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Supplemental Materials

Lactylation at H3K18 drives pathological angiogenesis via metabolic-epigenetic
crosstalk in ischemic retinopathy

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Data Online Supplement Figures S1-S7

Data Online Table S1-S4

Supplement Figures and Figure legends

Figure S1

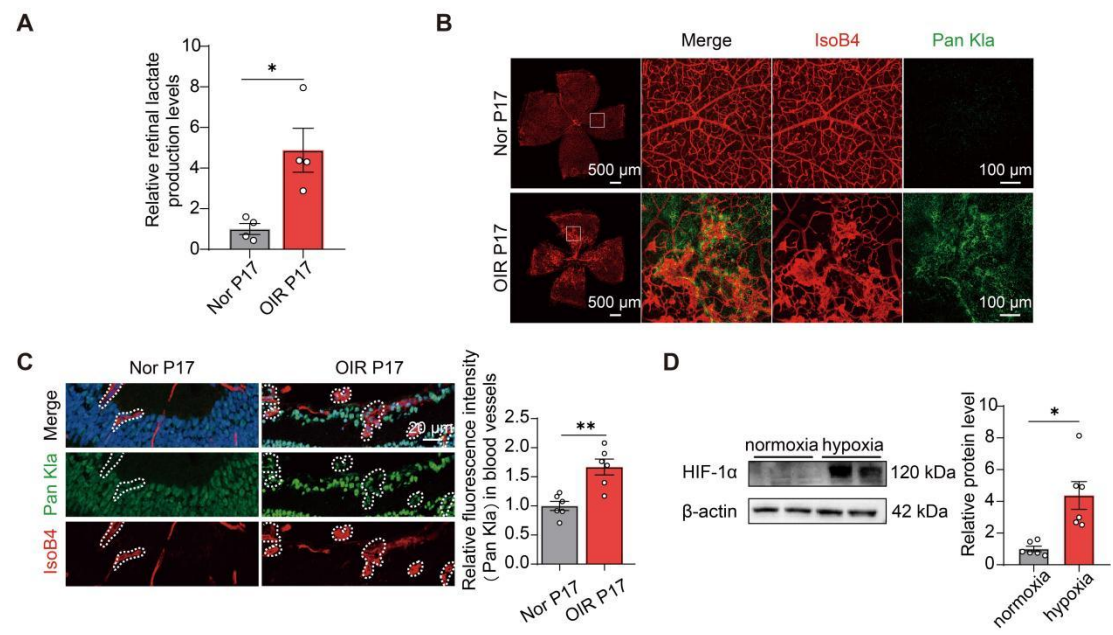


Figure S1. Elevated pan-lactylation levels exhibit significant association with retinal neovascularization in oxygen-induced retinopathy (OIR) models

A: Retinal lactate levels were significantly elevated in oxygen-induced retinopathy (OIR) mice on postnatal day 17 (P17) compared to normoxic controls (Nor P17) (n = 4).

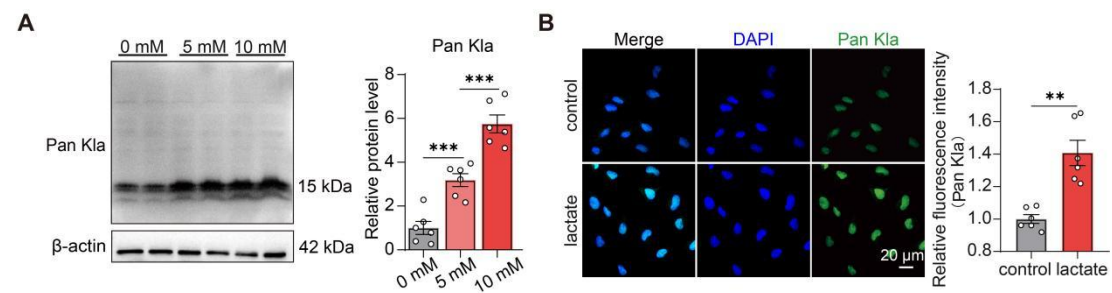
B: Representative retinal whole-mounts from Nor P17 and OIR P17 mice showing costaining with IsoB4 (red) and pan anti-lactyl-lysine antibodies (pan-Kla, green). Scale bars: 500 μ m (left panels); 100 μ m (right panels).

