

Figure S1. 3-oxoLCA affect Tfh cells differentiation in human CD4⁺ T cells.

A. The gating strategy to analyse Tfh cell subsets in cultured human CD4⁺ T cells.

B. Bcl6 expression in WT mouse naive CD4⁺ T cells stimulated with 3-oxoLCA by Western blot (n=3).

C. The gating strategy to analyse IL-21 positive cells in cultured human CD4⁺ T cells.

D. The proliferation of cultured human CFSE⁺CXCR5⁺ and CFSE⁺IL21⁺ T cells by CFSE assay (n=3).

The graphs show the data as mean ± SEM. **p* < 0.05, ***p* < 0.01, ****p* < 0.001, *****p* < 0.0001.

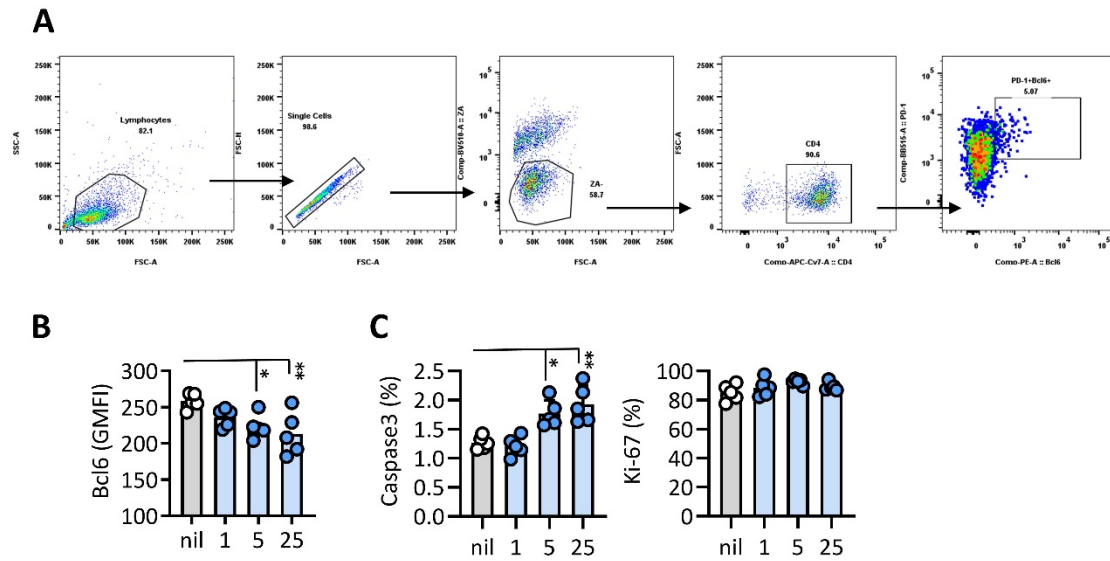


Figure S2. 3-oxoLCA affect Tfh cells differentiation in mouse CD4⁺ T cells.

A. The gating strategy to analyse Tfh cell subsets in cultured mouse CD4⁺ T cells.

B. The geometric mean fluorescence intensity (GMFI) of Bcl6 expression at different concentrations of 3-oxoLCA (n=5).

C. The percentage of Caspase-3 and Ki-67 of CD4⁺ T cells at different concentrations of 3-oxoLCA (n=5).

The graphs show the data as mean \pm SEM. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$.

