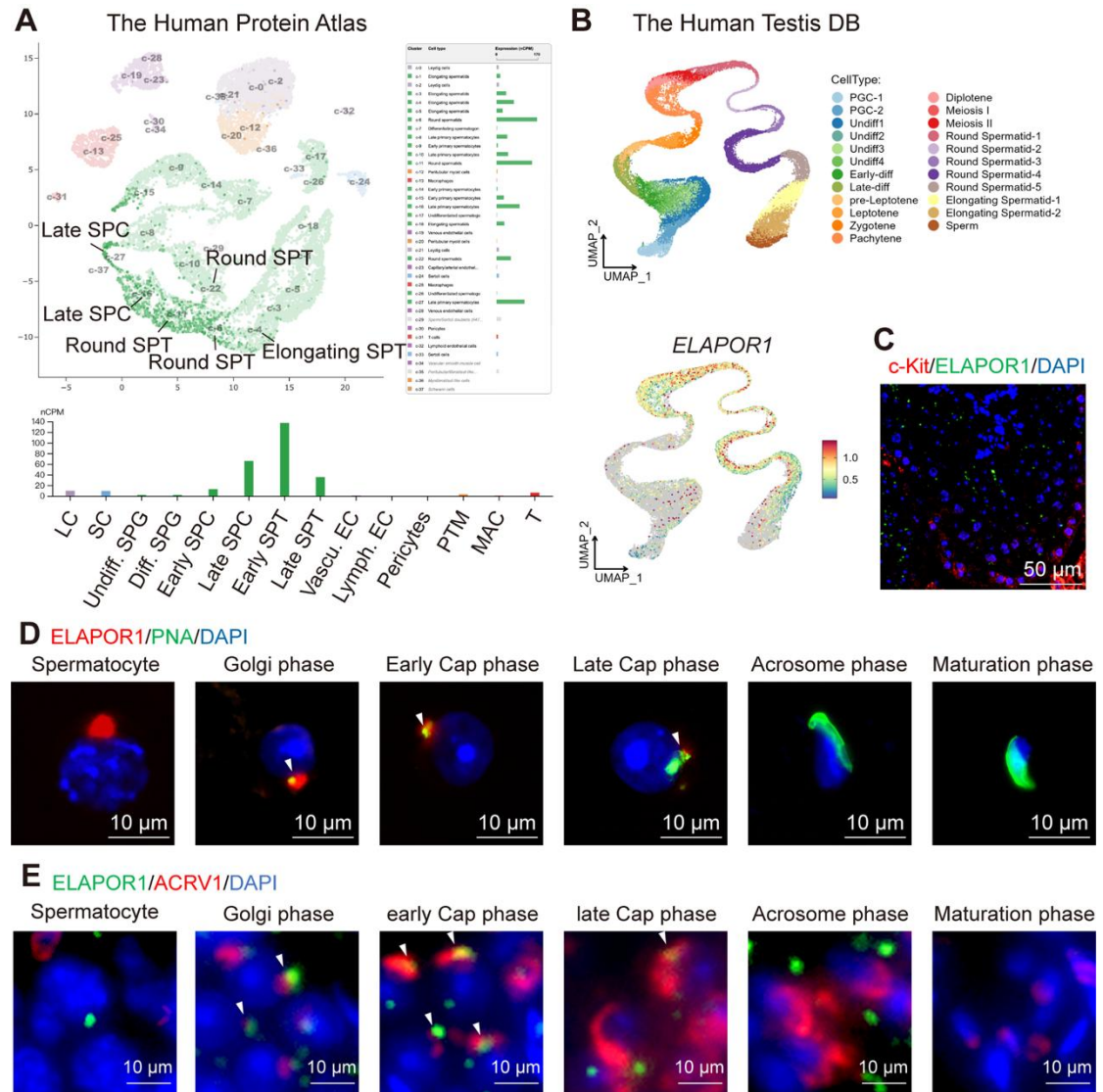


Figure S1. Additional evidence for ELAPOR1 expression in spermatocytes and spermatids.