C: Vascular networks in P17 retinal sections co-labeled with IsoB4 (red) and H3K18la (green) antibodies. Scale bar: 20 μ m (n = 6).

D: HIF-1 α protein expression was analyzed by western blotting using β -actin as a loading control (n = 6).

Data were analyzed using two-tailed unpaired Student's *t*-test. Data represent mean $\pm$ SEM. Significance: **P* < 0.05, ***P* < 0.01.

Figure S2



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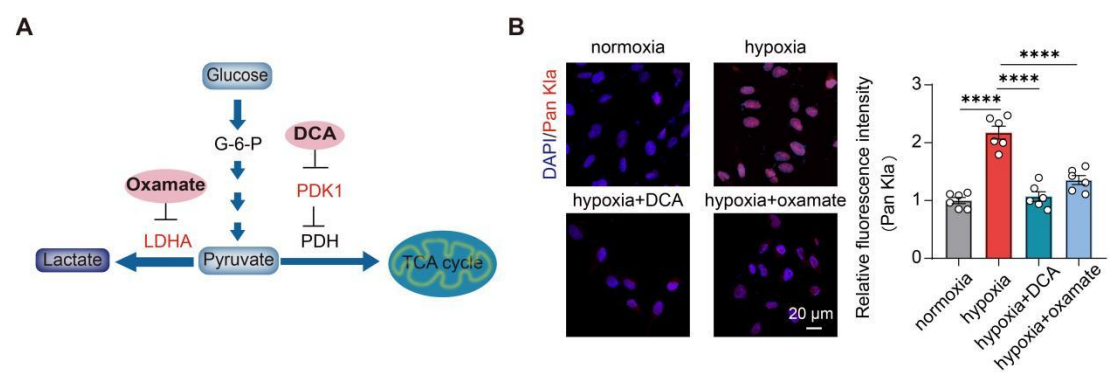
Figure S2. Lactate administration elevates global lactylation levels

A: HRMECs were treated with lactate (0, 5, and 10 mM) for 24 h. Lysates were immunoblotted with pan anti-lactyl-lysine antibody (pan-Kla), using β -actin as loading control (n = 6).

B: Immunofluorescence staining of HRMECs treated with or without 10 mM lactate, using the pan-Kla antibody (green) and nuclear counterstain DAPI (blue). Scale bar: 20 μ m (n = 6).

Data were analyzed using a one-way ANOVA with Tukey's multiple comparison test (A) or a two-tailed unpaired Student's *t*-test (B). Data presented as mean $\pm$ SEM (***P* < 0.01, ****P* < 0.001).

Figure S3



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Figure S3. Lactate inhibition reduces pan-lactylation levels

A: Dichloroacetate (DCA) activates pyruvate dehydrogenase (PDH) by inhibiting pyruvate dehydrogenase kinase 1 (PDK1), thereby shunting the glycolytic end product, pyruvate, into mitochondrial oxidative phosphorylation and reducing lactate production. Oxamate, a structural analog of pyruvate, functions as a competitive inhibitor of lactate dehydrogenase (LDHA). This inhibition suppresses lactate production from pyruvate, thereby disrupting the glycolytic flux.

B: HRMECs were exposed to (i) normoxia (21% O₂), (ii) hypoxia (1% O₂), (iii) hypoxia with 20 mM DCA, or (iv) hypoxia with 20 mM oxamate. Immunostaining with pan anti-lactyl-lysine antibody (pan-Kla, red) and nuclear counterstaining with DAPI (blue). Scale bar: 20 μm (n = 6).

Statistical analysis: One-way ANOVA with Tukey's multiple comparison test. Data are presented as the mean ± SEM (**** $P < 0.0001$).