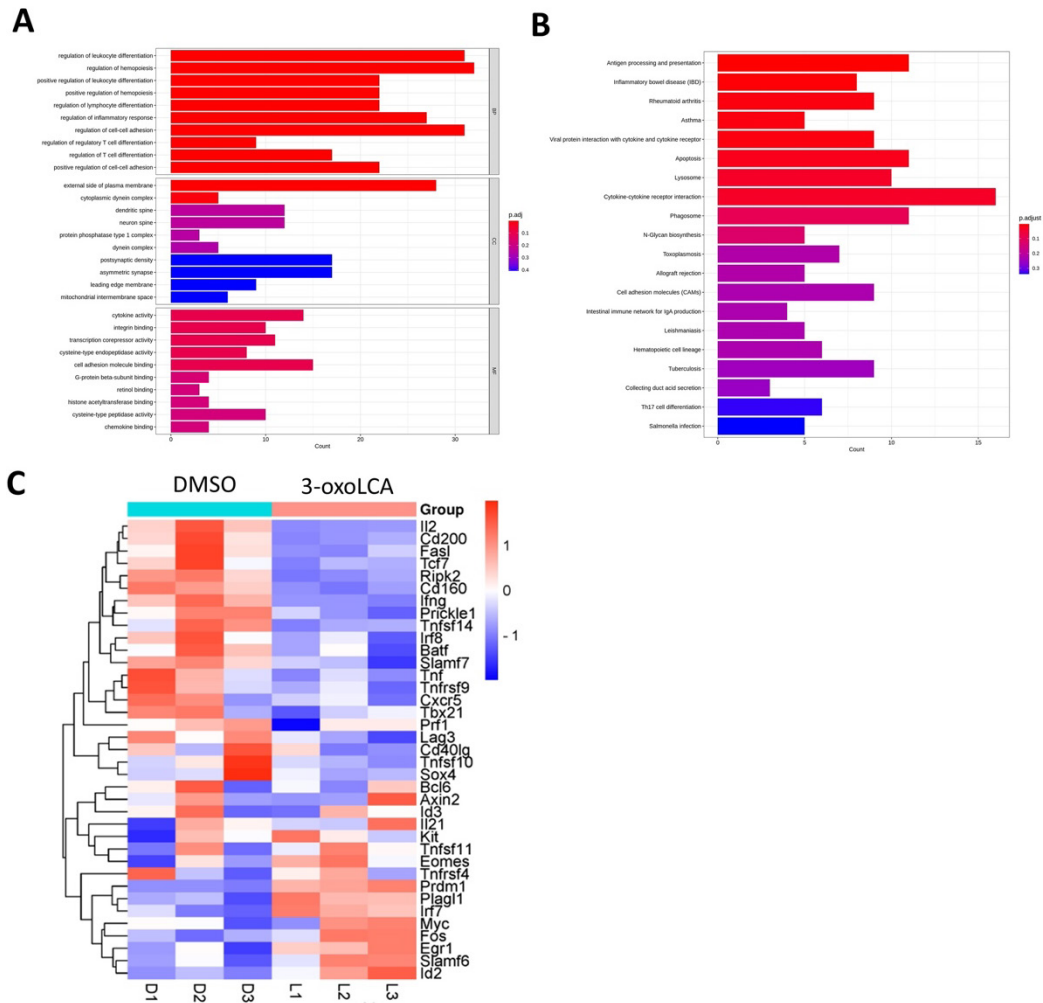


Figure S3. 3-oxoLCA inhibit Tfh cells through improve mitophagy.

A. Gene ontology enrichment analysis was performed on the DEGs that were differentially regulated by either 3-oxoLCA resulted in changes in the expression of genes involved in several biological processes.

B. Top enriched pathways of DEGs in 3-oxoLCA treated Tfh cells by Kyoto Encyclopedia of Genes and Genomes (KEGG) analysis.

C. Heatmap showing the RNA-seq analysis of Tfh-related genes in mouse naïve CD4⁺ T cells with anti-CD3/CD28 activation for 12h, and further treated with 3-oxoLCA or DMSO for 12 h.