(A) Single-cell RNA expression data for *ELAPOR1* in the cell type clusters identified in the human testis from the Human Protein Atlas database, visualized by a UMAP plot and bar plots. The labels on the UMAP plot indicated cell-type clusters with RNA Expression (nCPM) > 50. Late SPC: Late primary spermatocyte; Round SPT: Round spermatids; Elongating SPT: Elongating spermatids; LC: Leydig cell; SC: Sertoli cell; Undiff. SPG: Undifferentiated spermatogonia; Diff. SPG: Differentiated spermatogonia; Early SPC: Early primary spermatocyte; Early SPT: Early spermatids; Late SPT: Late spermatids; Vascu. EC: Vascular endothelial cells; Lymph. EC: Lymphatic endothelial cells; PTM: peritubular myoid cells; MAC: macrophages; T: T-cells. (B) Single-cell RNA expression data for *ELAPOR1* in the cell type clusters identified in the human testis from the Human Testis DB, visualized by UMAP plots. (C) Immunofluorescence staining of ELAPOR1 and

c-Kit (Kit proto-oncogene receptor tyrosine kinase) in *Elapor^{flax}* mouse testes. Bar = 50 μm . (D) Immunofluorescence staining for ELAPOR1, PNA as a marker of acrosomes, and DAPI as a marker of nuclei in isolated spermatocytes and spermatids from the mouse testes. Bar = 10 μm . White arrows indicate the colocalization of ELAPOR1 and PNA signals. Bar = 10 μm . (E) Immunofluorescence staining showing the colocalization (white arrows) of ELAPOR1 with acrosomal vesicle protein 1 (ACRV1). Bar = 10 μm .