Figure S4

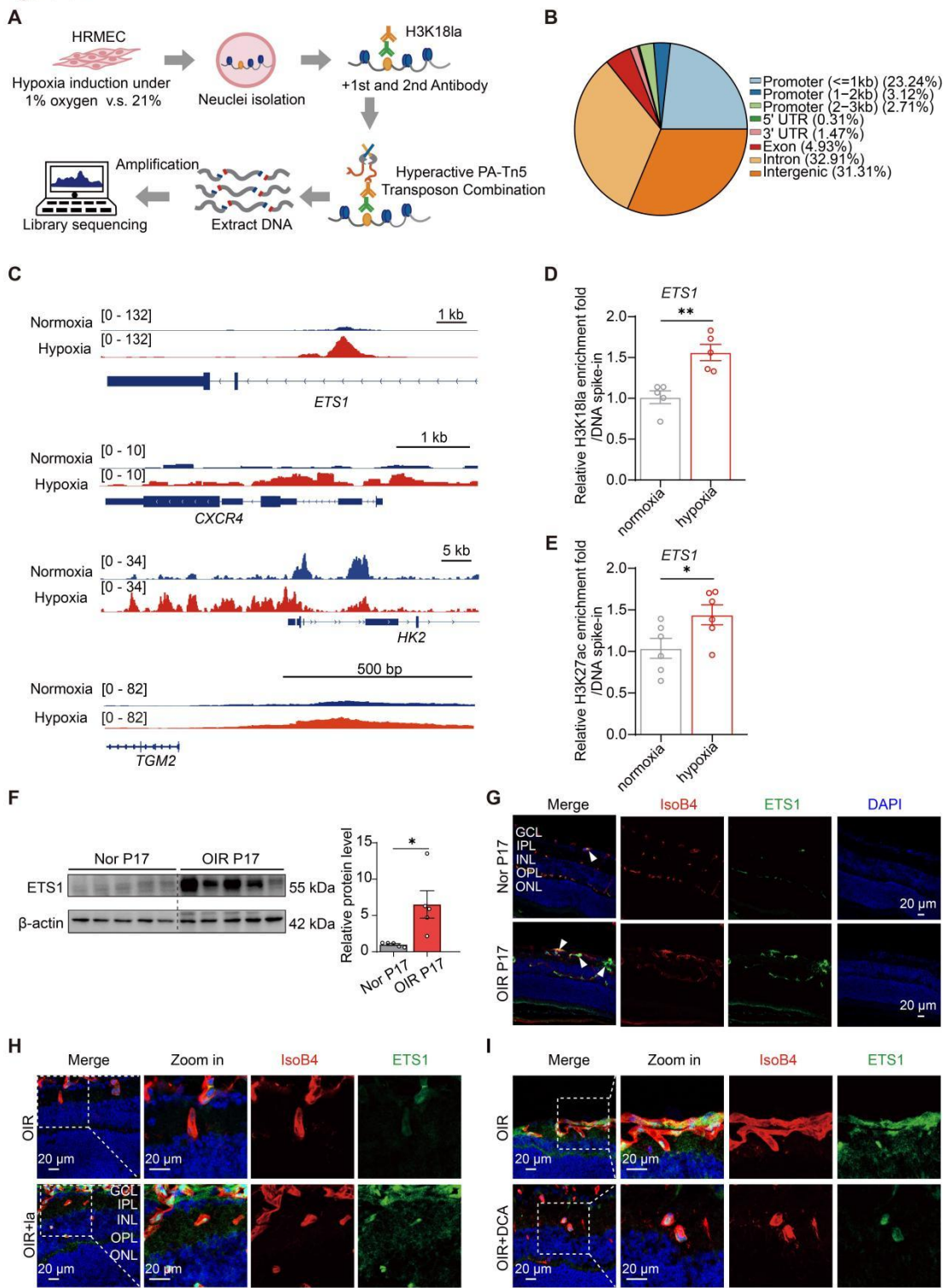


Figure S4. H3K18 lactylation in endothelial cells may regulate *ETS1* transcription

A: Experimental workflow for CUT&Tag profiling of H3K18 lactylation in HRMECs under normoxic (21% O₂) and hypoxic (1% O₂) conditions.

B: H3K18la peaks demonstrate predominant enrichment in enhancer regions.

C: IGV visualization of H3K18la occupancy at *CXCR4*, *ETS1*, *HK2*, and *TGM2* loci.

D,E: Quantitative CUT&RUN-qPCR confirms the hypoxia-induced co-occupancy of H3K18la (n = 5) and H3K27ac (n = 6) at *ETS1* enhancer (Chr11:128, 467, 536–128, 467, 935).