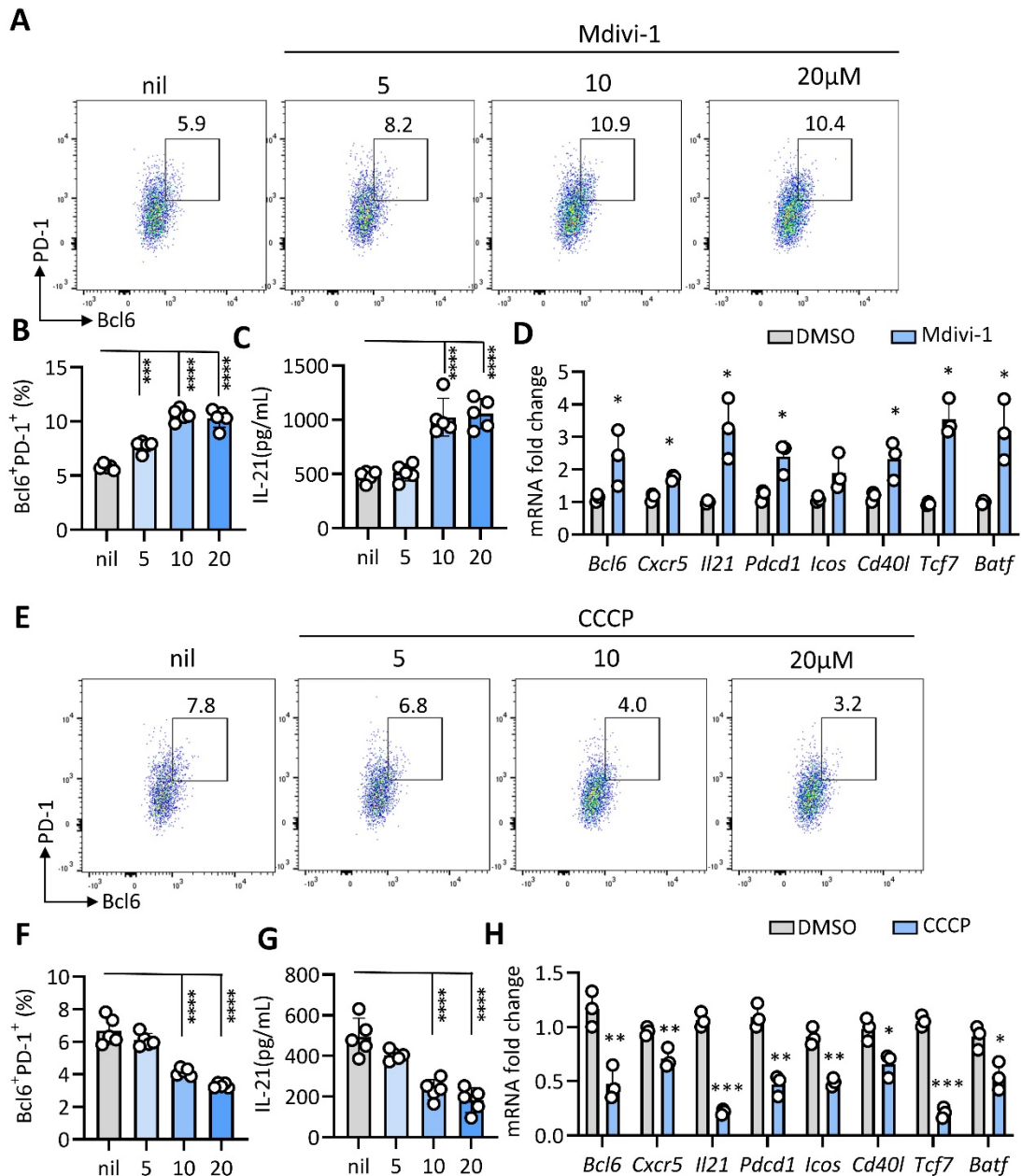


Figure S4. Modulation of intercellular mitophagy regulates Tfh cells differentiation.

A-B. Analysis of PD-1⁺Bcl6⁺ Tfh cells in cultured naïve CD4⁺ T cells from WT mice with mitophagy inhibitor (Mdivi-1) for 3days (n=5).

C. ELISA analysis of IL-21 production in naïve CD4⁺ T cells from WT mice with Mdivi-1 for 3days (n=5).

D. Tfh-related genes in naïve CD4⁺ T cells from WT mice treated with Mdivi-1 for 12h by real-time PCR (n=3).

E-F. Analysis of PD-1⁺Bcl6⁺ Tfh cells in cultured naïve CD4⁺ T cells from WT mice with mitophagy inducer (CCCP) for 3days (n=5).

G. ELISA analysis of IL-21 production in naïve CD4⁺ T cells from WT mice with mitophagy inducer (CCCP) for 3days (n=5).

H. Tfh-related genes in naïve CD4⁺ T cells from WT mice treated with CCCP for 12h by real-time PCR (n=5).

The graphs show the data as mean \pm SEM. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$.