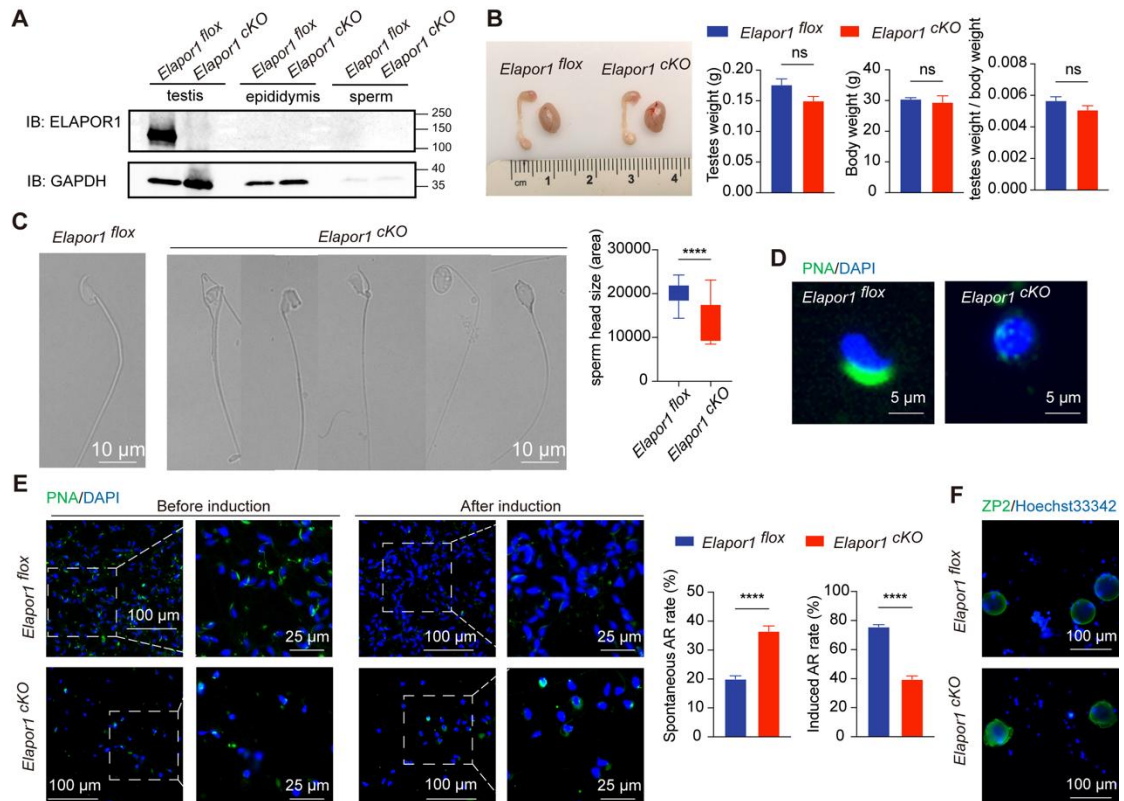


Figure S2. ELAPOR1 deficiency leads to nonfunctional acrosomes. (A) Immunoblot analysis of ELAPOR1 protein levels in testes, epididymis, and sperm from *Elapor1^{fllox}* and *Elapor1^{ckO}* mice. (B) Morphological images of the testes from *Elapor1^{fllox}* and *Elapor1^{ckO}* mice. Average individual testes weights, body weights, and testes/body weights of *Elapor1^{fllox}* and *Elapor1^{ckO}* mice (n = 3). (C) Representative images of sperm from *Elapor1^{fllox}* and *Elapor1^{ckO}* mice. Bar = 10 μ m. The sperm head sizes from *Elapor1^{fllox}* and *Elapor1^{ckO}* mice (n = 28). (D) Fluorescence staining of the acrosome with PNA of sperm from *Elapor1^{fllox}* and *Elapor1^{ckO}* mice. Bar = 5 μ m. (E) Acrosome reaction assessed by PNA staining of sperm from *Elapor1^{fllox}* and *Elapor1^{ckO}* before and after A23187 induction. Nuclei were stained with DAPI. Bar = 100 μ m in the main panels (left panels) and bar = 25 μ m in the magnified panels (right panels). Spontaneous acrosome reaction rates and induced acrosome reaction rates of *Elapor1^{fllox}* and *Elapor1^{ckO}* sperm (n = 6). (F) Fluorescence staining for ZP2 (pellucida sperm-binding protein 2) of eggs after binding with sperm from adult *Elapor1^{fllox}* and *Elapor1^{ckO}* mice. Nuclei were stained with Hoechst 33342. Bar = 100 μ m. The data are presented as the means \pm SEMs. Statistical analyses were conducted using Student's t-test (unpaired, two-tailed) for comparisons between two groups. ns = no significant difference. **** P < 0.0001.