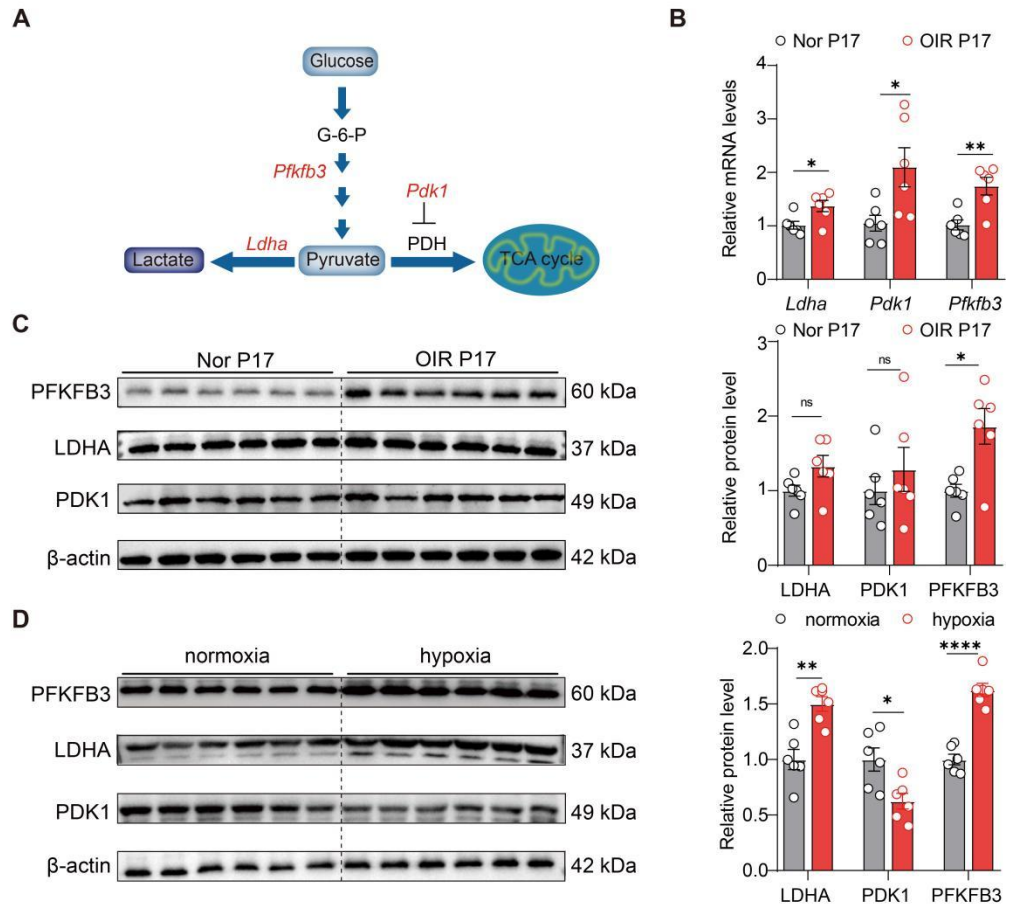
F: Western blot analysis confirming *ETS1* upregulation in OIR retinas (n = 5).

G: Immunofluorescence analysis revealed enhanced *ETS1* expression colocalized with the IsoB4 vasculature. Scale bar: 20 μm.

H,I: Immunofluorescence quantification reveals elevated *ETS1* protein expression in sodium lactate-treated OIR mice (200 mg/kg), whereas dichloroacetate (DCA, 200 mg/kg) attenuated the expression compared with OIR. Scale bar: 20 μm.

Data were analyzed using two-tailed unpaired Student's *t*-test. Data represent mean ± SEM. Significance: **P* < 0.05, ***P* < 0.01.