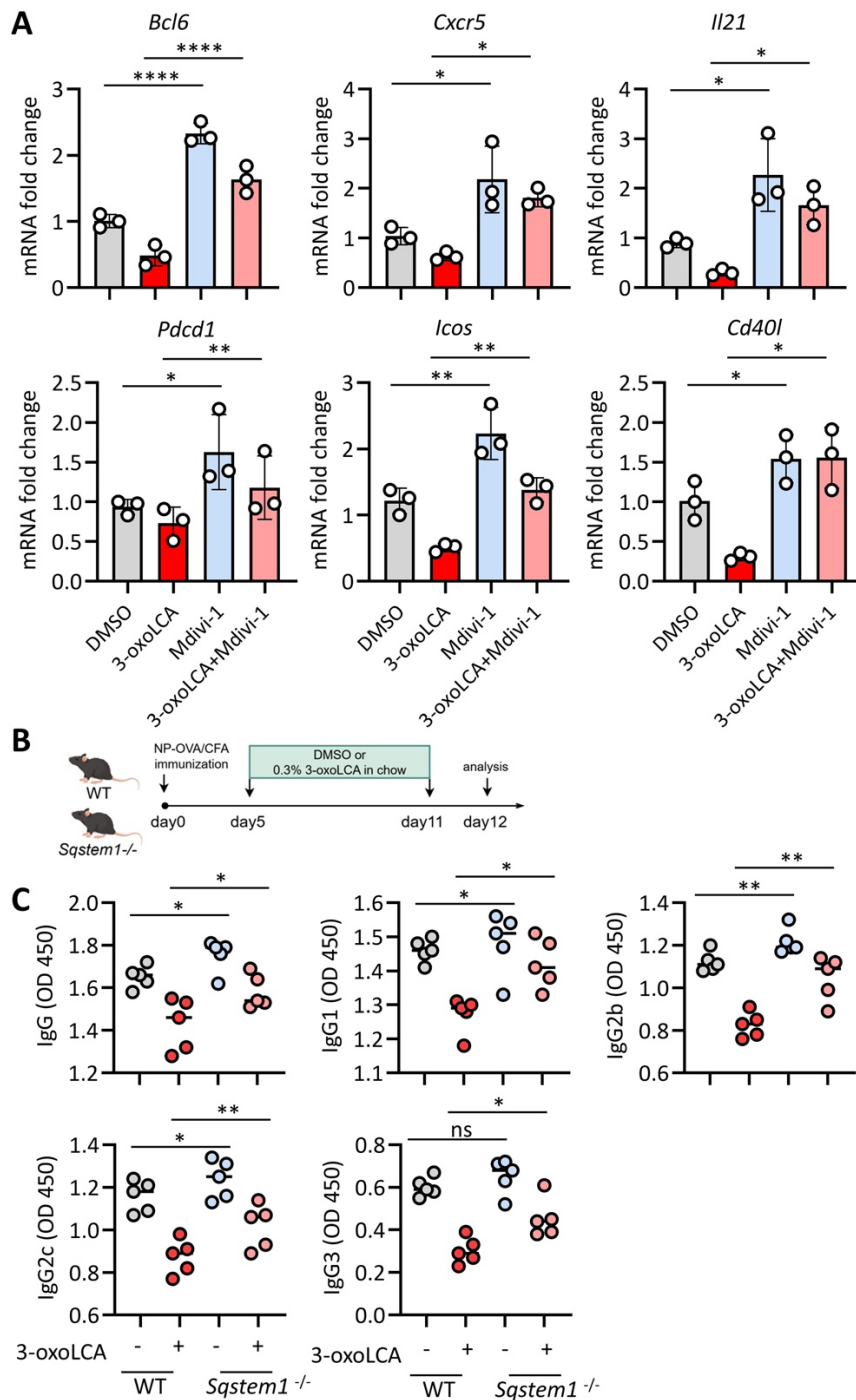


Figure S5. Inhibition of mitophagy restores 3-oxoLCA induced Tfh cells differentiation deficiency.

A. Real-time quantitative PCR analysis of mRNA abundance of Tfh cell-related genes in DMSO, 3-oxoLCA, Mdivi-1 and 3-oxoLCA + Mdivi-1 group (n=3).

B. Workflow of the experimental procedure of WT or *Sqstem1*^{-/-} C57BL/6 mice with NP-OVA/CFA immunization (subcutaneous), 3-oxoLCA (oral gavage) and analysis.

C. ELISA assay for serum NP-specific IgG1, IgG2b, IgG2c and IgG3 titers from in WT or *Sqstem1*^{-/-} C57BL/6 mice immunized with NP-OVA/CFA, followed by 3-oxoLCA treatment (n=5).

The graphs show the data as mean \pm SEM. * p < 0.05, ** p < 0.01, *** p < 0.001, **** p < 0.0001.

Figure S6. 3-oxoLCA3-oxoLCA reduces Tfh cells blasting associated with attenuated mTORC1 and STAT3 signaling pathway.

The phosphorylation of key signaling proteins (mTOR, p70S6K, S6, STAT3) in WT naïve CD4⁺ T cells was compared by Western blot following treatment with vehicle control or 25 μ M 3-oxoLCA (n=3).