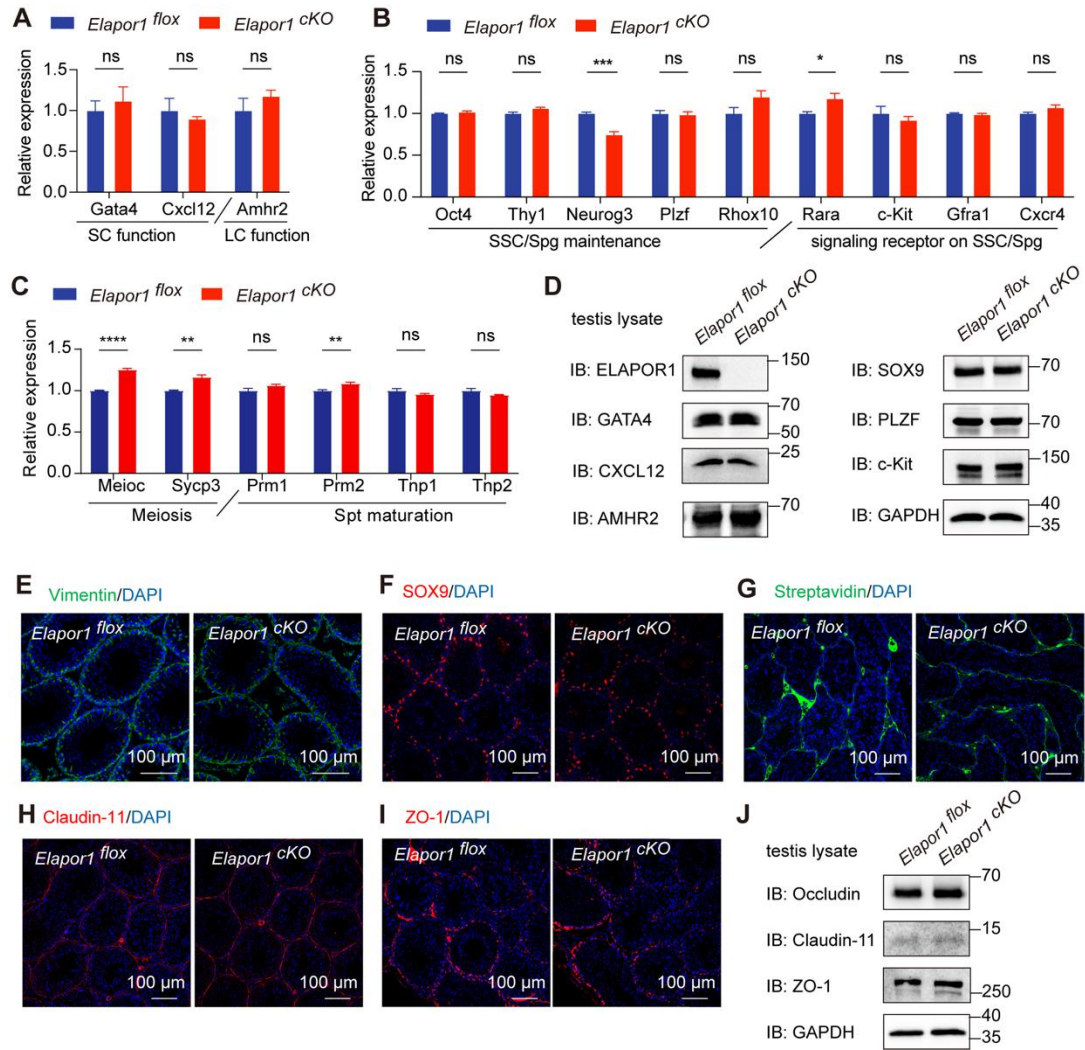


Figure S3. The impact of ELAPOR1 knockout on other spermatogenic processes and the integrity of the blood–testis barrier in the testes.

(A–C) RT–qPCR–based validation of the expression patterns of spermatogenic marker genes in *Elapor1*^{fllox} and *Elapor1*^{cKO} mouse testes (n = 4). SC: Sertoli cell; LC: Leydig cell; SSC, spermatogonial stem cell; Spg, spermatogonia; Spt, spermatids/sperm. (D) Immunoblot analysis of protein levels of spermatogenic marker genes in testicular lysates from *Elapor1*^{fllox} and *Elapor1*^{cKO} mice. (E–F) Immunofluorescence staining of Vimentin as a marker of blood–testis barrier (E) and SOX9 as a marker of Sertoli cells (F) in *Elapor1*^{fllox} and *Elapor1*^{cKO} mouse testes. Bar = 100 μ m. (G) Immunofluorescence staining of Streptavidin–488 showing the permeability through the blood–testis barrier in the seminiferous tubules from *Elapor1*^{fllox} and *Elapor1*^{cKO} mice by a biotin tracer 30 min after injection. Bar = 100 μ m. (H–I) Immunofluorescence staining for Claudin–11 (H) and ZO–1 (I) as markers of tight junctions in *Elapor1*^{fllox} and *Elapor1*^{cKO} mouse testes. Bar = 100 μ m. (J)

Immunoblot analysis of protein levels of marker genes of tight junctions in testicular lysates of *Elapor1^{fllox}* and *Elapor1^{CKO}* mice. The data are presented as the means \pm SEMs. Statistical analyses were conducted using Student's t-test (unpaired, two-tailed) for comparisons between two groups or one-way ANOVA for comparisons among three groups. ns = no significant difference. *P < 0.05; ** P < 0.01; *** P < 0.001; and **** P < 0.0001.