Figure S5



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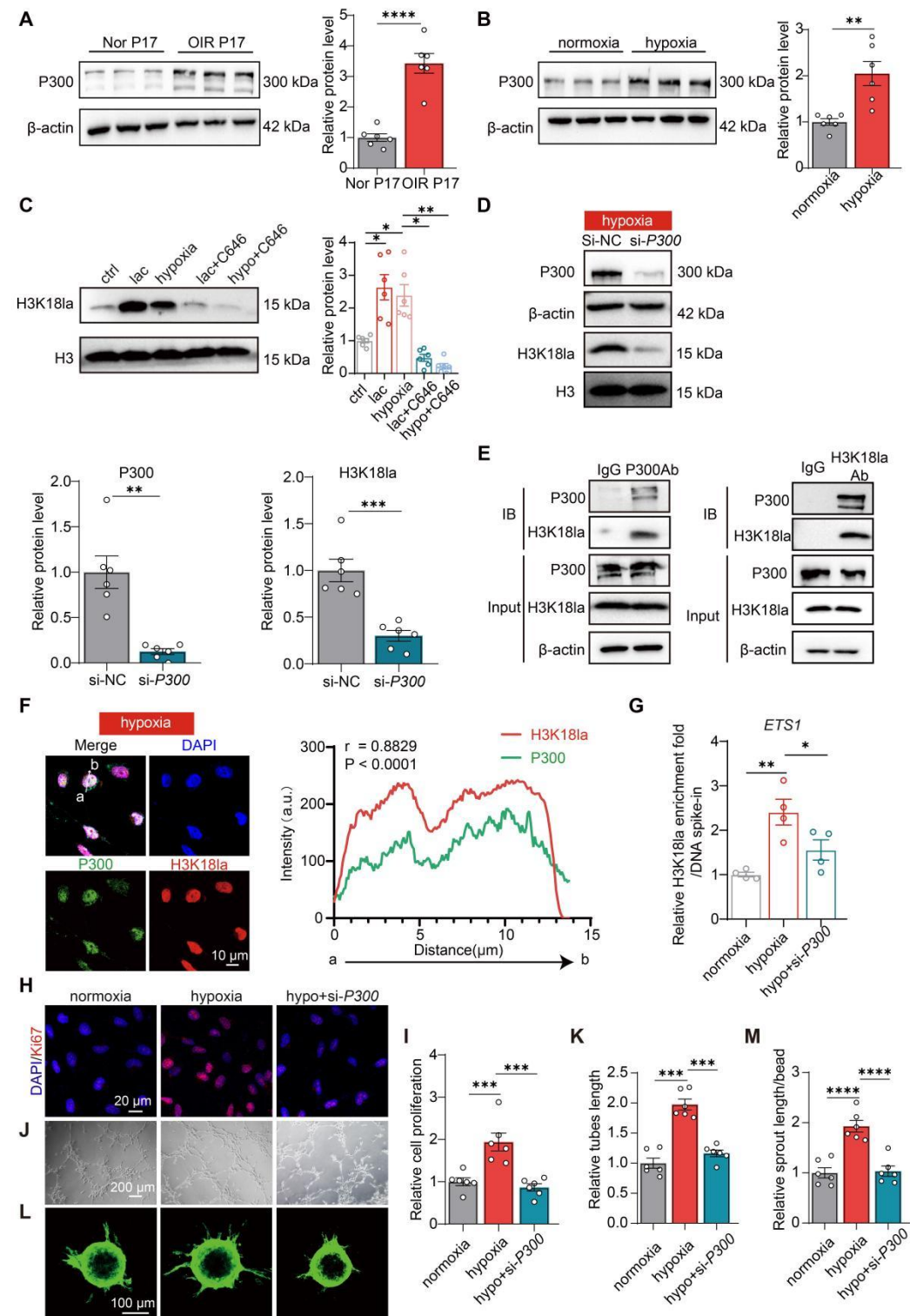
Figure S5. PFKFB3 expression is elevated in the OIR model and in hypoxic endothelial cells

A,B: Transcriptomic profiling identified robust upregulation of the glycolytic regulators *Pfkfb3*, *Pdk1*, and *Ldha* in OIR retinas compared with those into normoxic controls, independently validated by RT-qPCR (B).

C,D: Western blot demonstrating increased PFKFB3 expression in OIR retinal lysates (C) and hypoxic HRMECs (D).

Data were analyzed using two-tailed unpaired Student's *t*-test. Data represent mean $\pm$ SEM. Significance: **P* < 0.05, ***P* < 0.01, *****P* < 0.0001.

Figure S6



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Figure S6. P300-mediated H3K18 lactylation drives pathological retinal neovascularization

A: P300 expression in retinas of normoxic (Nor P17) and OIR mice at P17. β -actin loading control (n = 6).