The graphs show the data as mean \pm SEM. * p < 0.05, ** p < 0.01, *** p < 0.001, **** p < 0.0001.

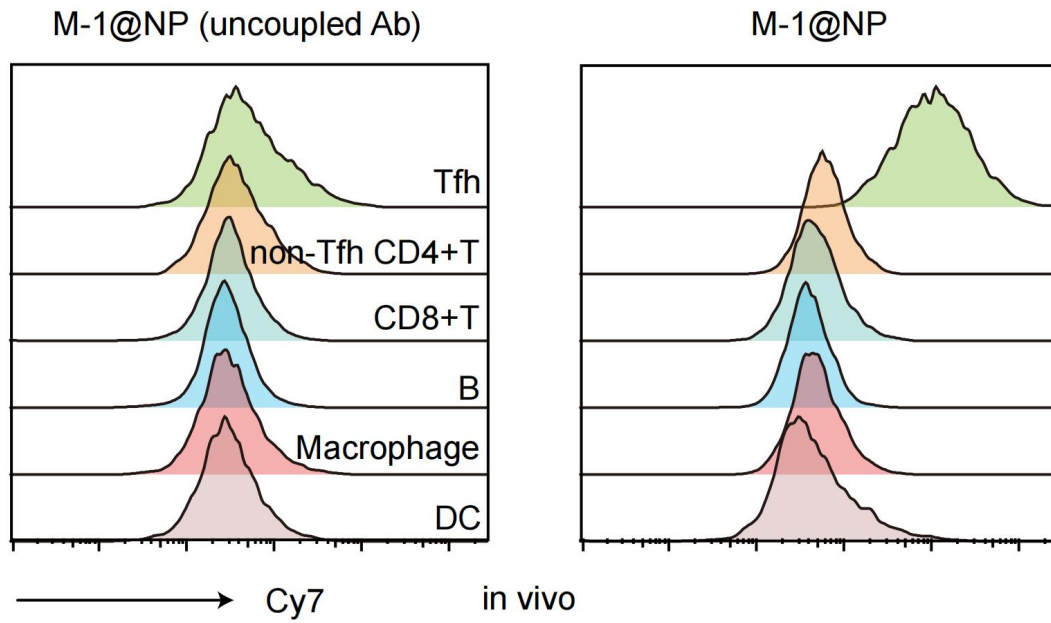


Figure S7. The distribution of M-1@NP in different lymphocyte subsets in vivo by flow cytometry.

The histogram of flow cytometry showing fluorescence intensity of M-1@NP in different immune cell subpopulations in mouse spleen.

The graphs show the data as mean \pm SEM. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$.