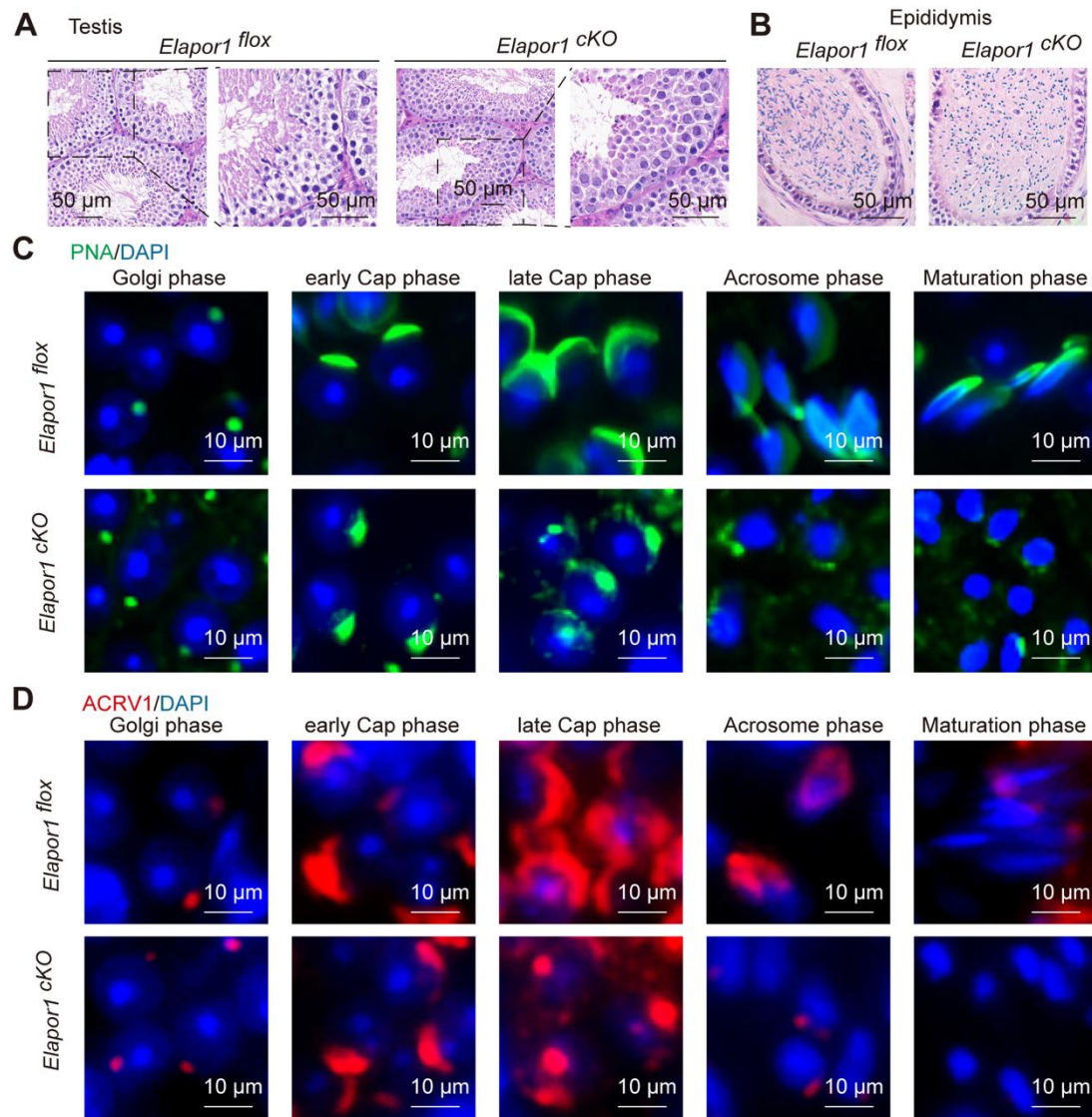


Figure S4. The abnormal morphology and defective acrosome formation of *Elapor1^{cKO}* sperm.

(A–B) H&E staining of the testicular (A) and epididymal (B) sections from *Elapor1^{flox}* and *Elapor1^{cKO}* mice. Bar = 50 μ m. (C–D) Fluorescence staining of adult testis sections from *Elapor1^{flox}* and *Elapor1^{cKO}* mice with PNA (C), ACRV1 (D), and DAPI. Bar = 10 μ m.