B: Hypoxia-induced (1% O₂, 24 h) P300 upregulation in HRMECs (n = 6).

C: H3K18 lactylation in HRMECs under: vehicle control/lactate (10 mM, 24 h)/hypoxia (1% O₂, 24 h)/ lac + C646 (10 μ M)/ hypo + C646 (10 μ M) (n = 6).

D: *P300* knockdown efficiency and H3K18la suppression using siRNA (50 nM) in hypoxic HRMECs (n = 6).

E: Co-immunoprecipitation validating P300-H3K18la interaction (n = 3).

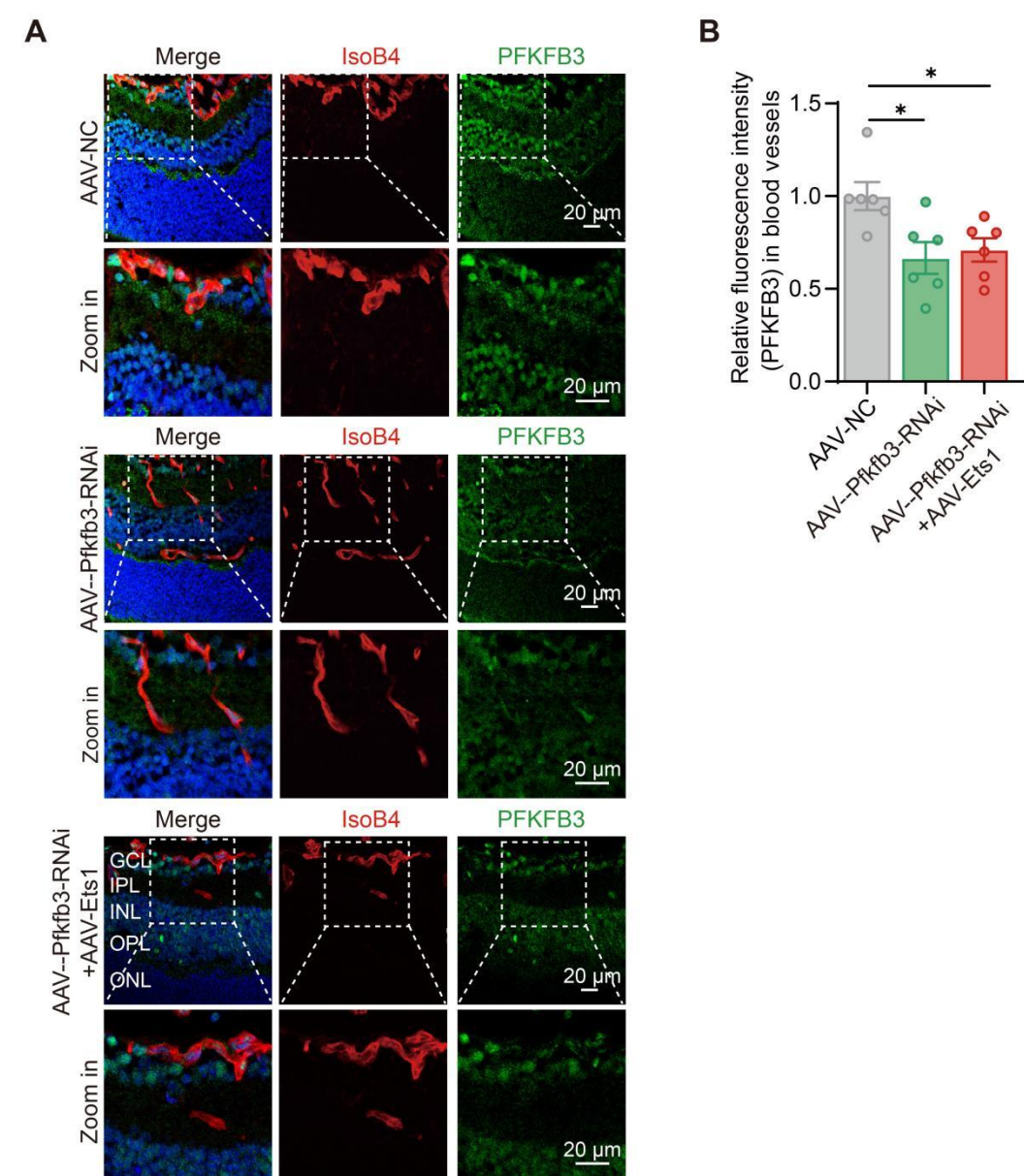
F: Confocal imaging of P300-H3K18la colocalization: P300 (green)/H3K18la (red). Pearson's r = 0.8829. Scale bar: 10 μ m.