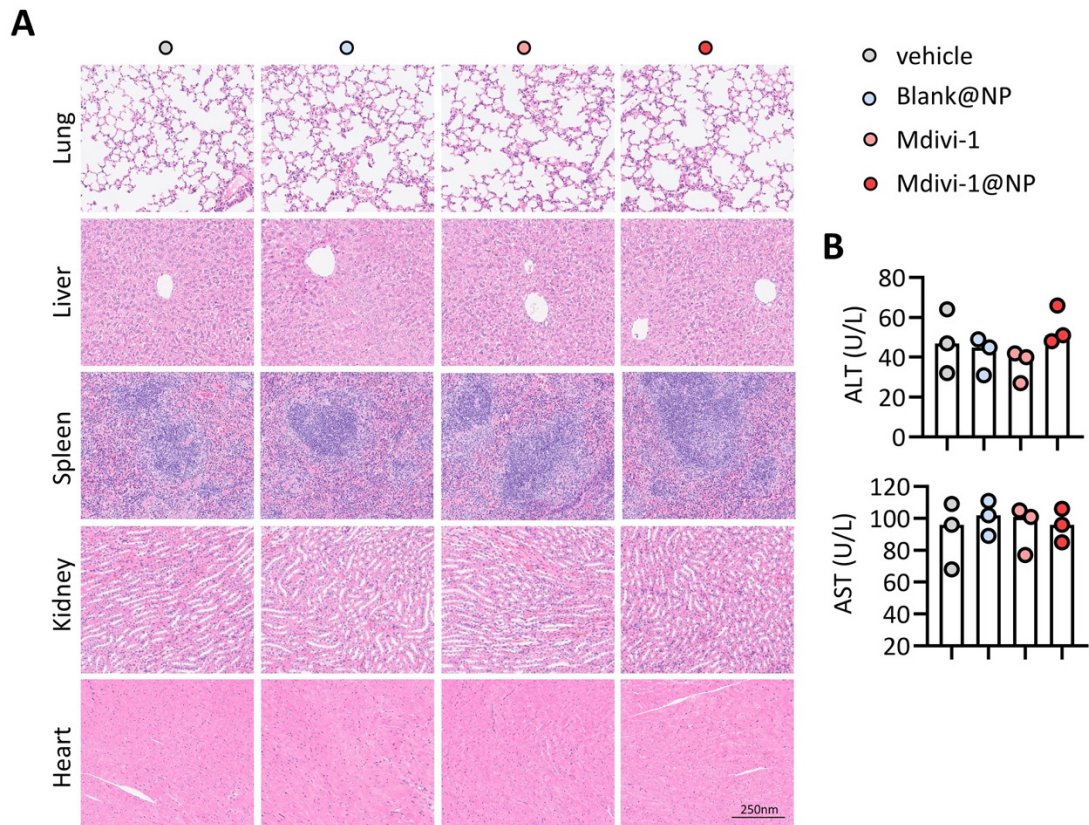


Figure S8. In vivo biocompatibility assessment.

A. Major organs of mice were taken for H&E staining at time points of 12 days after M-1@NP injection.

B. Serum glutathione transaminase and ghrelin levels in mice injected with the M-1@NP in the tail vein after 12 days (n = 3).

The graphs show the data as mean \pm SEM. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$.

Supplementary Table S1. PCR primers of mouse used in this study

genes	Forward Sequence	Reverse Sequence
<i>Bcl6</i>	CAGAGATGTGCCTCCATACTGC	CTCCTCAGAGAAACGGCAGTCA
<i>Cxcr5</i>	ATCGTCCATGCTGTTACGCCT	CAACCTTGGCAAAGAGGAGTTCC
<i>Il21</i>	GCCTCCTGATTAGACTTCGTCAC	CAGGCAAAAGCTGCATGCTCAC
<i>Pdcd1</i>	CGGTTTCAAGGCATGGTCATTGG	TCAGAGTGTCGTCCTTGCTTCC
<i>Icos</i>	GCAGCTTTCGTTGTGGTACTCC	TGTGTTGACTGCCGCCATGAAC
<i>Cd40l</i>	GAACTGTGAGGAGATGAGAAGGC	TGGCTTCGCTTACAACGTGTGC
<i>Tcf7</i>	CCTGCGGATATAGACAGCACTTC	TGTCCAGGTACACCAGATCCCA
<i>Batf</i>	CACAGAAAGCCGACACCCTTCA	GCTGCTCAGCACTGATGTGAAG
<i>Atg5</i>	CTTGCATCAAGTTCAGCTCTTCC	AAGTGAGCCTCAACCGCATCCT
<i>Atg7</i>	CCTGTGAGCTTGGATCAAAGGC	GAGCAAGGAGACCAGAACAGTG
<i>Ulk1</i>	GCAGCAAAGACTCCTGTGACAC	CCACTACACAGCAGGCTATCAG
<i>Becn1</i>	CAGCCTCTGAAACTGGACACGA	CTCTCCTGAGTTAGCCTCTTCC
<i>Pink1</i>	CGACAACATCCTTGTGGAGTGG	CATTGCCACCACGCTCTACT
<i>Bnip3</i>	GCTCCAAGAGTTCTCACTGTGAC	GTTTTTCTCGCCAAAGCTGTGGC
<i>Fundc1</i>	AGACACCACTGGTGGAATCGAG	CCTTCTGGAATAAAAATCCTGCAC

Supplementary Table S2. PCR primers of human used in this study.

genes	Forward Sequence	Reverse Sequence
<i>BCL6</i>	CATGCAGAGATGTGCCTCCACA	TCAGAGAAGCGGCAGTCACACT
<i>CXCR5</i>	TGAAGTCCGCAGTGACCTGTC	GAGGTGGCATTCTCTGACTCAG
<i>IL21</i>	CCAAGGTCAAGATCGCCACATG	TGGAGCTGGCAGAAATTCAGGG
<i>PDCD1</i>	AAGGCGCAGATCAAAGAGAGCC	CAACCACCAGGGTTTGAACTG
<i>ICOS</i>	CCCATAGGATGTGCAGCCTTTG	GGCTGTGTTCACTGCTCTCATG
<i>CD40L</i>	GCGGCACATGTCATAAGTGAGG	GTCCTTGTCTTTTAACGGTCAGC