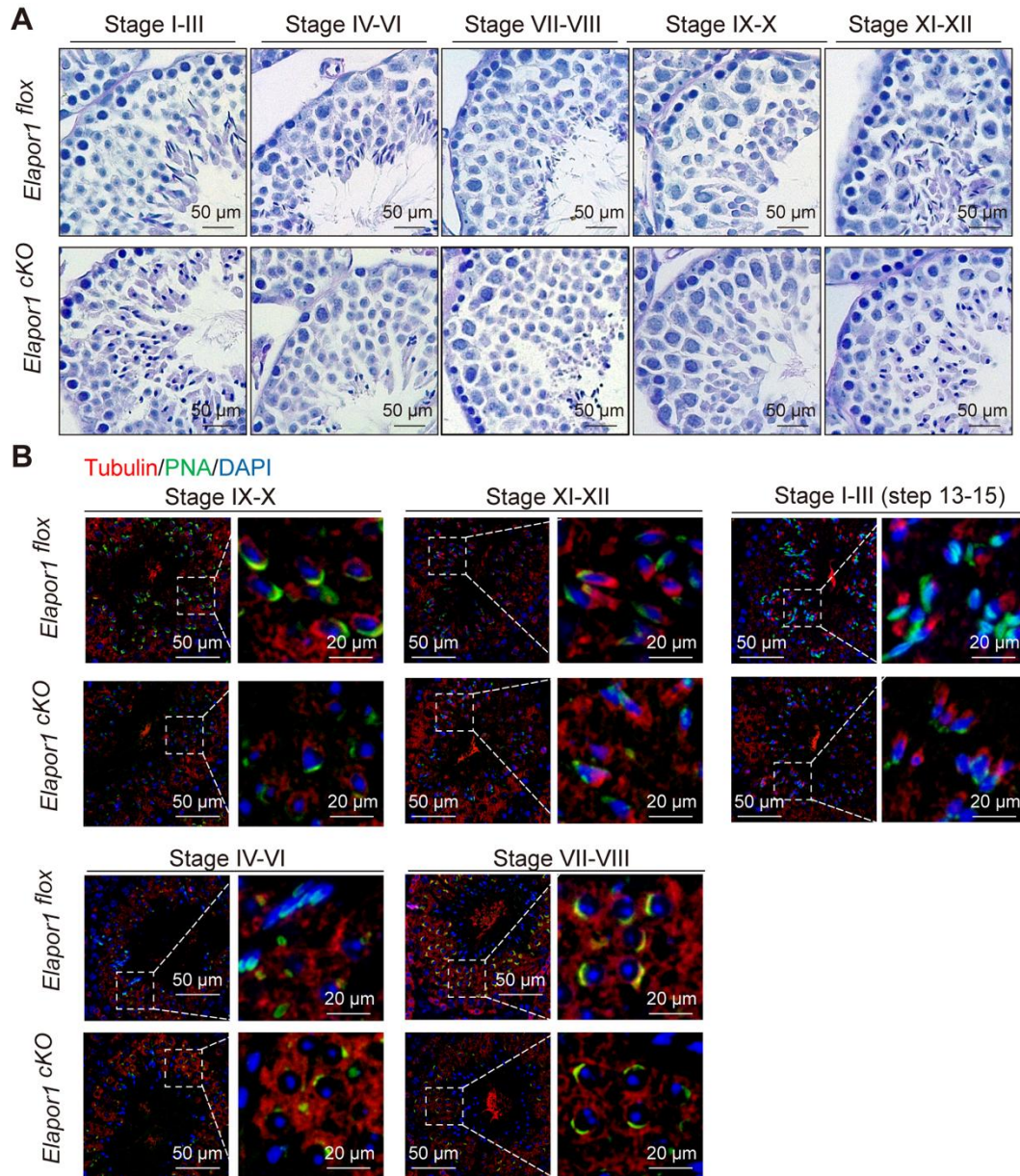


Figure S5. ELAPOR1 knockout disrupts the manchette during spermatogenesis. (A) The Periodic acid–Schiff (PAS) staining of seminiferous tubules of different stages from *Elapor1^{flx}* and *Elapor1^{cko}* mice. Bar = 50 μ m. (B) Immunofluorescence staining of tubulin as a marker of manchette formation, PNA as a marker of acrosome formation, and DAPI as a marker of nuclei in *Elapor1^{flx}* and *Elapor1^{cko}* mouse testes. Bar = 50 μ m in the main panels (left panels) and bar = 20 μ m in the magnified panels (right panels).

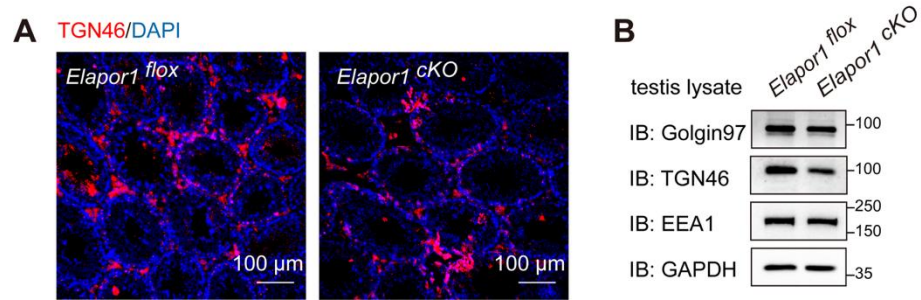


Figure S6. The disruptions of the TGN in *Elapor1^{cKO}* mouse testes. (A) A bigger panel of representative immunofluorescence staining images of TGN46 in the testes from *Elapor1^{flox}* and *Elapor1^{cKO}* mice. Bar = 100 μ m. (B) Immunoblot analysis of protein levels of vesicle transport marker genes in testicular lysates from *Elapor1^{flox}* and *Elapor1^{cKO}* mice.