G: Genome-wide H3K18la occupancy maps generated by CUT&RUN-qPCR in HRMECs subjected to normoxia (control), hypoxia (1% O₂, 24 h), or hypoxia with *P300* knockdown (n = 4).

H-M: Phenotypic recovery assays after post-*P300* inhibition under hypoxia: H, I: Ki67 proliferation; J, K: Tube formation; L, M: Spheroid sprouting (n = 6; scale bars labeled).

Data represent mean $\pm$ SEM. Unpaired two-tailed Student's *t*-tests (A, B, and D) or one-way ANOVA with Tukey's multiple comparison test (C, G, I, K, and M) were performed. Significance: **P* < 0.05, ***P* < 0.01, ****P* < 0.001, *****P* < 0.0001.

Figure S7



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Figure S7. PFKFB3 signals are diminished in knockdown retinas

A,B: Quantitative immunofluorescence demonstrating reduced PFKFB3 signal intensity in *Pfkfb3*-knockdown retinas. Scale bar: 20 μ m (n = 6).

124 **Supplementary Table 1: The sequences of siRNA**

siRNA	Sequence (5'-3')
si <i>ETS1-a</i> (Homo sapiens)	CGCUAUACCUCGGAUUACU
si <i>ETS1-b</i> (Homo sapiens)	CCACUAUUAACUCCAAGCA
si- <i>P300</i> (Homo sapiens)	CCGGUGAACUCUCCUAUAATT
si <i>PFKFB3</i> (Homo sapiens)	GAAGAGGATCAGTTGCTATGATT

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127 **Supplementary Table 2:Antibodies**

Target antigen	Vendor or Source	Catalog#	Working concentration	Persistent ID/URL
PFKFB3	ABclonal	A3934	1/1000(WB)	https://abclonal.com/catalog-antibodies/PFKFB3RabbitmAb/A3934
PDK1/PDHK1	ABclonal	A0834	1/1000(WB)	https://abclonal.com/catalog-antibodies/PDK1PDHK1RabbitpAb/A0834
LDHA	ABclonal	A1146	1/1000(WB)	https://abclonal.com/catalog-antibodies/LDHARabbitpAb/A1146
Anti-L-Lactyl-Histone H3(Lys18) Rabbit mAb	PTM BIO	PTM-1406 RM	1/2000(WB) 1/500(IF)	https://ptmbio.com/products/anti-l-lactyl-histone-h3-lys18-rabbit-mab/PTM-1406RM.htm
Anti-L-Lactyl-Histone H3(Lys14) Rabbit mAb	PTM BIO	PTM-1414	1/1000(WB)	https://ptmbio.com/products/anti-l-lactyl-histone-h3-lys14-rabbit-pab/PTM-1414.htm
Anti-L-Lactyl-Histone H4(Lys5) Rabbit mAb	PTM BIO	PTM-1407 RM	1/1000(WB)	https://ptmbio.com/products/anti-l-lactyl-histone-h4-lys5-rabbit-mab/PTM-1407RM.htm
Anti-L-Lactyl-Histone H4(Lys8) Rabbit mAb	PTM BIO	PTM-1415 RM	1/1000(WB)	https://ptmbio.com/products/anti-l-lactyl-histone-h4-lys8-rabbit-mab/PTM-1415RM.htm