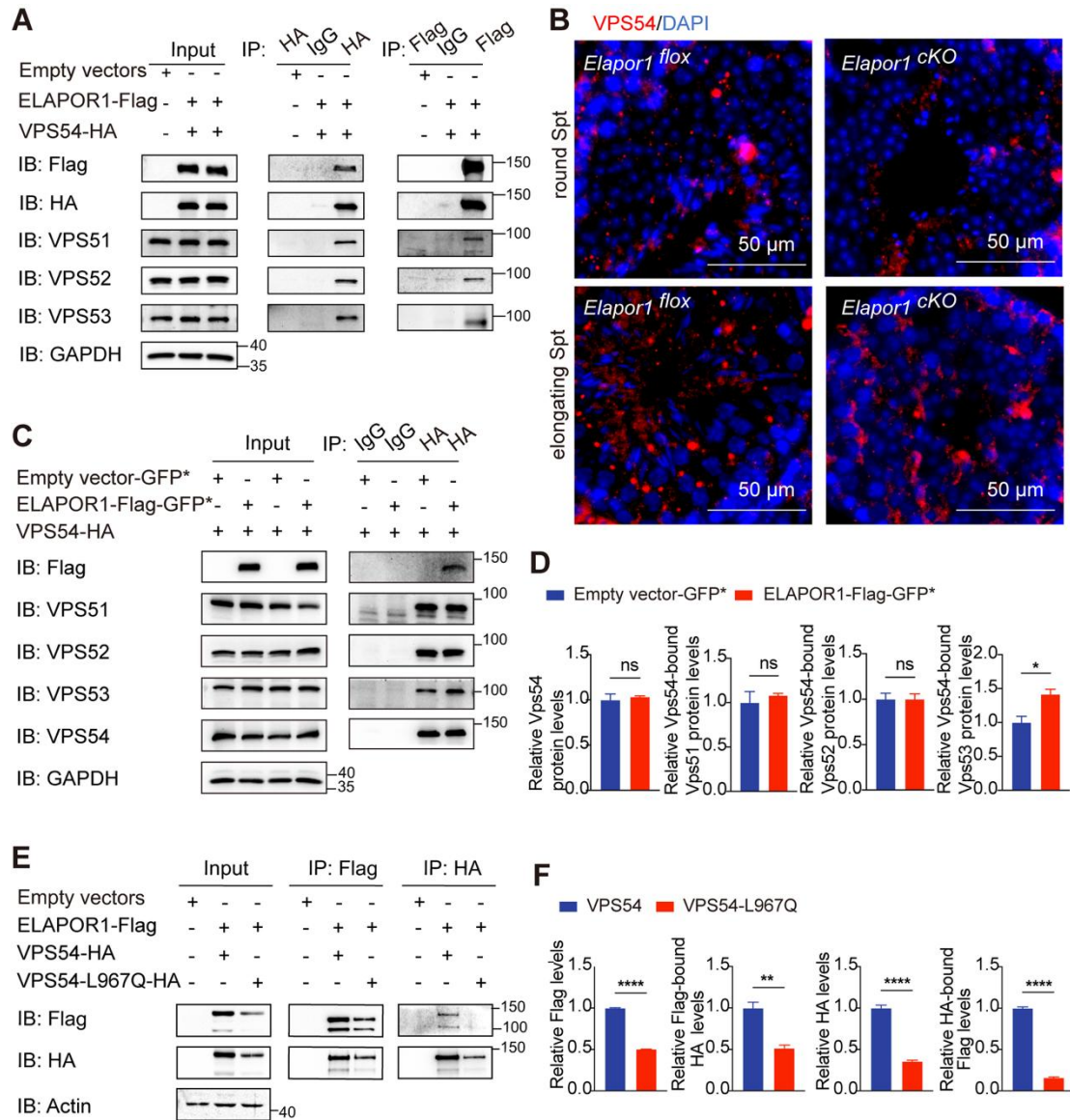


Figure S7. ELAPOR1 regulates the GARP complex assembly through interaction with VPS54.

(A) Co-IP of lysates from HEK293T cells transiently transfected with ELAPOR1-Flag and VPS54-HA plasmids using anti-Flag antibodies, anti-HA antibodies, and IgG control antibodies. Cells transfected with empty vectors were used as control samples. (B) Immunofluorescence staining of ELAPOR1 and VPS54 in round and elongating spermatids in *Elapor1^{flox}* and *Elapor1^{CKO}* mouse testes. Bar = 50 μ m. (C) Co-IP of lysates from HEK293T cell lines that constitutively expressed ELAPOR1-Flag-GFP* protein and were transfected with VPS54-HA plasmids using anti-HA antibodies and IgG control antibodies. Cell lines that constitutively expressed empty vectors-GFP* and were transfected with VPS54-HA plasmids were used as control samples. (D) Relative levels of VPS53, VPS52, and VPS51 proteins bound to VPS54 were evaluated (n = 3). (E) Co-IP of lysates

from HEK293T cells transfected with ELAPOR1-Flag and VPS54-HA or VPS54-L967Q-HA mutant plasmids. Cells transfected with empty vectors were used as control samples. (F) Relative levels of HA bound to Flag and Flag bound to HA (n = 3). The data are presented as the means \pm SEMs. Statistical analyses were conducted using Student's t-test (unpaired, two-tailed) for comparisons between two groups or one-way ANOVA for comparisons among three groups. ns = no significant difference. *P < 0.05; ** P < 0.01; and **** P < 0.0001.

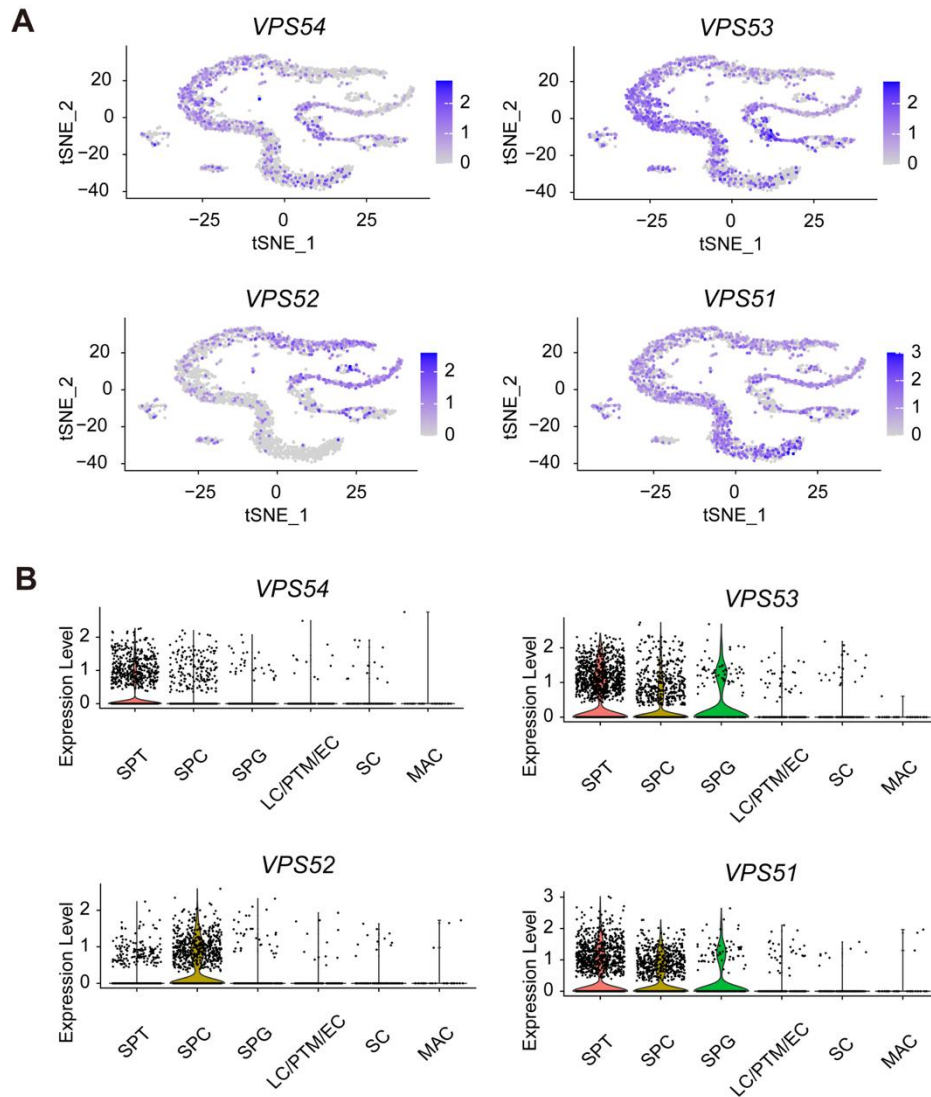


Figure S8. The expression patterns of the GARP complex subunits from human testes. (A) t-SNE plots of the expression patterns of *VPS54*, *VPS53*, *VPS52*, and *VPS51* in specific cell types from human testes. **(B)** Violin plots of the expression patterns of *VPS54*, *VPS53*, *VPS52*, and *VPS51* in specific cell types from human testes.

Movie S1. Video of sperm swimming capacity from *Elapor1^{fllox}* mice. Representative movie showing motility patterns of *Elapor1^{fllox}* mouse sperm.

Movie S2. Video of sperm swimming capacity from *Elapor1^{ckO}* mice. Representative movie showing motility patterns of *Elapor1^{ckO}* mouse sperm.

Table S1. Resources list.

Table S2. Primer list.

Table S3. Identified ELAPOR1 interacting protein list by MS.