Anti-L-Lactyl-Histone H4(Lys12) Rabbit mAb	PTM BIO	PTM-1411 RM	1/1000(WB)	https://ptmbio.com/products/anti-l-lactyl-histone-h4-lys12-rabbit-mab/PTM-1411RM.htm
Anti-L-Lactyl-Histone H3(Lys18) Rabbit mAb	PTM BIO	PTM-1427 RM	1/50 (CUT&Tag)	https://www.ptmbio.com/search/list.htm?searchtype=product&keyword=PTM-1427RM
Anti-Histone H3/HIST3H3 Monoclonal Antibody	Solarbio	K200033M	1/1000(WB)	https://www.solarbio.com/goodsInfo?id=9279
Anti-L-Lactyl Lysine Rabbit mAb	PTM BIO	PTM-1401 RM	1/1000(WB) 1/500(IF)	https://ptmbio.com/products/anti-l-lactyllysine-rabbit-mab/PTM-1401RM.htm
Ets-1(C-4)	SANTA CRUZ BIOTECHNOLOGY	sc-55581	1/1000(WB) 1/200(IF)	https://www.scbt.com/zh/p/ets-1-antibody-c-4
Ki67	Abcam	ab16667	1/200(IF)	https://www.abcam.cn/products/primary-antibodies/ki67-antibody-sp6-ab16667.html
DAPI	Abcam	ab104139	IF	https://www.abcam.cn/products/media/mounting-medium-with-dapi-aqueous-fluoroshield-ab104139.html
β-actin	Proteintech	66009-1-Ig	1/1000(WB)	https://www.ptgcn.com/products/Pan-Actin-Antibody-

				66009-1-Ig.htm
Anti-Histone H3 (acetyl K27) - ChIP Grade	Abcam	ab4729	1/50 (CUT&Tag)	https://www.abcam.cn/products/primary-antibodies/histone-h3-acetyl-k27-antibody-chip-grade-ab4729.html
PFKFB3	Abcam	ab181861	1/200(IF)	https://www.abcam.cn/products/primary-antibodies/pfkfb3-antibody-epr12594-ab181861.html

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129 **Supplementary Table3:Primer sequences used in RT-qPCR.**

	Gene	Primer	Primer Sequence (5'-3')	Length
Homo sapiens	<i>ETS1</i>	F	GCCAGTCATCTTTCAACAG	126
		R	CACGGTCCCGCACATAG	
	<i>ACTB</i>	F	AGCACAGAGCCTCGCCTTT	70
		R	ATCATCATCCATGGTGAGCTGG	
	<i>HK2</i>	F	AGGTCCTGATGCGGTTGG	154
		R	CGCTCCTCGCCTTTGTTC	
	<i>CXCR4</i>	F	CCACCAACAGTCAGAGGC	101
		R	AAAGATGAAGTCGGGAATAG	
	<i>TGM2</i>	F	ACAAATCCATCAACCGTTCC	160
		R	GCCAGTTTGTTTCAGGTGGTT	

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	Gene	Primer	Primer Sequence (5'-3')	Length
Mus musculus	<i>Ets1</i>	F	GAGATCCTGCAGAAAGAGGATG	178
		R	GGAGCGTCTGATAGGACTCTGT	
	<i>Actb</i>	F	GGCTGTATTCCCCTCCATCG	154
		R	CCAGTTGGTAACAATGCCATGT	
	<i>Pfkfb3</i>	F	GATCTGGGTGCCCGTCGATCACCG	147
		R	CAGTTGAGGTAGCGAGTCAGCTTC	
	<i>Ldha</i>	F	TGTCTCCAGCAAAGACTACTGT	155
		R	GACTGTACTTGACAATGTTGGGA	
	<i>Pdk1</i>	F	AGAGTCTGTGATCTTCCTCA	96
		R	CGATTCAAGTTCACGTCACAC	

	<i>Tgm2</i>	F	GCTGGACCAACAGGACAATGT	85
		R	CTCTAGGCTGAGACGGTACAG	
	<i>Hk2</i>	F	TGATCGCCTGCTTATTCACGG	112
		R	AACCGCCTAGAAATCTCCAGA	
	<i>Cxcr4</i>	F	GAAGTGGGGTCTGGAGACTAT	125
		R	TTGCCGACTATGCCAGTCAAG	

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132 **Supplementary Table 4:Primer sequences used in CUT&RUN-qPCR assays**

Sites	Primer Sequence (5'-3')
<i>ETS1</i> (Homo sapiens)	Forward primer: GCCCGTGGATGGTCTAAT
	Reverse primer: GATGTGGGTGTCGCTGGT

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